



RESOURCE ENGINEERING

RECEIVED
ENVIRONMENTAL AGENCY
1985 FEB -3 11:52
DALLAS, TEXAS

February 4, 1985

Mr. Don Porter
Environmental Protection Agency
First International Bldg.
1201 Elm Street
Dallas, TX 75270

001065

Re: French Ltd. Water Well

Dear Don:

Please find attached information relative to activities in addressing the old French water well found at the French Ltd. site. As you know, the well was in poor shape, and we attempted to develop it and obtain some useful data from it.

The attached information describes our attempt to clean it up and the data that was obtained from one obtained sample. The data reflects the water quality when the well was first cleaned out. Subsequent attempts to obtain a sample were fruitless.

Resource Engineering recommends that the well be completely closed by pulling the steel casing and completely cementing the hole. It appears that the outside of the casing could act as a conduit for migration of contaminants into the aquifer below the Beaumont Clay Formation. Please advise either Hazel Hoffman of the task force or myself if you concur with this recommendation.

3000 Richmond Avenue, Houston, Texas 77098 • (713) 520-8900

000069

001065

REI

Mr. Don Porter
Environmental Protection Agency
February 5, 1985
Page 2

Thank you for your assistance in this matter.

Very truly yours,

RESOURCE ENGINEERING

Coly
Jacob A. Scher
Project Manager

JAS/bw

cc: Mr. Alex Anjanow - TDWR
Mrs. Hazel Hoffman, Chairman French Ltd. Task Force

Attachments

001066



RESOURCE ENGINEERING

22 January, 1985

French Limited Task Group
c/o ARCO Chemical Co.
1200 Milam
Houston, TX 77001

Attention: Ms. Hazel Hoffman

Report of chemical analysis of grab samples taken December 12, 1984 from old supply well.

<u>Parameter</u>	<u>Concentration Measured, mg/l</u>
Arsenic	<0.05
Barium	<1.0
Cadmium	<0.01
Chromium	<0.01
Lead	<0.01
Mercury	<0.002
Selenium	<0.02
Silver	<0.01
Copper	<0.01
Manganese	0.35
Nickel	<0.01
Zinc	3.20
Molybdenum	<1.0
Iron	0.93
Total Organic Halogens	0.208

001067

LAB NO. 6037 cont.

3000 Richmond Avenue, Houston, Texas 77098 • (713) 520-9900



LAB NO. 6037 cont.

<u>Parameter</u>	<u>Concentration Measured, mg/l</u>
Total Organic Carbon	67.0
Total Fixed Solids	195.0
Volatile Solids	50.0
Total Residue	245.0
Chloride	63.0
Sulfate	26.0
Oil and Grease	6.0

Very truly yours,

RESOURCE ENGINEERING

Bo Blankfield
Laboratory Director

BB/vs

LAB NO. 6037
REI PROJ. NO. 275-06

001068

Summary of Activities Performed on
French Ltd. Supply Well No. LJ-65-08-806

Wednesday, 11/28/84- Work was begun on cleanup of French Ltd. Supply Well No. LJ-65-08-806. A waterwell drilling rig was set up over the well in order to drill out any obstructions that may be lodged in the casing. A metal obstruction was immediately encountered at 6 feet below grade. Attempts to drill through the obstruction failed.

Thursday, 11/29/84- A decision was made to cut the casing off below obstruction level. A trench was dug next to the well and it was discovered that the obstruction was due to the well casing itself; it was severely bent. The steel casing was cut off and replaced with 4 inch PVC. The PVC was attached and sealed to the casing with cement.

Friday, 11/30/84- Cleanup resumed smoothly until 35 feet below grade another metal object was encountered. After three hours of drilling, workers were only able to advance 3 inches.

0100



RESOURCE ENGINEERING

22 January, 1985

French Ltd.
REI Project No. 275-06

Attention:

Report of chemical analysis of grab sample taken December 12, 1984 from old supply well.

<u>Parameter</u>	<u>Concentration Measured, mg/l</u>
Arsenic	<0.05
Barium	<1.0
Cadmium	<0.01
Chromium	<0.01
Lead	<0.01
Mercury	<0.002
Selenium	<0.02
Silver	<0.01
Copper	<0.01
Manganese	0.35
Nickel	<0.01
Zinc	3.20
Molybdenum	<1.0
Iron	0.93
Total Organic Halogens	0.208

LAB NO. 6037 cont.

3000 Richmond Avenue, Houston, Texas 77098 • (713) 520-9900

001070

001070



LAB NO. 6037 cont.

<u>Parameter</u>	<u>Concentration Measured, mg/l</u>
Total Organic Carbon	67.0
Total Fixed Solids	195.0
Volatile Solids	50.0
Total Residue	245.0
Chloride	63.0
Sulfate	26.0
Oil and Grease	6.0

Very truly yours,

RESOURCE ENGINEERING

Bo Blankfield
Laboratory Director

BB/vs

LAB NO. 6037
REI PROJ. NO. 275-06

001071

CLIENT FRENCH LTD

PROJECT NO. 275-06

SAMPLED BY D. VALLANCE

DATE SAMPLED 12-12-84

RECEIVED _____

DATE COMPLETED _____



RESOURCE ENGINEERING INC.

LAB NO. 6037

RESULTS TO C. SCHER

PROJECT MANAGER C. SCHER

ANALYTICAL COST 425⁰⁰

MILEAGE _____ COST _____

HOURS _____ RATE _____ COST _____

SUBTOTAL _____

DISCOUNT _____

TOTAL COST \$ 425⁰⁰

SAMPLE DESCRIPTION / PARAMETER	010 sample well MW # 3																		
ARSENIC	<0.05	mg/L																	
BARIUM	<1.0																		
CADMIUM	<0.01																		
CHROMIUM	20.01																		
LEAD	20.01																		
MERCURY	<0.002																		
SELENIUM	<0.02																		
SILVER	20.01																		
COPPER	<0.01																		
MANGANESE	0.35																		
NICKEL	20.01																		
ZINC	3.20																		
MOLYBDENUM	<1.0																		
PTOX Total Organic Halogens	0.208																		
TOC " "	67.0	V																	
TSS (Total Suspended Solids)	19.5																		
VSS (Volatile Suspended Solids)	5.0																		
CHLORIDE	63	mg/L																	
SULFATE	26.0	mg/L																	
PH	6.0	mg/L																	
Fe	0.93	mg/L																	
TSS Total	24.5																		
TESTED BY	GM/BB																		
ANALYSIS COST																			

it was concluded that the metal obstruction was most likely the drop pipe of the submersible pump. A steel rod was dropped down-hole in an effort to tag bottom. Workers were able to lower rod to 200 feet. It was determined that the top of the submersible pump sits at 200 feet below surface. The drilling rig was taken off site.

Wednesday, 12/5/84- A service rig was set up over the well to try to pull out the drop pipe and submersible pump. Six attempts were made to pull the pump and drop pipe. Five tons of lifting force was ultimately applied to the assembly but drop pipe and pump would not move.

Thursday, 12/6/84- A 3/4 inch wash rod was set to the top of the pump. This rod will be used for sampling purposes. The well casing was cut off close to ground surface (0.9 ft. stickup) in case a drilling rig should have to set up over it again. A protective steel cover was installed over the well casing and secured with a padlock.

Wednesday, 12/12/84- Sampling of the well was attempted. Before purging the well, an initial set of water samples was bailed and collected by REI and a TDWR representative. After sampling was completed, a special well cover was installed. This well cover directed the water rising from the well during

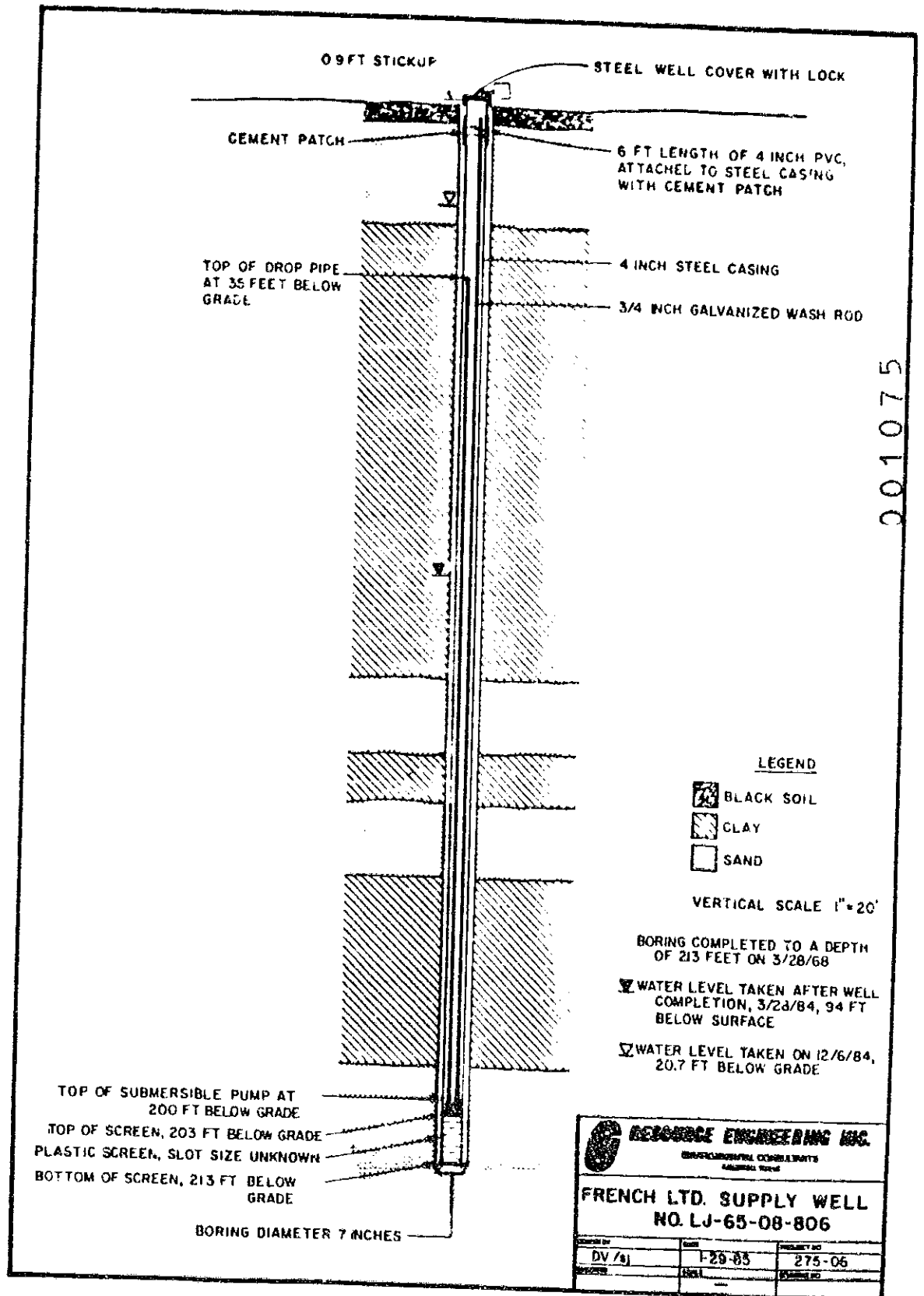
001073

purgin, thro a hose which ran to the lagoon. Purgin the well was accomplished by directing compressed air through the 3/4 inch wash rod. After approximately 1.5 well volumes of water had been removed, the well appeared to stop accepting the compressed air. It was believed that the wash rod became clogged, thus restricting the air. Bailing the well for sampling was then attempted; however, the bailer could not be dropped lower than 60 feet; the water level was at 88 feet.

Tuesday, 12/18/84- Another attempt was made to purge and sample the well; However, wash rod was still restricting air. Water level is at 76 feet below grade and cannot be reached by bailer.

Wednesday, 1/9/85- Water level is at 5.31 feet below grade.

00074

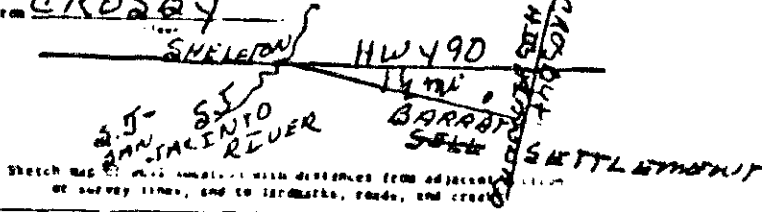


001075

1) OWNER
 Person having well drilled FRENCH LTD of HOUSTON INC. Address 11023 TELEPHON RD TX
 Landowner FRENCH LTD of HOUSTON INC. Address 11023 TELEPHON RD HOUSTON TX

2) LOCATION
 County HARRIS Labor _____ Abstract No. _____
 NE1/4 SW1/4 SE1/4 of Section _____ Block No. _____ Survey _____

Well is 1/2 mile direction from CROSBY
 of SE



3) TYPE OF WELL (Check)
 New well Jeopardy
 Reconditioning Plugging
 Proposed Use (Check)
 Domestic Industrial Municipal
 Irrigation Test Well Other
 Type of Well (Check)
 Cased Driven Auger Jetted Bored

6) WELL LOG
 Diameter of hole 7 in. Depth drilled 213 ft. Depth of completed well 213 ft. Date drilled 3-28

All measurements made from _____ ft. above ground level.

From (ft.)	To (ft.)	Description and color of formation material	From (ft.)	To (ft.)	Description and color of formation material
0	3	BLACK LAND			
3	24	SAND			
24	115	CLAY			
115	130	SAND			
130	140	CLAY			
140	155	SAND			
155	193	CLAY			
193	213	SALT + PEPPERS SAND			

7) COMPLETION (Check)
 Straight well Gravel packed Other
 Under reamed Open hole
 8) WATER LEVEL
 Static level 94 ft. below land surface Date _____
 Artesian pressure _____ lbs. per square inch Date _____

9) CASING
 Type old New Steel Plastic Other
 Cemented from _____ ft. to _____ ft.

10) SCREEN
 Type PLASTIC SCREEN
 Perforated Slotted

Diameter (Inches)	Setting		Gauge	Diameter (inches)	Setting		Slot Size
	From (ft.)	To (ft.)			From (ft.)	To (ft.)	
4"	0	20 3/4	MED.				

11) WELL TESTS
 Was a pump test made? Yes No If yes by whom Gal Amin Duma
 Yield: _____ gpm with _____ ft. drawdown after _____ hrs.
 Saline test _____ gpm with _____ ft. drawdown after _____ hrs.
 Artesian flow _____ gpm Date _____
 Temperature of water _____
 Was a chemical analysis made? Yes No
 Did any strata contain undesirable water? Yes No
 Type of water? _____ depth of strata _____

12) PUMP DATA
 Manufacturer's Name _____
 Type _____ H.P. _____
 Designed pumping rate _____ gpm _____ gph
 Type power unit _____
 Depth to bowls, cylinder, jet, etc., _____ below land surface.

I hereby certify that this well was drilled by me (or under my supervision) and that each one of the statements herein are true to the best of my knowledge and belief.
 Name J.W. GREAK Water Well Drillers Registration No. 617
 Address P.O. Box 92, HWY 90 LIBERTY TX
J.W. Greak T. EVANS

001076

Technical Report

for

RESOURCE ENGINEERING

ATTENTION: CHRIS ITIN

3000 RICHMOND AVENUE

HOUSTON, TX 77098

001077

Chain of Custody Data Required for ETC Data Management Summary Reports

<i>ETC Sample No.</i>	<i>Company</i>	<i>Facility</i>	<i>Sample Point</i>	<i>Date</i>	<i>Time</i>	<i>Elapsed Hours</i>
G4116	RESOURCE ENGINEERING	RES27506	WFRI	841212		

June A. Schaper
Denis C. K. Lin, Ph.D. *For*
Vice President
Research and Operations

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001078

Introduction

This report contains the analytical results on your water sample, WFR 84-12/12. It is designed to include comprehensive data from the entire analytical process in order to satisfy the needs of various levels of review.

The results obtained from your sample are presented in tabular format immediately following this introduction. Quality assurance data is tabulated along with the appropriate sample results for verification. Depending on the analyses ordered, the quality assurance data may include results from blank, spiked blank, spiked sample (i.e. matrix spike) and replicate sample as well as results from surrogate compound analyses. Quality assurance data for verification of proper instrument performance is also included where appropriate. The report appendices include the chain of custody record for your sample and, where appropriate, the gas chromatograms and mass spectra.

The procedures used in the analysis of the sample are described in this report's methodology section. All analytical procedures within our laboratory are performed within a strict, enforced Quality Assurance Protocol. A description of this Protocol is included in the report.

Results

Sample results, and associated quality assurance data, are always tabulated in one or more of this report's Quantitative Results Tables. The format of each table varies with the class of analysis.

Priority Pollutants

The priority pollutant compounds and elements are listed with their NPDES (National Pollution Discharge Elimination System) numbers, and the Method Detection Limit (MDL) published in the Federal Register, December 3, 1979. When a compound or element is present below its published MDL it is reported as BMDL (Below Method Detection Limit). When a compound or element is not present at any detectable concentrations it is reported as ND (Not Detected). Matrix spike and replicate analyses, where included, were performed on samples randomly chosen within each quality assurance batch and are therefore not necessarily spikes and replicates of this report's sample. Surrogate compound recovery data and instrument calibration data are included in the Method Performance Data Tables.

Qualitative Analysis

The GC/MS identification of compounds which are not included in the normal compound list for a given method is reported in a Quantitative Results Table. Each sample fraction is reported on an individual Quantitative Results Table. Wherever possible these compounds are reported with their molecular weight, empirical formula, and CAS number. The retention time and scan number is also provided to match the spectra included in the report appendices.

001079

TABLE 1: QUANTITATIVE RESULTS and QUALITY ASSURANCE DATA
Volatile Compounds - GC/MS Analysis Data (QR01)

Cham of Custody Data Required for ETC Data Management Summary Reports					
G4116	RESOURCE ENGINEERING	RES27506	WFRI	841212	
ETC Sample No.	Company	Facility	Sample Point	Date	Elapsed Hours

NPDES Number	Compound	Results		QC Replicate		QC Blank and Spiked Blank			QC Matrix Spik		
		Sample Concn. ug/l	MDL ug/l	First ug/l	Second ug/l	Blank Data ug/l	Concn. Added ug/l	% Recov	Unspiked Sample ug/l	Concn Added ug/l	% Recov
1V	Acrolein	ND	100	ND	ND	ND	1600	85	ND	1600	54
2V	Acrylonitrile	ND	100	ND	ND	ND	160	121	ND	160	115
3V	Benzene	12	10	17	18	ND	36	101	ND	36	91
4V	bis(Chloromethyl)ether	ND	10	ND	ND	ND	0	-	ND	0	-
5V	Bromoform	ND	10	ND	ND	ND	36	106	ND	36	94
6V	Carbon tetrachloride	ND	10	ND	ND	ND	36	105	ND	36	87
7V	Chlorobenzene	ND	10	ND	ND	ND	36	103	ND	36	91
8V	Chlorodibromomethane	ND	10	ND	ND	ND	36	105	ND	36	97
9V	Chloroethane	BMDL	10	ND	ND	ND	36	98	ND	36	90
10V	2-Chloroethylvinyl ether	ND	10	ND	ND	ND	36	168	ND	36	155
11V	Chloroform	ND	10	ND	ND	ND	36	106	ND	36	97
12V	Dichlorobromomethane	ND	10	ND	ND	ND	36	107	ND	36	98
13V	Dichlorodifluoromethane	ND	10	ND	ND	ND	36	66	ND	36	47
14V	1,1-Dichloroethane	ND	10	22	22	ND	36	108	ND	36	99
15V	1,2-Dichloroethane	ND	10	ND	ND	ND	36	109	28	36	77
16V	1,1-Dichloroethylene	ND	10	8	8	ND	36	103	ND	36	88
17V	1,2-Dichloropropane	ND	10	ND	ND	ND	36	107	ND	36	94
18V	cis-1,3-Dichloropropylene	ND	10	ND	ND	ND	36	104	ND	36	89
19V	Ethylbenzene	ND	10	ND	ND	ND	36	105	ND	36	90
20V	Methyl bromide	ND	10	ND	ND	ND	36	104	ND	36	86
21V	Methyl chloride	ND	10	ND	ND	ND	36	85	ND	36	72
22V	Methylene chloride	BMDL	10	10	7	BMDL	36	102	ND	36	111
23V	1,1,2,2-Tetrachloroethane	ND	10	ND	ND	ND	36	107	ND	36	100
24V	Tetrachloroethylene	BMDL	10	ND	ND	ND	36	102	ND	36	88
25V	Toluene	13	10	ND	ND	BMDL	36	94	ND	36	82
26V	1,2-Trans-dichloroethylene	ND	10	3650	4495	ND	36	106	ND	36	90
27V	1,1,1-Trichloroethane	ND	10	ND	ND	BMDL	36	86	ND	36	87
28V	1,1,2-Trichloroethane	ND	10	ND	ND	ND	36	106	ND	36	100
29V	Trichloroethylene	ND	10	114	112	ND	36	107	ND	36	89
30V	Trichlorofluoromethane	BMDL	10	ND	ND	ND	36	101	ND	36	92
31V	Vinyl chloride	ND	10	121	115	ND	36	86	ND	36	75
18V	trans-1,3-Dichloropropylene	ND	10	ND	ND	ND	36	101	ND	36	88

A. EPR published Method Detection Limit.
 B. Recovery normally variable using EPR Protocol Method G24.
 C. Recovery low due to sample matrix interference.

001080

TABLE 1: QUANTITATIVE RESULTS and QUALITY ASSURANCE DATA
Acid Compounds - GC/MS Analysis Data (QR02)

Chain of Custody Data Required for ETC Data Management Summary Reports					
G4116	RESOURCE ENGINEERING	RES27506	WFR1	841212	
ETC Sample No.	Company	Facility	Sample Point	Date	Time Elapsed Hours

NPDES Number	Compound	Results		QC Replicate		QC Blank and Spiked Blank			QC Matrix Spike		
		Sample Concn. ug/l	MDL ug/l *	First ug/l	Second ug/l	Blank Data ug/l	Concn. Added ug/l	% Recov	Unspiked Sample ug/l	Concn Added ug/l	% Recov
1A	2-Chlorophenol	ND	25	ND	ND	ND	150	82	ND	150	90
2A	2,4-Dichlorophenol	ND	25	ND	ND	ND	150	79	ND	150	83
3A	2,4-Dimethylphenol	ND	25	1	1	ND	150	88	ND	150	81
4A	4,6-Dinitro-o-cresol	ND	250	ND	ND	ND	250	90	ND	250	95
5A	2,4-Dinitrophenol	ND	250	ND	ND	ND	250	107	ND	250	118
6A	2-Nitrophenol	ND	25	5	4	ND	150	90	ND	150	73
7A	4-Nitrophenol	ND	25	ND	ND	ND	150	35.	ND	150	50.
8A	p-Chloro-m-cresol	ND	25	ND	ND	ND	150	114	ND	150	87
9A	Pentachlorophenol	BMDL	25	ND	ND	ND	150	82	ND	150	75
10A	Phenol	ND	25	45	42	ND	150	72	ND	150	85
11A	2,4,6-Trichlorophenol	ND	25	1	1	ND	150	97	ND	150	98

* EPA published Method Detection Limit.
B Recovery normally low using EPA Protocol Method 825.

001081

**TABLE 1: QUANTITATIVE RESULTS and QUALITY ASSURANCE DATA
BASE/NEUTRAL COMPOUNDS - GC/MS ANALYSIS DATA (OR03)**

Chain of Custody Data Required for ETC Data Management Summary Reports					
G4116	RESOURCE ENGINEERING	RES27506	WFR1	841212	
ETC Sample No.	Company	Facility	Sample Point	Date	Elapsed Time Hours

NPDES Number	Compound	Results		QC Replicate		QC Blank and Spiked Blank			QC Matrix Spike		
		Sample Concn. ug/l	MDL ug/l	First ug/l	Second ug/l	Blank Data ug/l	Concn. Added ug/l	% Recov	Unspiked Sample ug/l	Concn. Added ug/l	% Recov
1B	Acenaphthene	ND	10	ND	ND	ND	100	90	ND	100	92
2B	Acenaphthylene	ND	10	ND	ND	ND	100	88	ND	100	93
3B	Anthracene	ND	10	ND	ND	ND	100	92	ND	100	92
4B	Benzidine	ND	10	ND	ND	ND	100	32	ND	100	58
5B	Benzo(a)anthracene	ND	10	ND	ND	ND	100	90	ND	100	90
6B	Benzo(a)pyrene	ND	10	ND	ND	ND	100	84	ND	100	92
7B	Benzo(b)fluoranthene	ND	10	ND	ND	ND	100	79	ND	100	92
8B	Benzo(ghi)perylene	ND	10	ND	ND	ND	0	-	ND	0	-
9B	Benzo(k)fluoranthene	ND	10	ND	ND	ND	100	90	ND	100	87
10B	bis(2-Chloroethoxy)methane	ND	10	ND	ND	ND	100	88	ND	100	96
11B	bis(2-Chloroethyl) ether	ND	10	ND	ND	ND	100	88	ND	100	91
12B	bis(2-Chloroisopropyl)ether	ND	10	ND	ND	ND	100	89	ND	100	95
13B	bis(2-Ethylhexyl)phthalate	ND	10	ND	ND	ND	100	84	ND	100	94
14B	4-Bromophenyl phenyl ether	ND	10	ND	ND	ND	100	91	ND	100	84
15B	Butyl benzyl phthalate	ND	10	ND	ND	ND	100	80	ND	100	80
16B	2-Chloronaphthalene	ND	10	ND	ND	ND	100	84	ND	100	89
17B	4-Chlorophe 1 phenyl ether	ND	10	ND	ND	ND	100	89	ND	100	95
18B	Chrysene	ND	10	ND	ND	ND	100	90	ND	100	90
19B	Dibenzo(a,h)anthracene	ND	10	ND	ND	ND	0	-	ND	0	-
20B	1,2-Dichlorobenzene	ND	10	ND	ND	ND	100	82	ND	100	88
21B	1,3-Dichlorobenzene	ND	10	ND	ND	ND	100	75	ND	100	81
22B	1,4-Dichlorobenzene	ND	10	ND	ND	ND	100	80	ND	100	80
23B	3,3'-Dichlorobenzidine	ND	10	ND	ND	ND	100	55	ND	100	64
24B	Diethyl phthalate	ND	10	ND	ND	ND	100	9	ND	100	15
25B	Dimethyl phthalate	ND	10	ND	ND	ND	100	10	ND	100	11
26B	Di-n-butyl phthalate	ND	10	ND	ND	ND	100	69	ND	100	86
27B	2,4-Dinitrotoluene	ND	10	ND	ND	ND	100	102	ND	100	127
28B	2,6-Dinitrotoluene	ND	10	ND	ND	ND	100	112	ND	100	12
29B	Di-n-octyl phthalate	ND	10	ND	ND	ND	100	75	ND	100	9
30B	1,2-Diphenylhydrazine	ND	10	ND	ND	ND	100	91	ND	100	101
31B	Fluoranthene	ND	10	ND	ND	ND	100	78	ND	100	95
32B	Fluorene	ND	10	ND	ND	ND	100	86	ND	100	93

001082

**TABLE 1: QUANTITATIVE RESULTS and QUALITY ASSURANCE DATA
BASE/NEUTRAL COMPOUNDS - GC/MS ANALYSIS DATA (QR03)**

Chain of Custody Data Required for ETC Data Management Summary Reports					
ETC Sample No.	Company	Facility	Sample Point	Date	Time Elapsed Hours
G4116	RESOURCE ENGINEERING	RES27506	WFR1	841212	

NPDES Number	Compound	Results		QC Replicate		QC Blank and Spiked Blank			QC Matrix Spike		
		Sample Concn. ug/l	MDL ug/l ^a	First ug/l	Second ug/l	Blank Data ug/l	Concn. Added ug/l	% Recov	Unspiked Sample ug/l	Concn Added ug/l	% Recov
33B	Hexachlorobenzene	ND	10	ND	ND	ND	100	93	ND	100	85
34B	Hexachlorobutadiene	ND	10	ND	ND	ND	100	64	ND	100	72
35B	Hexachlorocyclopentadiene	ND	10	ND	ND	ND	0	-	ND	0	-
36B	Hexachloroethane	ND	10	ND	ND	ND	100	60	ND	100	70
37B	Indeno(1,2,3-c,d)pyrene	ND	10	ND	ND	ND	0	-	ND	0	-
38B	Isophorone	ND	10	ND	ND	ND	100	96	ND	100	100
39B	Naphthalene	BMDL	10	ND	ND	ND	100	85	ND	100	89
40B	Nitrobenzene	ND	10	ND	ND	ND	100	91	ND	100	93
41B	N-Nitrosodimethylamine	ND	10	ND	ND	ND	0	-	ND	0	-
42B	N-Nitrosodi-n-propylamine	ND	10	ND	ND	ND	100	95	ND	100	97
43B	N-Nitrosodiphenylamine	ND	10	ND	ND	ND	100	92	ND	100	106
44B	Phenanthrene	ND	10	ND	ND	ND	100	90	ND	100	91
45B	Pyrene	ND	10	ND	ND	ND	100	78	ND	100	95
46B	1,2,4-Trichlorobenzene	ND	10	ND	ND	ND	100	135	ND	100	191

^a EPA published Method Detection Limit.
^b Recovery normally low using EPA Protocol Method 825.
^c Recovery normally variable using EPA Protocol Method 825.

001083

TABLE 1: QUANTITATIVE RESULTS and QUALITY ASSURANCE DATA
Pesticide/PCB Compounds - GC/MS Analysis Data (QR04)

Chain of Custody Data Required for ETC Data Management Summary Reports

G4116	RESOURCE ENGINEERING	RES27506	WFR1	841212
ETC Sample No.	Company	Facility	Sample Point	Date
				Time Elapsed Hours

NPDES Number	Compound	Results		QC Replicate		QC Blank and Spiked Blank			QC Matrix Spike		
		Sample Concn. ug/l	MDL ug/l *	First ug/l	Second ug/l	Blank Data ug/l	Concn. Added ug/l	% Recov .	Unspiked Sample ug/l	Concn Added ug/l	% Recov .
1P	Aldrin	ND	10	ND	ND	ND	100	84	ND	100	49
2P	Alpha-BHC	ND	10	ND	ND	ND	100	17	ND	100	35
3P	Beta-BHC	ND	10	ND	ND	ND	100	68	ND	100	81
4P	Gamma-BHC	ND	10	ND	ND	ND	100	20	ND	100	41
5P	Delta-BHC	ND	10	ND	ND	ND	100	27	ND	100	14
6P	Chlordane	ND	10	ND	ND	ND	200	38	ND	200	67
7P	4,4'-DDT	ND	10	ND	ND	ND	100	79	ND	100	41
8P	4,4'-DDE	ND	10	ND	ND	ND	100	88	ND	100	39
9P	4,4'-DDD	ND	10	ND	ND	ND	100	94	ND	100	62
10P	Dieldrin	ND	10	ND	ND	ND	100	94	ND	100	76
11P	Endosulfan I	ND	10	ND	ND	ND	100	24	ND	100	25
12P	Endosulfan II	ND	10	ND	ND	ND	100	26	ND	100	21
13P	Endosulfan sulfate	ND	10	ND	ND	ND	100	54	ND	100	64
14P	Endrin	ND	10	ND	ND	ND	100	22	ND	100	74
15P	Endrin aldehyde	ND	10	ND	ND	ND	100	38	ND	100	20
16P	Heptachlor	ND	10	ND	ND	ND	100	87	ND	100	63
17P	Heptachlor epoxide	ND	10	ND	ND	ND	100	88	ND	100	77
18P	PCB-1242	ND	10	ND	ND	ND	0	-	ND	0	-
19P	PCB-1254	ND	10	ND	ND	ND	0	-	ND	0	-
20P	PCB-1221	ND	10	ND	ND	ND	0	-	ND	0	-
21P	PCB-1232	ND	10	ND	ND	ND	0	-	ND	0	-
22P	PCB-1248	ND	10	ND	ND	ND	0	-	ND	0	-
23P	PCB-1260	ND	10	ND	ND	ND	100	30	ND	100	41
24P	PCB-1016	ND	10	ND	ND	ND	0	-	ND	0	-
25P	Toxaphene	ND	10	ND	ND	ND	0	-	ND	0	-

* EPA published Method Detection Limit.
 † Recovery normally variable using EPA Protocol Method 825.

001084

TABLE 1: QUALITATIVE RESULTS

Tentatively Identified Organic Compounds - GC/MS Analysis Data - Volatile Fraction (QR06)

Chain of Custody Data Required for ETC Data Management Summary Reports						
G4116	RESOURCE ENGINEERING	RES27506	WFRI	841212		
ETC Sample No.	Company	Facility	Sample Point	Date	Time	Elapsed Hours

Compound Name	Data			Identifiers		Estim. Concent. ug/l		
	Scan Number	Retention Time (Min)	M W	CAS Number	Empirical Formula			
1 None identified	119	5.68	-	-	-	-		
2 None identified	162	7.34	-	-	-	-		
3 None identified	185	8.24	-	-	-	-		
4 1-Propene, 1-methoxy	223	9.71	72	7319166	C ₄ H ₈ O	2588		
5 None identified	238	10.29	-	-	-	-		
6 2-Butanone (MEK)	277	11.80	72	78933	C ₄ H ₈ O	2225		
7 None identified	299	12.66	-	-	-	-		
8 None identified	309	13.04	-	-	-	-		
9 2H-Pyran, tetrahydro-3-methyl	460	18.90	100	26093630	C ₆ H ₁₂ O	5.4		
10 Unknown peak	473	19.40	-	-	-	214		
11 Unknown peak	491	20.10	-	-	-	8		
12 Benzene-ethyl-	639	25.83	106	100414	C ₈ H ₁₀	10		

001085

TABLE 1: QUALITATIVE RESULTS

Tentatively Identified Organic Compounds - GC/MS Analysis Data - Acid Fraction (QR07)

Chain of Custody Data Required for ETC Data Management Summary Reports						
G4116	RESOURCE ENGINEERING	RES27506	WFR1	121284		
ETC Sample No.	Company	Facility	Sample Point	Date	Time	Elapsed Hours

Compound Name	Data			Identifiers		Estim. Concent. ug/l	
	Scan Number	Retention Time (Min)	M.W.	CAS Number	Empirical Formula		
1 2-Hexen-1-ol	45	3.9	100	928949	C ₆ H ₁₂ O	28	
2 None identified	151	-	-	-			

001086

TABLE 1: QUALITATIVE RESULTS

Tentatively Identified Organic Compounds - GC/MS Analysis Data - Base/Neutral Fraction (QR08)

Chain of Custody Data Required for ETC Data Management Summary Reports						
G4116	RESOURCE ENGINEERING	RES27506	WFRI	841212		
ETC Sample No.	Company	Facility	Sample Point	Date	Time	Elapsed Hours

Compound Name	Data			Identifiers		Estimated Concentration (ug/l)	
	Scan Number	Retention Time (Min)	M.W.	CAS Number	Empirical Formula		
1 Methyl-benzene	13	2.96	92	108883	C7H8	9	
2 Unknown	71	3.99	-	-	-	47	
3 2-Hexen-1-ol	92	4.36	100	928949	C6H12O	171	
4 Cyclohexanol	95	4.42	100	108930	C6H12O	11	
5 2-Cyclohexen-1-one	130	5.04	96	930687	C6H8O	11	
6 none identified	418	10.14	-	-	-	10	

001087

TABLE 2: METHOD PERFORMANCE DATA

TABLE 2: METHOD PERFORMANCE DATA
Surrogate Recovery Water- GC/MS Data (QR20)

Chain of Custody Data Required for ETC Data Management Summary Reports

G4116

ETC Sample No.	Company	Facility	Sample Point	Date	Time	Elapsed Hours
----------------	---------	----------	--------------	------	------	---------------

Compound	Amount Added ug	% Recovery	Control Limits *	
			Lower	Upper
<i>VOLATILE FRACTION</i>				
Toluene-D8	.250	106	86	119
Bromofluorobenzene	.250	105	85	121
1,2-Dichloroethane-D4	.250	88	77	120
<i>ACID FRACTION</i>				
Phenol-D5	100	30	15	103
2-Fluorophenol	100	66	23	121
2,4,6-Tribromophenol	100	81	10	130
<i>BASE/NEUTRAL FRACTION</i>				
Nitrobenzene-D5	50	72	41	120
2-Fluorobiphenyl	50	80	44	119
Terphenyl-D14	50	63	33	128
* (FE EPA Control Limits.				

001088

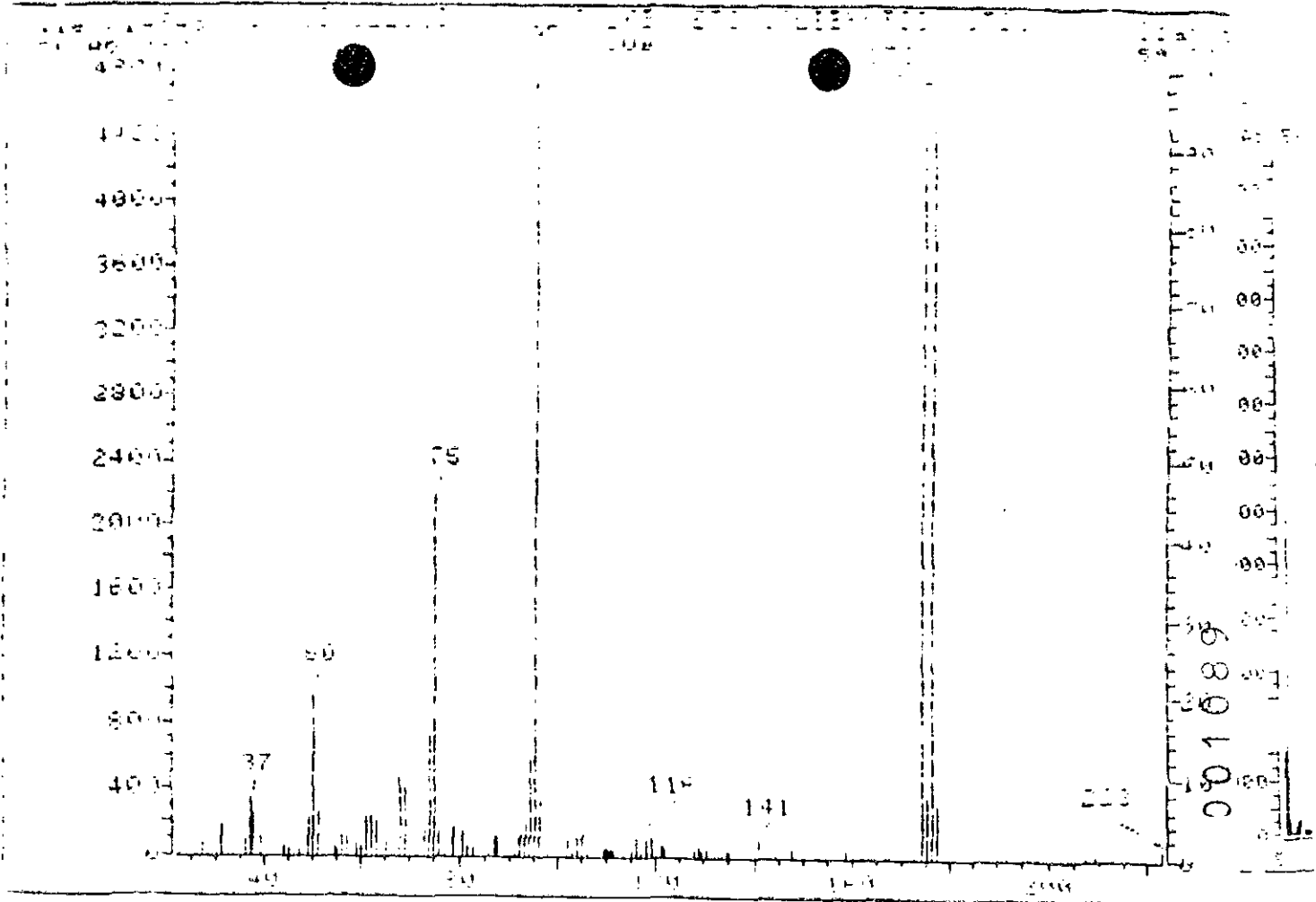


TABLE 2. METHOD PERFORMANCE DATA (CONT.)

NOTE: IUPAC Name: Benzofuran; CAS No. 100-04-1; Molecular Weight: 144.12

m/z	Ion Abundance Criteria	Relative Abundance Base Peak	Approximate Peak	Retention Time
37	0.40% of mass 95	20.30	20.30	01
50	0.70% of mass 95	25.05	25.05	01
95	Base peak, 100% relative abundance	100.00	100.00	01
96	5.0% of mass 95	5.00	5.00	01
173	Less than 1% of mass 95	0.00	0.00	01
174	Greater than 50% of mass 95	97.12	97.12	01
175	5.0% of mass 174	5.97	5.97	01
176	95-101% of mass 174	97.21	100.19	01
177	5.0% of mass 174	5.56	5.74	01

Injection Date: 12/26/84
 Injection Time: 04:30
 Run No.: 218546
 Spectrum No.: 54

Analyst: *J. Martin*
 Name: *Chengwan AO*
 QC Batch: *RV 2649*
 Sample: *41871, 41872, 41874, 41875, 41879, 41892, 41905, 41909*

B

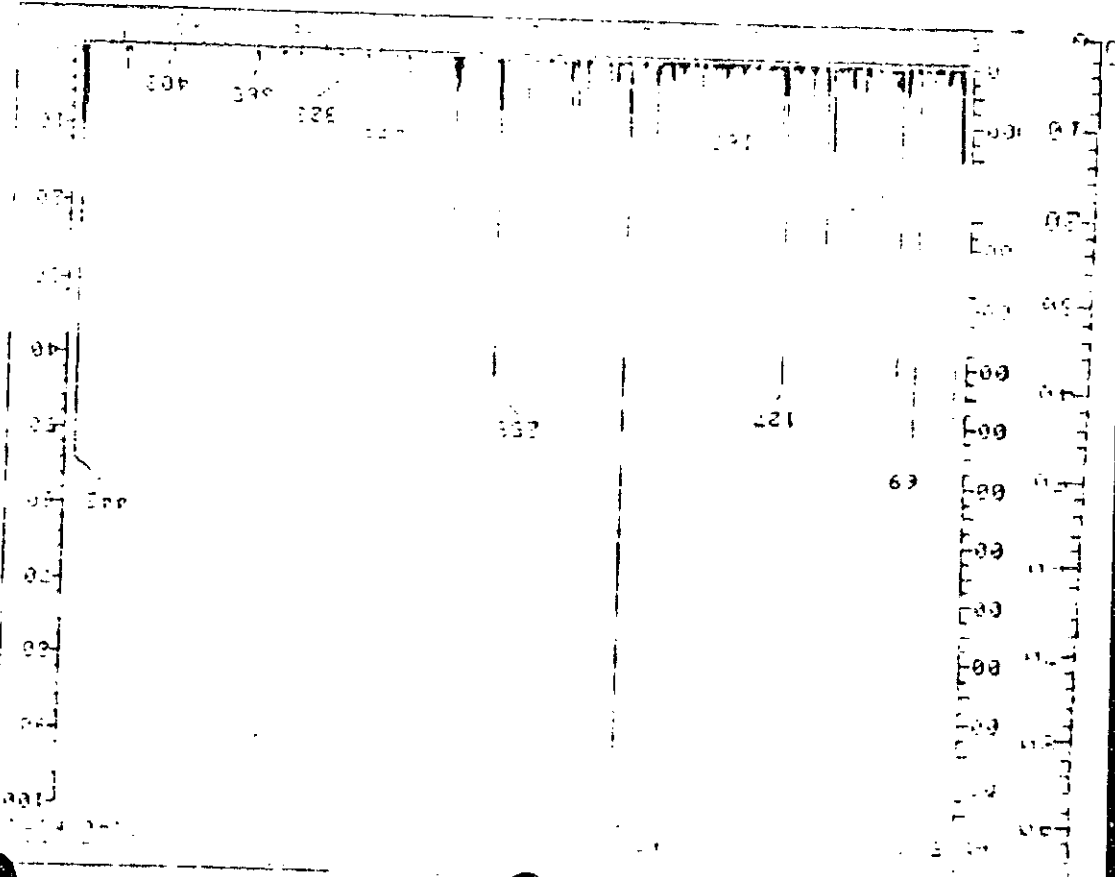
RB

60673, 61654
64116, 60668, 60669, 60672
62806, 63682, 63215
QA2525
John Marino
Richard Gray

Analyst:
QC Batch:
Sample:
Spectrum #: 258
Run No: F8271
Injection Date: 11/21/84

17.2%	0.79	18.22
Greater than 1%	2.65	28.16
Less than 1%	21.22	21.22
10-20% of mass	6.66	6.66
5-10% of mass	10.00	10.00
Base peak, 100% relative abundance	42.09	42.09
Less than 1% of mass	42.09	42.09
40-60% of mass	50.00	50.00
Less than 1% of mass	40.00	40.00

% Relative Abundance
Base Peak
Scan



001090

File 507 348103 ,G DFTPP SONG DFTPP STD. Scan 1085
 Spk Ab 57232 Job 10.53 min.

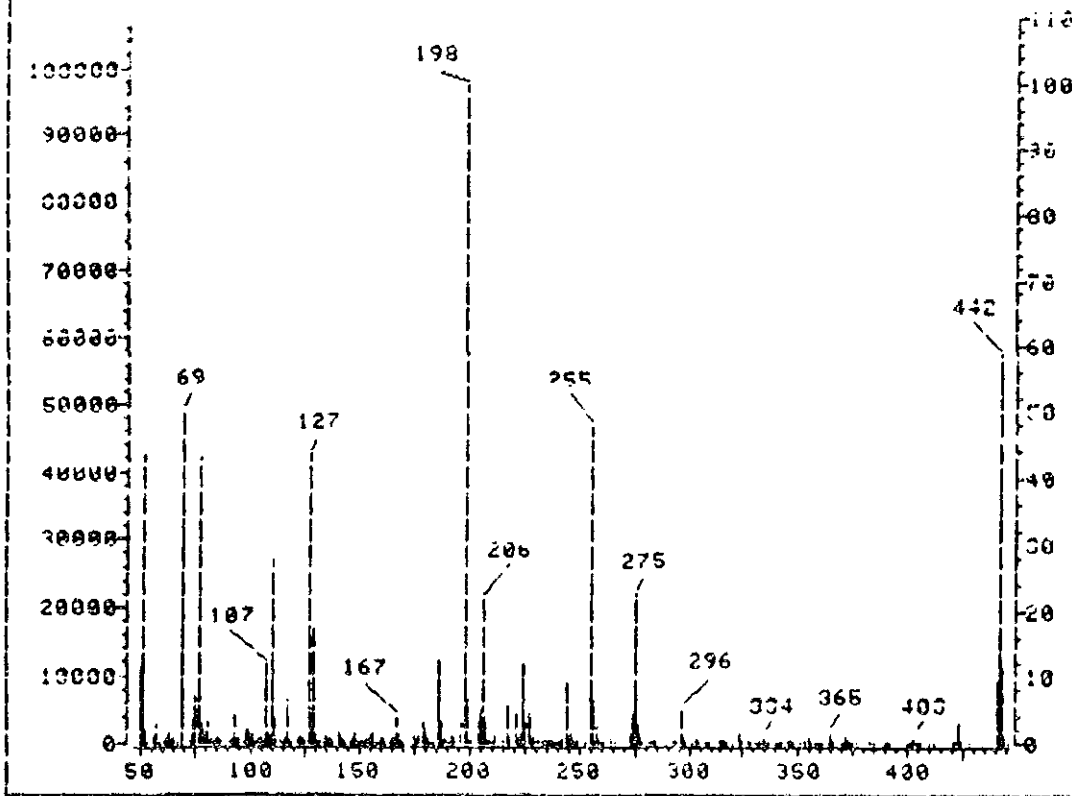


TABLE 2: METHOD PERFORMANCE DATA (UR23)

GC/MS Tuning Data - Decafluorotriphenylphosphine (DFTPP) for Base/Neutral Analysis

m/z	Ion Abundance Criteria	% Relative Abundance Base Peak	% Relative Abundance Appropriate Peak	Status
51	30-60% of mass 198	45.65	45.65	OK
68	Less than 2% of mass 69	0.00	0.00	OK
69	(reference only)	50.39	50.39	OK
70	Less than 2% of mass 69	0.00	0.00	OK
127	40-60% of mass 198	43.63	43.63	OK
197	Less than 1% of mass 198	0.00	0.00	OK
198	Base peak, 100% relative abundance	100.00	100.00	OK
199	5-9% of mass 198	6.62	6.62	OK
275	10-30% of mass 198	22.45	22.45	OK
365	Greater than 1% of mass 198	2.06	2.06	OK
441	Less than mass 443	9.68	83.91	OK
442	Greater than 40% of mass 198	58.67	58.67	OK
443	17-23% of mass 442	11.53	19.65	OK

Injection Date: 01/03/89
 Injection Time: 17:49
 Run No: >66507
 Spectrum No: 1085

Analyst: *[Signature]*
 Processor: *[Signature]*
 QC Batch: G82525
 Samples: G0669, G0672, G0673, F-1654, F8
 G-1019, F9625, F9626, G3682, G3215
 G4110, G0668

001091

Methodology for GC/MS Analysis of Priority Pollutant Compounds

The methods employed in the GC/MS analysis for priority pollutants are established EPA methods. Rigid compliance with the instrument parameters and performance criteria of the published methods was achieved. In some cases, the precise amounts of sample used and the sample handling procedures vary with the complexity of the sample matrix. Qualitative identification of the priority pollutants was performed using the relative retention times, the relative abundance of three characteristic ions and the abundance ratios. The entire mass spectrum was reviewed to confirm each identification. Quantitative analysis of detected compounds was performed by using a response factor generated by a major characteristic ion of the specific compound and an internal standard.

Compounds, in addition to those on the priority pollutant list, were identified through a computer-aided search of the NBS-EPA electronic library. After review, the identifications are included in a separate tabulation and labeled "tentatively identified".

Volatile Priority Pollutant Compounds

For the analysis of volatile priority pollutants, EPA Method 624 (Federal Register, December 3, 1979, page 69532) was used. The method can be summarized as follows: Heptane is added to the sample and the mixture is heated to a temperature of 40°C. The purgeable volatile organic compounds are transferred from the aqueous phase to the vapor phase. The vapor is swept through a sorbent column where the organic components are trapped. After the purge cycle is complete, the sorbent column is heated and backflushed with helium to desorb the organic purgeables onto a gas chromatographic column. The gas chromatograph is temperature programmed to separate the purgeable mixture. The separated purgeable components are then identified and quantitated using a computerized mass spectrometer.

Acid Base/Neutral and Pesticide Priority Pollutant Compounds

For the analysis of the Acid, Base/Neutral and Pesticide priority pollutants in an aqueous liquid matrix, EPA Method 600 (Federal Register, December 3, 1979, page 69540) was used. The method can be summarized as follows: A measured volume of sample, approximately 1 liter, is adjusted to a pH of 12 and extracted with methylene chloride. The pH of the sample is adjusted to a value less than 2 and extracted with an equal volume of fresh methylene chloride. A separator, funnel or continuous extractor is used to perform the extractions. The two extracts are dried and concentrated to a final volume. Each extract is injected into a GC/MS instrument specifically configured for the correct fraction.

001092

Neutral

Status

- UK
- UK
- UK
- UK
- UK
- UK
- UK
- UK
- UK
- UK
- UK
- UK
- UK

F84
3215

Summary of Quality Assurance/Quality Control Procedures (QA/QC)

ETC bases its quality assurance protocols on the following government guidelines.

- "Handbook for Analytical Quality Control in Water and Wastewater Laboratories", EPA-600/4-79-019, March 1979;
 - National Enforcement Investigation Center Policies, and Procedures manual: EPA-330/9, 79-001-R, October 1979;
 - the recommended guidelines for EPA Methods 624 and 625. (Federal Register, December 3, 1979, pp. 69532-69559);
 - "Manual of Analytical Methods for the Analysis of Pesticides in Humans and Environmental Samples," EPA 600/8-80-038, June 1980; and
 - "Determination of 2,3,7,8-TCDD in Soil and Sediment" EPA, Region VII, Kansas City, September 1983
- Organic Analysis: Multi-media, Multi-Concentration-IFB WA84-A267
- Dioxin Analysis: Soil/Sediment Matrix, Multi-Concentration, Selected Ion Monitoring with Jar Extraction Procedure-IFB WA84-A002

However, we have modified our protocols to provide a higher level of QA/QC than the guidelines require. For example, we analyze a higher than required number of quality control samples and we pay especially careful attention to the certification of the "reference standard" compounds we use in analysis. Below are listed the key QA/QC elements for the methods we used.

Analysis of Volatile Organic Compounds by Gas Chromatography/Mass Spectrometry

- Each batch of 13 samples consists of 9 customer samples (at a maximum), one blank sample, one spiked blank, one spiked sample and one replicate sample. This amounts to a 30% quality control factor
- Three surrogate compounds are added to each sample in the batch of 13.
- A blind quality control sample is introduced to the laboratory for analysis on a weekly basis
- Each GC/MS is checked and retuned, if necessary, at the beginning of each day to ensure that its performance on bromofluorobenzene (BFB) meets the EPA criteria.
- A calibration curve for quantitation is prepared using a mixture of Volatile Organic Priority Pollutant "standards" at a minimum of 3 different concentrations and using a mixture of 3 internal standards at a constant concentration.
- The calibration curve is verified with a mixture of priority pollutant standards every day. If the response factors vary greater than 25%, the instrument must be recalibrated

Analysis of Organic Compounds Extracted in Acid or Base/Neutral Solutions by Gas Chromatography/Mass Spectrometry

- Each batch of 20 samples consists of 16 customer samples (at a maximum), one blank sample, one spiked blank (for water matrices), one sample spiked with the priority pollutant standard mixture and a duplicate customer sample. This amounts to a 20% quality control factor.

- Three surrogate compounds are added to each sample in the batch for Base/Neutral analyses
- Three surrogate compounds are added to each sample in the batch for Acid analysis
- A blind quality control sample is introduced to the laboratory for analysis on a weekly basis
- Each GC/MS is checked and retuned, if necessary, at the beginning of each day to ensure that its performance on decafluorotriphenylphosphine (DFTPP) meets the EPA criteria
- A calibration curve for quantitation is prepared using a mixture of standards composed of either the Organic Acid or Base/Neutral Extractable Compounds at a minimum of 3 concentrations and using five internal standards for quantitation.

Analysis of Metals

All Samples

- New standards are prepared for each batch of samples.
- Normal calibration is performed using a blank sample and four standards that have been through the sample preparation procedure. A regression analysis is used to construct the calibration curve.
- All EP Toxicity samples and all samples determined by furnace atomic absorption are calculated by the "method of additions".
- For each sample analysis that requires the use of the "method of additions" technique, a three point calibration is performed using U.S. EPA "Methods for Chemical Analysis of Water and Wastes, 1979". Results are obtained using linear regression analysis. Any regression with a coefficient of correlation below 0.990 is considered suspect, necessitating review of calibration data or sample re-analysis.
- In constructing the normal calibration curves the lowest concentration levels we use are values greater than or equal to 5 times the Instrumental Detection Limit (IDL).
- All calibration standards are analyzed in duplicate, at a minimum.
- Independent reference standards are used to check the accuracy of calibration standards
- A check standard is analyzed every ten samples to validate the normal calibration curve
- One customer sample out of every ten is analyzed in triplicate.

Homogeneous Samples (except for Mercury analysis)

Samples are analyzed in batches of 30 or less. For batches in which the sample matrices are homogeneous, the QC program is a minimum of 25% and consists of analyzing:

- 3 sets of triplicate analyses;
- 2 Replicate spikes;
- 1 independent reference standard;
- 4 Calibration standards (processed using the sample preparation method);
- 4 Calibration standards (without sample preparation); and
- 1 Reagent Blank.

Heterogeneous Samples (except for Mercury analysis)

Samples are analyzed in batches of 30 or less. For batches in which the sample matrices are heterogeneous, the QC program is a minimum of 35% and consists of analyzing:

- 3 sets of triplicate analyses;
- 2 Replicate spikes,
- 1 Replicate independent reference standards;
- 4 Calibration standards (processed using the sample preparation method),
- 1 Procedural Blank;
- 4 Calibration standards (without sample preparation); and
- 1 Reagent Blank.

Analysis of Mercury

To analyze samples for mercury we group them by matrix in batches of 20 or less. QC program is a minimum of 30% and consists of analyzing:

- each of the 30 customer samples in duplicate;
- 3 sets of triplicate analyses;
- 2 Replicate spikes;
- 2 Replicate independent reference standards;
- 10 Calibration standards (processed using the sample preparation method); and
- 2 Procedural Blanks.

001095

Analysis of Pesticides, Herbicides and PCB's by Gas Chromatography

Pesticide, herbicide and PCB samples are grouped in batches of 16 customer samples or less according to the type of analysis to be performed. The QC program for each of these three types of analyses is a minimum of 20% and consists of analyzing:

- 1 procedural blank sample (a reagent blank is analyzed in the case of non-water matrices),
- 1 spiked blank sample (the spiked blank is eliminated in the case of non-water matrices),
- 1 replicate sample;
- 1 replicate spiked sample; and
- 1 known reference QC sample for at least each 100 samples analyzed.

The instrument is calibrated each run with three standards, and checked every 10 samples.

Analysis of Cyanides, Phenols, Fluoride, Chloride, Nitrate and Nitrite

- All parameters are analyzed using a Technicon Autoanalyzer II GT.
- 3 calibration standards are analyzed at the beginning and end of each batch.

- Each batch (up to 60 samples) consists of analyzing one blank, one spiked blank, one duplicate and spiked sample every 20 samples, and an unknown reference sample

Analysis of Total Organic Carbon (TOC)

TOC samples are analyzed on a daily basis with the number of samples analyzed per day dependent on the request for duplicate or quadruplicate analyses. The quality control program is designed to maintain the appropriate amount of QC and consists of the following elements.

- Daily instrument calibration
- One blank
- Standard recalibration every 10 samples
- Spiked samples at a low and high level
- Every sample is run in duplicate at a minimum

Analysis of Total Organic Halide (TOX)

- Blank reagent water for absolute carbon background must contain less than 5 ug/l of halide (as chloride).
- Using a trichlorophenol standard, the mean adsorption efficiency must be within +/- 15% of the standard value
- Calibration standards are run every 10 samples.
- Every sample is run in duplicate at a minimum.

001096

Analysis of 2,3,7,8-TCDD (Dioxin) by GC/MS (SIM)

- Each sample is dosed with a known quantity of $^{13}\text{C}_{12}$ -2,3,7,8-TCDD as internal standard and $^{37}\text{Cl}_4$ -TCDD as surrogate standard. The action limits for surrogate standard results is +/- 40% of the true value. Samples showing surrogate standard results outside of these limits are reextracted and reanalyzed.
- Two laboratory "method blanks" are run along with each set of 24 or fewer samples. The method blank is also dosed with the internal standard and surrogate standard.
- At least one per set of 24 samples is run in duplicate to determine intralaboratory precision.
- Qualitative Requirements. The following are met in order to confirm the presence of native 2,3,7,8-TCDD:
 - Isomer specificity must be demonstrated initially and verified once per 8-hour work shift. The verification consists of injecting a mixture containing TCDD isomers which elute close to 2,3,7,8-TCDD. The 2,3,7,8-TCDD must be separated from interfering isomers, with no more than 25% valley relative to the 2,3,7,8-TCDD peak.
 - The 320/322 ratio is within the range of 0.67 to 0.87.
 - Ions 320, 322, and 257 are all present and maximize together the signal to mean noise ratio must be 2.5 to 1 or better for all 3 ions.
 - The retention time is equal (within 3 seconds) the retention time for the isotopically labeled 2,3,7,6-TCDD.
 - At least one of the positives can be confirmed by obtaining partial scan spectra from mass 150 to mass 350. The partial scan guidelines are as follows.

- . the 320/322 ratio should be 1.58 +/- 0.16
- . the 257/259 ratio should be 1.03 +/- 0.10
- . the 194/196 ratio should be 1.54 +/- 0.15
- One sample is spiked with native 2,3,7,8-TCDD at a level of 1.0 PPB (for soil) for each set of 24 or fewer samples.
- In cases where no native 2,3,7,8-TCDD is detected, the actual detection limit is estimated and reported based on a signal to noise ratio of 2.5 to 1 at ions 320 and 322.
- For each sample, the internal standard is present with at least a 10 to 1 signal to noise ratio for both mass 332 and mass 334. Also, the internal standard 332/334 ratio must be within the range of 0.67 to 0.87.

Subcontractor QA/QC

Each subcontractor is required to maintain an appropriate level of quality control. To insure this, each subcontractor is required to submit to ETC the quality control data for all analyses it performs. This data is kept on file at ETC. In general, the amount of quality control required is one duplicate sample with one spiked sample for every ten analyses.

Chain-of-Custody

The chain-of-custody procedure is part of our quality assurance protocol. We believe our chain-of-custody record fully complies with the legal requirements of federal, state and local government agencies and of the courts of law. The record covers:

- labeling of sample bottles, packing the Sample Shuttle and transferring the Shuttle under seal to the custody of a shipper,
- outgoing shipping manifests,
- the chain-of-custody form completed by the person(s) breaking the Shuttle seal, taking the sample, resealing the Shuttle and transferring custody to a shipper,
- incoming shipping manifests;
- breaking the Shuttle's reseat;
- storing each labeled sample bottle in a secured area;
- disposition of each sample to an analyst or technician; and
- the use of the sample in each bottle in a testing procedure appropriate to the intended purpose of the sample.

The records show for each link in this process:

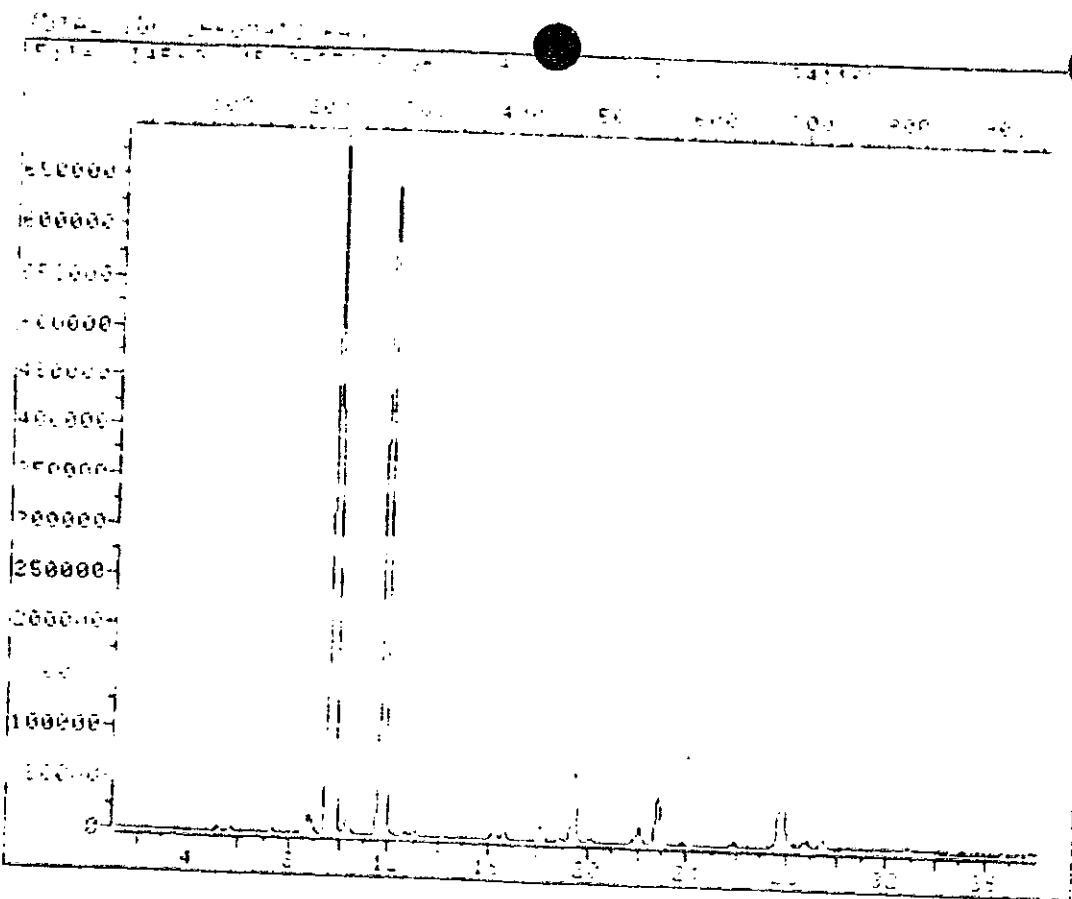
- the person with custody; and
- the time and date each person accepted or relinquished custody.

001097

Appendix A
Mass Spectral Data
for
Quantitated Compounds

- 1) A total ion chromatogram for each sample analysed by a GC/MS instrument
- 2) A mass spectrum and a reference spectrum for each priority pollutant compound detected in the sample

001098



Date File: 841218
 Name: 841218.DM.1
 Misc: 84118

Title: OFFICE BUILDING AIRBORNE POLLUTANTS
 Last Contention: 841218.DM.1
 Operator ID: 1361
 Quamt Time: 841218 23:18

rat:
 a. C.
 ie: 3
 ci: 6
 file
 ie:
 t. Ca
 --
 *2-
 --
 Ca
 or
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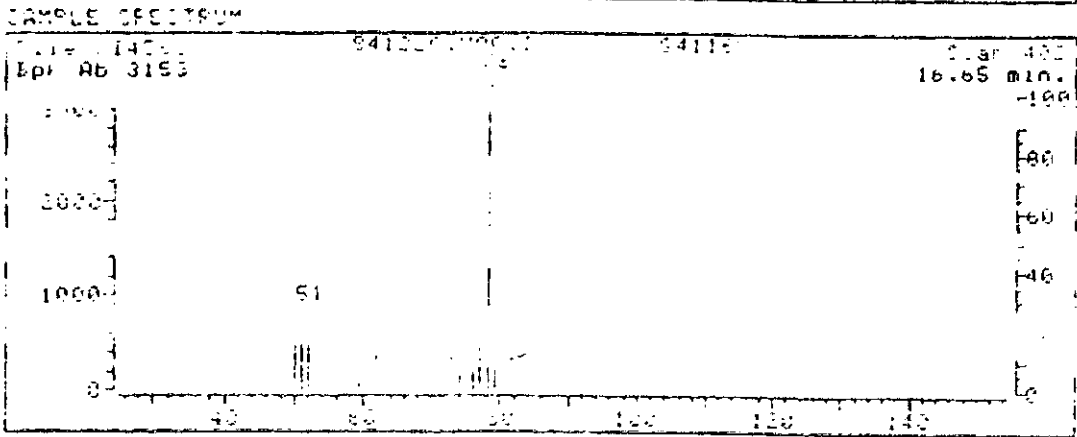
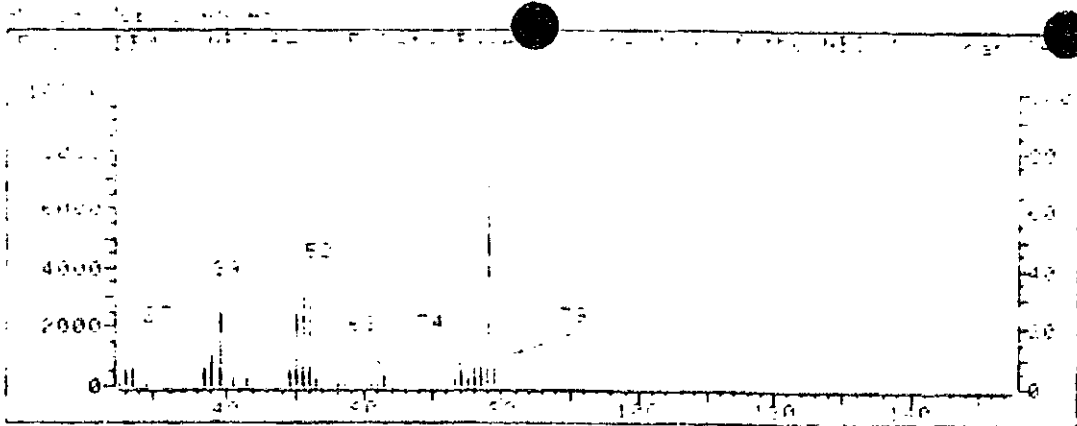
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 841026.MD-1
 841026

Injected at: 841026
 Injector Port: 100

Method: 1007
 IDFILE: PURGEABLE PHASE 17 FULCOTANTS.1
 Calibration: 84126 1987

Compound	R.T.	Scan#	Area	Conc	Units
2-Bromo-1-chloropropane	10.12	440	38356	200.00	ug
Carbon tetrachloride	12.07	307	865	41.91	ug
Chloroethane	11.72	280	2324	12.27	ug
1,2-Dichloroethane	11.72	280	2324	12.27	ug
1,1-Dichloroethane	11.72	280	2324	12.27	ug
Tetrachloroethene	11.72	280	2324	12.27	ug
Toluene	21.86	530	4281	20.73	ug
1,2-Trans-dichloroethene	21.76	529	33197	71.36	ug = 11 = 66
1,1,1-Trichloroethane	17.04	710	710	11.52	ug
Trichloroethylene	17.04	710	710	11.52	ug
Trichlorobromomethane	17.04	710	710	11.52	ug
meta-xylene	17.04	710	710	11.52	ug
meta-xylene	17.04	710	710	11.52	ug
ortho- and para-xylene	17.04	710	710	11.52	ug
1,2-Dichloroethane (d)	17.04	710	710	11.52	ug
Toluene-D8	22.31	561	11472	204.27	ug
o-Bromocyclohexane	22.31	561	11472	204.27	ug
1,4-Dichlorobenzene	22.31	561	11472	204.27	ug

001100



File: 1455
 Name: 84126.M (0001)
 Used: 84116

Compound Name: benzene
 Scan Number: 480
 Retention Time: 16.65 min.
 Event: 17%F
 Concentration: 0.152 ug

EF247

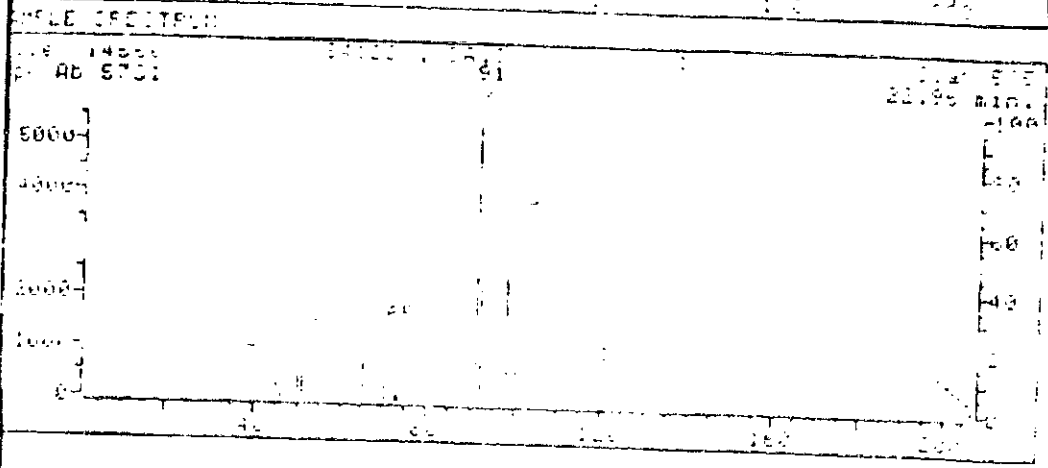
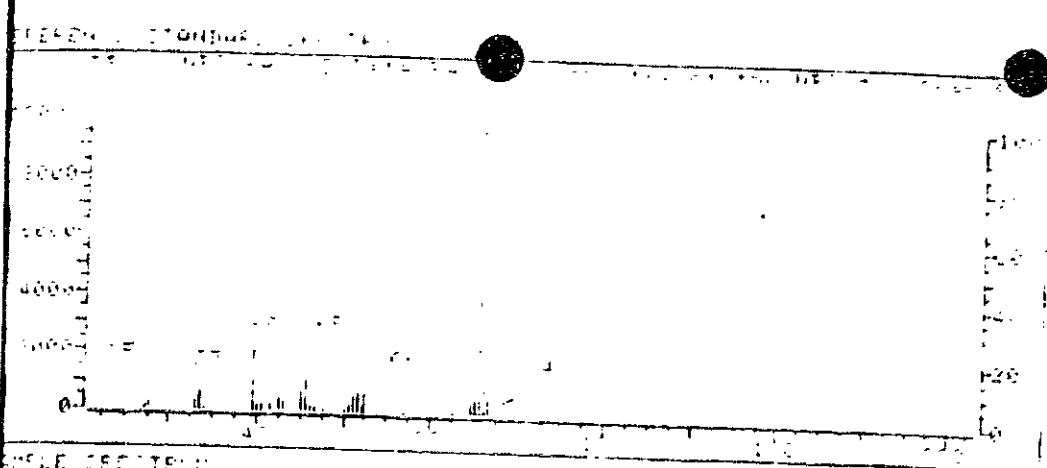
1000
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SAMPLE
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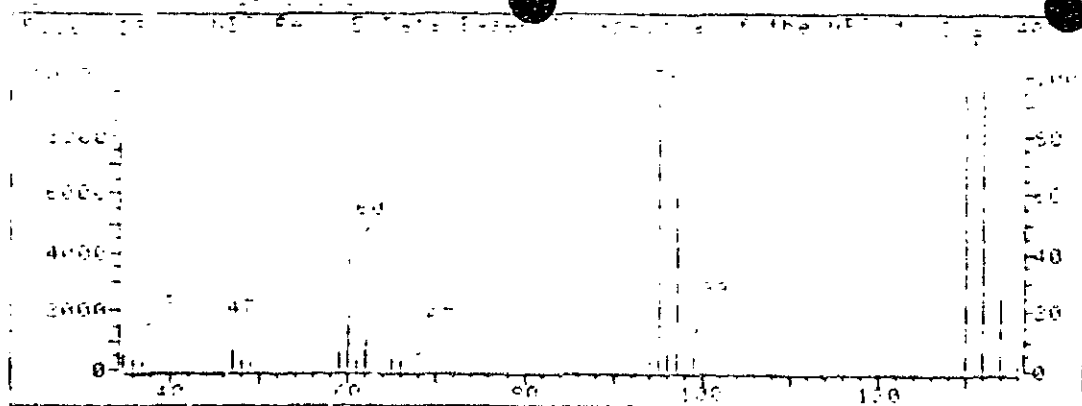
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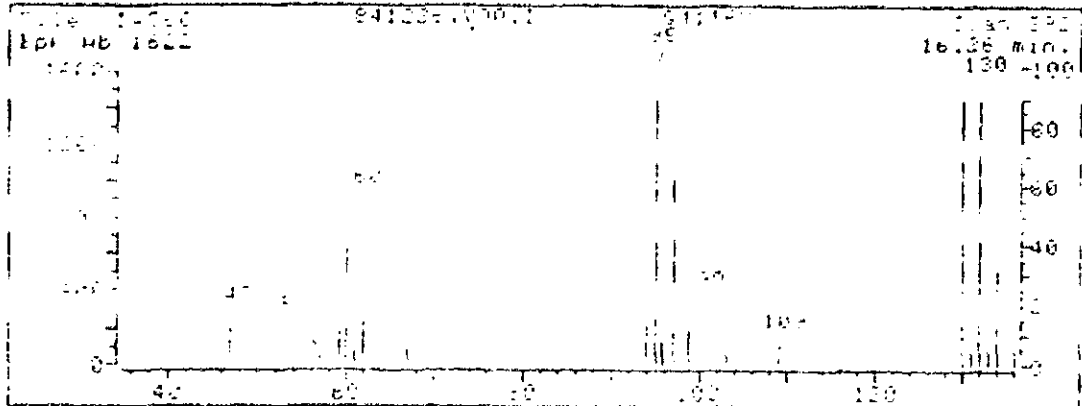


Name: B01226 (04/11)
 Misc: Gallic
 Compound Name: Gallic
 Retention Time: 20.95 min.
 Concentration: 2.35 ug

001102



SAMPLE SPECTRUM



Date: 5/1/80

Name: 84108A.V10.1

Mass: 84110

Compound Name: 1,4-DIOXANE

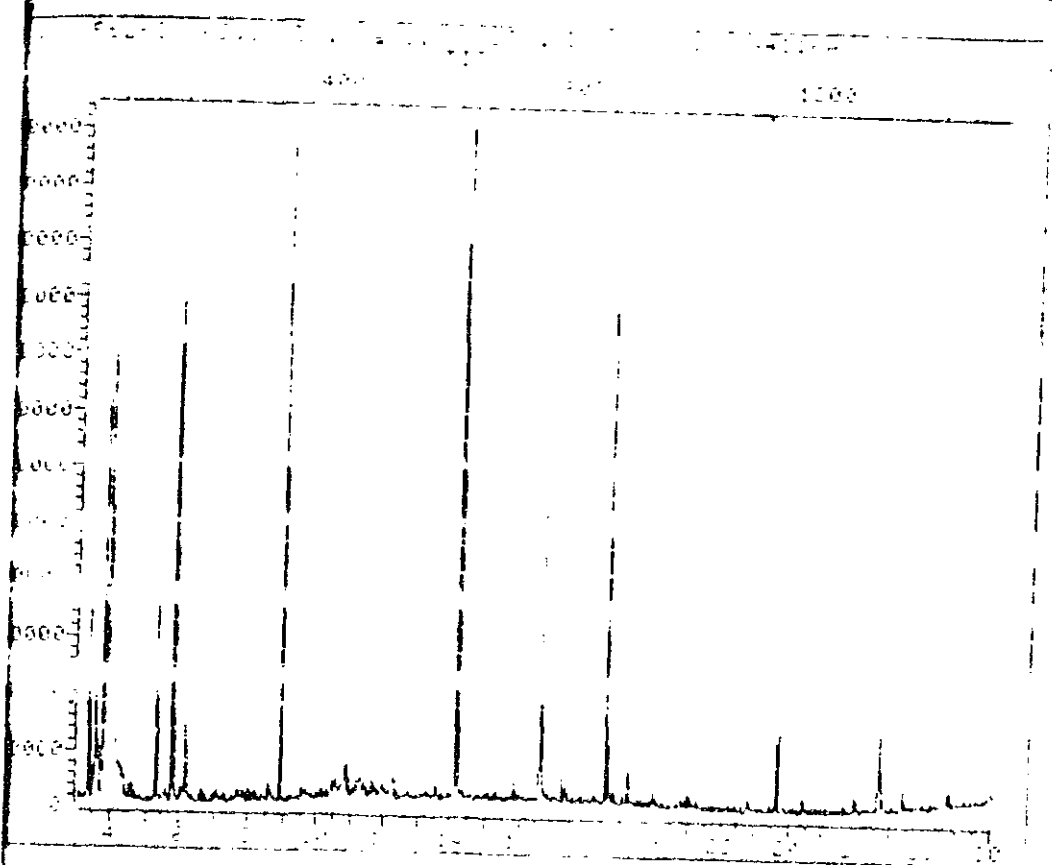
Scan Number: 199

Retention Time: 16.26 min.

Area: 5927

Concentration: 51.42 ug

001103



Data File: F000110
Name: S41271.F F1-1014

Id File: F000110
Title: HClO4 10% MeOH, F
Unit Cell: 1000.0000000000000

Operator: ID: 100576
Start Time: 10:00

001104

TOTAL
File

14000

12000

10000

8000

6000

4000

2000

001105

11/11/81

01 711170

Sample Name: 841231
Injected at: 12:01
Dilution Factor: 1.00

Date File: F02031:02
Name: 841231.S.METHOD
GALIB

ETL804

F 366BF

Method: F02031
Name: ACID.IDFILE.F
Last Calibration: 841231 12:01

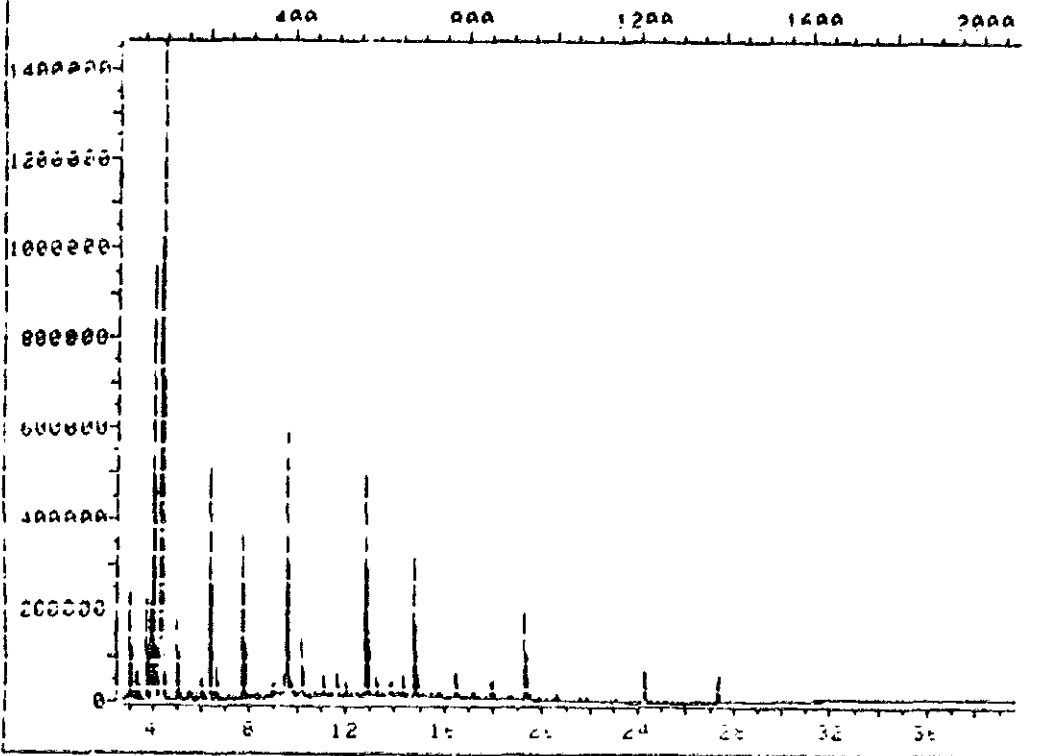
Compound	R.T.	Scan#	Area	Conc	Units
1) m4-1,4-Dichlorobenzene	5.74	150	224432	40.00	UG/ML
2) 2-Fluorophenol	7.74	20	121114	21.71	UG/ML
3) 2-Fluorophenol	3.74	41	8036	1.4	UG/ML
4) Phenol-09	5.26	123	136655	24.14	UG/ML
5) Phenol-09	5.74	151	1561	.27	UG/ML
6) m-cresol	6.87	320	6217	1.1	UG/ML
8) 2,4-Dichlorophenol	7.87	269	203	.36	UG/ML
9) 2,4-Dimethylphenol	8.23	292	989	1.8	UG/ML
11) p-Chloro-m-cresol	11.07	459	624	1.1	UG/ML
12) m-1,0-Naphthalene	14.13	619	331127	5.9	UG/ML
13) m-1,0-Naphthalene	14.13	619	47961	8.6	UG/ML
18) 2,4,6-Trichlorophenol	16.13	780	91224	16.4	UG/ML
19) Pentachlorophenol	16.31	887	1384	.25	UG/ML
20) M-CHLOROPHENOL	17.66	769	339	.6	UG/ML

* Compound vs. RET

57

TOTAL ION CHROMATOGRAM

FILE: 850105 DE 0-0000 0.0000 BIL-FST ON G. 850105 64116B
TIC



Data File: 850105:005
Name: BIL-FST ON G, 850105
Misc: 64116B

BIL#14

Id File: GENP
Title: BIL-FST TO FILE#6,11 30 04,00L
Last Calibration: 850104 22:45

Operator ID: 1105.6
Quant Time: 850105 02:06

001106

Date: 11/06/84
 Name: B/N-PEST 10 G, 850103
 Misc: 64100

Dilution Factor: 1.00

BIL#14

ID File: GENE
 Title: B/N-PEST 10 FILE06,11/30/84,WWC
 Last Calibration: 850104 22:45

Compound	R.T.	Scan#	Area	Conc	Units	
1) *d4-1,4-Dichlorobenzene	6.31	202	339017	40.00	UG/ML	271
2) N-Nitrosodimethylamine	3.23	28	2367	1.18	UG/ML	271
2) N-Nitrosodimethylamine	4.01	72	1012	5.1	UG/ML	271
2) N-Nitrosodimethylamine	4.12	78	1590	7.9	UG/ML	271
2) N-Nitrosodimethylamine	4.36	92	2341	4.1	UG/ML	271
4) 1,3-Dichlorobenzene	6.35	204	2279	3.2	UG/ML	281
5) 1,4-Dichlorobenzene	6.35	204	2279	3.2	UG/ML	291
6) 1,2-Dichlorobenzene	6.35	204	2279	3.9	UG/ML	311
7) Nitrobenzene-d4	7.70	266	260260	3.9	UG/ML	311
7) Nitrobenzene-d5	8.03	299	1133	3.1	UG/ML	311
7) Nitrobenzene-d6	8.35	317	1285	3.5	UG/ML	311
8) bis(2-Chloroisopropyl)ether	6.01	185	4851	6.7	UG/ML	311
8) bis(2-Chloroisopropyl)ether	6.33	203	9286	10.4	UG/ML	311
8) bis(2-Chloroisopropyl)ether	6.53	214	3727	4.4	UG/ML	311
8) bis(2-Chloroisopropyl)ether	8.07	301	4044	4.7	UG/ML	311
9) *d8-Naphthalene	9.51	382	633373	40.00	UG/ML	311
10) 2-Fluorobiphenyl	12.84	570	371318	37.5	UG/ML	311
11) 1-Fluoronaphthalene	9.49	381	803	0.3	UG/ML	311
11) 1-Fluoronaphthalene	10.13	417	1666	0.6	UG/ML	311
12) N-Nitrosodi-n-propylamine	7.70	260	38571	2.9	UG/ML	311
14) Nitrobenzene	8.32	315	1631	0.5	UG/ML	311
14) Nitrobenzene	8.46	323	782	0.1	UG/ML	311
14) Nitrobenzene	8.99	353	224	0.3	UG/ML	311
15) Isophorone	7.15	249	4122	0.4	UG/ML	311
15) Isophorone	8.35	317	1285	0.4	UG/ML	311
18) Naphthalene	8.17	307	2211	0.1	UG/ML	311
18) Naphthalene	9.15	361	800	0.6	UG/ML	311
18) Naphthalene	9.56	385	62322	4.3	UG/ML	311
18) Naphthalene	10.06	413	496	0.3	UG/ML	311
20) *d10-Acenaphthalene	14.78	679	211009	40.00	UG/ML	311
23) Dimethyl phthalate	13.45	604	485	0.8	UG/ML	311
23) Dimethyl phthalate	14.55	666	505	0.8	UG/ML	311
23) Dimethyl phthalate	14.80	680	39283	6.2	UG/ML	311
23) Dimethyl phthalate	15.24	705	714	0.1	UG/ML	311
24) Acenaphthylene	14.52	653	70	0.5	UG/ML	311
24) Acenaphthylene	15.00	691	508	0.5	UG/ML	311
24) Acenaphthylene	15.71	731	223	0.2	UG/ML	311
25) 2,6-Dinitrotoluene	14.55	666	2039	0.6	UG/ML	311
25) 2,6-Dinitrotoluene	14.78	679	26941	30.1	UG/ML	311
25) 2,6-Dinitrotoluene	15.00	691	844	0.4	UG/ML	311
25) 2,6-Dinitrotoluene	15.19	702	792	0.8	UG/ML	311
26) Acenaphthene	13.52	608	1681	0.2	UG/ML	311
26) Acenaphthene	13.77	623	2315	0.5	UG/ML	311
26) Acenaphthene	14.89	685	1711	0.2	UG/ML	311
26) Acenaphthene	16.12	754	1753	0.2	UG/ML	311

* Con

Compound	R.T.	Scan#	Area	Conc	Units
2,4-Dinitrotoluene	14.55	666	2039	2.52	UG/ML
2,4-Dinitrotoluene	14.78	677	26741	33.24	UG/ML
2,4-Dinitrotoluene	16.47	773	2962	3.16	UG/ML
2,4-Dinitrotoluene	16.76	790	1789	2.21	UG/ML
Diethyl phthalate	16.82	777	1367	1.6	UG/ML
Fluorene	16.45	773	1626	.28	UG/ML
N-Nitrosodiphenylamine	16.45	773	424	.4	UG/ML
N-Nitrosodiphenylamine	16.72	788	836	.28	UG/ML
N-Nitrosodiphenylamine	16.02	861	766	.26	UG/ML
d,10-Phenanthrene	19.24	930	225430	40.00	UG/ML
Phenanthrene	19.31	934	1839	.39	UG/ML
Anthracene	19.31	934	1839	.31	UG/ML
Di-n-butyl phthalate	21.50	1057	10513	1.80	UG/ML
Benzidine	24.26	1213	709	1.40	UG/ML
Alpha-BHC	17.02	809	1247	1.41	UG/ML
Alpha-BHC	17.39	826	724	.82	UG/ML
Alpha-BHC	16.10	810	925	1.1	UG/ML
Alpha-BHC	18.25	874	1022	1.45	UG/ML
Beta-BHC	18.18	870	925	2.06	UG/ML
Beta-BHC	18.25	874	1022	2.16	UG/ML
Beta-BHC	19.03	918	314	.28	UG/ML
Beta-BHC	19.74	958	269	.60	UG/ML
Gamma-BHC	18.18	870	925	1.1	UG/ML
Gamma-BHC	18.25	874	1022	1.40	UG/ML
Gamma-BHC	19.03	918	314	.45	UG/ML
Gamma-BHC	19.74	958	269	.37	UG/ML
Delta-BHC	16.25	814	1022	1.1	UG/ML
Delta-BHC	19.03	918	314	.29	UG/ML
Delta-BHC	19.74	958	269	.50	UG/ML
d,12-Chrysene	27.32	1386	57235	40.00	UG/ML
Dieldrin	23.07	1143	414	.2	UG/ML
Dieldrin	23.21	1154	362	.5	UG/ML
Dieldrin	24.26	1213	310	.49	UG/ML
Dieldrin	25.19	1271	361	.5	UG/ML
Endrin	24.26	1213	247	2.15	UG/ML
Endrin	24.63	1234	230	2.07	UG/ML
Endrin	25.20	1266	180	1.62	UG/ML
Endrin aldehyde	24.26	1213	247	.75	UG/ML
Endrin aldehyde	24.63	1234	230	.65	UG/ML
Endrin aldehyde	26.46	1337	233	.39	UG/ML
Endrin aldehyde	26.78	1355	284	1.89	UG/ML
4,4'-DDT	27.32	1386	2616	4.81	UG/ML
Terphenyl-D14	24.26	1213	60875	31.77	UG/ML
Butyl benzyl phthalate	26.00	1311	183	.45	UG/ML
Butyl benzyl phthalate	27.16	1377	272	.42	UG/ML
bis(2-Ethylhexyl)phthalate	27.91	1419	3507	2.10	UG/ML
Di-n-octyl phthalate	29.70	1520	2214	.95	UG/ML

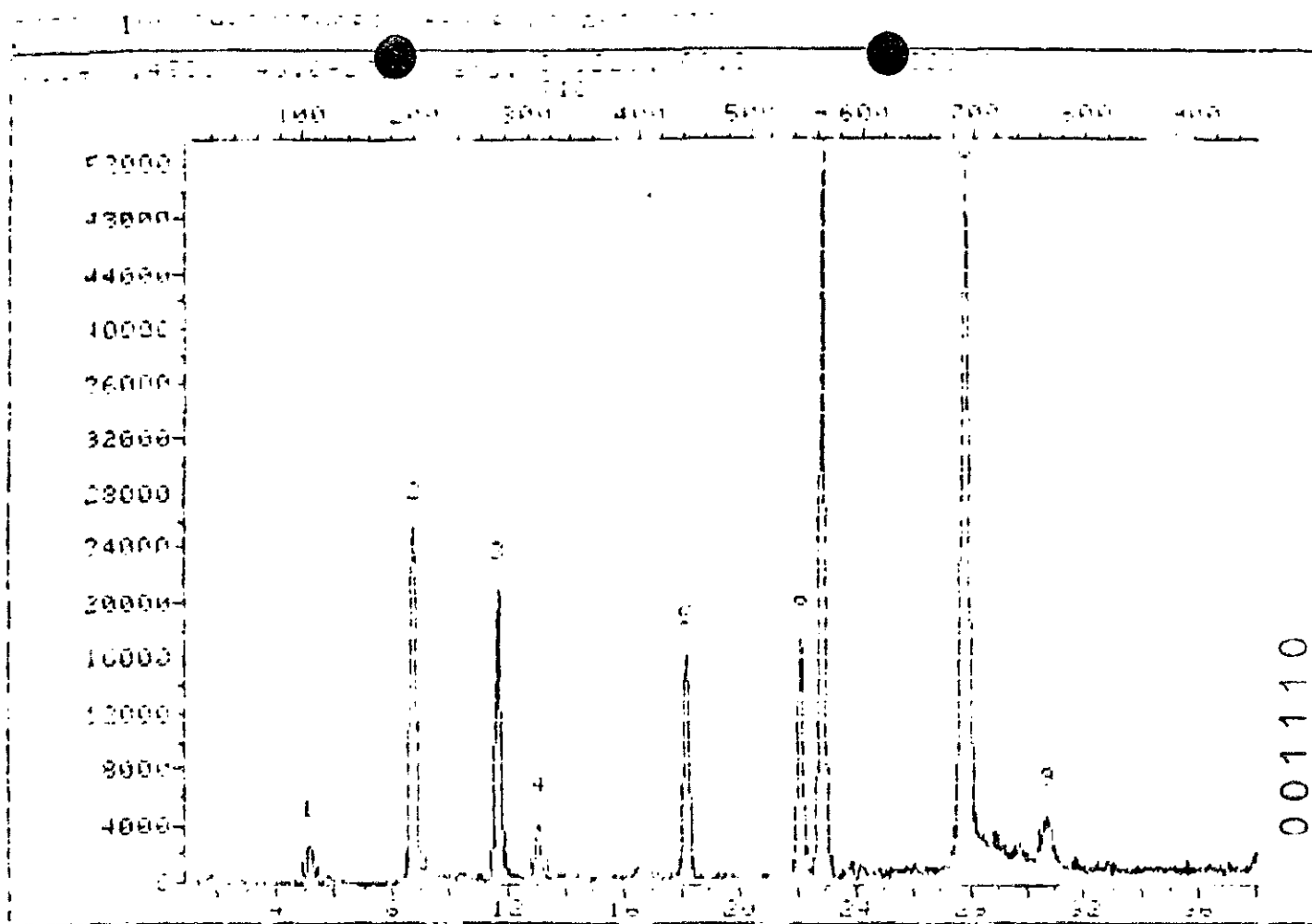
001108

* Compound is ISTD

Appendix C1
GC/MS Subsidiary Data

001109

325



001110

Data File: 001110
 Name: 0011106.D
 Misc Data: 001110

11/15/77
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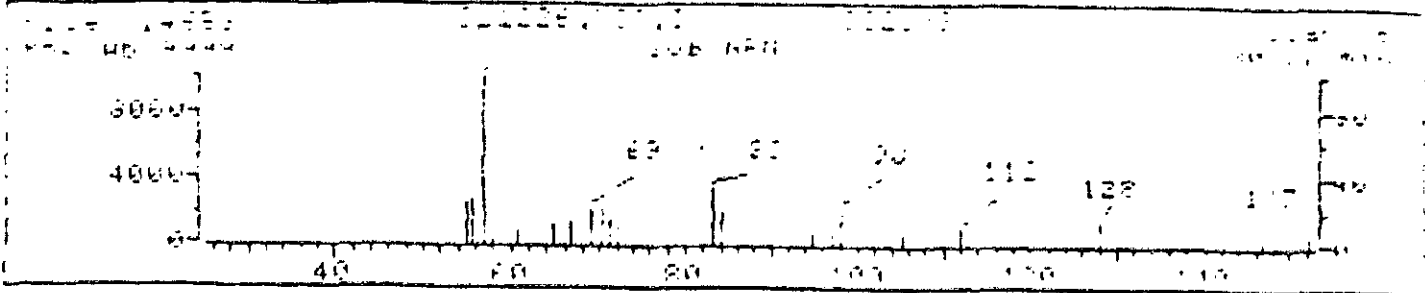
Retention Time	Compound	Height	Area	Conc	Unit
1.1	1,1,1-Trichloroethane	441	43194	100.00	NS
1.2	Carbon tetrachloride	708	284		
1.3	Carbon tetrachloride	100	100		
3.1	Chlorobenzene	596	1218		
4.1	Toluene	10	10		
2.1	Toluene	757	5081	10.00	NS
1.0	1,1,1-Trichloroethane	100	100	10.00	NS
7.1	1,1-Dichlorobenzene-04	378	21711	100.00	NS
7.2	Toluene-02	517	101994	100.00	NS
7.3	1,1-Dichlorobenzene	100	100	10.00	NS
7.4	1,1-Dichlorobenzene	544	16891	100.00	NS

* Compound is 1,1,1

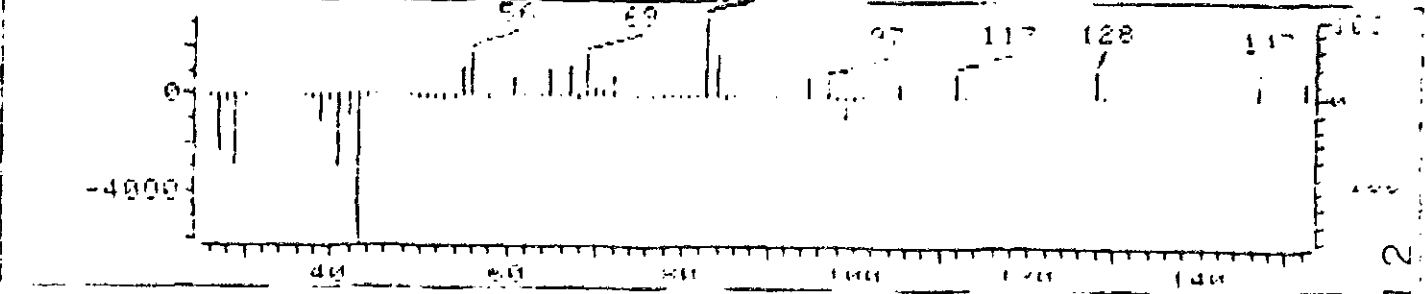
001111

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 2 4
 3 7
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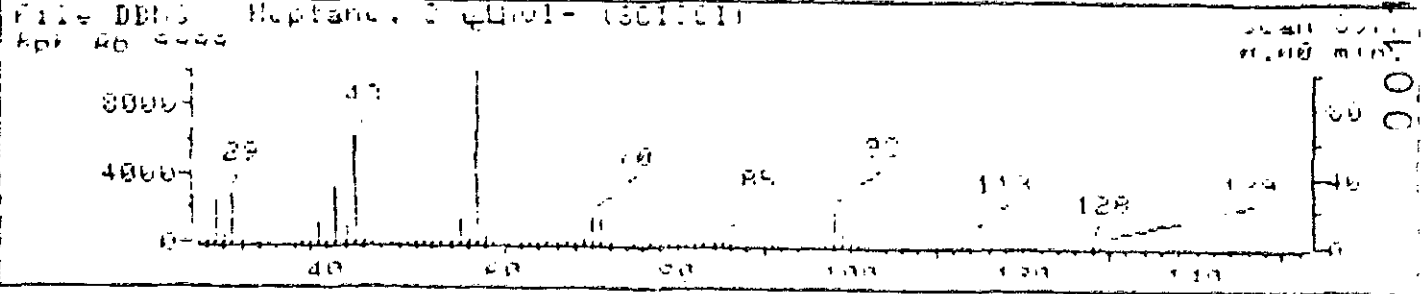
DIFFERENCE SPECTRUM



DIFFERENCE



LIBRARY REFERENCE SPECTRUM



File DB05 Heptane, 3-ethyl- (801901)
 Exp No 6666
 Scan 777
 Area 16507
 Semi-quantitative Conc 36.73 NG

Data File: 11557 Scan Number: 777
 Search Speed: 3 Titling option: 3 Number of ion ranges searched: 10

Peak	Library	K	dK	100%	111%
1	Heptane, 3-ethyl- (801901)	15869804	28	70	2 0
2	4-Pentenal, 2-ethyl (801901)	5204202	30	76	1 0
3	3-Undecene, 6-methyl (801901)	74430527	39	48	2 0

File: F010
 841231.B (FACID.F)

Injected at: 841231 11:55
 Dilution Factor: 1.00

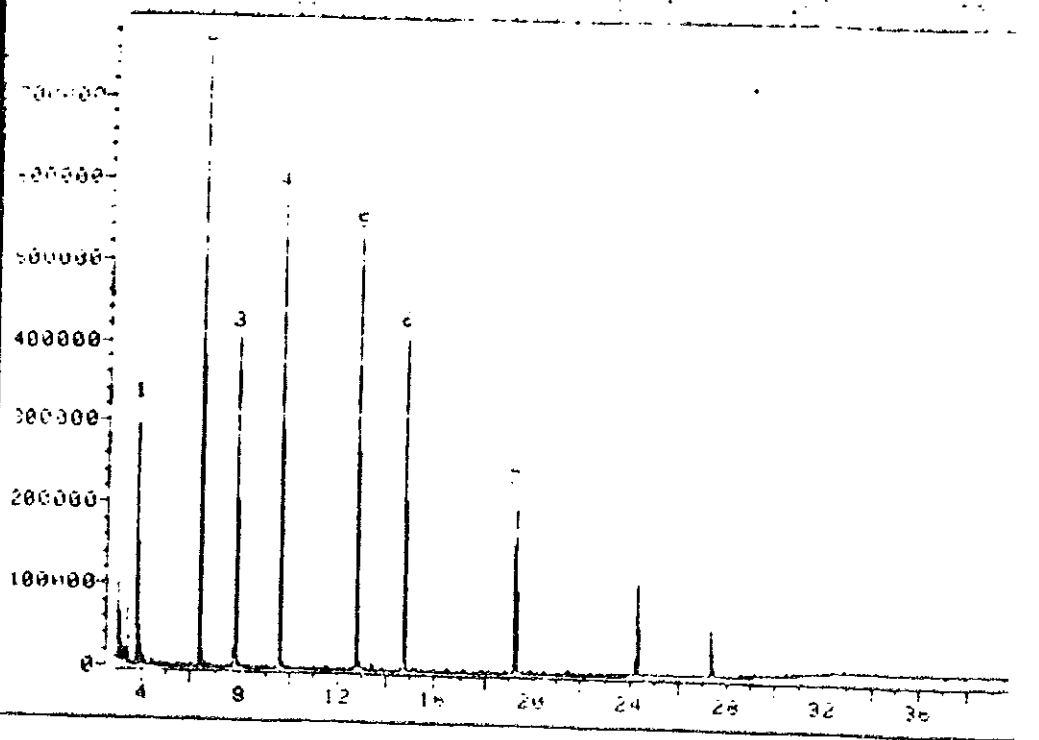
ETAC

Ⓢ F366BC

File: F010
 : ACID IDFILE.F
 Calibration: 841231 11:01

Compound	R.T.	Scan#	Area	Conc	Units
d4-1,4-Dichlorobenzene	5.80	153	201100	40.00	UG/ML
1-Fluorobenzene	4.01	57	410	0.08	UG/ML
Phenol	6.10	111	387	0.08	UG/ML
Phenol-2D	6.17	112	107310	3.14	UG/ML
Phenol-3D	6.17	111	510	0.01	UG/ML
d8-Naphthalene	8.90	153	956	0.19	UG/ML
d10-Chloronaphthalene	8.90	317	814247	40.00	UG/ML
d10-Fluoranthene	10.10	619	394360	4.00	UG/ML
d10-Fluoranthene	10.10	320	793	0.02	UG/ML
2-Chloro-1-methylnaphthalene	10.50	787	181310	10.00	UG/ML
2-Fluoro-1-methylnaphthalene	10.50	787	100	0.00	UG/ML

001114



Data File: >Gc512:103
Name: BN PST ON G. 87-103
Misc Data: 0009296

BTL# 5

001115

REPORT

Operator: IU: TR0576

Quant Rev: 3

Quant time: 870104 22:57

Injected at: 870104 18:04

Data File: >66512:103

Dilution Factor: 1.00

Name: BN/PSI ON G, 850103
Misc: WC25278

BILE

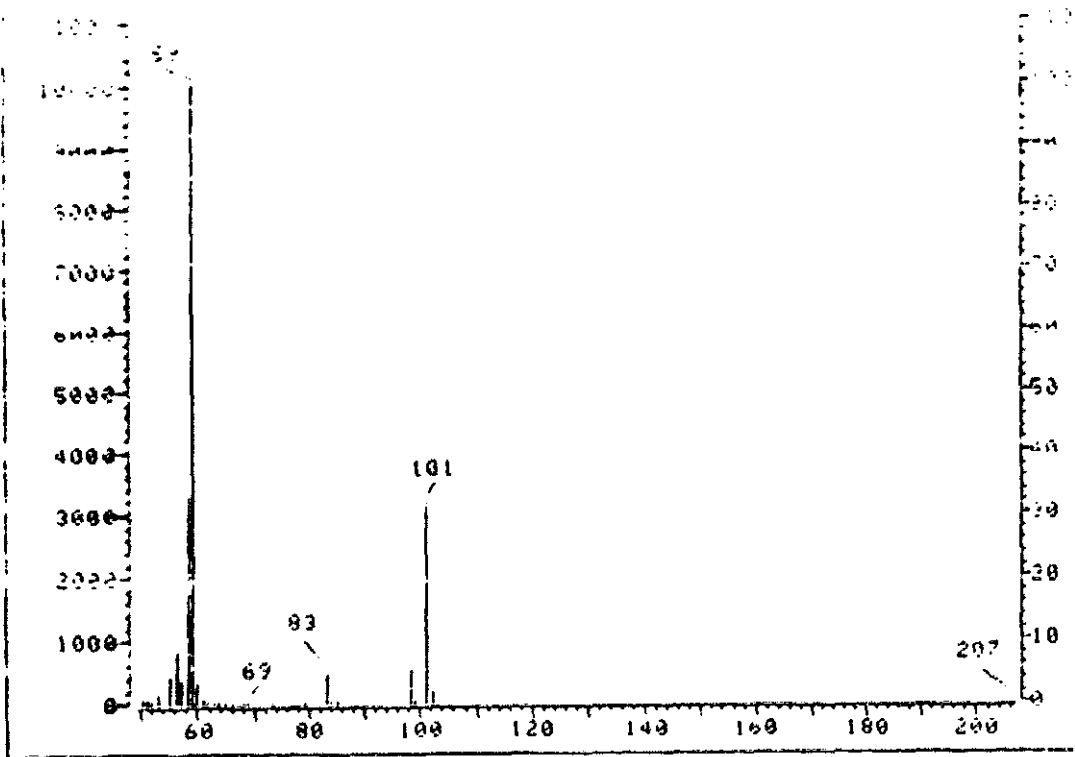
ID File: G8NP

Title: B/N+PEST ID FILE#6, 11/30/84, WWC

Last Calibration: 870104 22:45

70)
70)
71)
71)
72)
73)
73)

Compound	R.T.	Scan#	Area	Conc	Units	* C
1) *d4-1,4-Dichlorobenzene	6.31	199	388523	40.00	UG/ML	
2) N-Nitrosodimethylamine	2.76	10	783	7.74	UG/ML	
2) N-Nitrosodimethylamine	4.36	89	828	3.6	UG/ML	
3) bis(2-Chloroethyl) ether	5.92	177	386	.06	UG/ML	
4) 1,3-Dichlorobenzene	6.77	225	925	.13	UG/ML	16
5) 1,4-Dichlorobenzene	6.77	225	925	.12	UG/ML	17
6) 1,2-Dichlorobenzene	6.77	225	925	.14	UG/ML	17
7) Nitrobenzene-d5	7.69	277	275761	65.40	UG/ML	5
8) bis(2-Chloroisopropyl) ether	5.60	159	586	.60	UG/ML	
8) bis(2-Chloroisopropyl) ether	6.31	199	10041	40.34	UG/ML	0
8) bis(2-Chloroisopropyl) ether	7.41	261	670	1.69	UG/ML	
8) bis(2-Chloroisopropyl) ether	7.69	277	1321	1.36	UG/ML	
9) *d8-Naphthalene	9.48	378	733278	40.00	UG/ML	
10) 2-Fluorobiphenyl	12.77	564	401287	36.87	UG/ML	
12) N-Nitrosodi-n-propylamine	7.69	277	39832	8.83	UG/ML	
17) 1,2,4-Trichlorobenzene	9.41	374	598	1.0	UG/ML	
18) Naphthalene	9.53	381	1974	.12	UG/ML	
19) Hexachlorobutadiene	10.10	453	223	1.09	UG/ML	
20) *d10-Acenaphthalene	14.70	673	237602	40.00	UG/ML	
23) Dimethyl phthalate	14.70	673	43560	4.17	UG/ML	
25) 2,6-Dinitrotoluene	14.45	679	434	.43	UG/ML	
25) 2,6-Dinitrotoluene	14.70	673	30040	2.85	UG/ML	
26) Acenaphthene	14.80	679	497	.07	UG/ML	
27) 2,4-Dinitrotoluene	14.45	679	434	.48	UG/ML	
27) 2,4-Dinitrotoluene	14.70	673	30040	2.92	UG/ML	
33) *d10-Phenanthrene	19.17	926	240928	40.00	UG/ML	
38) Di-n-butyl phthalate	21.47	1056	7207	1.16	UG/ML	
40) Benzidine	24.24	1212	1340	2.47	UG/ML	
48) *d12-Chrysene	27.28	1384	56189	40.00	UG/ML	
53) Dieldrin	24.24	1212	567	.91	UG/ML	
54) Endrin	23.44	1167	749	6.87	UG/ML	
54) Endrin	25.12	1262	470	4.31	UG/ML	
54) Endrin	25.97	1310	554	5.08	UG/ML	
54) Endrin	26.24	1325	438	4.62	UG/ML	
57) Endrin aldehyde	25.12	1262	470	1.83	UG/ML	
57) Endrin aldehyde	25.97	1310	554	2.16	UG/ML	
57) Endrin aldehyde	26.24	1325	438	1.71	UG/ML	
57) Endrin aldehyde	26.66	1349	741	2.89	UG/ML	
58) 4,4'-DDT	27.28	1384	2407	4.51	UG/ML	
61) Terphenyl-D14	24.22	1211	105429	56.04	UG/ML	
62) Butyl benzyl phthalate	25.99	1311	229	1.19	UG/ML	
66) bis(2-Ethylhexyl)phthalate	27.87	1417	1160	.71	UG/ML	
68) Benzo(b)fluoranthene	31.36	1614	416	.31	UG/ML	
68) Benzo(b)fluoranthene	31.41	1617	261	1.9	UG/ML	
69) Benzo(k)fluoranthene	31.36	1614	416	.33	UG/ML	
69) Benzo(k)fluoranthene	31.41	1617	261	.21	UG/ML	



Data File: >Go512::103
 Name: BN-PST ON G, 89 103
 Misc Data: QC25256
 RT (min): 3.74
 Scan: 54
 Area: 402122
 Semi-quantitative Conc: 10.74 UG/ML

BTL# 5

No PM hits for this scan.

001118

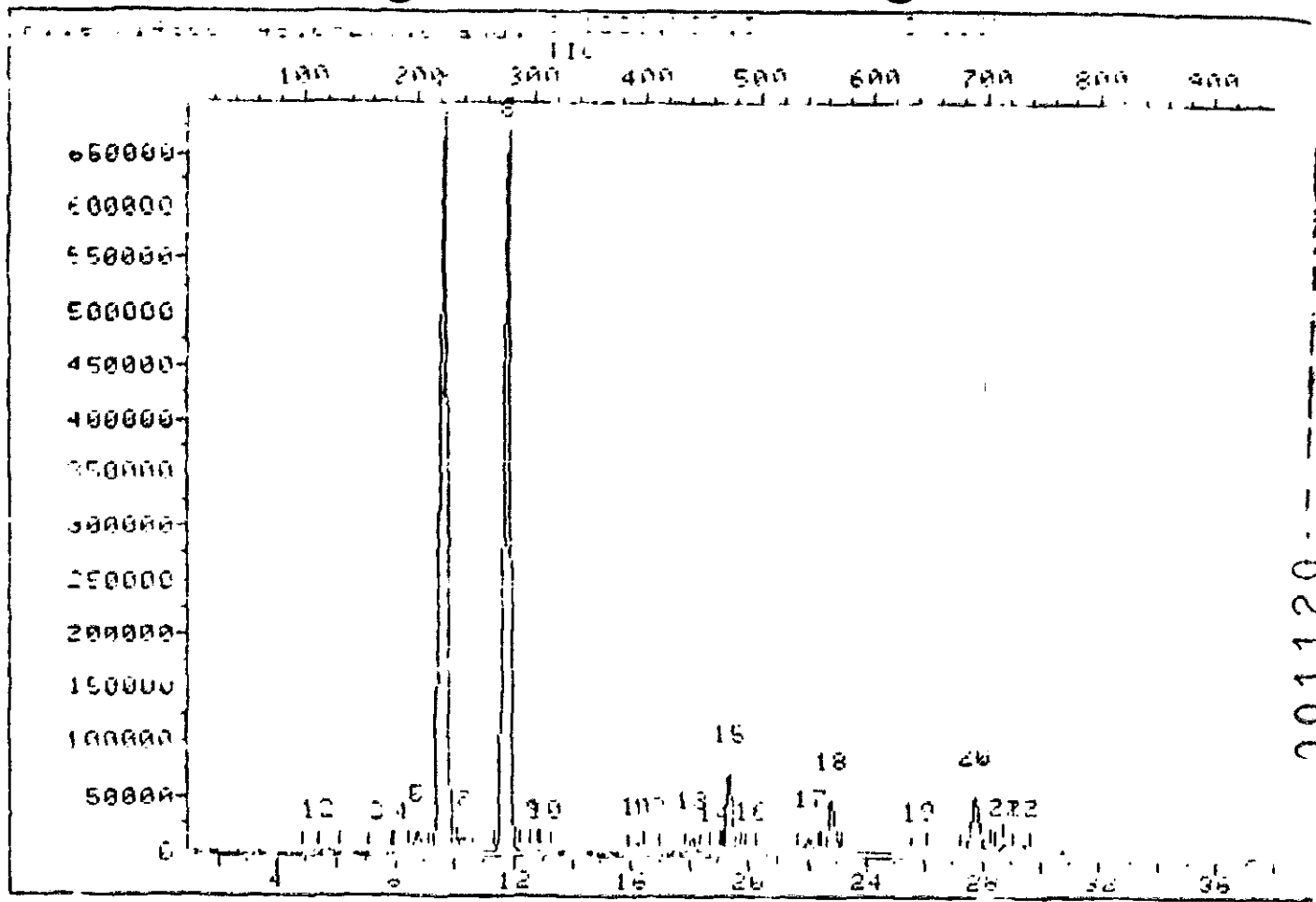
Appendix C
Mass Spectral Data
for
Tentatively Identified Compounds

001119

1. For each tentatively identified compound a mass spectrum of the detected compound, a reference mass spectrum for that compound and a plot of the spectral differences are provided

001119

TOTAL ION CHROMATOGRAPHY ANALYSIS



001120

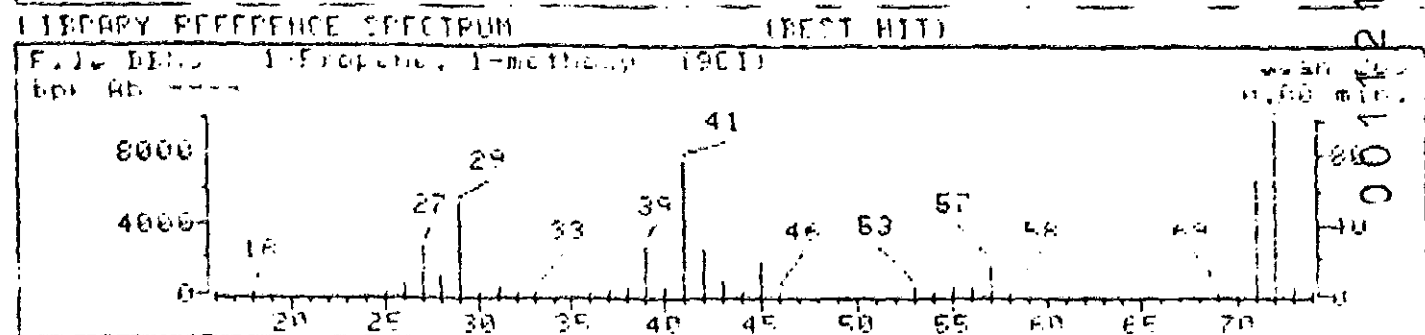
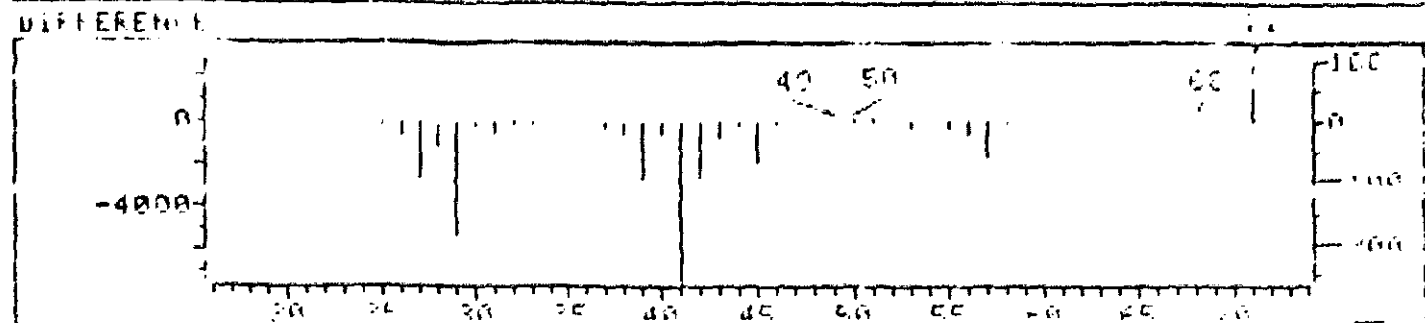
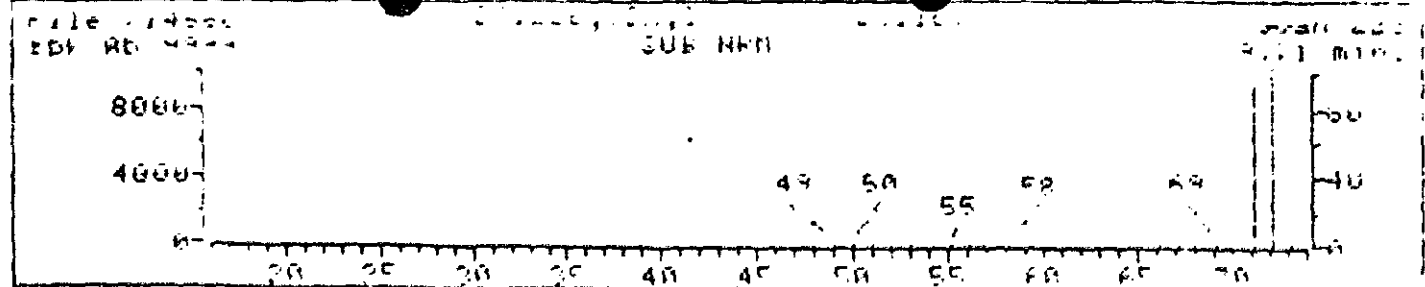
Data File: 011000.D
 Name: 011006.U0A.T
 Misc Data: 011160

D
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Data File Search:

1. 1-
P-
1

COMPIE SPECTRUM (BACKGROUND SUBTRACTED)



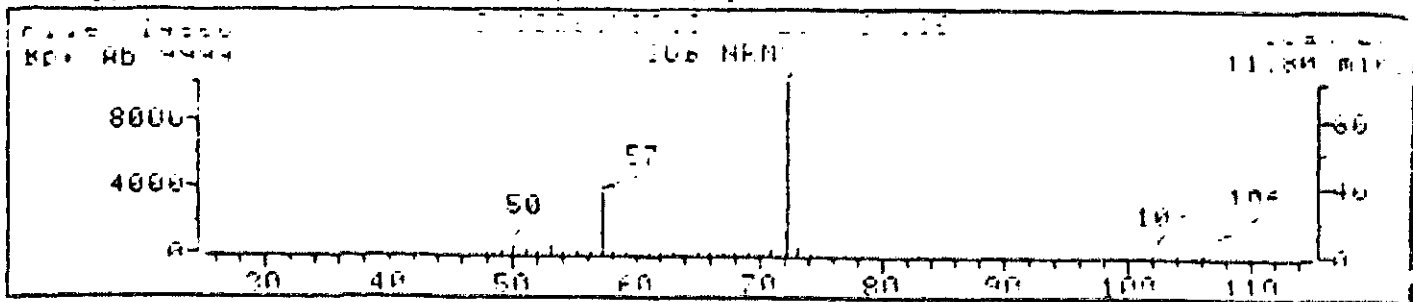
Data File: 14560
 Name: 011200.0001
 Misc Data: 14560
 RT (min): 9.51
 Scan: 207
 Area: 5026740
 Semi-quantitative Conc: 12945.00 mg

Data File: 14560 Scan Number: 207
 Search Speed: 2 Tilt option: 0 Number of ion ranges searched: 10

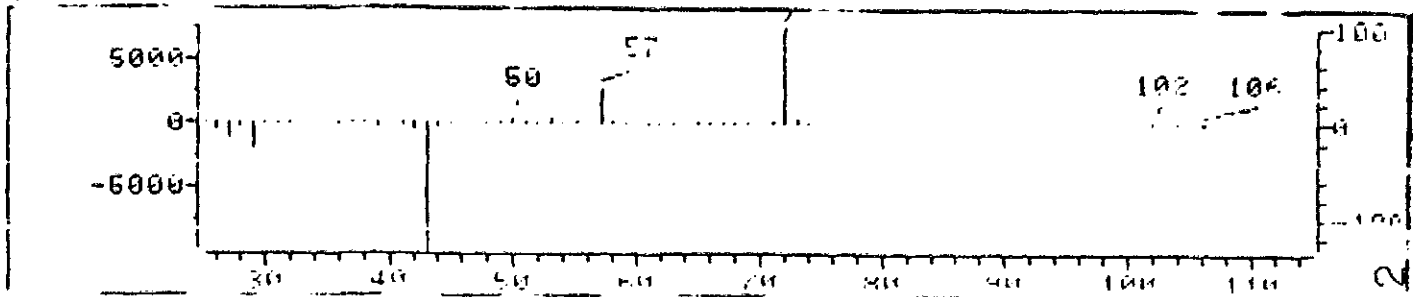
1. 1-Propene, 1-methoxy (901) 72.6480

Prob.	Conf	K	dK	#Ions	Tilt
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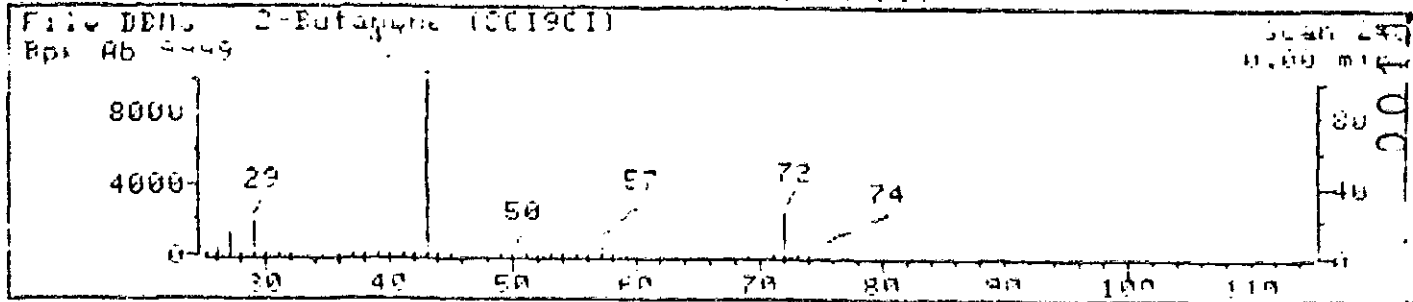
SAMPLE SPECTRUM (BASE PEAKS SUBTRACTED)



DIFFERENCE



LIBRARY REFERENCE SPECTRUM (BEST HIT)



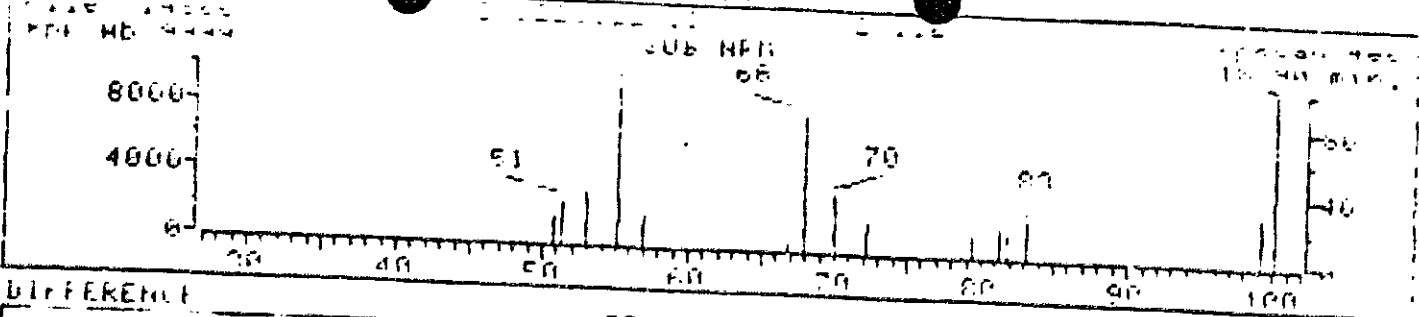
Data File: M14560
 Name: 041226.V00A.T
 Misc Data: 011160
 RT (min): 11.80
 Scan: 277
 Area: 4321481
 Semi-quantitative Conc: 11126.94 NG

Data File: M14560 Scan Number: 277
 Search Speed: 2 Titling option: 0 Number of ion ranges searched: Data Bear

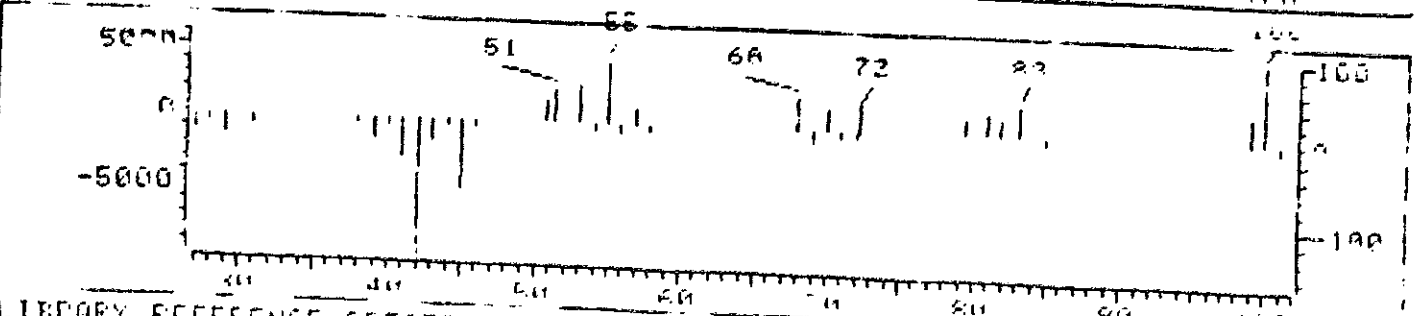
- 1. 2-Butanone (001901) 72 04800 1.
- 2. Acetaldehyde, methylhydrazone (001901) 72 030882 2.
- 3. Formaldehyde, dimethylhydrazone (001901) 72 074002

Peak	Comp	K	d	off	hit	
1.	60	78933	23	46	0	0
2.	60	17167736	20	74	0	0
3.	52	2035894	23	53	1	0

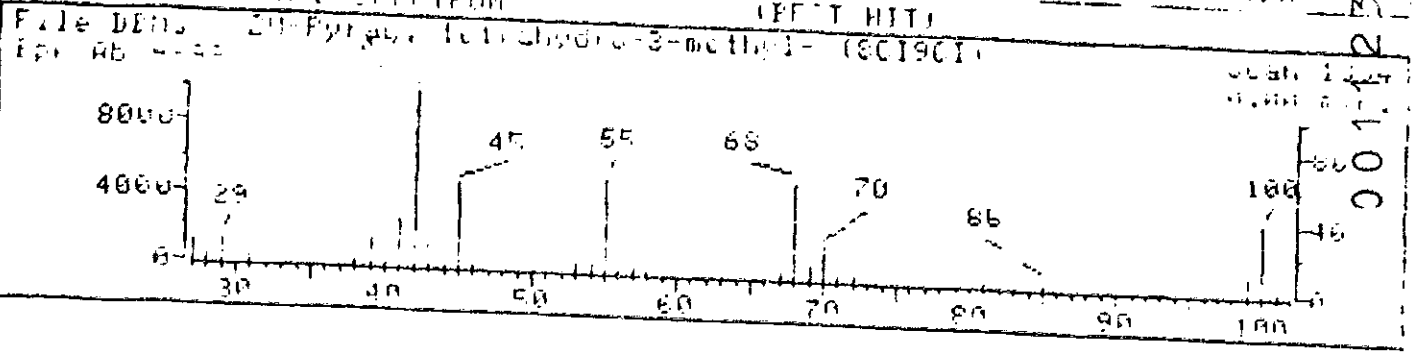
SAMPLE SPECTRUM (BASE POINT SUBTRACTED)



DIFFERENCE



LIBRARY REFERENCE SPECTRUM (BEST HIT)



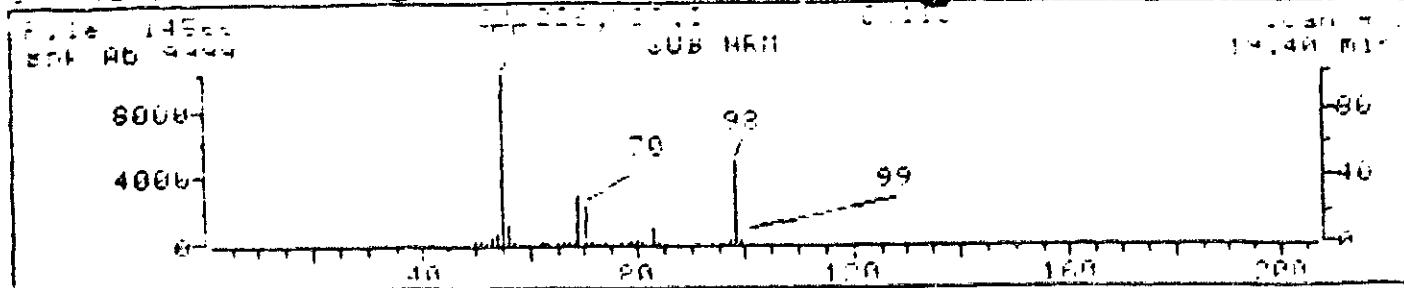
Date File: 214570
 Name: 841007 000 1
 Misc Data: 001140
 RT (min): 18.90
 Scan: 440
 Area: 10440
 Semi-quantitative Data: 24.78 NG

Data File: 214570 Scan Number: 440
 Search Speed: 2 Titrating option: 5 Number of ion ranges searched: 10

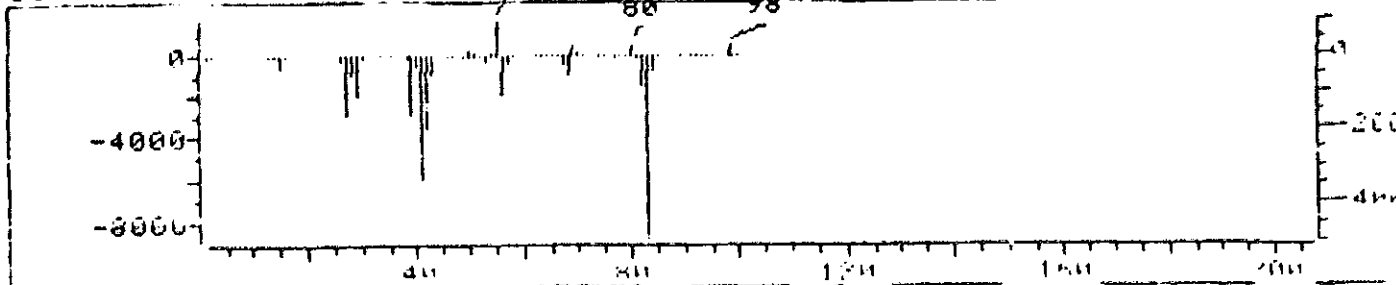
- 1. 2H-Pyran, tetrahydro-3-methyl- (8C19C1) 100 C6H12O
- 2. 2-Butenoic acid, 2-methyl- (9C1) 100 C5H8O2

	Prob.	Cart	V	dK	#Flg	Tilt
1.	27	26.093630	20	82	2	0
2.	24	13201462	24	84	3	0

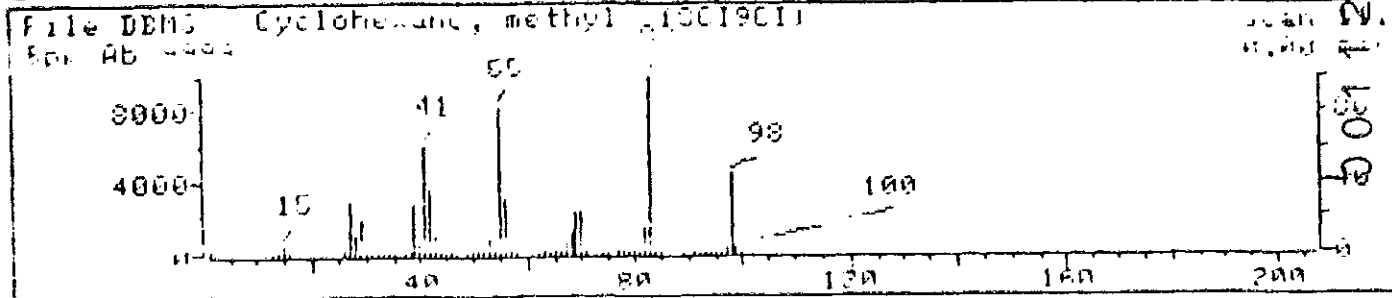
SAMPLE SPECTRUM (BEFORE FOUND SUBTRACTED)



DIFFERENCE



LITERARY REFERENCE SPECTRUM (BEST HIT)



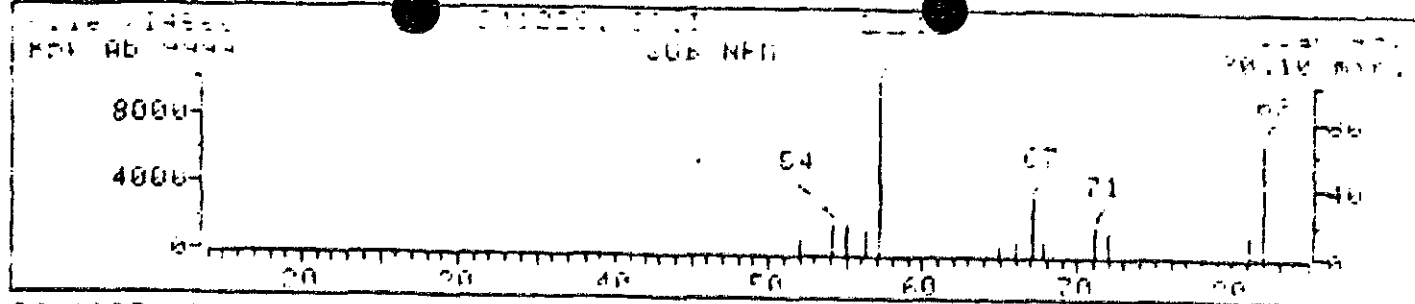
Data File 14560
Name 84122A,000,1
Misc Data 011150
PT (min): 19.40
Scan 473
Area 415679
Semi-quantitative Conc 1070.00 NG

Data File 14560 Scan Number 473
Search Speed 2 Titling option 9 Number of 100 ranges searched 1

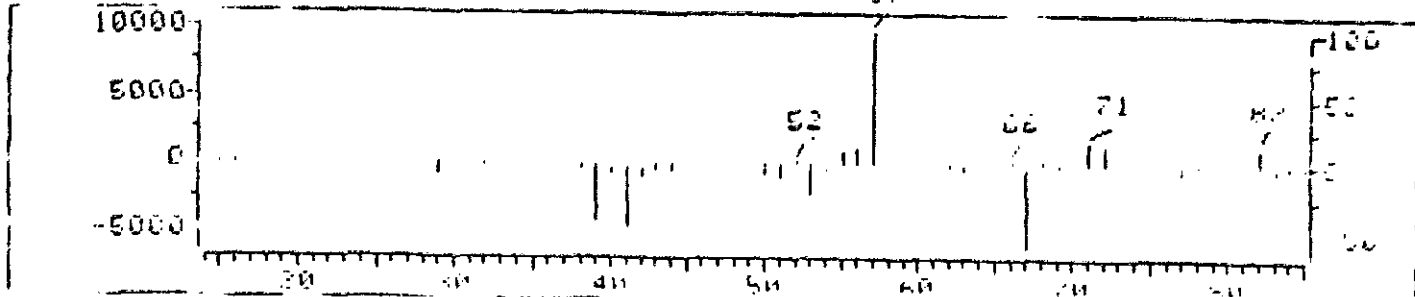
1. Cyclohexane, methyl (C6H11) 98 C7H14 1
2. 2-Pentene, 2,3-dimethyl- (C6H12) 98 C7H14
3. 3-Hexene, 3-methyl, (E) (C6H12) 98 C7H14

Peak	Comp	Int	dk	#Cl	Tilt
1.	70	100872	26	77	3 0
2.	52	1057475	37	68	0 0
3.	52	3899363	25	73	3 0

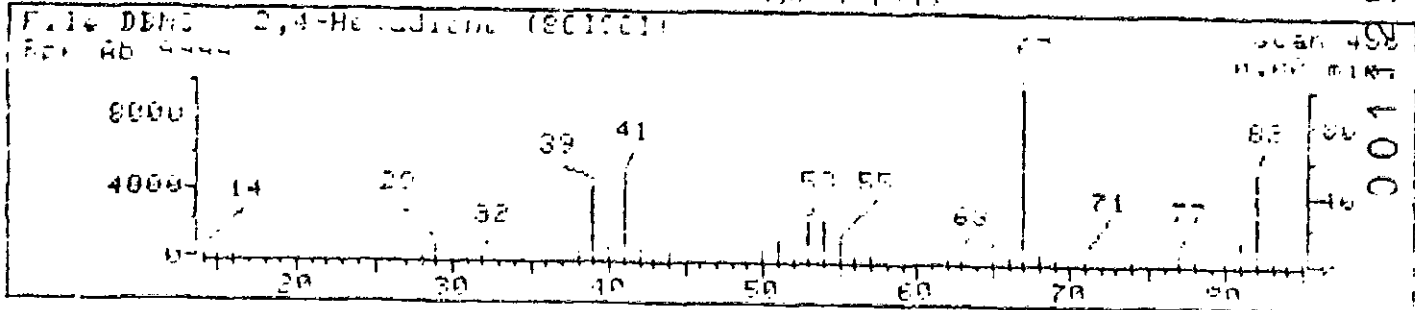
SAMPLE SPECTRUM (BACKGROUND SUBTRACTED)



DIFFERENCE



LIBRARY REFERENCE SPECTRUM



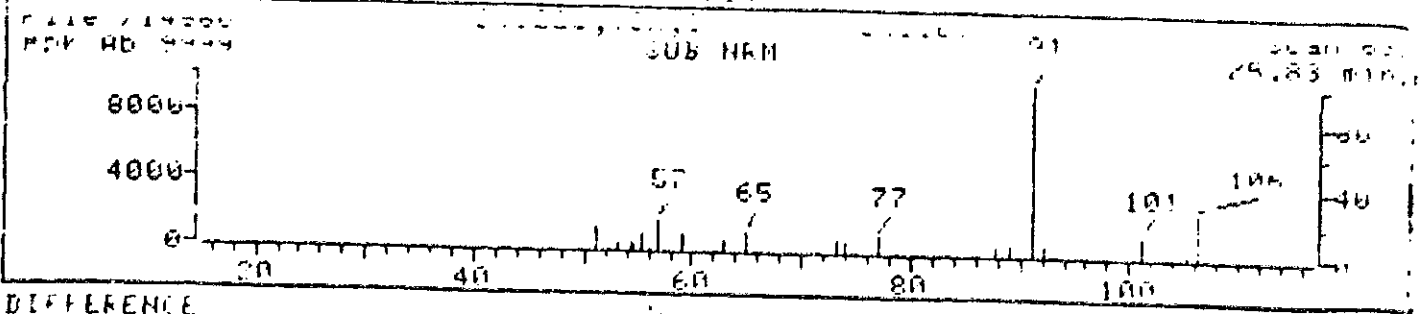
Name: 841026,006,1
 Misc Data: CAS160
 RT (min): 20.10
 Scan: 491
 Area: 1956
 Semi-quantitative Conc: 40.03 NG

Data File: 214560 Scan Number: 491
 Search Speed: 2 Tilt option: 5 Number of 10m ranges searched: 60

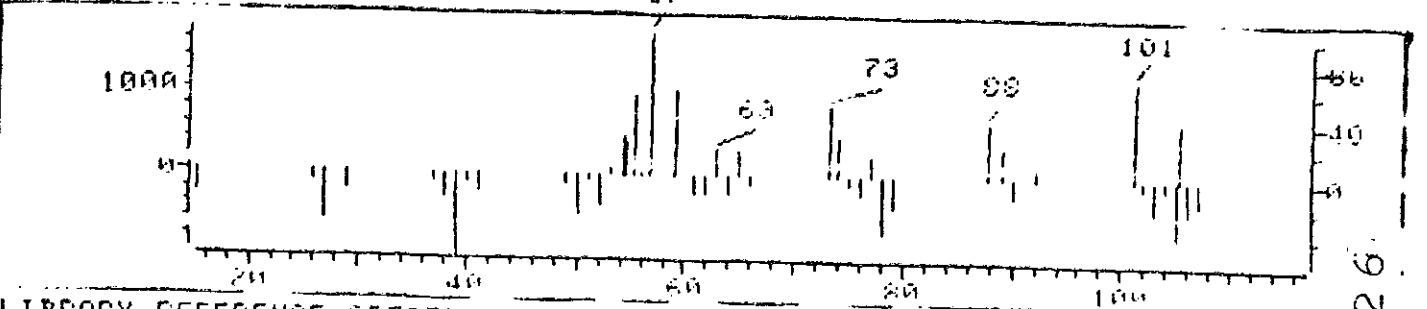
1. 2,4-Hexadiene (801901) 89.06H10

Prob.	Fact	K	dR	#Flg	Tilt
1	20	500461	22	75	3 0

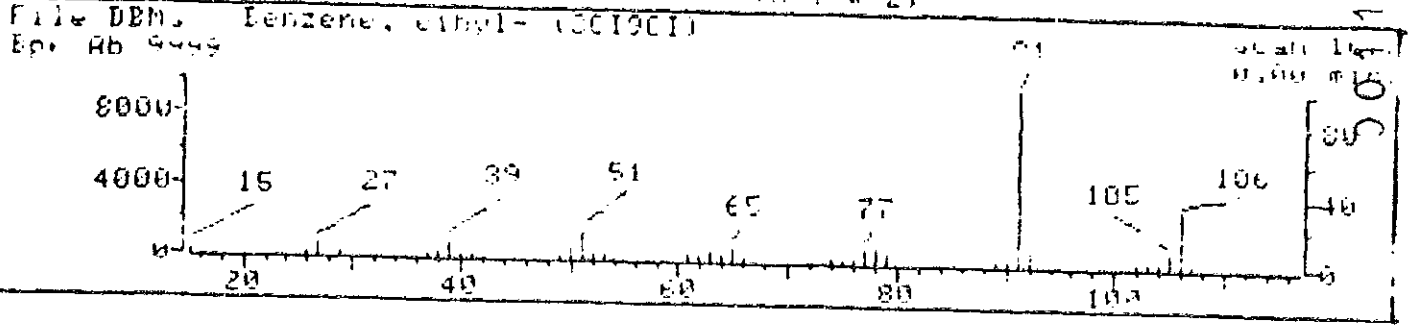
SAMPLE SPECTRUM (BACKGROUND SUBTRACTED)



DIFFERENCE



LIBRARY REFERENCE SPECTRUM



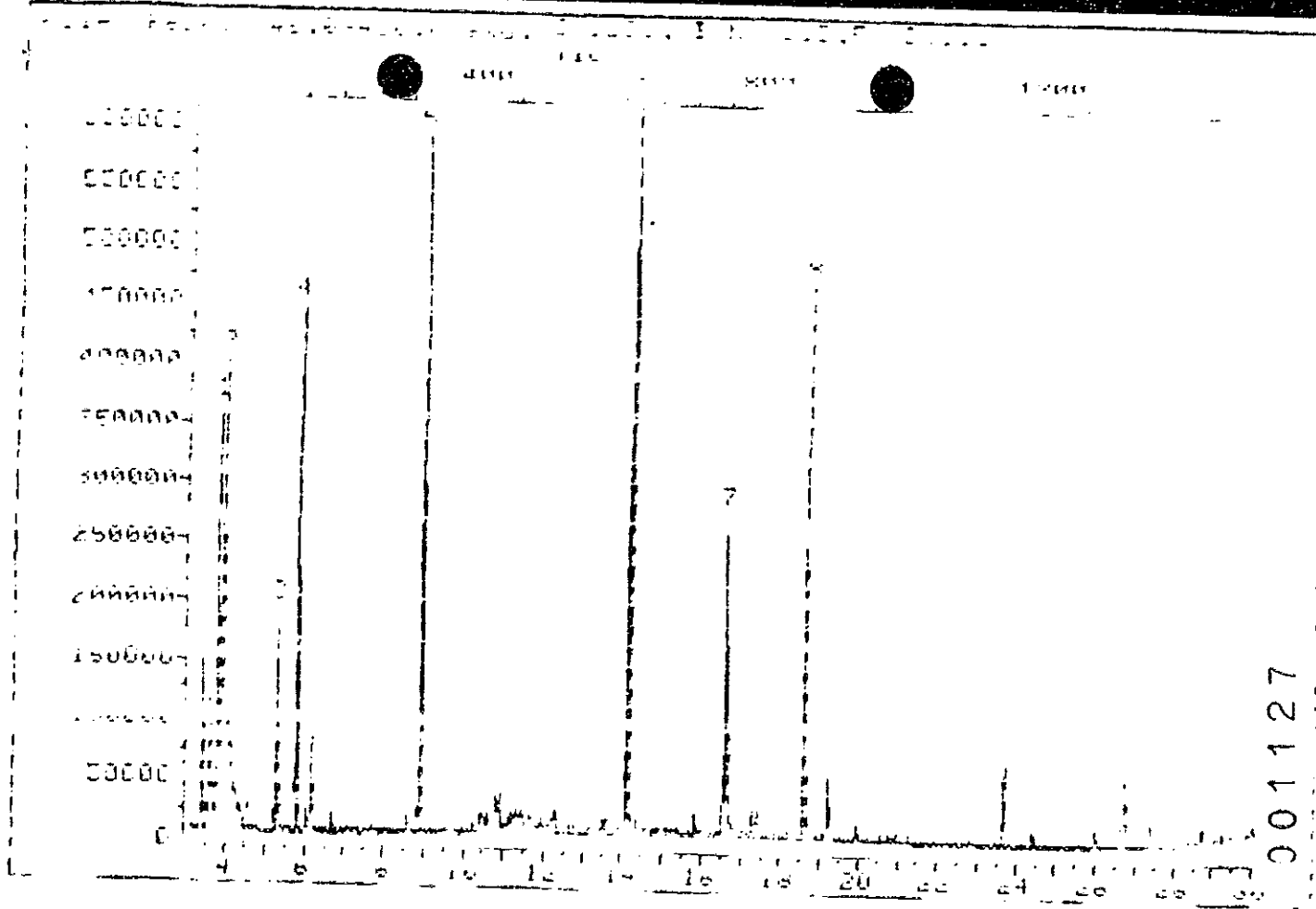
Data File: 714500 111
Name: 041026,00A.T
Misc Dir: 04116V
RT (min): 25.83
Scan: 470
Area: 20064
Semi-quantitative Conc: 51.66 NG

Data File: 714500 Scan Number: 470
Search Speed: 2 Tilt option: S Number of ion ranges searched: 2

- 1. Benzene, 1,2 dimethyl- (C10H14) 10% CBH10
- 2. Benzene, ethyl- (C10H12) 10% CBH10

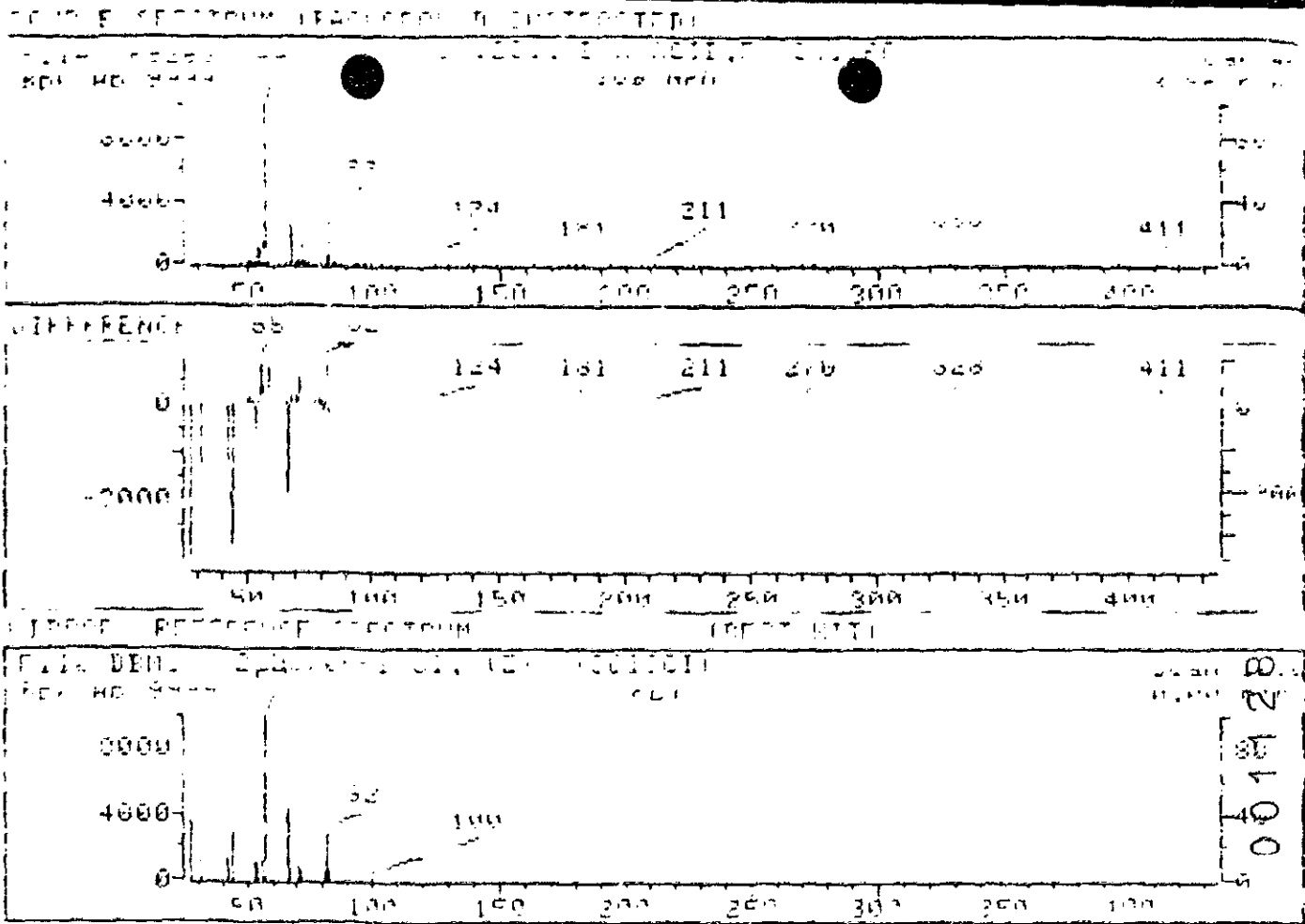
Prob.	Conf	K	dK	#Flg	Tilt
1.	49	95476	41	51	2 0
2.	47	100414	37	47	2 0

100
80
60
40
20
0
100
80
60
40
20
0



001127

DATA FILE: 001127.D
NAME: 001127.D (N-ACETYL)

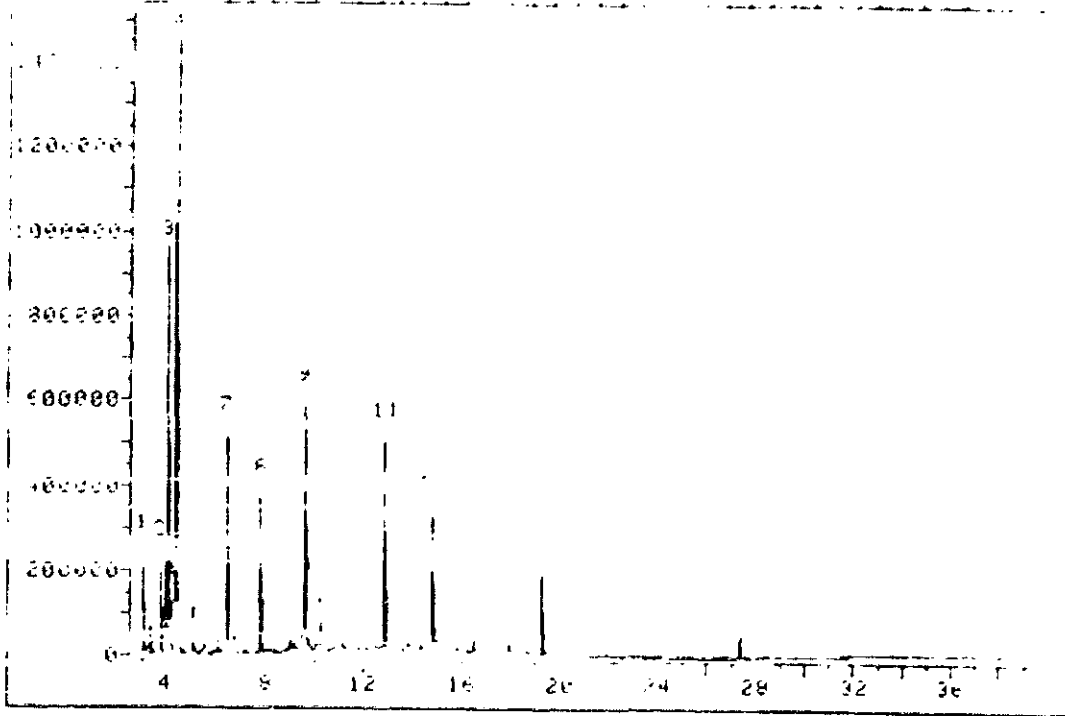


Data File: 11263
 Name: 041231 (N-411)
 RT (min): 1.00
 Scan: 41
 Area: 100000
 Semi-quantitative Conc: 26.51 (10/MI)

Data File: 11263 Scan Number: 41
 Search Speed: 1 Title: option 5 Number of top peaks: 3

- 1. 2-Hexan-1-ol (C6H14O) 100.00%
- 2. Cyclohexanol (C6H12O) 100.00%
- 3. Pentane-2,4-dione (C5H8O2) 100.00%

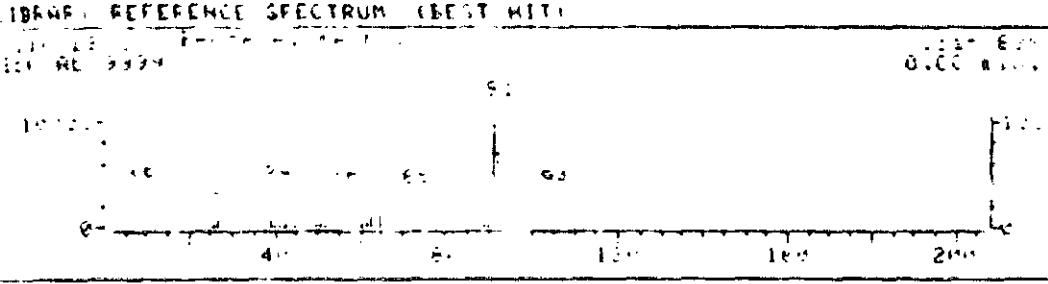
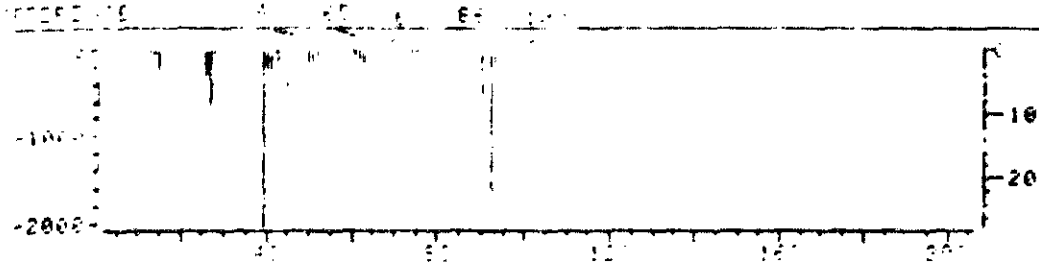
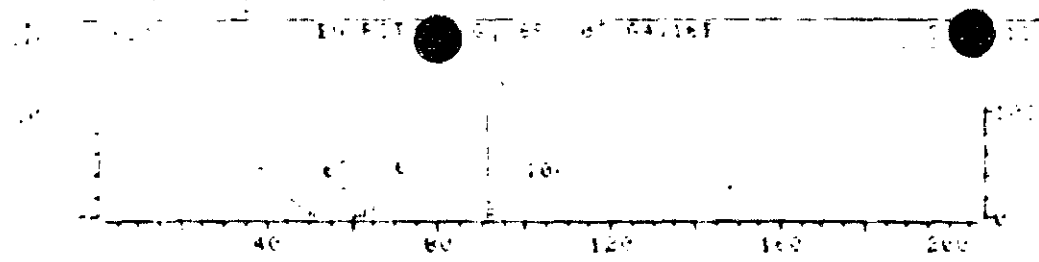
Peak	Area	K	RF	File	Alt
1	620000	10	51	2	0
2	150000	24	51	0	0
3	100007	23	61	3	0



Data File: 250521:U3
 Name: E. P. G. G. G.
 Misc Data: 6411.6

001129

0-1
 0-2
 1
 1
 2
 3



Data File: GC001:117
 Name: Benzene, 1,4-dibromobenzene
 Misc Data: GC114E
 RT (min): 2.70
 Peak: 1
 Mass: 235.96
 Semi-quantitative Comp: 0.75 0.6 1a

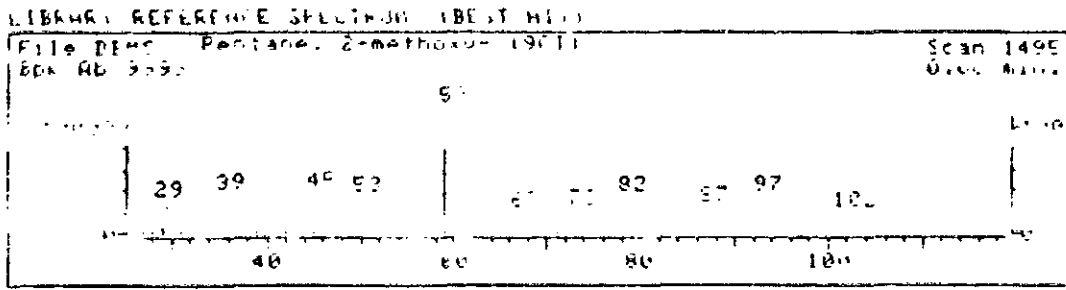
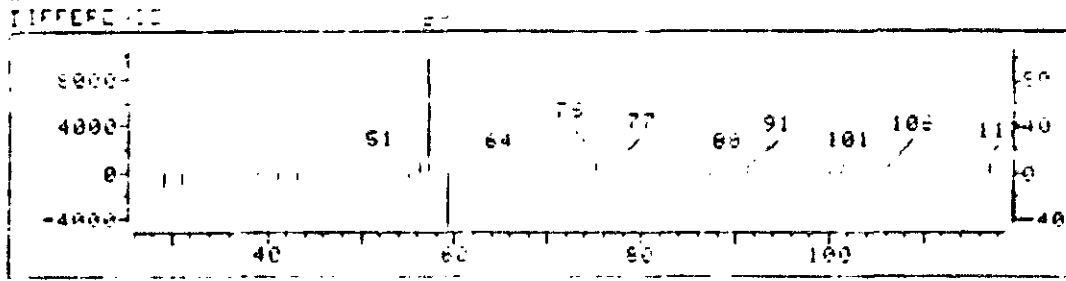
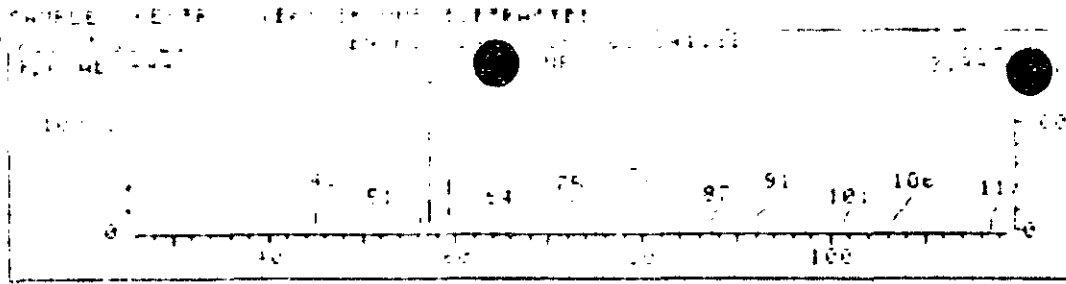
Data File: GC001 Scan Number: 13
 Peak Number: 1

1. Benzene, 1,4-dibromobenzene (100% 901)
2. Benzene, 1,3-dibromobenzene (100% 901)
3. Benzene, 1,2-dibromobenzene (100% 901)

Prob.	CH#	I	g	#lg	Tilt
1.	94	108883	76	14	1 0
2.	67	14469631	51	50	0 -2
3.	86	13637055	54	50	0 -2

001130

BT

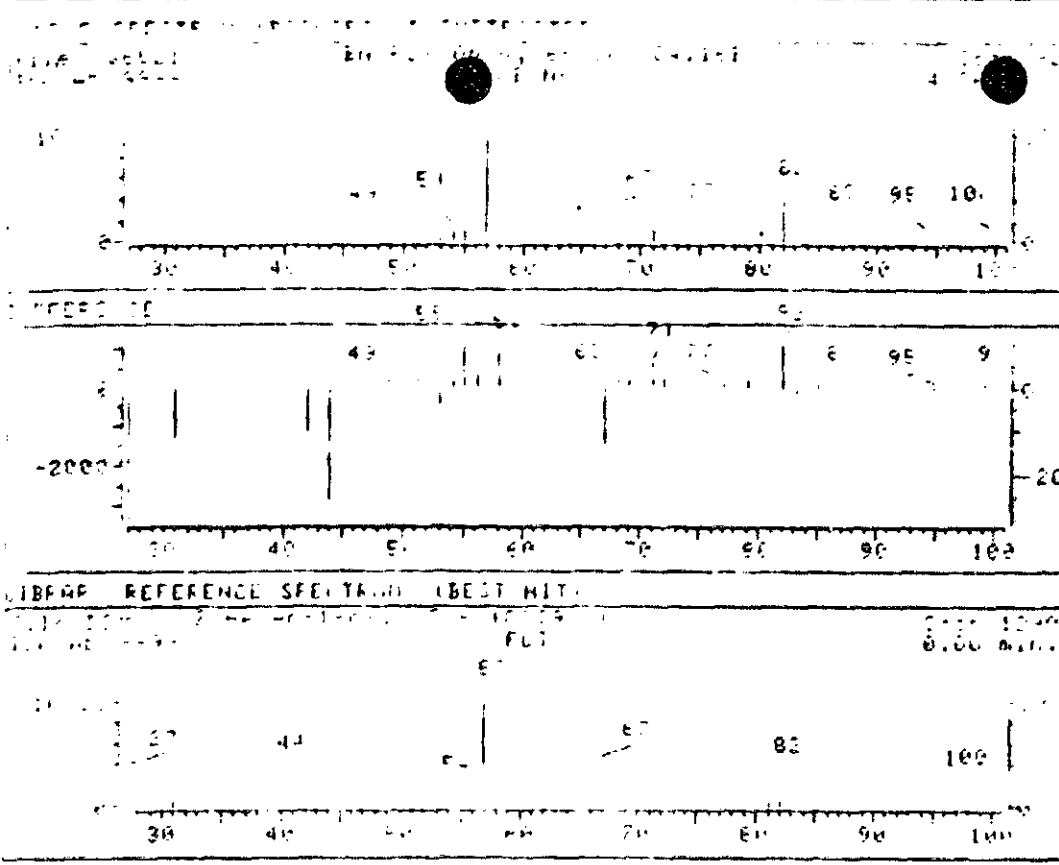


Data File: >60821:03
 Name: BL PFI GR G. SF 100
 Misc Data: 641 5E
 RT (min): 3.91
 Scan: 71
 Semi-quantitative Data: 40 59 25 10

Data File: >60821 Scan Number: 71
 Search Speed: 2 Titling Method: E Number of Libraries Searched: 100

1.	Prob.	Case#	Score	Library	Weight	Count	Ident	Library Name	Search
1.	11	6795826	98	53	2	0		Pentane, 2-methoxy- (9011)	1.

001131



Data File: >G6521::UP
 Instrument: BM-FST-04-G, RT-100
 Mass Data: G-1100
 RT (min): 4.70
 Scan: 92
 Semi-quantitative Conc: 1.1E-06 M

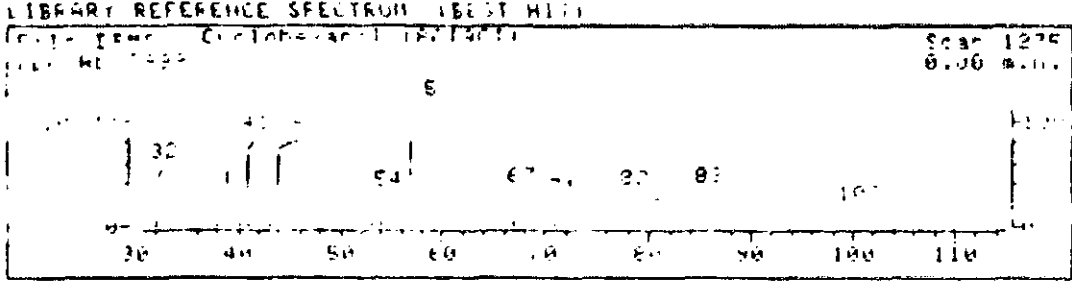
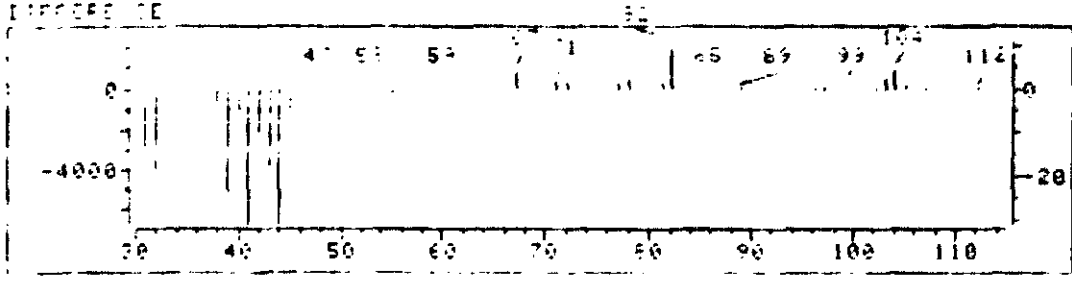
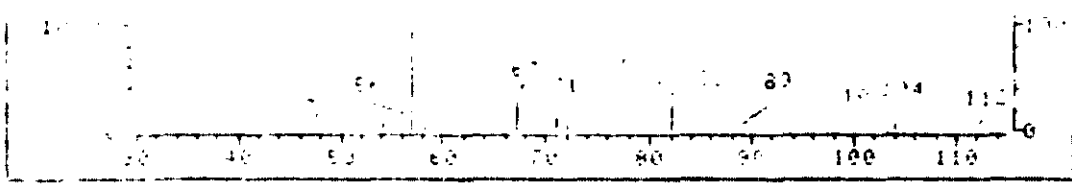
18 Files: >G6521 Scan Number: 92
 Search Method: Full Scan Search Number of Ion Pairs Selected: 5

1. 2-Nitrophenol, (2-N) (801901) 100.0% ID
2. 4-Nitrophenol, (4-N) (801902) 97.0% ID
3. Aziridine, 1-methyl- (801903) 97.0% ID

Prob.	Case#	i	d	sig	Tilt
1.	48	928949	36	62	1 0
2.	33	106950	36	70	1 0
3.	29	1072442	34	56	0 0

001132

File: 16521
 Date: 11/11/83



Data File: 16521:113
 Name: Cyclohexanol (871901)
 Misc Data: 64.166
 RT (min): 4.42
 Scan: 1275
 Semi-quantitative Conc: 100.93 ug/ml

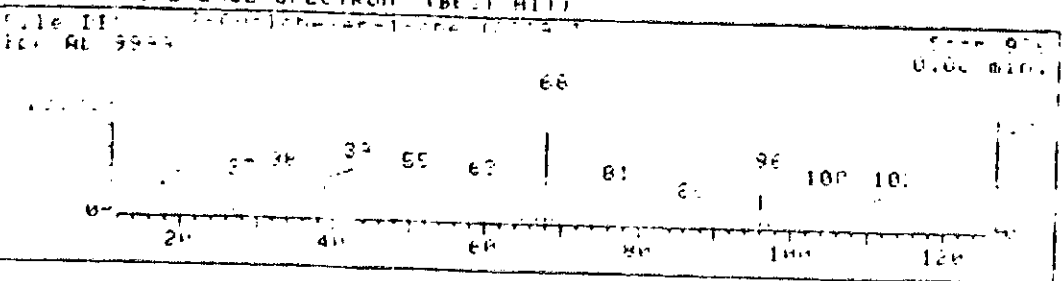
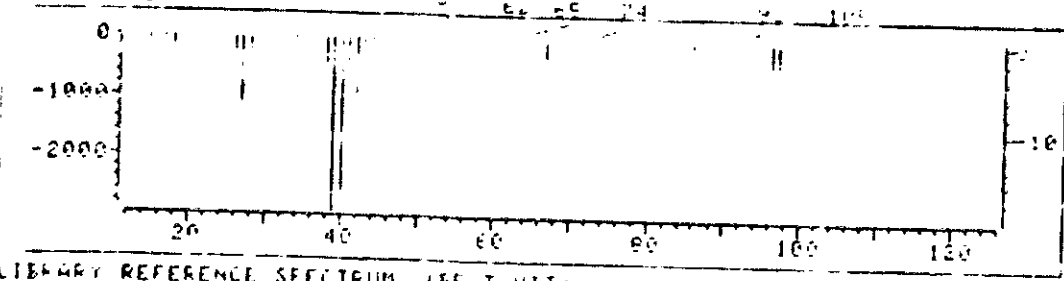
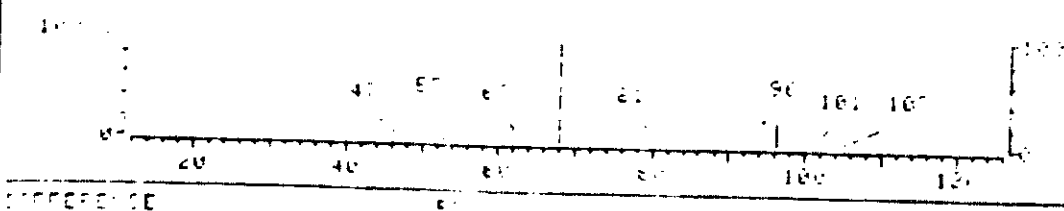
Data File: 16521 Scan Number: 95
 Search Speed: 2 Tilt Ang: 0.000 Number of 100 ranges selected: 10

- 1. Cyclohexanol (871901) 100.0000
- 2. 1,4-Dioxane (1072442) 0.0000
- 3. Aziridine, 1-methyl- (001901) 0.0000

Prob.	Ca#	F	PK	4Fig	Tilt
1.	51	108930	35	67	0 0
2.	35	4791402	37	61	3 0
3.	32	1072442	25	50	0 0

001133

INSTRUMENT: FTIR-2000
 SAMPLE: 9944
 DATE: 01/10/93
 TIME: 10:00



Data File: 200511407
 Name: 2-Fluorobenzoic acid
 Misc Data: 0.000
 RT (min): 1.04
 Peak: 170
 Semi-quantitative Conc: 10.7% UC NL

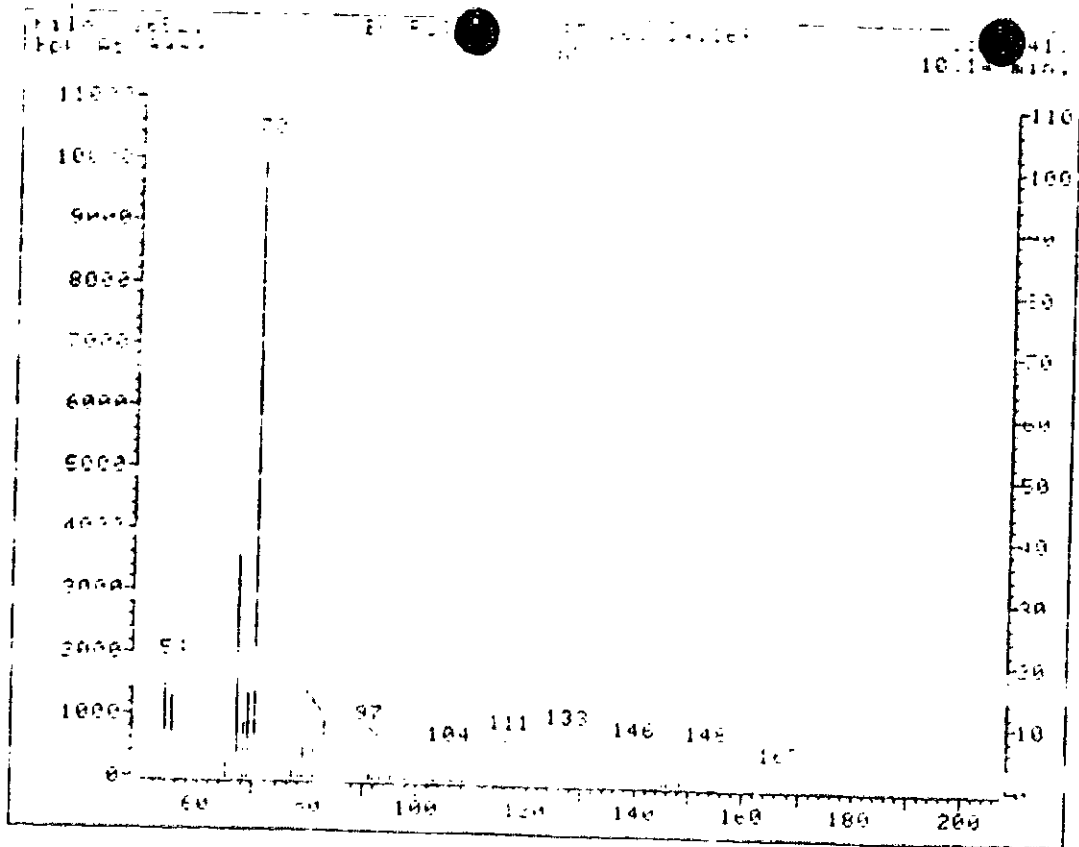
2-Fluorobenzoic acid

Data File: 200511407 Scan Number: 130
 Peak: 170 Total Area: 1.5e5 Number of Ion Range: 170

1. 2-Fluorobenzoic acid (01901) 96.5%

Prob.	Case	i	d	+ lg	Tilt
1.	03	97.687	44	41	2 0

001134



Data File: >Gc021:00
 Sample: F0.PE3.M.S. 000007
 Misc Data: G4160
 RT (min): 10.14
 Scan: 418
 Semi-quantitative Data: 10.21 GC NL

001135

6.17.11

Use RT file for this scan.

Appendix E

Chain-of Custody Forms

- 1) A field Chain-of-Custody form (CC1) is included for all samples shipped by ETC shuttle
- 2) An in-house sample Chain-of-Custody form is included for the period the sample was in ETC's possession.
- 3) A subcontractor's Chain-of-Custody form is included for any analytical work not performed within ETC's laboratory.
- 4) Any additional Chain-of-Custody material provided by a client or by a client's sampling agent is also included

001136

CHAIN OF CUSTODY FORM (CCF)

See Date

Prepared By

6/28/84
J. Trotter
64116/mj

Company

RESOURCE ENGINEERING

Facility Name

11 GREENWAY PLAZA

Attn

Coby Sher

Address

Houston, TX 77406

Phone

() -

SAMPLE IDENTIFICATION

Facility Site Code

RES11271501A

Source Codes

We (W) River/Stream (R) Surface Impoundment (I) Lake/Ocean (L)
 Soil (S) Bottom Sediment (D) Pretreatment Facility (P) Treatment Facility (T)
 Outfall (C) Generation Point (G) Leachate Collector Sys (C) Other (O)

W

FR-11

12/12/84

Source Code (1st 3 digits)

Year Sample Point ID

Start Date (mo/day/yr)

Start Time (2400 hr. clock)

Elapsed Time (complete)

Example

W

1101J263P

05/15/81

09:30

1 hr

40
5

SAMPLE CONTAINER

Sample Bottle	Container	Sample Bottle	Condition
E94/45	E-E4		00
E94/45	V-V2		00
T94/45	T6	hdsp	

ETC # changed to per of Ellen L. 64116 request

CHAIN OF CUSTODY CONTINUED

1. Shuttle Opened By (print): DEAN GORE Date: 6/29/84 Time: 8:30 AM
 Signature: [Signature] Sec # 0018125 Inoc: YES

2. I have received these materials in good condition from the above person.
 Name: Derrick Vallance Signature: [Signature]
 Date: 12/12/84 Time: 10:00 Remarks:

3. I have received these materials in good condition from the above person.
 Name: S.C. Itin Signature: [Signature]
 Date: 12-14-84 Time: 3:00 PM Remarks:

4. I have received these materials in good condition from the above person.
 Name: _____ Signature: _____
 Date: _____ Time: _____ Remarks: _____

5. Shuttle Sealed By (print): S.C. Itin Date: 12-14-84 Time: 3:15 PM
 Signature: [Signature] Sec # 0018126

ETC USE ONLY Opened By: Jacobs Date: 12-17-84 Time: 9:30
 Sec # 18126 Condition: ok

E9445
ETC Sample No.

SAMPLE POINT INFORMATION FORM (CC2)

FIELD MEASUREMENT DATA

Select up to 3 parameters to be recorded by entering the appropriate code letter in the first space for each of the 3 data entry fields provided. Enter the actual measurement data (in the units specified) for the three parameters you have selected.

PARAMETERS	ACTUAL (let just fly)	EXAMPLE
Flow (CFS) A	<input type="text"/>	C 2.5 <input type="text"/>
Volume (Gals) B	<input type="text"/>	F 5.0 <input type="text"/>
Sample Depth (ft) C	<input type="text"/>	G 1 1 0.3 <input type="text"/>
Depth to Water (ft) D	<input type="text"/>	
Discharge Rate (GPM) E	<input type="text"/>	
Depth to Bottom (ft) F	<input type="text"/>	
Event Time (2400-Hr Clock) G	<input type="text"/>	
Depth to Screen (ft) H	<input type="text"/>	

FIELD TEST DATA

DC (Mg/L) Sample Temp. (°C)

pH
 Single Measurement 2nd of Quadruplicate 3rd of Quadruplicate 4th of Quadruplicate

Specific Conductance (uMOS/cm)
 Single Measurement 2nd of Quadruplicate 3rd of Quadruplicate 4th of Quadruplicate

THE FOLLOWING DATA IS FOR YOUR RECORDS ONLY

SAMPLING METHOD (choose one)

- | | | |
|----------------------------------------|--------------------------------------------|--------------------------------------|
| <input type="checkbox"/> AIR-LIFT PUMP | <input type="checkbox"/> PERISTALTIC PUMP | <input type="checkbox"/> TRIEF |
| <input type="checkbox"/> AUGER | <input type="checkbox"/> PETERSEN | <input type="checkbox"/> TRIER |
| <input type="checkbox"/> BAULER | <input type="checkbox"/> PISTON PUMP | <input type="checkbox"/> VEHMEYER |
| <input type="checkbox"/> BOTTLE | <input type="checkbox"/> SCOOP / SHOVE | <input type="checkbox"/> OTHER _____ |
| <input type="checkbox"/> COLUMBIA | <input type="checkbox"/> SQUEEZE PUMP | _____ |
| <input type="checkbox"/> DIPPER | <input type="checkbox"/> SUBMERSIBLE PUMP | _____ |
| <input type="checkbox"/> KEMMERER | <input type="checkbox"/> SUCTION LIFT PUMP | _____ |
| <input type="checkbox"/> NISSEN | <input type="checkbox"/> SURBER | _____ |

SAMPLE TYPE (choose one)

GRAB () COMPOSITE () OTHER ()

_____ (describe) _____ (describe)

WEATHER

SAMPLE DESCRIPTION (e.g., color, odor)

_____ (describe) _____ (describe)

Form Prepared By: _____ name (print) Employer: _____

X-MS Analysis Custody Log

DATE 8/22/84 SHIFT 12/00
 FRACTION VOA
 INSTRUMENT J
 SEQUENCE FILE 8
 METHOD FILE VOAT
 ID FILE VOA3
 ANALYST W. M. ...
 Supervisor M. S. ...
 BATCH #'s QV 26490

(PLEASE INITIAL)

CURRENT CSUS STATUS		STANDARDS UPDATED	
ICR	ST	DATE	RM 12/22
IP	ST	BY	

STANDARD	PPR	NO.	VOL.
OBEG	50	8103	1
ISTD	40	9140	3
SOLC	25	8989	10
ABC	18	8047	10
		7402	

U1, U2 REF 2

NAME	DATA FILE	UL INJ	UL F	BIL	TAPE 0	SPECIALS (WRITE A-TYPE)	PLU Y/M
OBEG	2I4546				I00130		
QC 2649V	2I4547				NG	5 μmol (455)	
QC 2649V5	2I4548				CP	10 UL ABC	
QC 2649V	2I4551				NG		
QC 2649V	2I4552						
QC 2649V	2I4553				OK 36102		
G1891V5	2I4554	5ml		4	OK R	10 UL ABC	
G1891V	2I4555	5ml		5	OK S		
G4242V	2I4556	1		2	T	Return 1:25	
G4242VR	2I4557			3	U	Return 1:25	
G4243V	2I4558			4	OK V		
G4244V	2I4559			5	OK W		
G4116V	2I4560			6	OK X	3:11	
G1879V	2I4561			7	OK 362AA		
G1892V	2I4562			8	OK B		
G1905V	2I4563			9	OK C		
G1409V	2I4564	V		10	OK D		
G4242V	2I4565		1:25	4	OK		
G4242VR	2I4566		1:25	5	OK		

Nov 11-12-84

Sample ID	Lab List	Vol (ml)	BN	ACID
F9625	721E	810	1.0	/
F9626	↓	980	1.0	/
61019	7284	1000	1.0	/
F8475	7075	930	1.0	/
62866	7442	1000	/	10
63682	7601	995	1.0	50
63215	7603	1000	1.0	10
64116	7600	935	1.0	10
60668	7631	970	1.0	10
60669		1000	1.0	10
60672		1000	1.0	10
60673	↓	970	1.0	10
G1654	7615	1000	1.0	10
QC 2525		1000	1.0	10
QC 2525 S		1000	1.0	10
G0669 S		1000	1.0	10
G1654 R		985	1.0	10

QC Batch # 2525

Analysis PP/TOTAL

REPEATS

LMPD/BN ; CMPD/ACID

Matrix H2O

Turnaround ASAP

Date Start 12/27/24

BN _____

ACID _____

Extraction Method: _____

Sep. Funnel _____

Continuous _____

Other _____

COMMENTS:

REPEATS:

SAMPLE #	FRACTION	QC #	COMMENT
F9625	BN	2416	low score
F9626	↓	↓	low
61019	BN	2419	NO SCORE REC.
F8475	BN	2433	SPECIAL LMS NOT AVOID FOR REPEAT
62866	ACID	2462	low score

* 9 caps @ 150 ug/ml
2 caps @ 250 ug/ml

** 16 caps @ 100 ug/ml
chlor @ 200 ug/ml

001140

Spike/Sum. spiked: 5 ml water 12/27/24

FRACTION	SPIKE			SURROGATES		
	Amt (ml)	Conc.	Lot #	Amt (ml)	Conc.	Lot #
BN	1.0	100 ug/ml	9209			
ACID	1.0	*	9013			
PEST	1.0	**	9502			
Other AR 1260	1.0	100 ug/ml	9262			
LMPD/ACID (9.0 ml)	1.0	150 ug/ml	9379			
CMPD/BN	1.0	100 ug/ml	9163			
GERANTY & MILER BN SPIKE						
QA				1.0	100 ug/ml	24-96

Set-up: _____ BN Conc. Done - Patrick 12/27

Supervisor: _____ ACID Conc. Done - Koleb 12/27/24

GC-15 Analysis

DATE: 8-4-73 SHIFT

FRACTION: BN + Acids

INSTRUMENT: F

SEQUENCE FILE: JM

METHOD FILE: BURE/ACIDF

ID FILE: FBIP/FAID

ANALYST: [Signature]

SUPERVISOR: [Signature]

BATCH #'S: QA2508

QA2525, QA2506

(PLEASE INITIAL)

CURRENT LAB STATUS		STANDARDS UPDATED	
AS	2/12	DATE	
WS	8/12	BY	

12-31 UPDATED

NAME	UL INJ	DATA FILE	TYPE	REMARKS
NETPP	25			9295
Acid Calib 1	60			9010
Acid Calib 2	100			9011
Acid Calib 3	300			9212
BN Calib 1	60			9526
BN Calib 2	100			9527
BN Calib 3	300			9528
M-Chlorophane	60			9382
M-Chlorophane	100			9381
M-Chlorophane	300			9380

NAME	DATA FILE	UL INJ	TYPE	SPECIALS (WRITE N-TYPE)
NETPP	#1 7F6238			T 00131
BN Calib X 3	7F6240	1		
BN Calib 2 2	41	2		
BN Calib 8 1	42	3		
Acid Calib 3	43	4		
Acid Calib 2	44	5		
Acid Calib 1	45	6		
QC 2508 AB	46	7		
F 9154 AB	47	8		
F 9155 AB	48	9		
F 9156 AB	49	10		
F 9157 AB	50	11		
F 9149 AB	51	12		
F 9150 AB	52	13		
F 9148 AB	53	14		
Acid Calib 2	54	15		
300ppm M-chlorophane	55	16		
100 " "	56	17		
60 " "	57	18		
QC 2525 AS	58	19		
G-0669 AS	59	20		
QC 2525 A	60	21		
G-1654 AR	61	22		
G-1654 A	62	23		
G-4116 A	63	24		

Chemp company (w/olows)
wrote only computer printout
as report
Such BN+15

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Analysis Custody Log

03 SHIFT
 NAME BN 2525
 NUMBER 7
 SOURCE FILE 7A
 SOURCE FILE BNDC
 LE 8300
 VISIT 1/17/84
 HOURS 7:30-5:25

(PLEASE INITIAL)

STANDARD	STANDARDS UPDATED
<u>7A</u>	DATE BY

STANDARD	QTY	QTY	QTY
DETPP	25	4534	2nd
Calib 1	1	4526	1st
Calib 2	10	4537	
Calib 3	300	9538	
Calib 4	150	4524	

NAME	DATA FILE	UL INJ	QTY	DIL	TAPE #	SPECIALS (WRITE A-TYPE)	PLUS Y/N
DETPP	766507	2nd					
Calib 1	766508		1				
Calib 2	766509		2				
Calib 3	766510		3				
Calib 4	766511		4				
QC2525B	766512		5			BN+1	
QC2525BS	766513		6				Y
G-0669BS	766514		7				
G-9625B	766515		8				
F9626B	766516		9				
G-1019B	766517		10				
F-975B	766518		11				
G-3682B	766519		12				
G-37215B	766520		13				
G-4116B	766521		14				Y
G-0668B	766522		15				Y
DETPP	766523		16				Y
Calib 2	766524		17				
G-0669B	766525		18				Y
G-0672B	766526		19	(83)			Y
G-0673B	766527		20				Y
G-1654B	766528		21				Y
G-1654BR	766529		22				
300 100/100m	766530						
60 100 "	766531						
300 "	766532						

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