

1 **Matrigel inhibits elongation and drives endoderm differentiation in**
2 **aggregates of mouse embryonic stem cells**

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20 **Abstract**

21 Modelling peri-implantation mammalian development using the self-organising properties of stem cells is a
22 rapidly growing field that has advanced our understanding of cell fate decisions occurring in the early embryo.
23 Matrigel, a basement membrane matrix, is a critical substrate used in various protocols for its efficacy in
24 promoting stem cell growth and self-organization. However, its role in driving stem cell lineage commitment,
25 and whether this effect is driven by biochemical or physical cues is not being clearly defined. Here, we grow
26 embryoid bodies in suspension, Matrigel, and agarose, an inert polysaccharide, to attempt to decouple the
27 physical and biochemical roles of Matrigel and better understand how it drives stem cell differentiation. We
28 show that stem cell aggregates in Matrigel are hindered in their ability to elongate compared to those grown
29 in agarose or in suspension indicating that prohibitive role in self-organisation. Aggregates in Matrigel are
30 also driven to differentiate into endoderm with ectoderm differentiation inhibited. Furthermore, these
31 effects are not due to the physical presence of Matrigel as the same effects are not witnessed in aggregates
32 grown in agarose. Our results thus indicate that Matrigel has a significant and complex effect on the
33 differentiation and morphology of embryoid bodies.

34

35 **Keywords**

36 embryonic morphogenesis, embryonic stem cells, stembyro, basement membrane, extracellular matrix
37 Matrigel

38

39 Introduction

40 In mammals the establishment of the body plan begins with gastrulation, a process initiated in the posterior
41 region of the embryo through the appearance of the primitive streak (PS) that establishes the formation of
42 the endoderm, mesoderm, and ectoderm [1-4]. Studying these early, key developmental milestones is made
43 challenging in mammals as implantation into the uterine lining renders the embryo inaccessible for direct
44 experimentation and manipulation. However, understanding these early self-organising events remains a
45 fundamental field of study in developmental biology.

46 Recent advances in stem cell culture techniques have revolutionised the study of early embryonic
47 development as we are now able to study early embryonic milestones using *in vitro* stem cell-based embryo
48 models, so-called 'stembryos' [5-13]. A variety of 'stembryo' techniques have been developed in a short
49 space of time and have been shown to be able to resemble the blastocyst [14-17], undergo gastrulation [18,
50 19], develop an anterior-posterior axis [20-22], develop a primitive neural tube [23-25], or even undergo the
51 early stages of somitogenesis [23, 26] and even cardiogenesis [27, 28].

52 During the development of these protocols one of the critical factors found to drive stem cell self-
53 organisation has been the addition of Matrigel. Matrigel is an extracellular matrix (ECM) derived from
54 Engelbreth-Holm-Swarm mouse sarcoma [29, 30] that contains glycoproteins, proteoglycans, and growth
55 factors and has been found to support embryo culture [31]. Used at different concentration the addition of
56 Matrigel can drive 'stembryo' elongation, somitogenesis and neural tube development [23, 26]. These
57 findings suggest that Matrigel plays a key role in driving stem cell differentiation in 'stembryos'. However,
58 Matrigel has a complex and an unstandardised composition, and its composition can vary from batch to batch
59 [32]. This variability may affect the reproducibility of experiments using Matrigel. Furthermore, Matrigel
60 provides both structural support as well as a host of growth factors and signalling molecules making it unclear
61 whether the advanced morphologies noted in 'stembryo' cultures with Matrigel are due to the presence of
62 mechanical or biochemical cues. To uncouple these components, an inert material is needed to evaluate the
63 influence of just mechanical cues on stem cell morphogenesis. Here, we investigate the morphological and
64 gene expression changes induced in embryoid bodies (EBs), the simplest three-dimensional aggregates of

65 stem cells used to study their differentiation, when grown in suspension, Matrigel, and agarose, an inert
66 polysaccharide. In doing so we help to dissect the role of structural support on the fate of mouse embryonic
67 stem (mES) cells.

68 **Materials and Methods**

69 **mES cell culture**

70 In order to culture 129/Ola mouse embryonic stem (mES) cells, a feeder layer of inactive murine embryonic
71 fibroblast cells was cultured in 6-well plates coated with 0.1% gelatin (Sigma-Aldrich G7041-100G) at a
72 density of 1.6×10^4 cells/cm² in a base medium containing DMEM (1x) +GlutaMAX™ (Gibco 10566016), 15%
73 FBS (Gibco 10493106), 0.05 mM β-mercaptoethanol (Gibco 21985023), and 1% PenStrep (100 U penicillin/0.1
74 mg/ml streptomycin, Gibco 15140122). Two h before seeding the mES cells, the medium was changed to 2i
75 + LIF medium consisting of base medium supplemented with CHIR99021 (Chiron, 3 μM, Sigma-Aldrich
76 SML1046-5MG), PD0325901 (1 μM, Sigma-Aldrich PZ0162-5MG), and Leukaemia inhibitory factor (10 ng/ml,
77 LIF, Thermo Fisher Scientific A35934). The mES cells were then seeded onto the feeder layer at a density of
78 7×10^3 cells/cm², and the medium was changed daily, with cells being passaged every other day.

79 **Cell aggregates in suspension**

80 The aggregate formation method was adopted from Bailie Johnson *et al.* [33]. mES cells colonies were treated
81 with Dispase II (5mg/ml, Sigma-Aldrich D4693-1G) at 37°C for 20 min, followed by inactivation of Dispase and
82 transfer of the cell suspension to a 15 ml tube. The cells were spun down and resuspended in PBS to remove
83 residual medium. Following the PBS washes, the cells were resuspended in N2B27 medium which consists of
84 base medium supplemented with 1% N2 supplement (Gibco 17502048) and 1% B27 plus supplement. (Gibco
85 A3582801). The solution was diluted in N2B27 to give 5×10^4 cells per 5ml of medium for one 96-well plate.
86 The suspension was then added in 40μl droplets/well to a non-adhesive 96-well U-bottom plate (Greiner Bio-
87 One 650185). The plate was incubated at 37°C and 5% CO₂ for 48hrs after which a Chiron pulse (CHIR99021,
88 3μM) was added to the aggregates and the plate was incubated for an additional 24hrs. The following day,

89 the Chiron pulse was removed and fresh N2B27 was added. The medium was changed every day until the
90 required endpoint and not more than 168hrs.

91 **Cell aggregates on agarose**

92 The 96-well flat bottom plates were coated with 30µl of 1.2% agarose solution (1.2% w/v of Lonza Bioscience
93 50004 agarose added to deionised water and sent for autoclaving) and left to dry for 10min at RT. mES cell
94 aggregates were made as above, however, instead of adding the cell suspension to 96-well U-bottom plates,
95 40µl drops were placed in each well of the agarose-coated plate. The plate was incubated at 37°C and 5% CO₂
96 for 48hrs after which a Chiron pulse (3µM) was added to the aggregates, and they were incubated for an
97 additional 24hrs. The following day, the Chiron pulse was removed and fresh N2B27 was added. The medium
98 was changed every day until the required endpoint and no longer than 168hrs.

99 **Cell aggregates embedded in Matrigel**

100 mES cells were lifted using Dispase II after a 20 min incubation period and the pellet was washed twice in PBS
101 before counting. The cells were counted, and the appropriate volume was aliquoted and spun down again to
102 give a concentration of 3x10⁴ cells/ml. The pellet was resuspended in 1ml of Matrigel (Corning, 356234).
103 Droplets of 20µl were evenly placed on a 10 cm culture plate and Matrigel was left to solidify for 5 min at
104 37°C and subsequently covered with N2B27 medium. The Chiron pulse was added to the medium in the dish
105 on day 2 of the culture and removed after 24hrs. The medium was changed every second day until the
106 required endpoint, but not more than 168hrs.

107 **RNA extraction and cDNA synthesis**

108 RNA was collected from aggregates grown in suspension, agarose or Matrigel 144 hrs post aggregation using
109 the High Pure RNA Isolation Kit (Roche 11828665001) following the manufacturer's instructions. Due to the
110 limited amount of material obtained from each experiment, two experiments were combined to form one
111 biological replicate. This was done for a total of two biological replicates. The ImProm-II™ Reverse
112 Transcription System (Promega) was utilised for cDNA synthesis. A minimum of 1 µg of RNA was combined

113 with 1 μ l of Oligo (dT)15 primer (Promega, C110B) and nuclease-free water to make up a total of 5 μ l and
114 incubated at 70°C and 4°C for 5 min each. PCR master mix made up of 6.1 μ l nuclease-free water, 4 μ l 5x
115 reaction buffer (Promega, M289A), 2.4 μ l MgCl₂ (Promega, A351H), 1 dNTP mix (Promega, C114B), 0.5 μ l
116 Recombinant RNasin® Ribonuclease Inhibitor (Promega, N251A), 1 μ l Reverse Transcriptase (Promega,
117 M314A), was added to each sample. Thermal cycling was as follows: 25°C for 5 min, 42°C for 60 min and 70°C
118 for 15 min.

119 **qPCR**

120 The StepOnePlus™ Real-Time PCR System was used for quantitative PCR with SYBR green PCR Master-Mix
121 (Thermo-Fisher Scientific 4368708). Primer sequences are available in supplementary material Table S1.
122 Briefly, 2 μ l of cDNA (diluted 1:1 with nuclease-free water) was mixed with 8 μ l master which consisted of: 5
123 μ l SYBR green Master-Mix, 0.4 μ l of 10 μ M forward and reverse primer mixture (5 μ l of 100 μ M stock with 40
124 μ l nuclease-free water), and 2.6 μ l nuclease-free water. Each reaction had three technical replicates. The run
125 parameters can be found in supplementary material Table S2. *Gapdh* was used as a housekeeping gene to
126 calculate relative expression via the 2- $\Delta\Delta$ Ct method. Expression was normalised to cell aggregates cultured
127 in base medium that did not receive a Chiron pulse. Data analysis was performed using MS Excel, while
128 statistical analysis and graph generation were done using GraphPad Prism8.

129 **Immunofluorescence**

130 Cell aggregates were fixed at 4°C in 4% PFA for 1 h on a shaker. Subsequently, the aggregates were washed
131 three times for 5-10 minutes with PBST (PBS, 0.05% Tween-20) on a rotating shaker and were permeabilised
132 with 0.5% Triton-X-100 in PBS at room temperature for 1 h. After a brief washing step with PBST (Three times
133 for 5-10 minutes), the aggregates were blocked with PBS, 10% FBS, and 0.2% Triton-X-100 at room
134 temperature for 1 h. Primary antibody incubation was performed overnight at 4°C in blocking buffer. The
135 following day, the aggregates were washed three times quickly with PBS, followed by three 20 min washes.
136 Secondary antibody incubation was performed overnight at 4°C in blocking buffer, after which the washing
137 procedure was repeated. Hoechst was added to the 20 min washes following secondary antibody incubation

138 and the samples were left to incubated overnight at 4°C. Prior to imaging, aggregates mounted onto Mowiol
139 drops in glass-bottomed dishes. Antibody details and dilutions used are provided in Table S3.

140 **Image analysis**

141 The EVOS™ M5000 Imaging System microscope (Thermo Fisher Scientific, USA) was used to capture images
142 at a magnification of 10X. ImageJ was used to optimise the brightness and contrast of all images and for
143 measurements. The outlines of the entire aggregate were traced manually to determine the perimeter, while
144 the minor and major axial lengths were determined using the line tool. The elongation index was calculated
145 by dividing the major axial length by the minor axial length. Multichannel images were taken using a Zeiss
146 LSM 880 Airyscan confocal microscope (Zeiss, Germany) using a 20x or a 40x water-immersion objective. Z-
147 stack slices were spaced at 0.4µm and the final image was deconvolved and displayed at maximum projection
148 using the open-source ImageJ2 platform.

149 **Statistical analysis**

150 Statistical analyses were carried out using the functions provided by GraphPad Prism 8.4.2 (679). Because of
151 the scarcity of biological material, two biological replicates (each consisting of two pooled experiments) were
152 used for statistical analysis. When appropriate, an unpaired, nonparametric Mann-Whitney test was
153 performed. Alternatively, a Two-way Anova was performed followed by a Fisher's Least Significant Difference
154 test. In all cases, the error bars represent the mean ± standard error of the mean (mean ± s.e.m.). P values
155 are represented as follows: ns = $p \leq 0.1234$, * = $p \leq 0.0332$, ** = $p \leq 0.0021$, *** = $p \leq 0.0002$, **** = $p < 0.0001$.

156 **Results**

157 **The effect of Matrigel and agarose on the morphology of mES cell aggregates**

158 To examine the role of physical support in mES cell differentiation, we grew aggregates of ES cells in
159 suspension, on agarose, and in Matrigel (Fig. 1A). Under all conditions embryoid bodies (EBs) were subjected
160 to a pulse of Chiron at 48 h for 24 h and shape changes were quantified over time by measuring the aspect
161 ratio of the aggregates (the longest axis of the aggregate, the major axis, divided by the distance

162 perpendicular to the midpoint of the longest axis, the minor axis) (Fig. 1A). Aggregates were also classified
163 based on visual characteristics and categorised as spherical, ovoid, tear-shaped, budding, or elongating based
164 on their appearance and aspect ratio (Fig. S1). While aggregates grown in suspension (Fig. 1B and E) or
165 agarose (Fig. 1C and F) started to elongate at 72 h, following a pulse of Chiron, those grown in Matrigel
166 primarily retained their spherical or ovoid structures at the same time (Fig. 1D and G). By 144 h, all aggregates
167 in suspension (Fig. 1B and E) and 8/20 of those in agarose (Fig. 1C and F) were able to elongate and resemble
168 the cup-shaped morphology of the embryo. However, aggregates in Matrigel retained a more oval shape by
169 the end of the culture period and did not elongate as easily (Fig. 1D and G). It was also noted that agarose
170 aggregates had a unique, temporary balloon-like morphology (Fig. S1) that appeared at 24 h (16/20) and
171 were mostly no longer present by 96 h (1/20) (Fig. 1F).

172 Aspect ratio analyses revealed that suspension aggregates were the only group that underwent a rapid period
173 of elongation at 72 h and an additional increase in axial length at 144 h (Fig. 1H). Interestingly, there was a
174 dip in aspect ratio at 120 h under both suspension and agarose conditions, meaning that the aggregates grew
175 in width rather than length. The suspension aggregates had the highest rate of growth in perimeter, followed
176 by the agarose aggregates, and the Matrigel embedded aggregates showed the smallest increase in
177 perimeter during the 144 h period (Fig. 1I).

178 **Matrigel drives endoderm differentiation in mES cell aggregates**

179 Physical support from the maternal environment plays an essential role in early embryonic development
180 during induction of the PS, when cells first undergo an epithelial-to-mesenchymal transition (EMT) and form
181 mesodermal and endodermal progenitors [34]. In this study, we sought to determine the effects of physical
182 constraints on the development of the PS and in driving EMT in EBs. To investigate this, we measured the
183 expression of EMT and PS markers using qPCR in EBs grown in suspension, agarose, and Matrigel with a
184 Chiron pulse, and compared this to EBs grown in suspension without any exposure to a Wnt agonist.

185 The posterior epiblast marker *Wnt3* was significantly increased in suspension aggregates compared to
186 agarose and Matrigel aggregates while there was little variation in the expression of *β-catenin* (Fig. 2).

187 *Brachyury*, a marker of the PS and mesoderm, was also upregulated at 144 h under suspension conditions,
188 with no increase in expression noted in agarose or Matrigel conditions (Fig. 2). However, another PS and
189 mesoderm marker, *Nodal*, was significantly up-regulated in both agarose and Matrigel conditions (Fig. 2).
190 The mesendoderm marker *Eomes* was up-regulated in EBs in suspension, as well as in Matrigel (Fig. 2), while
191 another mesendoderm marker *Mixl1* was significantly up-regulated significantly in suspension and agarose
192 conditions but not in Matrigel (Fig. 2). Interestingly, the early endoderm marker, *Gata6*, had increased levels
193 of expression in Matrigel aggregates compared to the control (although not significantly), while a marker of
194 the definitive endoderm marker, *Sox17*, was significantly increased in Matrigel compared to the other
195 conditions, suggesting that Matrigel may be directing differentiation towards the endoderm lineage. A key
196 EMT marker, *Snai1* was down-regulated significantly under suspension and agarose conditions compared to
197 the control (Fig. 2) and *E-cadherin*, an epithelial marker, was down-regulated in suspension compared to the
198 other two conditions (Fig. 2). The anterior epiblast marker *Pou3f1* had higher levels of expression in all three
199 conditions compared to controls, albeit with a significant increase only observed in suspension (Fig. 2).
200 Similarly, *Slc7a3*, another anterior epiblast marker, had increased expression levels in all three conditions
201 compared to controls with a significant increase in suspension and agarose conditions (Fig. 2). The
202 anterior/neuroectoderm marker *Sox2* had an increased relative level of expression in the three conditions,
203 while *Pax6*, a late neuroectoderm marker, was only up-regulated in suspension aggregates. This suggests that
204 neuroectoderm formation may be at a more advanced stage in suspension aggregates exposed to a Chiron
205 pulse.

206 **Aggregates in Matrigel form pro-amniotic-like cavities**

207 The effects that physical constraints had on mesoderm and endoderm gene expression next led us to
208 investigate the spatial expression of the mesoderm marker *Brachyury* and the endoderm marker *Sox17* in
209 our aggregates using immunofluorescence. When in suspension very few aggregates expressed *Brachyury*.
210 When they did, this expression was either restricted to one pole of the aggregate (Fig. 3A, 1/50) or dispersed
211 throughout the entire aggregate (Fig. 3B, 2/50). A highly similar pattern was observed in the agarose
212 aggregates with only a few aggregates being positive for *Brachyury*, and when they were, this staining was

213 either localised (Fig. 3C, 2/46) or dispersed (Fig. 3D, 2/46). Aggregates in Matrigel, however, have significantly
214 more Brachyury positive aggregates with 18/60 showing a dispersed pattern of expression (Fig. 3E).
215 Interestingly, all of these also developed a Brachyury lined lumen in their centers (Fig. 3F, 18/60). All
216 aggregates in Matrigel were found to have formed a similar cavity, even those without any Brachyury-positive
217 staining. The cells surrounding these cavities appeared as epithelial-like and were brightly stained with E-
218 cadherin (Fig. 3G). The suspension and agarose aggregates clearly lacked such cavities, but they had regions
219 that were negative for E-cadherin (Fig. 3H and 3I) suggesting that EMT has occurred.

220 Endoderm staining was observed infrequently in all conditions investigated. In suspension aggregates, it was
221 noted as either small pockets of Sox17-positive cells near the periphery (Fig. 3J, 7/46) or dispersed across the
222 surface of the aggregate (Fig. 3K, 7/46). A similar expression pattern was observed in agarose aggregates
223 with small pockets of Sox17-positive cells (Fig. 3L, 3/49) or a dispersed expression pattern (Fig. 3M, 5/49).
224 Aggregates in Matrigel were noted to express Sox17 in dispersed groups across the surface of the aggregate
225 (Fig. 3N, 5/60).

226 **Discussion**

227 The emergence and subsequent development of 'stembryo' culture systems has generated a non-invasive,
228 scalable, and novel way to investigate early mammalian embryonic morphogenesis and cell fate decisions
229 gastrulation and germ layer formation in mammals. One of the key aspects missing from 'stembryo'
230 structures is the presence of physical support provided by the ECM normally generated by the basement
231 membrane of the visceral endoderm. To address this issue, many of these systems incorporate the
232 commercial basement membrane substitute Matrigel. Although Matrigel can promote stem cell self-
233 organisation and provides some degree of mechanical constraint, it is ill-defined and suffers from lot-to-lot
234 variability. In addition, its effect on organoid structure and differentiation is influenced by both mechanical
235 and biochemical cues, and the specific component/s responsible for the observed morphologies remain
236 unknown. Matrigel also does not support any controlled modifications of its stiffness or components, and
237 thus there is a need to use an inert and manipulatable scaffold to separate the effects of physical and
238 chemical cues on stem cell self-organisation in this burgeoning field. In this study, we used agarose, an inert

239 polysaccharide, to test the role of mechanical cues on the fate of embryoid bodies and to try to create similar
240 physical constraints to those provided by the uterus.

241 We found that EB morphology was significantly influenced by the physical constraints present during culture,
242 with suspended aggregates exhibiting more effective growth and elongation than counterparts grown in the
243 presence of Matrigel or agarose. At 48 h, some suspension and agarose aggregates began to exhibit tear-
244 shaped or budding structures, indicating an intrinsic breaking of symmetry. At 72 h, there was a sudden
245 increase in the variety of morphologies, including balloon-like structures, which were observed as
246 intermediates in the elongation process. Interestingly, we found that the presence of Matrigel inhibited
247 elongation, causing aggregates to maintain their oval or spherical shape for longer. When we measured
248 aspect ratios, we found that aggregates in suspension underwent a drop in aspect ratio at 120 h, followed by
249 a sudden increase at 144 h. This suggests that cell proliferation occurs before elongation, along the major
250 axis. Aggregates grown in agarose exhibited a similar dip and rise in aspect ratio, but the same pattern was
251 not found in aggregates grown on Matrigel. Overall, we found that early self-patterning events in stem cell
252 aggregates in culture are hindered by the presence of Matrigel. These aggregates neither grow nor elongate
253 as effectively as those in suspension or in agarose, with Matrigel playing a prohibitive role in self-organisation.

254 Previous studies have shown that after 144 h in culture 'stembryo' should start to undergo a gastrulation-
255 like process [20, 23, 35, 36]. Although we noted elongation in EBs in both suspension and agarose conditions
256 by this time point, there were marked differences in the expression of key markers between the two
257 conditions. Aggregates in suspension had increased levels of *Brachyury* suggesting that the PS had been
258 initiated. In contrast, aggregates grown in agarose did not have this same up-regulation in *Brachyury*
259 expression. They did, however, have up-regulated β -*catenin*, an upstream regulator of *Brachyury*. This could
260 indicate a delay in the gastrulation process due to the differences in the physical properties of the
261 environments as the more restrictive agarose may present a less conducive environment for elongation and
262 subsequent gastrulation. Aggregates in Matrigel had similarly low levels of *Brachyury* and β -*catenin*, further
263 supporting the theory that physical support hinders stem cell driven *in vitro* gastrulation-like events.
264 Aggregates in suspension also had the lowest relative levels of *E-cadherin*, an epithelial marker, and the

265 highest levels of mesenchymal marker *Snai1*, suggesting that these aggregates are likely undergoing EMT, a
266 critical process in gastrulation. This is further support for suspension aggregates being more developmentally
267 advanced than aggregates in either agarose or Matrigel.

268 However, the expression patterns of *Nodal* and *Wnt3* in our aggregates indicate that the developmental
269 landscape in our conditions is more complex than this. *Nodal* and *Wnt3* are known to work together to induce
270 the expression of *Brachyury* and *Eomes*, another marker of gastrulation [1]. We observed that *Nodal* and
271 *Wnt3* have opposing expression patterns in suspension aggregates, with elevated levels of *Wnt3* and low
272 levels of *Nodal*. Interestingly, the opposite pattern, high levels of *Nodal* and relatively low levels of *Wnt3* was
273 observed in aggregates in agarose and Matrigel. These results would indicate that the aggregates under these
274 conditions are more advanced in terms of mesoderm differentiation. As *Eomes* has been shown to precede
275 *Brachyury* expression [37], and based on the high levels of *Eomes* we can assume that *Brachyury* expression
276 would increase in both suspension and Matrigel aggregates if cultured further. As both aggregates in Matrigel
277 and agarose show a similar pattern of expression in this case, we hypothesise that it is principally mechanical
278 cues driving this expression pattern and this warrants further investigation.

279 Analysing the expression of key markers of PS formation and EMT transition, it appears that our aggregates
280 most closely resemble mouse embryos at E5.5-E6.5, between the emergence of the germ layers and the
281 formation of the anterior-posterior axis (A-P axis), which occurs at approximately 120-144 h of
282 development[6]. It is noted that this is significantly delayed compared to other 'stembyro' culture processes,
283 where aggregates can undergo a gastrulation-like process after 96 h and exhibit similarities to the E8.5 mouse
284 embryo [20, 23, 35, 38].

285 Despite this apparent developmental delay, suspension aggregates had high levels of
286 anterior/neuroectoderm markers, *Pou3f1*, *Slc7a3*, *Sox2*, indicating the initiation of the anterior-posterior(A-
287 P) axis. We observed a distinct increase in *Pax6*, a late neuroectoderm marker, in suspension aggregates
288 similar to what is observed in E8.5 mouse embryos and gastruloids (one of the most robust 'stembyro'
289 systems) at 120 h [38]. This is possibly due to the high levels of *Sox2* present in mES cells prior to

290 differentiation, allowing rapid neural differentiation [39], provided they were able to differentiate without
291 any confounding effects, such as physical constraints.

292 Aggregates in Matrigel demonstrated the highest expression of endoderm markers compared to the other
293 two conditions. *Gata6* is typically a marker for cardiac mesoderm and DE progenitors [40], while *Sox17* is a
294 well-known DE marker [41]. *Gata6* mRNA was up-regulated in Matrigel aggregates, followed by up-regulation
295 of *Sox17*, indicating that this condition likely favoured endoderm formation. It is unclear if this preference is
296 due to physical restrictions imposed by Matrigel or the presence of certain signalling factors. However,
297 evidence suggests that laminin, a major component of Matrigel, can direct human ES cells toward the
298 endoderm lineage [42]. Furthermore, studies have demonstrated that culturing EPI stem cells in Matrigel
299 results in up-regulated levels of endoderm-related transcription factors such as *Sox17* [43], further
300 supporting the notion that Matrigel influences lineage commitment towards endoderm.

301 Immunofluorescent staining for Brachyury revealed that the positive cells were located exclusively at the
302 elongated pole of the aggregates in both suspension and agarose, as observed in other studies [20, 35, 38,
303 44], although the frequency of this pattern was significantly lower than reported by others. Aggregates in
304 Matrigel, however, had many more aggregates that were Brachyury-positive, and these cells were principally
305 in the centre of the aggregate surrounding the lumen. Studies have demonstrated that Matrigel can increase
306 the expression of Wnt3 antagonists such as *Dkk1* and *Sfpr1* [45]. Therefore, it is plausible that cells located
307 in the periphery of our Matrigel aggregates express Wnt3 agonists, resulting in the localisation of Wnt3
308 activity in the centre of the aggregates, which leads to the up-regulation of Brachyury expression. By
309 embedding aggregates in Matrigel, we were able to observe cavity formation that resembled the pro-
310 amniotic cavity. This was likely due to the cells pulling apart, facilitated by E-cadherin localisation around the
311 membrane. Matrigel is used as a substitute for ECM derived from extra-embryonic tissue [18] and the
312 interaction between cells and Matrigel via integrin receptors facilitates cell polarisation and cavity formation
313 [46]. In the embryo, cavity formation is mediated by the interaction between laminin and the $\beta 1$ -integrin
314 receptor between embryonic and extra-embryonic tissue [46]. The same mechanism may operate in Matrigel

315 aggregates, allowing them to form pro-amniotic cavity-like structures. To test the establishment of polarity
316 in our aggregates, future work should involve immunofluorescence staining for aPKC and Par6 [47, 48].

317 Overall, our results suggest that Matrigel has a significant and complex effect on the differentiation and
318 morphology of mES cells in culture. It can hinder stem cell self-organisation and neural differentiation, drive
319 endoderm differentiation, and induce the formation of a pro-amniotic cavity. Its effects are not driven simply
320 by the mechanical force it provides as its effects on stem cells are not mimicked by using agarose. Although
321 Matrigel is a critical component of ‘stembryo’ cultures, caution should be taken when viewing it as a
322 substitute for an ECM. Future research in the field should focus on finding a more defined and manipulatable
323 replacement for Matrigel in order to advance the field and the accuracy of *in vitro* embryo models.

324

325 **Author contributions**

326 M.G. conceived the project. A.A carried out the experiments. A.A. and M.G. wrote the paper.

327

328 **Declaration of competing interest**

329 The authors have no financial or competing interests to declare.

330

331 **Data availability**

332 Data will be made available upon request.

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335

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356 **Figure captions**

357 **Fig. 1: Comparative analysis of the effect of various physical constraints on ES cell aggregate morphology.**

358 (A) ES cell aggregates were cultured in suspension, on agarose or embedded in Matrigel. All aggregates
359 received a 24 h Chiron pulse on the third day of culture. To calculate the elongation of aggregates, the major
360 axis (the longest axis; blue line) was measured and divided by the minor axis- the line perpendicular to the
361 major axis at its midpoint (red line). (B) Morphology of aggregates in suspension. Aggregates had elongated
362 by 144 h. (C) Morphology of aggregates cultured on agarose. At 24 h the aggregates had already showed
363 signs of asymmetry and by 144 h the aggregates had elongated. (D) Morphology of aggregates embedded in
364 Matrigel. The aggregates were relatively smaller than the other two conditions and did not elongate
365 significantly. (E) Characteristics of n = 20 aggregates grown in suspension for the period of 144 h. Major
366 morphological changes occurred at 72 h and at 144 h all the aggregates had elongated. (F) Characteristics of
367 n = 20 aggregates grown on agarose for the period of 144 h. The aggregates had a unique balloon-like
368 morphology in the first 96 h of culture and by 144 h some aggregates were still in the budding stage and had
369 not elongated fully. (G) Characteristics of n = 20 aggregates grown in Matrigel for the period of 144 h.
370 Aggregates had a delayed growth and only showed signs of asymmetry at 96 h. (H) Aspect ratio of aggregates
371 in suspension (black line, empty circle), on agarose (grey line, dotted circle) and in Matrigel (dotted line, solid
372 circle) for n = 20 aggregates per condition. (I) Perimeter of aggregates in suspension (black line, empty circle),
373 on agarose (grey line, dotted circle) and in Matrigel (dotted line, solid circle) for n = 20 aggregates per
374 condition. Error bars indicate mean \pm SEM. The aspect ratio within the same condition between two
375 consecutive time points was evaluated for significance. An unpaired, nonparametric Mann-Whitney test was
376 performed and * = $p \leq 0.0332$, ** = $p \leq 0.0021$, *** = $p \leq 0.0002$, **** = $p < 0.0001$. Scale bar: 100 μ m.

377 **Fig. 2: Effects of various physical constraints on gene expression of aggregates.** QPCR was performed on

378 reverse transcribed RNA extracted at 144 h. Primitive streak markers: *Wnt3*, *β -catenin*, *Brachyury* and *Nodal*.

379 Mesendoderm markers: *Eomes* and *Mixl1*. Endoderm markers: *Gata6* and *Sox17*. EMT markers: *Snai1* and *E-*

380 *cadherin*. Anterior/neurectoderm markers: *Pou3f1*, *Slc7a3*, *Sox2* and *Pax 6*. Relative fold expression to cell

381 aggregates without a Chiron pulse. Error bars indicate mean \pm SEM. An unpaired, nonparametric Mann-
382 Whitney test was performed and * = $p \leq 0.0332$, ** = $p \leq 0.0021$, *** = $p \leq 0.0002$, **** = $p < 0.0001$.

383 **Fig. 3: Immunofluorescence staining of Brachyury, Sox 17 and E-cadherin in aggregates cultured under**
384 **various physical constraints.** (A) Localised Brachyury positive cells at the elongating tip of suspension
385 aggregates. (B) Brachyury positive cells spread throughout suspension aggregates that remained spherical.
386 (C) Localised Brachyury positive cells in agarose aggregates. (D) Dispersed Brachyury-positive cells in agarose
387 aggregates. (E) Brachyury positive cells dispersed throughout Matrigel aggregates. (F) Brachyury positive cells
388 lining the lumen of Matrigel aggregates. (G) Epithelial-like cells surrounding the cavity in Matrigel aggregates
389 stained positive for epithelial marker E-cadherin. (H) Suspension aggregates stained for E-cadherin had
390 regions with reduced expression that may represent EMT. (I) Agarose aggregates stained for E-cadherin had
391 regions with reduced expression that may represent EMT. (J) Pockets of Sox17 positive cells in the periphery
392 of suspension aggregates. (K) Sox17 positive cells dispersed across the surface of suspension aggregates. (L)
393 Pockets of Sox17 positive cells in aggregates cultured on agarose. (M) Sox17 positive cells dispersed across
394 the surface of aggregates grown on agarose. (N) Sox17 positive cells in dispersed groups across the surface
395 of aggregates in Matrigel. Scale bar: 25 μ m.

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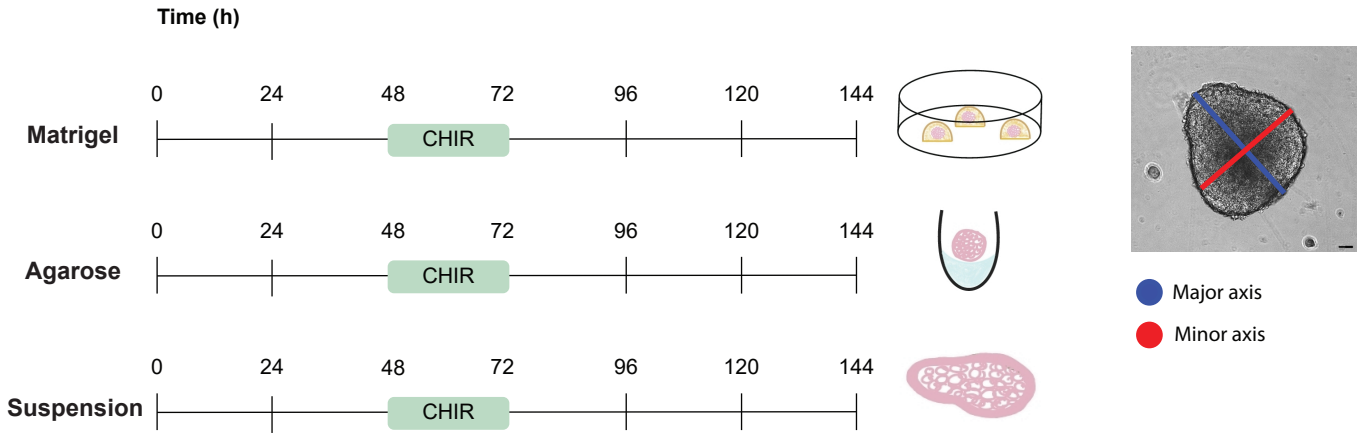
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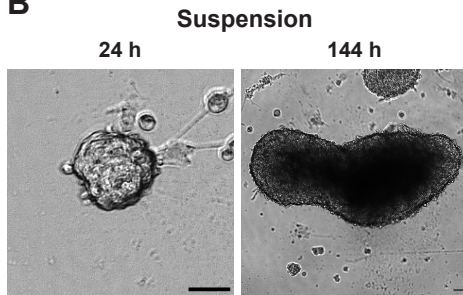
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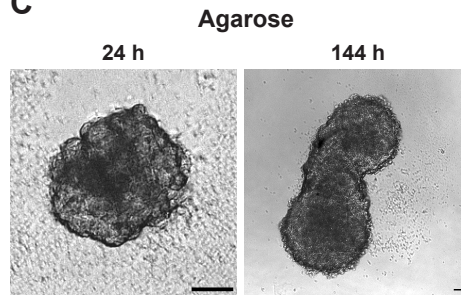
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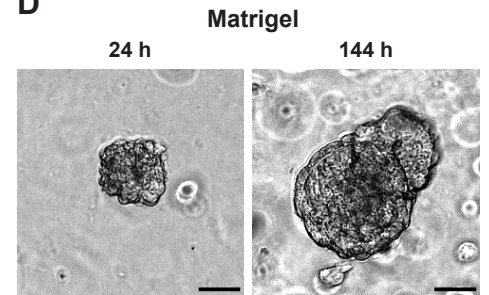
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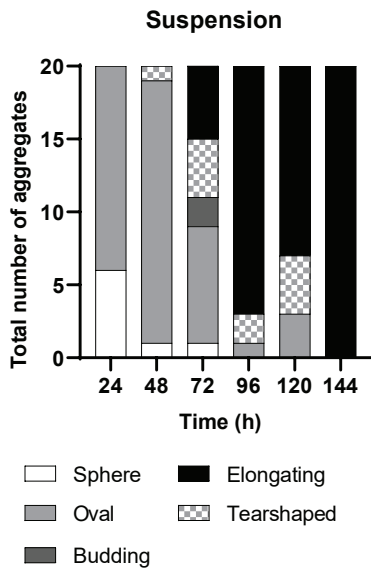
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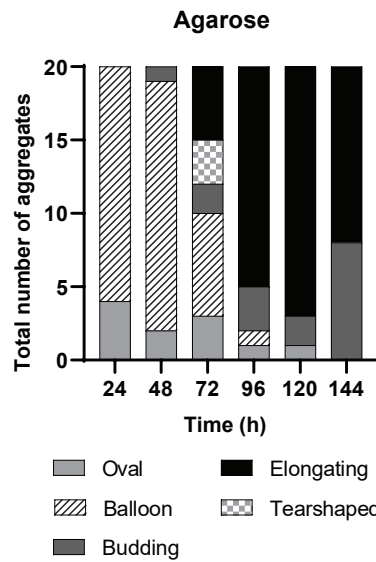
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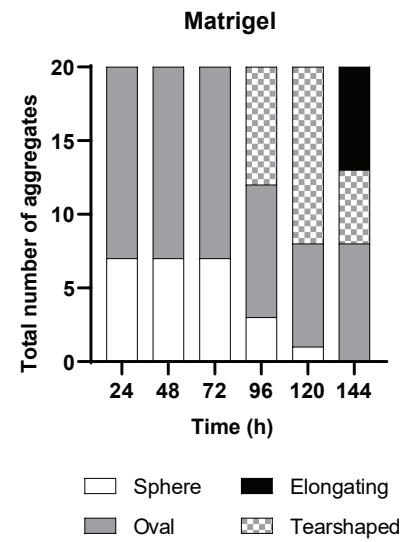
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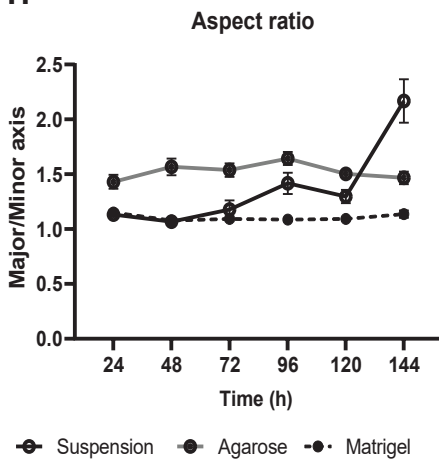
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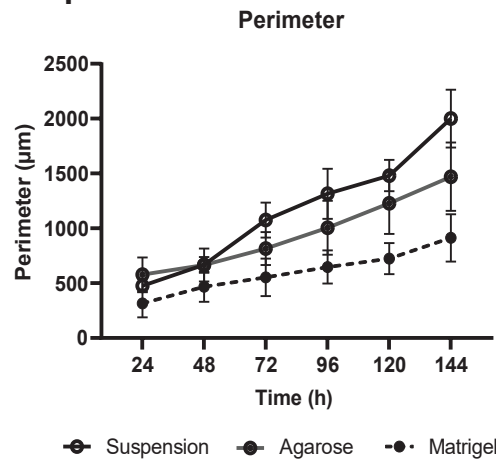
G



H



I



Wnt3

β-catenin

Brachyury

Nodal

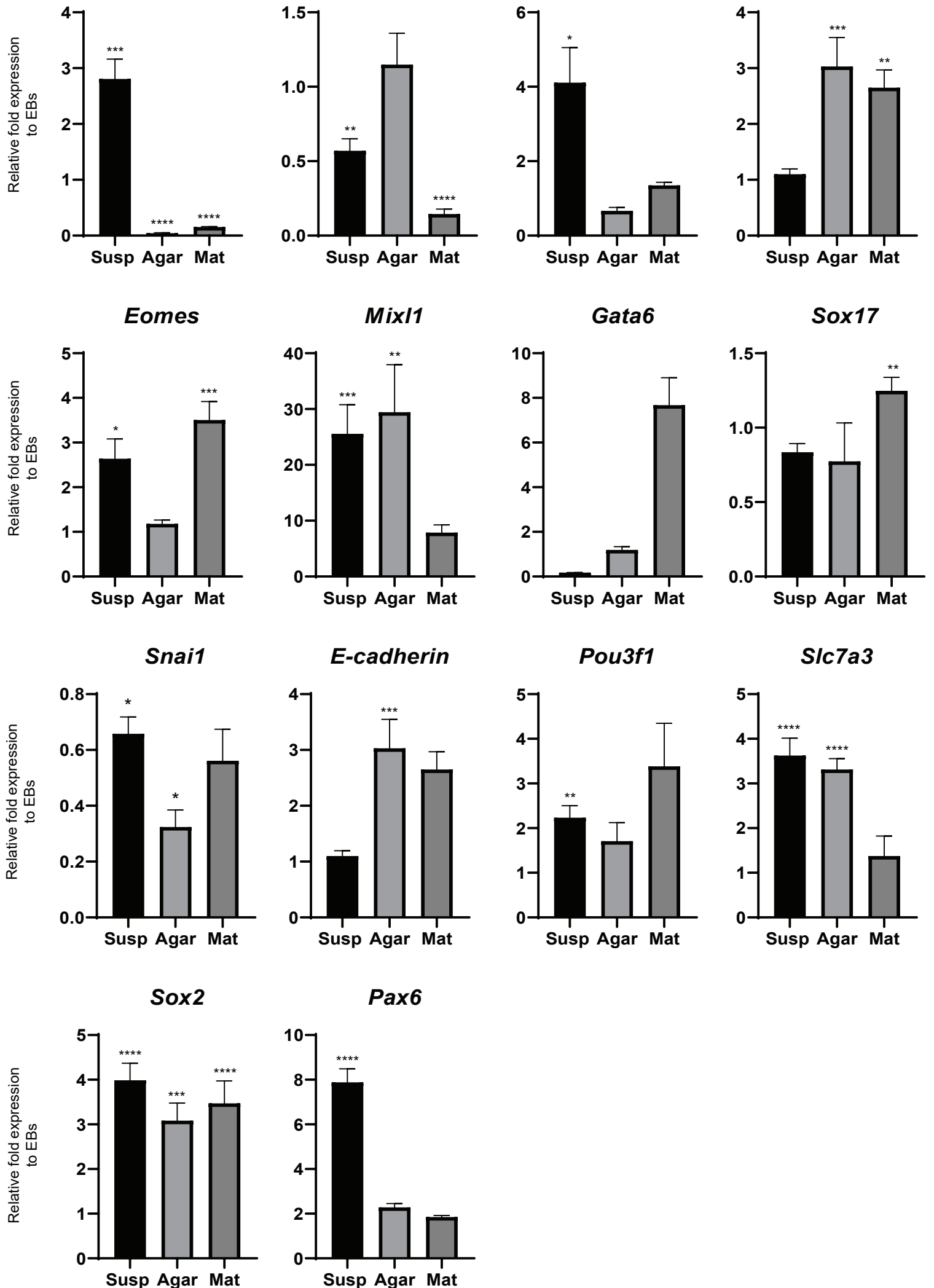


Fig. 2

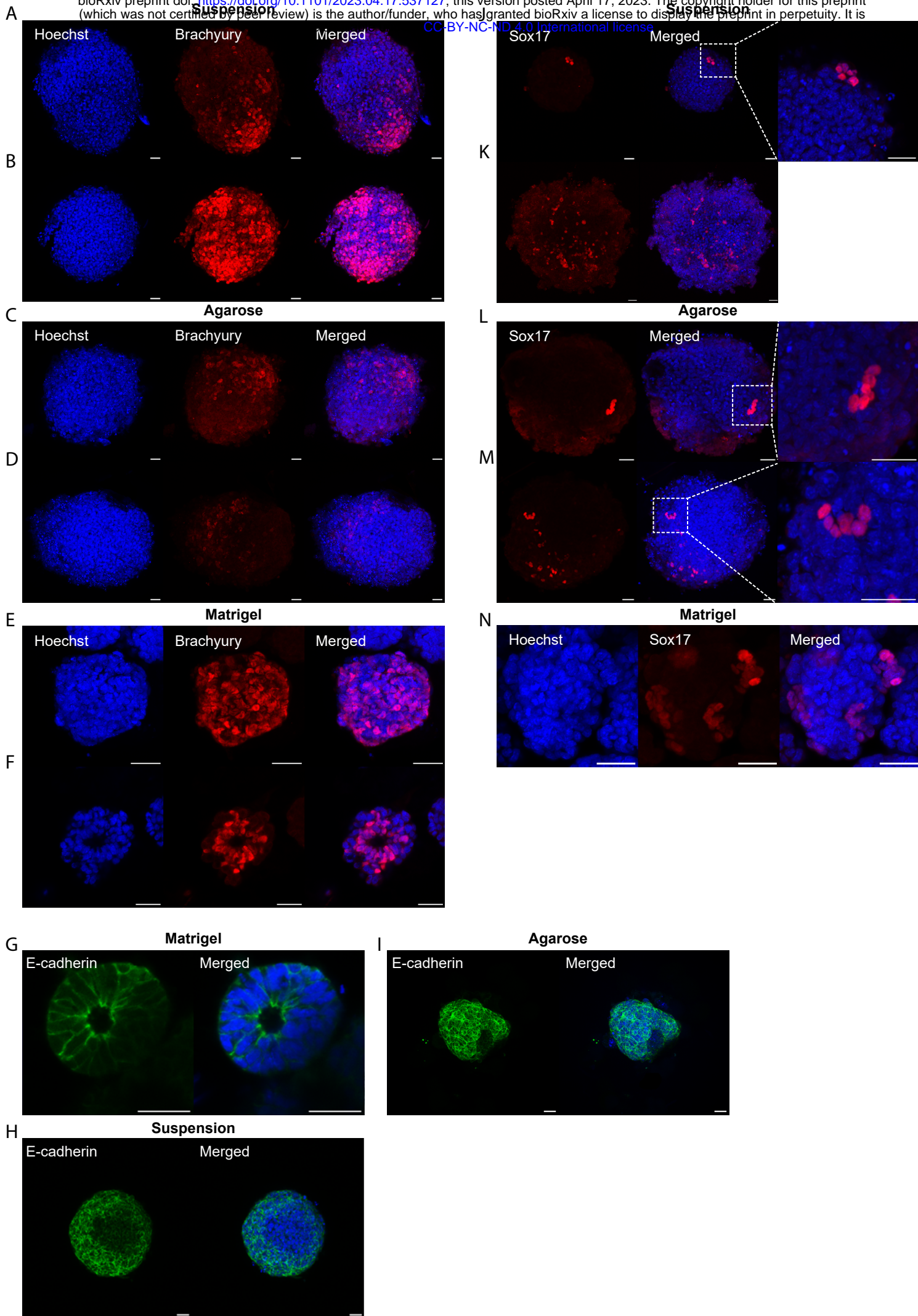


Fig. 3

404 **References:**

- 405 1. Arnold, S.J. and E.J. Robertson, *Making a commitment: cell lineage allocation and axis patterning in*
406 *the early mouse embryo*. Nature Reviews Molecular Cell Biology, 2009. **10**(2): p. 91-103. DOI:
407 10.1038/nrm2618.
- 408 2. Stower, M.J. and F. Bertocchini, *The evolution of amniote gastrulation: the blastopore-primitive streak*
409 *transition*. WIREs Developmental Biology, 2017. **6**(2): p. e262. DOI:
410 <https://doi.org/10.1002/wdev.262>.
- 411 3. Sheng, G., A. Martinez Arias, and A. Sutherland, *The primitive streak and cellular principles of building*
412 *an amniote body through gastrulation*. Science, 2021. **374**(6572): p. abg1727. DOI:
413 doi:10.1126/science.abg1727.
- 414 4. Tam, P.P.L. and R.R. Behringer, *Mouse gastrulation: the formation of a mammalian body plan*.
415 *Mechanisms of Development*, 1997. **68**(1): p. 3-25. DOI: [https://doi.org/10.1016/S0925-](https://doi.org/10.1016/S0925-4773(97)00123-8)
416 [4773\(97\)00123-8](https://doi.org/10.1016/S0925-4773(97)00123-8).
- 417 5. Amel, A., S. Rossouw, and M. Goolam, *Gastruloids: A Novel System for Disease Modelling and Drug*
418 *Testing*. Stem Cell Rev Rep, 2023. **19**(1): p. 104-113. DOI: 10.1007/s12015-022-10462-5.
- 419 6. Arias, A.M., Y. Marikawa, and N. Moris, *Gastruloids: Pluripotent stem cell models of mammalian*
420 *gastrulation and embryo engineering*. Dev Biol, 2022. **488**: p. 35-46. DOI:
421 10.1016/j.ydbio.2022.05.002.
- 422 7. Veenvliet, J.V., et al., *Sculpting with stem cells: how models of embryo development take shape*.
423 *Development*, 2021. **148**(24). DOI: 10.1242/dev.192914.
- 424 8. van den Brink, S.C. and A. van Oudenaarden, *3D gastruloids: a novel frontier in stem cell-based in*
425 *vitro modeling of mammalian gastrulation*. Trends Cell Biol, 2021. **31**(9): p. 747-759. DOI:
426 10.1016/j.tcb.2021.06.007.
- 427 9. Zylicz, J.J., *Defined Stem Cell Culture Conditions to Model Mouse Blastocyst Development*. Current
428 *Protocols in Stem Cell Biology*, 2020. **52**(1): p. e105. DOI: <https://doi.org/10.1002/cpsc.105>.

- 429 10. Taniguchi, K., I. Heemskerk, and D.L. Gumucio, *Opening the black box: Stem cell-based modeling of*
430 *human post-implantation development*. J Cell Biol, 2019. **218**(2): p. 410-421. DOI:
431 10.1083/jcb.201810084.
- 432 11. Shahbazi, M.N. and M. Zernicka-Goetz, *Deconstructing and reconstructing the mouse and human*
433 *early embryo*. Nature Cell Biology, 2018. **20**(8): p. 878-887. DOI: 10.1038/s41556-018-0144-x.
- 434 12. Simunovic, M. and A.H. Brivanlou, *Embryoids, organoids and gastruloids: new approaches to*
435 *understanding embryogenesis*. Development, 2017. **144**(6): p. 976-985. DOI: 10.1242/dev.143529.
- 436 13. Turner, D.A., P. Baillie-Johnson, and A. Martinez Arias, *Organoids and the genetically encoded self-*
437 *assembly of embryonic stem cells*. BioEssays, 2016. **38**(2): p. 181-191. DOI: 10.1002/bies.201500111.
- 438 14. Rivron, N.C., et al., *Blastocyst-like structures generated solely from stem cells*. Nature, 2018.
439 **557**(7703): p. 106-111. DOI: 10.1038/s41586-018-0051-0.
- 440 15. Sozen, B., et al., *Self-Organization of Mouse Stem Cells into an Extended Potential Blastoid*.
441 *Developmental Cell*, 2019. **51**(6): p. 698-712.e8. DOI: 10.1016/j.devcel.2019.11.014.
- 442 16. Yu, L., et al., *Blastocyst-like structures generated from human pluripotent stem cells*. Nature, 2021.
443 **591**(7851): p. 620-626. DOI: 10.1038/s41586-021-03356-y.
- 444 17. Heidari Khoei, H., et al., *Generating human blastoids modeling blastocyst-stage embryos and*
445 *implantation*. Nat Protoc, 2023. DOI: 10.1038/s41596-023-00802-1.
- 446 18. Harrison, S.E., et al., *Assembly of embryonic and extraembryonic stem cells to mimic embryogenesis*
447 *in vitro*. Science, 2017. **356**(6334): p. eaal1810. DOI: 10.1126/science.aal1810.
- 448 19. Sozen, B., et al., *Self-assembly of embryonic and two extra-embryonic stem cell types into gastrulating*
449 *embryo-like structures*. Nat Cell Biol, 2018. **20**(8): p. 979-989. DOI: 10.1038/s41556-018-0147-7.
- 450 20. van den Brink, S.C., et al., *Symmetry breaking, germ layer specification and axial organisation in*
451 *aggregates of mouse embryonic stem cells*. Development, 2014. **141**(22): p. 4231-42. DOI:
452 10.1242/dev.113001.
- 453 21. Turner, D.A., et al., *Anteroposterior polarity and elongation in the absence of extraembryonic tissues*
454 *and spatially localised signalling in Gastruloids, mammalian embryonic organoids*. Development,
455 2017. **144**(21): p. 3894-3906. DOI: 10.1242/dev.150391.

- 456 22. Beccari, L., et al., *Multi-axial self-organization properties of mouse embryonic stem cells into*
457 *gastruloids*. Nature, 2018. **562**(7726): p. 272-276. DOI: 10.1038/s41586-018-0578-0.
- 458 23. Veenvliet, J.V., et al., *Mouse embryonic stem cells self-organize into trunk-like structures with neural*
459 *tube and somites*. Science, 2020. **370**(6522): p. eaba4937. DOI: 10.1126/science.aba4937.
- 460 24. Libby, A.R.G., et al., *Axial elongation of caudalized human organoids mimics aspects of neural tube*
461 *development*. Development, 2021. **148**(12). DOI: 10.1242/dev.198275.
- 462 25. Berenger-Currias, N.M., et al., *A gastruloid model of the interaction between embryonic and extra-*
463 *embryonic cell types*. J Tissue Eng, 2022. **13**: p. 20417314221103042. DOI:
464 10.1177/20417314221103042.
- 465 26. van den Brink, S.C., et al., *Single-cell and spatial transcriptomics reveal somitogenesis in gastruloids*.
466 Nature, 2020. **582**(7812): p. 405-409. DOI: 10.1038/s41586-020-2024-3.
- 467 27. Rossi, G., et al., *Capturing Cardiogenesis in Gastruloids*. Cell Stem Cell, 2021. **28**(2): p. 230-240.e6.
468 DOI: 10.1016/j.stem.2020.10.013.
- 469 28. Olmsted, Z.T. and J.L. Paluh, *A combined human gastruloid model of cardiogenesis and neurogenesis*.
470 iScience, 2022. **25**(6): p. 104486. DOI: 10.1016/j.isci.2022.104486.
- 471 29. Kleinman, H.K. and G.R. Martin, *Matrigel: Basement membrane matrix with biological activity*.
472 Seminars in Cancer Biology, 2005. **15**(5): p. 378-386. DOI:
473 <https://doi.org/10.1016/j.semcancer.2005.05.004>.
- 474 30. Kleinman, H.K., et al., *Basement membrane complexes with biological activity*. Biochemistry, 1986.
475 **25**(2): p. 312-318. DOI: 10.1021/bi00350a005.
- 476 31. Carnegie, J., et al., *Can Matrigel substitute for Vero cells in promoting the in-vitro development of*
477 *mouse embryos?* Hum Reprod, 1995. **10**(3): p. 636-41. DOI:
478 10.1093/oxfordjournals.humrep.a136002.
- 479 32. Hughes, C.S., L.M. Postovit, and G.A. Lajoie, *Matrigel: A complex protein mixture required for optimal*
480 *growth of cell culture*. PROTEOMICS, 2010. **10**(9): p. 1886-1890. DOI: 10.1002/pmic.200900758.

- 481 33. Baillie-Johnson, P., et al., *Generation of Aggregates of Mouse Embryonic Stem Cells that Show*
482 *Symmetry Breaking, Polarization and Emergent Collective Behaviour In Vitro*. J Vis Exp, 2015(105).
483 DOI: 10.3791/53252.
- 484 34. Gattazzo, F., A. Urciuolo, and P. Bonaldo, *Extracellular matrix: a dynamic microenvironment for stem*
485 *cell niche*. Biochim Biophys Acta, 2014. **1840**(8): p. 2506-19. DOI: 10.1016/j.bbagen.2014.01.010.
- 486 35. Turner, D.A., et al., *Anteroposterior polarity and elongation in the absence of extra-embryonic tissues*
487 *and of spatially localised signalling in gastruloids: mammalian embryonic organoids*. Development,
488 2017. **144**(21): p. 3894-3906. DOI: 10.1242/dev.150391.
- 489 36. Beccari, L., et al., *Generating Gastruloids from Mouse Embryonic Stem Cells*. Protocol Exchange, 2018.
490 DOI: 10.1038/protex.2018.094.
- 491 37. Tomic, J., et al., *Eomes and Brachyury control pluripotency exit and germ-layer segregation by changing*
492 *the chromatin state*. Nature Cell Biology, 2019. **21**(12): p. 1518-1531. DOI: 10.1038/s41556-019-0423-
493 1.
- 494 38. Beccari, L., et al., *Multi-axial self-organization properties of mouse embryonic stem cells into*
495 *gastruloids*. Nature, 2018. **562**(7726): p. 272-276. DOI: 10.1038/s41586-018-0578-0.
- 496 39. Thomson, M., et al., *Pluripotency Factors in Embryonic Stem Cells Regulate Differentiation into Germ*
497 *Layers*. Cell, 2011. **145**(6): p. 875-889. DOI: 10.1016/j.cell.2011.05.017.
- 498 40. Morrisey, E.E., et al., *GATA-6: a zinc finger transcription factor that is expressed in multiple cell*
499 *lineages derived from lateral mesoderm*. Dev Biol, 1996. **177**(1): p. 309-22. DOI:
500 10.1006/dbio.1996.0165.
- 501 41. Choi, E., et al., *Dual Lineage-Specific Expression of Sox17 During Mouse Embryogenesis*. STEM CELLS,
502 2012. **30**(10): p. 2297-2308. DOI: 10.1002/stem.1192.
- 503 42. Wong, J.C., et al., *Definitive endoderm derived from human embryonic stem cells highly express the*
504 *integrin receptors alphaV and beta5*. Cell Adh Migr, 2010. **4**(1): p. 39-45. DOI: 10.4161/cam.4.1.10627.
- 505 43. Inamori, S., et al., *Modeling early stages of endoderm development in epiblast stem cell aggregates*
506 *with supply of extracellular matrices*. Development, Growth & Differentiation, 2020. **62**(4): p. 243-
507 259. DOI: 10.1111/dgd.12663.

- 508 44. Girgin, M.U., et al., *Gastruloids generated without exogenous Wnt activation develop anterior neural*
509 *tissues*. Stem Cell Reports, 2021. **16**(5): p. 1143-1155. DOI: 10.1016/j.stemcr.2021.03.017.
- 510 45. Yamanaka, Y., et al., *Reconstituting human somitogenesis in vitro*. Nature, 2023. **614**(7948): p. 509-
511 520. DOI: 10.1038/s41586-022-05649-2.
- 512 46. Bedzhov, I., et al., *Developmental plasticity, cell fate specification and morphogenesis in the early*
513 *mouse embryo*. Philos Trans R Soc Lond B Biol Sci, 2014. **369**(1657). DOI: 10.1098/rstb.2013.0538.
- 514 47. Izumi, Y., et al., *An Atypical PKC Directly Associates and Colocalizes at the Epithelial Tight Junction with*
515 *ASIP, a Mammalian Homologue of Caenorhabditis elegans Polarity Protein PAR-3*. Journal of Cell
516 Biology, 1998. **143**(1): p. 95-106. DOI: 10.1083/jcb.143.1.95.
- 517 48. Joberty, G., et al., *The cell-polarity protein Par6 links Par3 and atypical protein kinase C to Cdc42*. Nat
518 Cell Biol, 2000. **2**(8): p. 531-9. DOI: 10.1038/35019573.
- 519