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EDITORIAL

Surgeon's perspective on short bowel syndrome: Where are we?

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Abstract

Short bowel syndrome (SBS) is due to a massive loss of small bowel: the reduction of gut function is below the minimum necessary to maintain health (in adults) and growth (in children) so intravenous supplementation is required. Parenteral nutrition represents the milestone of treatment and surgical attempts should be limited only when the residual bowel is sufficient to increase absorption, reducing diarrhea and slowing the transit time of nutrients, water and electrolytes. The surgical techniques lengthen the bowel (tapering it) or reverse a segment of it: developed in children, nowadays are popular also among adults. The issue is mainly represented by the residual length of the small bowel where ileum has shown increased adaptive function than jejunum, but colon should be considered because of its importance in the digestive process. These concepts have been translated also in intestinal transplantation, where a colonic graft is nowadays widely used and the terminal ileum is the selected segment for a living-related donation. The whole replacement by a bowel or multivisceral transplant is still affected by poor long term outcome and must be reserved to a select population of SBS patients, affected by intestinal failure associated with irreversible complications of parenteral nutrition.

Key words: Parenteral nutrition; Bowel rehabilitation; Surgical rescue; Intestinal transplantation; Short bowel syndrome

Core tip: Short bowel syndrome represents a surgical dilemma: parenteral nutrition is considered the gold standard of care and any surgical attempt must be limited by the universal principle "first do not harm." The surgical rehabilitation should be pursued when there are enough residual intestines to obtain a better bowel function: lengthening the intestine or reversing a loop of it with different techniques should have the only aim of slowing the transit while increasing the absorptive surface. When intestinal failure is associated to life-threating parenteral nutrition complications, bowel transplantation should be considered as an option.

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INTRODUCTION

Short bowel syndrome (SBS) results from a reduced length of the small intestine. A "normal small bowel length," measured from the duodeno-jejunal flexure to ileocolic valve, is estimated at 250 cm ± 40 cm at birth, and the growth is maximal during the first year of life[1]. In adults, the small bowel length varies from 275 cm to 850 cm, with a mean of 350 cm ± 60 cm, depending on the method used, radiologic, surgical, or per autopsy[2]. The massive loss of small bowel represents the most frequent mechanism of intestinal failure, defined by the European Society for Clinical Nutrition and Metabolism as "the reduction of gut function below the minimum necessary for the absorption of macronutrients and/or water and electrolytes, such that intravenous supplementation is required to maintain health and/or growth"[3]. Among children "the minimum necessary for the absorption" is a residual small bowel length of more than 25% of the expected for gestational age[3], in adults SBS usually appears when the small bowel length is less than 200 cm (67% of the normal length)[4]. Malabsorption and diarrhea represent the classical symptoms, associated to deficit of growth in the pediatric population. Wilmore *et al*[5] first demonstrated long-term survival with parenteral nutrition (PN) in a child affected by SBS. Nowadays home PN represents the standard of care in patients affected by massive loss of small bowel with excellent long term results[6-12]. PN does not replace physiologically the bowel function because uses the intravenous route to supplement nutrients and it could be affected by several life-threating complications. Under this perspective, a surgical rehabilitation in case of SBS should be represented by: (1) the possibility to slow the transit and obtain an adequate absorptive surface of the remnant intestine through lengthening procedures and (2) whole replacement of the massive intestinal loss with a bowel transplant. It is worthwhile to analyze briefly the main reported studies on the issue in international literature, in order to develop an updated perspective under the surgical point of view.

OVERVIEW OF THE LITERATURE

SBS is mainly, but not only, a matter of length. In children, the massive resection of the small bowel could lead to a "very short bowel syndrome" (≤ 40 cm)[13,14], "ultra-short bowel syndrome" (between < 30 and < 10 cm)[15-17] or "no gut syndrome" (only the duodenum is left)[18-20]. Adults with less than 200 cm but more than 75 cm of small bowel[21] have a potentially functional intestine especially if the colon (and specifically the ileocolic valve) is preserved in continuity. Among SBS patients, the role of the colon in the process of digestion has been demonstrated since the '90s[22-25]. The presence of remaining colon is associated with a lower dependency on PN[26,27] and there is agreement that the remaining small bowel after massive intestinal loss is supported by the colon (if in continuity) for completion of the digestion process. On the other hand, jejunum and ileum have different roles in digestion and ileum has probably a greater adaptive potential than jejunum[28]. A remnant ileum (especially in continuity with the colon) could probably guarantee a faster weaning from PN. Clinical experience shows that patients with a jejuno-colonic anastomosis (SBS type II), even better with a jejuno-ileo-colonic anastomosis (SBS type III), have an improved absorption with time after a period of intestinal rehabilitation, whereas patients with end-terminal jejunostomy without colon (SBS type I) do not show that. When the colon is missing, among adults 115 cm of small bowel with an end enterostomy are considered the limit before SBS.

SURGEON'S PERSPECTIVE

In SBS the remaining small bowel may dilate. This is important for surgeons in order to lengthen the intestine, tapering it. It has been shown that the extent of dilation is associated with the bowel length, and both are related to enteral autonomy[29]. Two surgical procedures are popular in order to lengthen the bowel: Bianchi and Serial Transverse Enteroplasty Procedure (STEP). The Bianchi procedure, summarized by Bianchi in 1997[30], is also known as longitudinal lengthening and tailoring (LILT). The small bowel mesentery is separated as two leaves with a GI anastomosis stapler to create a tunnel, and then the two resulting small bowel segments of smaller diameter are connected with an end-to-end anastomosis in an iso-peristaltic fashion. In the STEP, first described by Kim in 2003[31], the dilated small bowel is narrowed by serial transverse applications of the GI stapler from opposite directions, creating a new lengthened small intestine (zig-zag channel). This procedure does not require an intestinal anastomosis and the mesenteric vascular supply is untouched. Since its first description, STEP has become a widespread procedure, sometimes repeated on the same patient (re-STEP) to obtain a longer intestinal segment. Bianchi and STEP procedures have been performed at first in children and more recently also in adults[32-35]. Most of the studies are on STEP: while enteral autonomy (median time: 21 mo) is eventually possible in some patients[36], improved enteral tolerance can be achieved in a majority[37,38]. STEP can be performed on shorter intestinal segments or intricate segments such as the duodenum, which is technically not feasible for Bianchi procedure, and it seems to have a lower mortality but an overall progression to transplantation[39]. The spiral intestinal lengthening and tailoring procedure is a new

surgical technique based on a spiral shape incision of the dilated intestine (at 45°-60° to its longitudinal axis), and re-tubularization in a longer but narrower fashion. It does not alter the orientation of the muscle fibers like STEP, offering minimal mesenteric handling compared to Bianchi procedure. It has been reported in a 3-year-old girl[40] where, 6 mo after the procedure, PN was weaned off. Another manuscript described the technique in a 10-month-old child[41] showing at 1-year follow-up a growth on the 15-25th centile on 82% oral calories and 18% PN, passing 2-3 daily stools. Three children with "no gut" syndrome and dilated duodenum underwent a novel surgical procedure of "duodenal lengthening" combined with a technical modification of STEP[18]: duodenal tapering was performed with sequential transverse applications of an endoscopic stapler on the anterior and posterior wall of the duodenum, avoiding bilio-pancreatic injury. Two patients weaned PN off at 12 mo post-surgery and the last one's PN caloric requirements decreased by 60%. The surgical rescue of "no gut" syndrome has been reported in adults as well. Bueno *et al*[20] demonstrated the feasibility of lengthening a dilated duodenum in a patient where his mega-duodenal stump was tapered by STEP, restoring his digestive continuity through an end-to-side duodeno-colonic anastomosis. After 24 mo of follow-up, the time on daily PN was shortened from 24 to 9 h and the volume and calorie requirements were reduced by half.

Since lengthening procedures slow the bowel transit time, a "reversed anti-peristaltic segmental bowel loop" has been proposed with the same aim: this procedure can be indicated in patients with an adequate remnant bowel length. Median oral autonomy was described up to 100% ± 38% with a lower amount of parenteral calories, as well as PN dependence[42]. In another report[43] 56% of patients improved their enteral autonomy.

The different graft types used in intestinal transplantation are the isolated small bowel, combined liver-intestine, multivisceral and modified multivisceral ones[44]: liver-containing grafts have shown the longest survivals. Apart from cadaveric donation, living-related intestinal transplantation has been pursued especially in a pediatric setting[45]: terminal ileum represents the used graft, because of technical feasibility and its greater adaptive potential than jejunum[28]. Short term results of intestinal transplantation have recently improved in terms of survival and digestive autonomy, due to advances in surgery and immunosuppression. Immunosuppressive therapy has evolved significantly over the past 20 years: the tacrolimus-based therapy as maintenance, preceded by induction with anti-thymocyte globulin or an interleukin-2 blocker, is the main used protocol worldwide. A "secondary" agent like steroids, azathioprine, mycophenolate mofetil or an mTOR inhibitor is recommended after an episode of rejection. Innovative cross match strategies and optimizing organ allocation could improve the long-term outcome, but the main causes of death and graft loss remain sepsis and rejection. Challenges for long-term results are chronic rejection and immunosuppressant-related complications[46,47]. According to Intestinal Transplant Registry reports[44], 1611 children were transplanted worldwide between 1985 and 2013, with an overall patient survival rate of 51%. In the 2014-2016 Scientific Registry of Transplant Recipients[48], the 6 American centers that in 2016 performed 10 or more intestinal transplants in adults reported a 1-year graft survival from 61% to 83% and a 3-year graft survival from 29% to 73%. In an earlier report from 2008 to 2010, the 1-year graft survival in adults was 71%, illustrating the relatively modest gains achieved[47]. Intestinal transplantation should be suggested to a very select subset of SBS patients with severe and irreversible complications of PN and no hope of intestinal rehabilitation. In conclusion, among SBS patients the surgical rehabilitation (Table 1) of the remnant bowel must be performed to slow the intestinal transit time increasing at the same time the absorptive surface: only in cases of irreversible intestinal failure with PN life-threating complications, intestinal transplantation could represent a therapeutic option even if still encumbered by suboptimal long term results.

Table 1 Surgical rehabilitation of short bowel syndrome

SBS surgical rehabilitation
Lengthening procedures
Bianchi
STEP
SILT
Duodenal lengthening
Reversed anti-peristaltic segmental bowel loop
Intestinal transplantation

SBS: Short bowel syndrome; STEP: Serial transverse enteroplasty procedure; SILT: Spiral intestinal lengthening and tailoring.

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REFERENCES

1 **Siebert JR**. Small-intestine length in infants and children. *Am J Dis Child* 1980; **134**: 593-595 [PMID: 7386434]
2 **Gondolesi G**, Ramisch D, Padin J, Almau H, Sandi M, Schelotto PB, Fernandez A, Rumbo C, Solar H. What is the normal small bowel length in humans? first donor-based cohort analysis. *Am J Transplant* 2012; **12 Suppl 4**: S49-S54 [PMID: 22702412 DOI: 10.1111/j.1600-6143.2012.04148.x]
3 **Pironi L**. Definitions of intestinal failure and the short bowel syndrome. *Best Pract Res Clin Gastroenterol* 2016; **30**: 173-185 [PMID: 27086884 DOI: 10.1016/j.bpg.2016.02.011]
4 **Fung JJ**. William Hunter Harridge lecture: The changing face of short-gut syndrome management. *Am J Surg* 2017; **213**: 448-451 [PMID: 28159115 DOI: 10.1016/j.amjsurg.2017.01.018]
5 **Wilmore DW**, Dudrick SJ. Growth and development of an infant receiving all nutrients exclusively by vein. *JAMA* 1968; **203**: 860-864 [PMID: 4965871]
6 **Pironi L**, Goulet O, Buchman A, Messing B, Gabe S, Candusso M,

Bond G, Gupte G, Pertkiewicz M, Steiger E, Forbes A, Van Gossum A, Pinna AD; Home Artificial Nutrition and Chronic Intestinal Failure Working Group of ESPEN. Outcome on home parenteral nutrition for benign intestinal failure: a review of the literature and benchmarking with the European prospective survey of ESPEN. *Clin Nutr* 2012; **31**: 831-845 [PMID: 22658443 DOI: 10.1016/j.clnu.2012.05.004]

7 **Scolapio JS**, Fleming CR, Kelly DG, Wick DM, Zinsmeister AR. Survival of home parenteral nutrition-treated patients: 20 years of experience at the Mayo Clinic. *Mayo Clin Proc* 1999; **74**: 217-222 [PMID: 10089988 DOI: 10.4065/74.3.217]

8 **Messing B**, Lémann M, Landais P, Gouttebel MC, Gérard-Boncompain M, Saudin F, Vangossum A, Beau P, Guédon C, Barnoud D. Prognosis of patients with nonmalignant chronic intestinal failure receiving long-term home parenteral nutrition. *Gastroenterology* 1995; **108**: 1005-1010 [PMID: 7698566]

9 **Colomb V**, Dabbas-Tyan M, Taupin P, Talbotec C, Révillon Y, Jan D, De Potter S, Gorski-Colin AM, Lamor M, Herreman K, Corriol O, Landais P, Ricour C, Goulet O. Long-term outcome of children receiving home parenteral nutrition: a 20-year single-center experience in 302 patients. *J Pediatr Gastroenterol Nutr* 2007; **44**: 347-353 [PMID: 17325556 DOI: 10.1097/MPG.0b013e31802c6971]

10 **Pironi L**, Joly F, Forbes A, Colomb V, Lyszkowska M, Baxter J, Gabe S, Hébuterne X, Gambarara M, Gottrand F, Cuerda C, Thul P, Messing B, Goulet O, Staun M, Van Gossum A; Home Artificial Nutrition & Chronic Intestinal Failure Working Group of the European Society for Clinical Nutrition and Metabolism (ESPEN). Long-term follow-up of patients on home parenteral nutrition in Europe: implications for intestinal transplantation. *Gut* 2011; **60**: 17-25 [PMID: 21068130 DOI: 10.1136/gut.2010.223255]

11 **Amiot A**, Messing B, Corcos O, Panis Y, Joly F. Determinants of home parenteral nutrition dependence and survival of 268 patients with non-malignant short bowel syndrome. *Clin Nutr* 2013; **32**: 368-374 [PMID: 22992308 DOI: 10.1016/j.clnu.2012.08.007]

12 **Messing B**, Crenn P, Beau P, Boutron-Ruault MC, Rambaud JC, Matuchansky C. Long-term survival and parenteral nutrition dependence in adult patients with the short bowel syndrome. *Gastroenterology* 1999; **117**: 1043-1050 [PMID: 10535866]

13 **Capriati T**, Giorgio D, Fusaro F, Candusso M, Schingo P, Caldaro T, Laureti F, Elia D, Diamanti A. Pediatric Short Bowel Syndrome: Predicting Four-Year Outcome after Massive Neonatal Resection. *Eur J Pediatr Surg* 2018; **28**: 455-463 [PMID: 28719916 DOI: 10.1055/s-0037-1604113]

14 **Norsa L**, Artru S, Lambe C, Talbotec C, Pigneur B, Ruemmele F, Colomb V, Capito C, Chardot C, Lacaille F, Goulet O. Long term outcomes of intestinal rehabilitation in children with neonatal very short bowel syndrome: Parenteral nutrition or intestinal transplantation. *Clin Nutr* 2018; : [PMID: 29478887 DOI: 10.1016/j.clnu.2018.02.004]

15 **Tannuri U**, Barros F, Tannuri AC. Treatment of short bowel syndrome in children. Value of the Intestinal Rehabilitation Program. *Rev Assoc Med Bras (1992)* 2016; **62**: 575-583 [PMID: 27849236 DOI: 10.1590/1806-9282.62.06.575]

16 **Batra A**, Keys SC, Johnson MJ, Wheeler RA, Beattie RM. Epidemiology, management and outcome of ultrashort bowel syndrome in infancy. *Arch Dis Child Fetal Neonatal Ed* 2017; **102**: F551-F556 [PMID: 28866623 DOI: 10.1136/archdischild-2016-311765]

17 **Dore M**, Junco PT, Moreno AA, Cerezo VN, Muñoz MR, Galán AS, Sánchez AV, Prieto G, Ramos E, Hernandez F, Martínez LM, Santamaria ML. Ultrashort Bowel Syndrome Outcome in Children Treated in a Multidisciplinary Intestinal Rehabilitation Unit. *Eur J Pediatr Surg* 2017; **27**: 116-120 [PMID: 28052307 DOI: 10.1055/s-0036-1597812]

18 **Bueno J**, Redecillas S, GarcÍa L, Lara A, Giné C, Molino JA, Broto J, Segarra O. Duodenal lengthening in short bowel with dilated duodenum. *J Pediatr Surg* 2015; **50**: 493-496 [PMID: 25746715 DOI: 10.1016/j.jpedsurg.2014.11.047]

19 **Jain V**, Huerta S. More on 'No Gut Syndrome': A case report. *Int J Surg Case Rep* 2016; **19**: 35-37 [PMID: 26708947 DOI: 10.1016/j.ijscr.2015.12.014]

20 **Bueno J**, Burgos R, Redecillas S, López M, Balsells J. Duodenal lengthening in an adult with ultra-short bowel syndrome. A case report. *Rev Esp Enferm Dig* 2018; **110**: 59-62 [PMID: 29106286 DOI: 10.17235/reed.2017.5187/2017]

21 **Lauro A**, Cirocchi R, Cautero N, Dazzi A, Pironi D, Di Matteo FM, Santoro A, Pironi L, Pinna AD. Reconnection surgery in adult post-operative short bowel syndrome < 100 cm: is colonic continuity sufficient to achieve enteral autonomy without autologous gastrointestinal reconstruction? Report from a single center and systematic review of literature. *G Chir* 2017; **38**: 163-175 [PMID: 29182898]

22 **Cosnes J**, Gendre JP, Le Quintrec Y. Role of the ileocecal valve and site of intestinal resection in malabsorption after extensive small bowel resection. *Digestion* 1978; **18**: 329-336 [PMID: 750260 DOI: 10.1159/000198220]

23 **Nordgaard I**. What's new in the role of colon as a digestive organ in patients with short bowel syndrome. *Nutrition* 1998; **14**: 468-469 [PMID: 9614315]

24 **Jeppesen PB**, Mortensen PB. Colonic digestion and absorption of energy from carbohydrates and medium-chain fat in small bowel failure. *JPEN J Parenter Enteral Nutr* 1999; **23**: S101-S105 [PMID: 10483907 DOI: 10.1177/014860719902300525]

25 **Smith KH**, Saunders JA, Nugent KP, Jackson AA, Stroud MA. Reduced parenteral nutrition requirements following anastomosis of a short residual colonic segment to a short jejunum. *JPEN J Parenter Enteral Nutr* 2011; **35**: 732-735 [PMID: 22042049 DOI: 10.1177/0148607111406504]

26 **Nguyen BT**, Blatchford GJ, Thompson JS, Bragg LE. Should intestinal continuity be restored after massive intestinal resection? *Am J Surg* 1989; **158**: 577-9; discussion 579-80 [PMID: 2511774]

27 **Thompson JS**. Reoperation in patients with the short bowel syndrome. *Am J Surg* 1992; **164**: 453-6; discussion 456-7 [PMID: 1443368]

28 **Kong W**, Wang J, Ying R, Li Y, Jin H, Mao Q, Yao D, Guo M. A potential anatomic subtype of short bowel syndrome: a matched case-control study. *BMC Gastroenterol* 2016; **16**: 12 [PMID: 26822147 DOI: 10.1186/s12876-016-0425-4]

29 **Ives GC**, Demehri FR, Sanchez R, Barrett M, Gadepalli S, Teitelbaum DH. Small Bowel Diameter in Short Bowel Syndrome as a Predictive Factor for Achieving Enteral Autonomy. *J Pediatr* 2016; **178**: 275-277.e1 [PMID: 27587075 DOI: 10.1016/j.jpeds.2016.08.007]

30 **Bianchi A**. Longitudinal intestinal lengthening and tailoring: results in 20 children. *J R Soc Med* 1997; **90**: 429-432 [PMID: 9306995]

31 **Kim HB**, Fauza D, Garza J, Oh JT, Nurko S, Jaksic T. Serial transverse enteroplasty (STEP): a novel bowel lengthening procedure. *J Pediatr Surg* 2003; **38**: 425-429 [PMID: 12632361 DOI: 10.1053/jpsu.2003.50073]

32 **Fan S**, Li Y, Zhang S, Wang J, Li J. Success of serial transverse enteroplasty in an adult with extreme short bowel syndrome: a case report. *Int Surg* 2015; **100**: 626-631 [PMID: 25875543 DOI: 10.9738/INTSURG-D-14-00177.1]

33 **Yaprak M**, Erdoğan O, Oğus M. The STEP procedure in an adult patient with short bowel syndrome: a case report. *Turk J Gastroenterol* 2011; **22**: 333-336 [PMID: 21805426]

34 **Bellolio R F**, Klaassen L J, Pulgar B D, Molina P ME, Pinedo M G, Zúñiga D A. Serial transverse enteroplasty for short bowel syndrome. Case report. *Rev Med Chil* 2010; **138**: 478-482 [PMID: 20668797]

35 **Yannam GR**, Sudan DL, Grant W, Botha J, Langnas A, Thompson JS. Intestinal lengthening in adult patients with short bowel syndrome. *J Gastrointest Surg* 2010; **14**: 1931-1936 [PMID: 20734155 DOI: 10.1007/s11605-010-1291-y]

36 **Jones BA**, Hull MA, Potanos KM, Zurakowski D, Fitzgibbons SC, Ching YA, Duggan C, Jaksic T, Kim HB; International STEP Data Registry. Report of 111 consecutive patients enrolled in the International Serial Transverse Enteroplasty (STEP) Data Registry: a retrospective observational study. *J Am Coll Surg* 2013; **216**: 438-446 [PMID: 23357726 DOI: 10.1016/j.jamcollsurg.2012.12.018]

37 **Oh PS**, Fingeret AL, Shah MY, Ventura KA, Brodlie S, Ovchinsky

N, Martinez M, Lobritto SJ, Cowles RA. Improved tolerance for enteral nutrition after serial transverse enteroplasty (STEP) in infants and children with short bowel syndrome--a seven-year single-center experience. *J Pediatr Surg* 2014; **49**: 1589-1592 [PMID: 25475799 DOI: 10.1016/j.jpedsurg.2014.07.019]

38 **Mercer DF**, Hobson BD, Gerhardt BK, Grant WJ, Vargas LM, Langnas AN, Quiros-Tejeira RE. Serial transverse enteroplasty allows children with short bowel to wean from parenteral nutrition. *J Pediatr* 2014; **164**: 93-98 [PMID: 24094877 DOI: 10.1016/j.jpeds.2013.08.039]

39 **Frongia G**, Kessler M, Weih S, Nickkholgh A, Mehrabi A, Holland-Cunz S. Comparison of LILT and STEP procedures in children with short bowel syndrome -- a systematic review of the literature. *J Pediatr Surg* 2013; **48**: 1794-1805 [PMID: 23932625 DOI: 10.1016/j.jpedsurg.2013.05.018]

40 **Cserni T**, Biszku B, Guthy I, Dicso F, Szaloki L, Folaranmi S, Murphy F, Rakoczy G, Bianchi A, Morabito A. The first clinical application of the spiral intestinal lengthening and tailoring (silt) in extreme short bowel syndrome. *J Gastrointest Surg* 2014; **18**: 1852-1857 [PMID: 24957255 DOI: 10.1007/s11605-014-2577-2]

41 **Alberti D**, Boroni G, Giannotti G, Parolini F, Armellini A, Morabito A, Bianchi A. "Spiral intestinal lenghtening and tailoring (SILT)" for a child with severely short bowel. *Pediatr Surg Int* 2014; **30**: 1169-1172 [PMID: 25119303 DOI: 10.1007/s00383-014-3583-x]

42 **Layec S**, Beyer L, Corcos O, Alves A, Dray X, Amiot A, Stefanescu C, Coffin B, Bretagnol F, Bouhnik Y, Messing B, Panis Y, Kapel N, Joly F. Increased intestinal absorption by segmental reversal of the small bowel in adult patients with short-bowel syndrome: a case-control study. *Am J Clin Nutr* 2013; **97**: 100-108 [PMID: 23151533 DOI: 10.3945/ajcn.112.042606]

43 **Thompson JS**. Reversed Intestinal Segment Revisited. *Transplant Proc* 2016; **48**: 453-456 [PMID: 27109977 DOI: 10.1016/j.transproceed.2015.09.072]

44 **Grant D**, Abu-Elmagd K, Mazariegos G, Vianna R, Langnas A, Mangus R, Farmer DG, Lacaille F, Iyer K, Fishbein T; Intestinal Transplant Association. Intestinal transplant registry report: global activity and trends. *Am J Transplant* 2015; **15**: 210-219 [PMID: 25438622 DOI: 10.1111/ajt.12979]

45 **Garcia Aroz S**, Tzvetanov I, Hetterman EA, Jeon H, Oberholzer J, Testa G, John E, Benedetti E. Long-term outcomes of living-related small intestinal transplantation in children: A single-center experience. *Pediatr Transplant* 2017; **21**: [PMID: 28295952 DOI: 10.1111/petr.12910]

46 **Celik N**, Mazariegos GV, Soltys K, Rudolph JA, Shi Y, Bond GJ, Sindhi R, Ganoza A. Pediatric intestinal transplantation. *Gastroenterol Clin North Am* 2018; **47**: 355-368 [PMID: 29735029 DOI: 10.1016/j.gtc.2018.01.007]

47 **Matsumoto CS**, Subramanian S, Fishbein TM. Adult intestinal transplantation. *Gastroenterol Clin North Am* 2018; **47**: 341-354 [PMID: 29735028 DOI: 10.1016/j.gtc.2018.01.011]

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REVIEW

Complement-mediated renal diseases after kidney transplantation - current diagnostic and therapeutic options in *de novo* and recurrent diseases

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Abstract

For decades, kidney diseases related to inappropriate complement activity, such as atypical hemolytic uremic syndrome and C3 glomerulopathy (a subtype of membranoproliferative glomerulonephritis), have mostly been complicated by worsened prognoses and rapid progression to end-stage renal failure. Alternative complement pathway dysregulation, whether congenital or acquired, is well-recognized as the main driver of the disease process in these patients. The list of triggers include: surgery, infection, immunologic factors, pregnancy and medications. The advent of complement activation blockade, however, revolutionized the clinical course and outcome of these diseases, rendering transplantation a viable option for patients who were previously considered as non-transplantable cases.

Several less-costly therapeutic lines and likely better efficacy and safety profiles are currently underway. In view of the challenging nature of diagnosing these diseases and the long-term cost implications, a multidisciplinary approach including the nephrologist, renal pathologist and the genetic laboratory is required to help improve overall care of these patients and draw the optimum therapeutic plan.

Key words: Complement-related diseases; Kidney transplantation; *De novo*; Recurrent diseases

Core tip: The recent progress in our understanding of the pathophysiology of complement-mediated diseases is gaining considerable popularity. Complement dysregulation due to inherited or acquired factors is currently the culprit mechanism. Several constitutional abnormalities usually trigger the process of recurrence, with a subsequent high rate of graft loss. The development of the terminal complement inhibitor "eculizumab" is a breakthrough in controlling abnormal complement activation. While diagnosing complement abnormalities is one challenge, treatment cost with this new agent is another major hurdle in any health care system. New lines of promising therapies are currently in the pipeline.

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INTRODUCTION

The complement components can be seen in biopsies of almost all types of glomerulonephritis, which can be broadly divided into two main groups: (1) "complement over-activation" includes IgA nephropathy (IgAN) and immune complex membranoproliferative glomerulonephritis (MPGN); and (2) "complement dysregulation" that encompasses atypical hemolytic uremic syndrome (aHUS) and C3 glomerulopathy (C3G)[1]. While complement activation is triggered by immune complex formation in the former group, genetic mutations are the driver of complement over-activation in the latter one. This explains why the disease process in the former class is potentially modifiable by immunosuppression in the post-transplantation period, which is not the case in the latter class. Our understanding of the biogenetic causes of C3G and aHUS/thrombotic microangiopathy (TMA) has been expanding. The mechanisms of these diseases not only affect their clinical history, but also affect the recurrence rate[2]. The role of complement in C3G evolution is now well-recognized[3]. Recent progress in understanding the pathophysiology of MPGN led to newer classifications of MPGN into immune complex-mediated and complement-mediated subtypes. The hallmark of complement-mediated MPGN is the deposition of C3 and other complement products in glomerular tissues[4]. This is caused by dysregulation and loss of control of the AP complement pathway[5]. The AP is tightly regulated under physiological conditions. It can be disrupted through either inherited (mutations/polymorphisms) or acquired (autoantibodies) interferences to the regulating components. Histological staining using immunofluorescence (IF) is currently the best determinant technique, and C3G is defined by dominant C3 with dispersed, reduced or absent immunoglobulin (Ig). Based on electron microscopy (EM) examination, C3G subdivides into complement three glomerulonephritis (C3GN) and dense deposit disease (DDD). In C3GN, discrete deposits can be seen in the mesangium and capillary walls (subendothelial and subepithelial regions). On the contrary, DDD deposits are large in size, extremely dense (osmiophilic) and intramembranous, which leads to a characteristic thickening of the glomerular basement membrane (GBM)[5]. The term aHUS is applied to a heterogenous group of diseases (Figure 1) that share TMA manifestations with an associated decline in renal function (classically, no IF staining of C3 or any other complement components). In aHUS, complement abnormalities (either genetic mutations or acquired autoantibodies) are well-recognized mechanisms with a clearly associated complement-mediated TMA[1]. In this article, we will discuss various types of complement-mediated renal diseases after kidney transplantation and their current therapeutic options.

Methodology

In view of the lack of prospective controlled trials concerned with complement-mediated diseases post-kidney transplant, we tried to shed the light in this review on the most recent expert opinions, with regard to the best tools of management for these devastating diseases.

CLINICAL PRESENTATION

Salient features of C3G

DDD and C3GN share some salient features that include proteinuria, hematuria and increased serum creatinine concentration[6,7]. Recurrence of C3G is typically encountered one to two years after transplant[7]. C3G comprises a spectrum of diseases that result from aberrant control of complement activation, deposition and dysregulation, leading to C3 glomerular deposition with characteristic electron-dense deposits (EDD) in EM (Table 1).

Table 1 Morphological features of C3 glomerulopathy

	Morphological features of C3G
Light microscopy	Active lesions
	Mesangial expansion with or without hypercellularity
	Endocapillary hypercellularity including monocytes and/or neutrophils
	Capillary wall thickening with double contours (combination of capillary wall thickening + mesangial increase is referred to as a membranoproliferative pattern)
	Fibrinoid necrosis
	Cellular/fibrocellular crescents
	Chronic lesions
	Segmental or global glomerulosclerosis
	Fibrous crescents
IF microscopy	Typically dominant C3 staining
Electron microscopy	DDD: Dense osmiophilic mesangial and intramembranous electron dense deposits.
	C3GN: Amorphous mesangial with or without capillary wall deposits including subendothelial, intramembranous and subepithelial EDD
	Subepithelial "humps" may be seen in both DDD and C3GN

Adapted from Goodship *et al*[12]. C3G: C3 glomerulopathy; DDD: Dense deposit disease; C3GN: C3 glomerulonephritis; EDD: Electron dense deposits, fibrinoid necrosis.

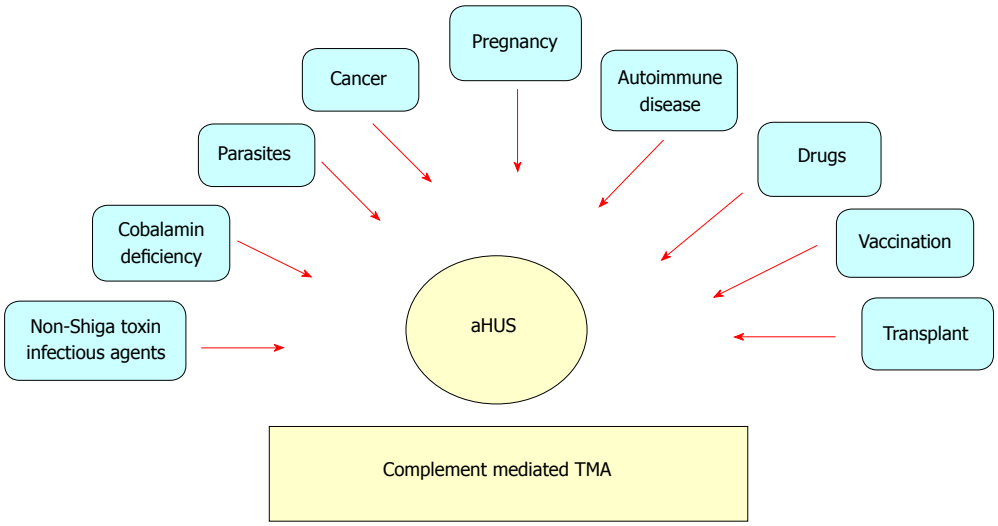

Figure 1 Heterogeneity of atypical hemolytic uremic syndrome. Adapted from Salvadori *et al*[1]. TMA: Thrombotic microangiopathy; aHUS: Atypical hemolytic uremic syndrome.

Pathology

Renal biopsy is crucial for C3G diagnosis. LM is not helpful, due to its extremely diverse appearance. IF is the mainstay for diagnosis. A unique criterion in IF studies is the presence of dominant C3 staining, which is twice as intense as any other immunoreactant (IgG, IgM, IgA, and C1q)[8]. Ninety percent of DDD patients, but fewer C3GN patients, can be diagnosed through applying this criterion[8]. Repeated biopsy may be required to confirm the diagnosis. As C3G may present in acute infection, C3 can be observed with post-infectious GN. Humps are no longer pathognomonic criteria of post-infectious GN, however they can also be encountered in C3G. However, the presence of double contours in the GBM raises the possibility of C3G diagnosis. To differentiate DDD from C3GN, EM studies should be accomplished, as it has pivotal clinical implications. Moreover, staining for IgG as well as light chains on pronase-digested paraffin should be applied for all cases of C3GN on standard IF, particularly in adults (Figure 2 and Table 1)[9,10].

Salient features of TMA

TMA is mostly presented 3-6 mo post-transplant, but it can occur at any time after renal transplantation[13]. Presentation of TMA is not universal, ranging from the renal-limited form up to a complete systemic picture with its classic triad of thrombocytopenia, microangiopathic hemolytic anemia (MAHA) and decline in renal function[14]. MAHA is defined as increased LDH, decline in HB and haptoglobin, and appearance of schistocytes in peripheral blood smears. On the other hand, localized (renal-limited) TMA usually presents later in the post-transplant course. In the acute stage, evidence of endothelial injury with platelet aggregation (thrombosis), fibrinoid necrosis, as well as

Table 2 Morphological features in microangiopathy

Active lesions	Chronic lesions
Glomeruli: Thrombi - Endothelial swelling or denudation - Fragmented RBCs - Subendothelial flocculent material. EM: Mesangiolysis - Microaneurysms	Glomeruli: LM: Double contours of peripheral capillary walls, with variable mesangial interposition - EM: New subendothelial basement membrane - Widening of the subendothelial zone
Arterioles: Thrombi - Endothelial swelling or denudation - Intramural fibrin - Fragmented red blood cells - Intimal swelling - Myocyte necrosis	Arterioles: Hyaline deposits
Arteries: Thrombi - Myxoid intimal swelling - Intramural fibrin - Fragmented red blood cells	Arteries: Fibrous intimal thickening with concentric lamination (onion skin)

Adapted from Goodship *et al*[12]. EM: Electron microscopy; LM: Light microscopy.

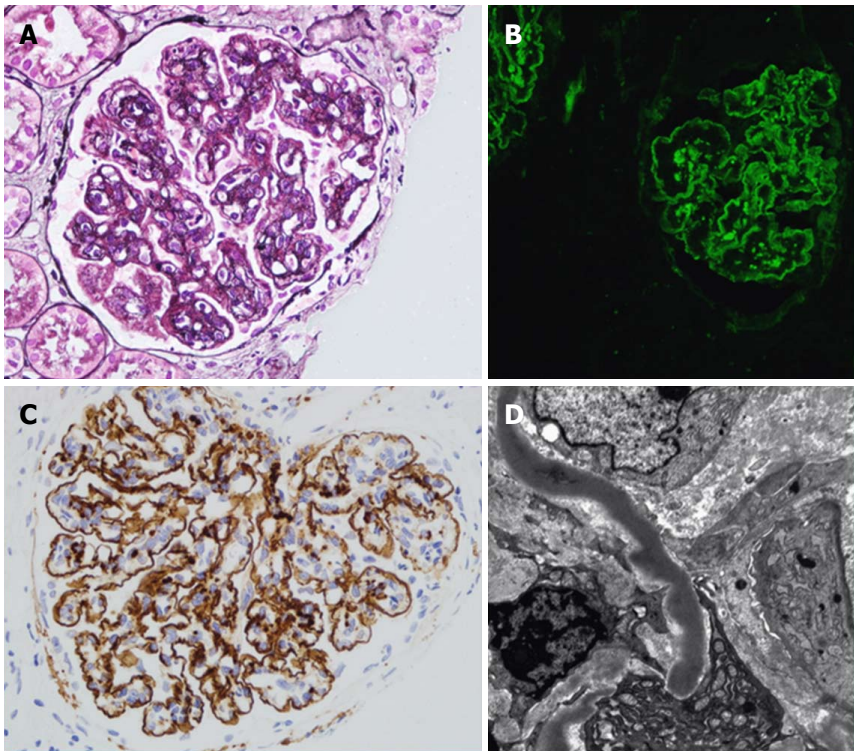

Figure 2 Renal histology in individuals with dense deposit disease. A: Light microscopy with silver stain showing a membranoproliferative glomerulonephritis pattern with double contours of the glomerular basement membrane; B: Immunofluorescence; C: Immunohistochemistry with immunoperoxidase showing strong capillary wall staining of C3 and some granular mesangial C3; D: Characteristic sausage-like, intramembranous, osmiophilic deposits on electron microscopy. Adapted from Barbour *et al*[11].

glomerular ischemia can be seen. On the other hand, chronic lesions show duplication and multilayering of the GBM, with clustering of the matrix layers and vessel wall cells leading to the characteristic onion skin shape appearance (Table 2)[15]. As TMA is not always present with full-blown systemic pictures, genetic studies to unmask the underlying complement defect are ultimately mandated, particularly if no other clear cause has been associated (*e.g.*, AMR-associated TMA). AMR can give a TMA-like picture, as it is an antibody interaction with the endothelium. This is also a fundamental maneuver to differentiate *de novo* from recurrent disease (positive genetic testing), with consequent clinical therapeutic implications[16].

Extrarenal manifestations of aHUS and C3G

Twenty percent of aHUS patients express extrarenal manifestations. Their relation to complement activation and TMA evolution is unclear. Drusen is rarely seen in TMA[17]. Drusen formation, which represents an accumulation of lipids and complement-rich proteins between Bruch's membrane and the retinal pigmentary epithelium, is commonly reported in age-related macular degeneration but present at a much earlier age with C3G[18]. In C3G, retinal drusen and acquired partial lipodystrophy have been commonly reported. The latter is most commonly encountered with C3 nephritic factors. Factor D, an essential agent for C3 convertase formation, is highly concentrated in adipocytes that undergo C3 nephritic factor-induced complement-dependent lysis[19] (Table 3).

Pathogenesis and classification of C3G

The new classification of MPGN encompasses two

Table 3 Extrarenal manifestations reported in atypical hemolytic uremic syndrome, dense deposit disease, and C3 glomerulonephritis

aHUS	DDD/C3GN
Digital gangrene, skin	Retinal drusen
Cerebral artery thrombosis/stenosis	Acquired partial lipodystrophy
Extracerebral artery stenosis	
Cardiac involvement/myocardial infarction	
Ocular involvement	
Neurologic involvement	
Pancreatic, gastrointestinal involvement	
Pulmonary involvement	
Intestinal involvement	

Adapted from Goodship *et al*[12]. aHUS: Atypical hemolytic uremic syndrome; C3GN: C3 glomerulonephritis; DDD: Dense deposit disease.

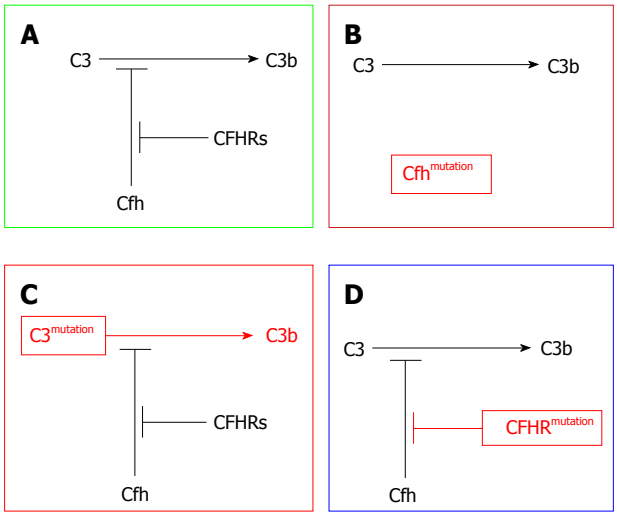

Figure 3 Disease mechanisms in C3 glomerulopathy, based on genetic defects identified in family studies. A: Physiological regulation of C3 activation to C3b *via* the alternative pathway is mediated by complement factor H (CFH) (Cfh). Competitive inhibition of CFH by CFHR proteins is termed CFH deregulation; B: Homozygous deficiency or dysfunction of CFH results in excessive C3 activation; C: Hyper-functional C3 produces excessive C3 activation despite normal CFH activity; D: Abnormal CFHR proteins enhance CFH deregulation, leading to excessive C3 activation. Adapted from Barbour *et al*[11].

subtypes: the immune complex-mediated GN (ICGN) and complement-mediated GN (CGN), recently named (C3G). The former is characterized by both Ig as well as complement component deposition in kidney tissues as recognized by IF studies. The latter is characterized by dominant complement deposition with smaller amounts of Ig deposition. Further subdivision of C3G into C3GN and DDD can be attained through EM studies[20]. Both subtypes are triggered through dysregulation of any part of the AP. For example, patients may develop the C3 convertase-stabilizing factor called C3NeF, which leads to uncontrolled complement activation. Loss-of-function mutations in complement regulatory proteins (CFH or CFI)[20-23] or gain-of-function mutations in C3 leads to CFH resistance, which has been postulated as an underlying mechanism (Figure 3).

Pathophysiology and recurrence of C3G

Pathophysiology of AP activation in DDD and C3GN is nearly the same. In both disorders, disturbance of the fluid phase is triggered as a result of aberrant gene mutations or the presence of autoantibodies. However, the presence of C3 nephritic factor (C3NeF) is by far the most commonly acquired complement defect. C3NeF has the ability to block CFH-mediated decay by stabilizing C3 convertase[5,24]. By binding to C3 convertase, C3NeF has the ability to trigger it approximately ten times[25,26]. C3 convertase can also block the action of CFH, CR1, as well as decay-accelerating factor (DAF).

C3NeF is prevalent in 50%-80% C3G patients[27]. Other autoantibodies have also been found (*e.g.*, autoantibodies against factor B[28], CFH[29,30] and C3 convertase)[28]. In C3G, CFH mutations have been frequently reported. Different forms of mutations can be presented as defective or completely absent protein H. These mutations can be seen in homozygous or heterozygous forms[31,32]. C3NeF can also be encountered, which denotes the clustering of different risk factor varieties. More recently, genetic mutations involving the CFHR gene have been reported in the C3G cohort of patients[33]. CFHR group genetic mutations[34], deletions[35], duplications[36], as well as hybrid genes[37] have also been observed in C3G patients, either in an isolated manner or in a familial cohort. Malik and his associates[38] reported that members of one family can develop C3G as an result of aberrant copies of CFHR3 and CFHR1 loci. The presence of familial C3G underscores the genetic basis of several C3G varieties and their relation to AP dysregulation.

To summarize, complement dysregulation is the specific etiology of C3G, which could be genetic or acquired. While genetic causes encompasse complement gene mutations, acquired causes include the C3NeFs, which have the ability to impede normal complement regulation[1]. Moreover, genetic varieties constitute the pathophysiologic basis of C3G and aHUS evolution (Table 4). Recently, a robust correlation between CFH-related proteins and a variety of complement-mediated diseases have been documented. Functional parameters (*e.g.*, complement regulators and CFH competitors) have recently attained significant popularity[39].

TMA or C3G?

Both TMA and C3G have a common underlying

Table 4 Overview of mutations in complement factor H-related protein genes

Genetic defect	Phenotypical expression
Duplication in *CFHR5* gene	C3 glomerulopathy (CFHR5 nephropathy)
Duplication in *CFHR1* gene	C3 glomerulopathy
Hybrid *CFHR3/CFHR1*	C3 glomerulopathy
Hybrid *CFHR2/CFHR5*	C3 glomerulopathy
Hybrid *CFH/CFHR1*	aHUS
Hybrid *CFH/CFHR3*	aHUS

Adapted from Salvadori *et al*[1]. aHUS: Atypical hemolytic uremic syndrome; CFH: Complement factor H.

causation: AP dysregulation. However, the question that arises is "which factors influence the evolution of one disease rather than the other?"[40]. The prevalence of the fluid phase complement activation dysregulation in animal models suggests that C3G is the responsible factor. On the other hand, complement activation involving capillary walls can result in TMA evolution[41]. Furthermore, absolute CFH deficiency is in favor of an activation of the fluid phase complement with subsequent C3G evolution, while the lack of an aberrant CFH binding region is in favor of TMA evolution[41]. It has also been postulated that CFH and CFH/CFHR mutations induce aHUS to inhibit CFH-binding to many cell surfaces, while C3G-associated mutations in CFHRs cannot inhibit CFH binding to endothelial cell surfaces[42]. The prevalence of familial C3G mutations serves as a robust indicator of the genetic base of C3G recurrence[1].

Risk of DDD recurrence

Despite the well-known DDD variants of C3, its pathogenesis has only recently been recognized. The five-year graft survival rate was only 50% in one retrospective study of 75 children[6]. In adults, a majority of the recipients developed recurrence in post-transplant periods, with 25% of them losing their allografts[43]. In another broader cohort that included eighty adults and children with C3G, Medjeral-Thomas *et al*[44], reported histological recurrence in all six DDD recipients. Graft loss had resulted in 50% of his cases. For recipients who developed DDD recurrence, the ten-year graft survival rate has been reported to be up to 57.5% in an UNOS review[45]. Risk factors for DDD recurrent disease and graft loss are not well-recognized. However, the histological recurrence rate was reported to be more than 70%[46,47]. Recurrence may present spontaneously in post-transplant periods, though it may take several years to manifest[47]. This discrepancy raises some questions, such as the impact of the longevity of follow-ups, the need for tissue diagnosis, and the real rate of DDD recurrence.

Risk of C3GN recurrence

There is no documented relation between mode of presentation, C3 serum levels, or C3NeF levels and C3GN recurrence[48]. The only trustworthy risk factor correlated with C3 recurrence is the presence of heavy proteinuria, with two thirds of C3 patients showing vulnerability to recurrence and a high incidence of graft loss[5,7,27]. All the available data about recurrence are based on case series, with the largest by Zand *et al*[7] that failed to reveal robust evidence of recurrence risk. This observation is partially explained by the heterogeneity of complement defects implicated in C3GN evolution. Early reports postulated HLA-B8 DR3 and living related donation as possible risk factors for recurrence[49]. However, the more recent reports suggested the following: (1) history of graft loss owing to recurrence[50]; (2) aggressive histopathological alterations in native kidney biopsy; and (3) hybrid CFHR3 1 gene-related C3GN. Wong *et al*[51] have recently reported a high rate of C3G recurrence (five patients received a total of eight kidney transplants). Four (50%) renal allografts had disease recurrence, of which three had biopsy-proven recurrence, with time to recurrence ranging from as early as 2 wk following living-related donor transplantation, to 93 and 101 mo for the two remaining allografts, respectively[51].

Diagnosis of C3G recurrence

The declining appearance of proteinuria, hematuria or eGFR is a strong indicator of C3G recurrence. Final diagnosis is usually made through LM, IF, and EM studies of kidney biopsy. After histopathological examination, a thorough evaluation of any genetic mutation in the AP should be accomplished, especially if these studies were not previously fulfilled with the native kidney disease.

Diagnosis of C3G/TMA recurrence

A robust work-up of analytic studies including genetic, biochemical and pathological evaluation should be instituted, including the following: (1) complement components and complement regulatory protein levels; (2) peripheral WBC MCP levels; (3) screening for antibodies to CFH and C3NeFs; and (4) mutation screening of CFH, CFI, CFB, C3, and MCP. Furthermore, recombination in the CFHR region should be tested[52].

Prognosis of DDD/C3GN

In both DDD and C3GN, recurrent disease is usually associated with allograft loss[6,44,53]. The one-year allograft survival was reported to be 94%, with 69% at five years, and 28% at ten years. Three predictive criteria for progression to ESRD were recognized: (1) crescentic GN; (2) severe arteriolar sclerosis by LM; and (3) decline of renal function at the time of first biopsy[44].

Prognosis of TMA

Compared to recurrent TMA, the prognosis of *de novo* TMA is quite poor. Fifty percent of patients may lose their graft within a couple of years after diagnosis[54,55]. Many reports were in favor of this attitude[54-56]. Before

Table 5 Recommended therapy approach for C3 glomerulopathy based on small prospective trial, case reports, and expert opinion

All patients	Moderate disease	Severe disease
Lipid control	Urine protein > 500 mg/24 h despite supportive therapy, or	Urine protein > 2000 mg/24 h despite immunosuppression and supportive therapy or
Optimal BP control (< 90% in children and ≤ 120/80 mm Hg in adults)	Moderate inflammation on renal biopsy or	Severe inflammation represented by marked endo- or extracapillary proliferation with/without crescent formation despite immunosuppression and supportive therapy or
Optimal nutrition for both normal growth in children and healthy weight in adults	Recent increase in serum creatinine suggesting risk for progressive disease	Increased S. Cr suggesting risk for progressive disease at onset despite immunosuppression and supportive therapy
	Recommendation	Recommendations
	Prednisone	Methylprednisolone pulse-dosing as well as other anti-cellular immune suppressants have had limited success in rapidly progressive disease
	Mycophenolate mofetil	Data are insufficient to recommend eculizumab as a first-line agent for the treatment of rapidly progressive disease

Adapted from Goodship *et al*[12].

the era of eculizumab (EZ), Schwimmer *et al*[54] reported that 54% of systemic TMA can develop dialysis requiring AKI, and about 38% lost their allograft. However, no one patient with localized TMA has complicated with TMA-related allograft loss or a need for dialysis. Nevertheless, both systemic and localized forms may experience unfavorable long-term graft survival[54,57].

THERAPY OF COMPLEMENT DYSREGULATION-RELATED DISEASES

Treatment of de novo C3G

The therapeutic approach for *de novo* C3G therapy is similar to that of recurrent disease. Very minimal information is available regarding *de novo* C3G[58].

Treatment of recurrent C3G

In light of the paucity of data from controlled studies, some experts have suggested an approach that depends on disease severity (*i.e.,* mild, moderate and severe) based on the degree of proteinuria and the magnitude of allograft dysfunction (Table 5): (1) conservative measures, as with other glomerulotides, including RAS blockade and lipid-lowering agents; (2) glucocorticoids, MMF, rituximab and PE have been used with variable success[59,60]. In selected patients, MMF has been reported to be effective in C3GN controls in a retrospective study[12,61]; and (3) EZ was firstly reported by Bomback *et al*[62], in treating six patients with C3G (three with DDD and three with C3GN) in an open-labelled trial. EZ dose is guided by previous experience in aHUS and used for one year. Improved kidney function was observed in two patients; one patient showed partially improved proteinuria, while another patient showed better histological and laboratory findings[62]. Notably, elevated serum membrane attack complex (MAC) levels were associated with clinical improvement[63]. Duration of therapy is not yet defined. The beneficial effects of EZ in DDD recurrence[46] and C3GN recurrence[64] have been shown in case reports[65]. However, histopathological evidence of disease progression has been observed in subsequent biopsies. This highlights the fact that there is no standard accepted biomarker for disease monitoring, which can be used to assess the patient's response to treatment and predict better renal function.

In 2018, Garg *et al*[66] described the spectrum of C3 pathophysiology and its clinical implications. The observed variability of the degrees of upstream (site of C3 convertase) and downstream (site of C5 convertase) complement dysregulation may result in variable phenotypic differences[67,68]. Consequently, the nature of this spectrum will be reflected clinically on disease progress in two ways: firstly, the variability in response to EZ therapy (Figure 4)[66]. In C3G, if the dominant process focused on activation of C5 convertase (resulting in increased soluble C5b-9 levels), EZ will be of therapeutic benefit. On the other hand, patients with the dominant process focused on dysregulation at the level of C3 convertase (increased C3 split product levels), the impact of EZ therapy will be less impressive, and the process of uncontrolled complement dysregulation will persist with consequent ongoing renal injury. Secondly, future application of "soluble C5b-9" as well as "C3 degradation product" measurements will be feasible in monitoring EZ therapy (and other newly introduced C3 convertase inhibitors agents) and, thereby, will help in predicting its response[66]: (1) compstatin is a C3 inhibitory peptide that can block C3 and its convertase interaction, so that all of the three complement pathways are activated; (2) CP40 is a compstatin analog with a selective C3 inhibitor property. CP40 can prevent *in vitro* complement-mediated hemolysis induced by C3GN patient sera. Moreover, it can abort dysregulated AP activation induced by autoantibodies and genetic

A

Underlying defect	Lab response		Tissue response (histopathology)	Reference
	SCr	PCR		
None	○	●	●	Bomback *et al*, 2012[62]
C3Nef	●	●	Not performed	McCaughan *et al*, 2012[46]
C3Nef	●	●	●	Sa'nchez-Moreno *et al*, 2014[85]
None	●	●	●	Le Quintrec *et al*, 2015[80]

B

Underlying defect	Lab response		Tissue response (histopathology)	Reference
	SCr	PCR		
C3Nef	○	○	●	Bomback *et al*, 2012[62]
C3Nef, CD46 mutation	●	Non-proteinuric throughout	○	Bomback *et al*, 2012[62]
C3Nef, CFH mutations	○	● > ●	● (Increased fibrosis and continuously active C3GN)	Gurkan *et al*, 2013[64]
CFH and CFI mutations	●	●	● Improved tubulointerstitial injury, recovered ischemic injury; presistent 2-3 C3 deposition	Garg N *et al*, 2018[66]

● = Improved ○ = No change ● = Worsened ● > ● = Improved, then worsened

Figure 4 Response of complement 3 glomerulopathy subtypes to eculizumab therapy based on laboratory parameters and tissue (histopathological) response. A: Dense deposit disease response to eculizumab therapy[66]; B: Complement 3 glomerulonephritis response to eculizumab therapy[66]. CFH: Complement factor H; CFI: Complement factor I; C3Nef: C3 nephritic factor.

mutations[63]. Since C3d is the major complement fragment deposited in C3GN and DDD, CP40 represents a promising therapeutic agent. CP40 has been evaluated in paroxysmal nocturnal hemoglobinuria and hemodialysis-induced inflammation[69,70]. If CP40 is able to offer a disease-specific targeted therapy, this agent may represent a breakthrough in C3G control; (3) other novel therapeutics: antibody-based agents targeting complement function by blocking particular components of C3 convertase to hamper its formation and/or function (*e.g.*, anti-C3b monoclonal antibodies reported by Paixao-Cavalcante *et al*[71], anti-FB antibodies as described by Subias[72], and anti-properdin antibodies as professed by Pauly *et al*[73] targeting complement blockade are all under thorough evaluation[74]). Soluble complement receptor1 (CR1): a robust regulator of complement activity *in vitro,* soluble CR1 can prevent dysregulation of the AP C3 convertase. The safety and efficacy of the soluble CR1 in normalizing complement activity in pediatric patients with ESRD have been reported. With its ability to breakdown active C3b, soluble CR1 infusion can induce clinical improvement in C3GN as well as in the serum levels of MAC in patients with DDD recurrence[37].

Methods of achieving C3GN control are summarized in Table 5[34,75-86]. Until enough data from randomized control trials become available, the guidelines related to complement blockade therapy of C3GN should be based on those applied in aHUS (Table 6)[12].

Renal transplantation for C3G

Minimal data is available concerning renal transplantation for C3G. The available recommendations (Table 7) are currently based on expert opinion. Recurrence post-transplant is common, with about half of the patients with C3G at risk of losing their grafts[12].

TREATMENT OF POST-TRANSPLANT TMA

For cases of TMA secondary to medication, switching of the culprit drug to another agent (mTOR or CNI) is associated with a better response[88-90]. The first line of therapy of *de novo* TMA should encompass withdrawal of the offending drug, an essential step that is usually associated with correction of the hematological profile[57].

Plasmapheresis (PE) and intravenous immunoglobulins (IVIG) (particularly with AMR-associated TMA):

Table 6 Monitoring eculizumab therapy

CH50 (total complement activity)	AH50 (alternative pathway hemolytic activity)	Eculizumab trough	Alternative assays
Measures the combined activity of all of the complement pathways	Measures combined activity of alternative and terminal complement pathways	May be a free or bound level	The following assays are under investigation
Tests the functional capability of serum complement components to lyse 50 % of sheep erythrocytes in a reaction mixture	Tests functional capability of alternate or terminal pathway complement components to lyse 50% of rabbit erythrocytes in a Mg^{2+}-EGTA buffer	ELISA: using C5-coated plates, patient sera, and an anti-human IgG detection system	Free C5
Low in congenital complement deficiency (C1-8) or during complement blockade	Will be low in congenital C3, FI, FB, properdin, FH, and FD deficiencies or during terminal complement blockade	Not affected by complement deficiencies	*In vitro* human microvascular endothelial cell test
Normal range: Assay dependent	Normal range is assay-dependent.	Recommended trough level during complement blockade: 50-100 μg/mL	SC5b-9 (also referred to as sMAC and TCC) remain detectable in aHUS remission, so not recommended as a monitoring tool
Recommended goal during therapeutic complement blockade: < 10% of normal	Recommended goal during complement blockade: < 10% of normal		

Adapted from Goodship *et al*[12]. aHUS: Atypical hemolytic uremic syndrome; C3: Complement component 3; C5: Complement component 5; EGTA: Ethyleneglycol tetraacetic acid; ELISA: Enzyme-linked immunosorbent assay; FB: Complement factor B; FD: Complement factor D; FH: Complement factor H; FI: Complement factor I; sC5b-9: Soluble C5b-9; sMAC: Soluble membrane attack complex; TCC: Terminal complement complex.

Table 7 Transplant considerations in C3 glomerulopathy[1]

Timing	Donor selection	Risk reduction
Avoid transplantation during acute period of renal loss	No specific recommendation can be made on donor choice. When considering living donors, high risk of recurrence should be weighed against presumed risk of waiting on cadaveric donor list	C3G histological recurrence is as high as 90%[7,87]
Avoid transplantation during acute inflammation		Limited data suggest: rapid progression to ESRD in native kidneys increases recurrence risk[87]
No data supporting whether specific complement abnormalities (*e.g.*, high titer C3Nef, low C3 or high soluble C5b-9) predict increased risk for relapse		There are no known strategies to reduce recurrence risk of C3G
		Clinical recurrence should drive decision to treat[7]
		In absence of clinical trials, use of anti-complement therapy is based solely on a small open-label trial and positive case reports[62] (the impact of publication bias is unknown)
		C3G associated with monoclonal gammopathy has a high rate of recurrence[7]

[1]Based on limited retrospective cohort data. Adapted from Goodship *et al*[12]. C3: Complement component 3; C3G: C3 glomerulopathy; C3Nef: C3 nephritic factor; ESRD: End-stage renal disease.

fresh-frozen plasma (FFP) is advised as a reposition fluid, which must be type-specific, ordered in advance and thawed before use, despite the high risk of reactions; however, it replaces all plasma constituents and is appropriate for patients with TMA[91]. Before the era of EZ, the following supportive explanations have been provided: (1) proven efficacy in TTP[92]; (2) a graft salvage rate of more than 80%, as reported by Karthikeyan *et al*[13]. He addressed two possible benefits for this type of therapy: clearance of the platelet aggregation factors (*e.g.*, thromboxane A2) and replenishment of the deficient agents (*e.g.*, PGI2-stimulating factor)[13]; (3) with frequent possibility of the presence of underlying complement dysregulation, commencing PE therapy will also be beneficial in two ways: clearance of the aberrant complement components, and replacement with normally functioning complement proteins[93]; (4) clearance of the anti-HLA antibodies in AMR-associated aHUS improved patient outcome[55,94]; (5) PE/IVIG therapy was successfully associated with a 100% response rate in five solid organ transplants complicated by a systemic form of TMA. There was no evidence of relapse after cessation of the culprit drug (*e.g.*, tacrolimus) in a recent report[57].

Belatacept, a fusion protein composed of the Fc fragment of human IgG1 linked to the extracellular domain of cytotoxic T-lymphocyte-associated antigen 4 (CTLA-4), selectively inhibits T cell activation through a co-stimulatory blockade[95].

EZ, an anti-C5 agent that blocks lytic C5b-9 MAC generation, not only revolutionized aHUS therapy but was also effective in preventing its recurrence[96]. The role of complement activation in TMA evolution has been recognized in a majority of *de novo* TMA patients. Chua *et al*[97], for example, reported deposition of C4d in all biopsies of post-transplant TMA. Efficacy of this

Table 8 Eculizumab dosing in atypical hemolytic uremic syndrome based on dosing goal

Minimal dose	Discontinuation
Desire to continue dosing with the minimal dose required to achieve a pre-identified level of complement blockade[1]	Desire to discontinue complement blockade
Dose reduction or interval extension Goal CH50 < 10% (recommended) Goal AH50 < 10% (recommended) Goal eculizumab trough >100 μg/mL	No consensus exists regarding tapering of dose

[1]Additional monitoring may be required during intercurrent events (*e.g.*, infection, surgery, vaccination) to detect unblocked complement activity. Adapted from Goodship *et al*[12]. AH50: Alternative pathway hemolytic activity; CH50: Total complement activity.

agent has also been documented in the management of resistant cases of medication-associated *de novo* TMA, including those with unidentified genetic mutations[98-103]. Moreover, efficacy of EZ has been also shown in some cases of resistant AMR-associated TMA[103-111]. However, Loupy *et al*[112] reported a similar graft survival (95.8% *vs* 89.7% at two years post-transplant, respectively) and estimated GFR (52.6 mL/min *vs* 46.7 mL/min) in comparing PE-treated recipients with the EZ-treated group. Considering the high cost of this drug, utilization of this agent is better confined to PE-dependent patients, AMR-associated TMA and to cases with refractory hemolysis.

Treatment of recurrent TMA

Minimal work-up of genetic studies should include: CFH, CFI, CFHR, CFB, MCP and C3[113]. All cases with suspected TMA should be screened for all complement components and its related proteins. Cases with isolated membrane cofactor protein (MCP) mutations (not combined with other gene defects) may be safe for kidney donation. Cases with documented TMA and with a lack of definitive genetic defects may proceed with kidney transplantation under the umbrella of intensive PE therapy[114]. Polygenic patterns of TMA should be dealt with cautiously in case of living donation[115].

Prevention of aHUS

Avoid trigger factors that stimulate complement activity (*e.g.*, ischemia-reperfusion injury, viral infection and culprit medications)[52]. Immunosuppressive regimens devoid of medications related to TMA evolution[116] are advised. PE therapy alone is not sufficient for TMA cure and prevention, with the following explanations postulated: (1) PE alone frequently failed to prevent TMA recurrence[117]; (2) TMA regression cannot be preserved after cessation of therapy; and (3) recipients treated with PE showed an evidence of "subclinical" disease[118], which declares that PE has no influence on complement activity. Prophylactic use of rituximab proved to be beneficial as an anti-CFH-antibody[119], and this effect can be augmented with the addition of PE therapy[120,121]. The anti-C5 monoclonal antibody EZ has been reported to be successful in preventing TMA recurrence in recipients with CFH, CFH/CFHR1 hybrid gene mutations as well as in C3 gene mutations[122-125].

Prophylactic complement blockade

Eighty percent of kidney transplantation recipients with TMA proved to be associated with genetic mutations[126]. Based on the fact that a TMA episode is suspected with trigger factor (*e.g.*, surgery), a robust suggestion is to protect the patient with complement blockade, if not already instituted[127]. Unfortunately, this suggestion lacks appropriate evidence[12].

Therapeutic protocols for aHUS recurrence

Given a clear role of complement blockade in the management of TMA, two regimens have been suggested: (1) minimal dosage to achieve complement blockade; and (2) a dose withdrawal scheme (Table 8)[84]. EZ monitoring, however, is mandated for better response (Table 6)[128-131].

HOW TO MONITOR COMPLEMENT BLOCKADE - TABLE 6 DESCRIBES EZ THERAPY MONITORING

Duration of therapy

There is not enough data supporting life-long therapy. However, sustaining EZ seems to be reasonable in certain situations. Figure 5 represents a small guide, meanwhile early biomarkers of disease recurrence and complement activation became available.

Unanswered questions

The lacunae in satisfactory data still present as proper dosage, dose intervals, and duration of therapy[132], as well as the impact of this type of therapy on transplant spectrum[133].

Cessation of therapy

Figure 5 represents a guiding scheme suggested for EZ withdrawal[12].

Is EZ therapy the end of the road?

In 2013, Verhave *et al*[118] reported the feasibility of successful kidney transplantation without EZ therapy in four patients with high-risk aHUS. Patients received living donor kidneys with a therapeutic regimen consisting of: Basiliximab for induction, tacrolimus in low dosage, prednisone, and MMF for maintenance

Table 9 Risk of atypical hemolytic uremic syndrome recurrence according to the implicated genetic abnormalities

Gene mutation	Location	Functional Impact	Mutation frequency in aHUS (%)	Recurrence after transplantation (%)
CFH	Plasma	Loss	20-30	75-90
CFI	Plasma	Loss	2-12	45-80
CFB	Plasma	Gain	1-2	100
C3	Plasma	Gain	5-10	40-70
MCP	Membrane	Loss	10-15	15-20
THBD	Membrane	Loss	5	One case
Homozygous *CFHR1* del (3%-8%)	Circulating	Undetermined	14-23 (> 90% with anti-CHF AB)	NA

Adapted from Salvadori *et al*[1]. aHUS: Atypical hemolytic uremic syndrome; NA: Not available; CFH: Complement factor H; CFI: Complement factor I; CFB: Complement factor B; C3: Complement 3; MCP: Membrane cofactor protein; THBD: Thrombomodulin.

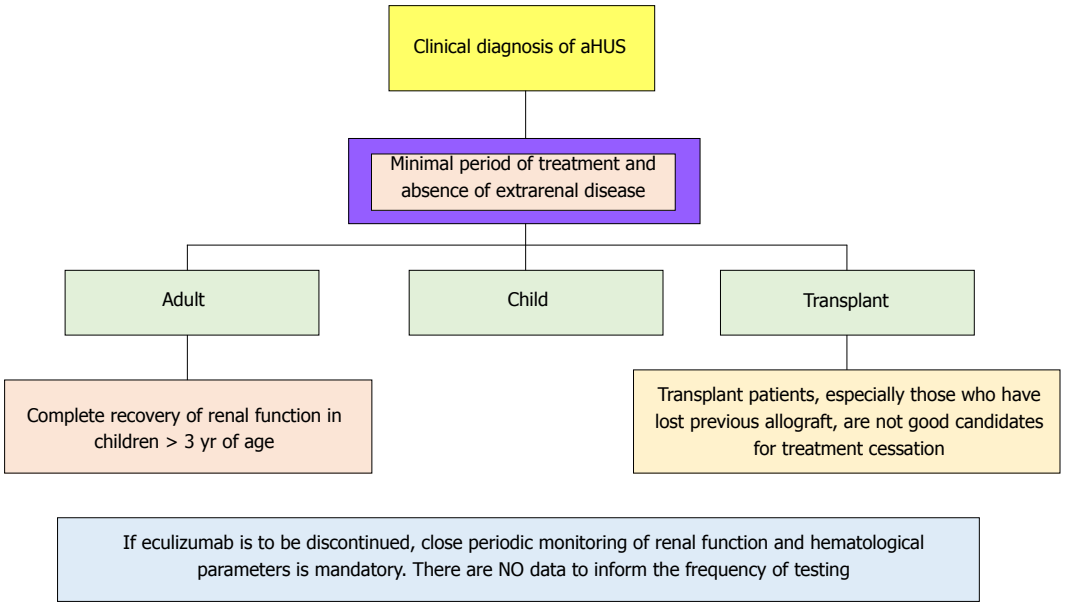

Figure 5 Recommendations for cessation of treatment with complement inhibitors. There are no prospective controlled studies in patients with atypical hemolytic uremic syndrome (aHUS) to define criteria for discontinuation of eculizumab therapy. This flow diagram is based on expert opinion[134-137]. Discontinuation can be considered on a case-by-case basis in patients after at least 6-12 mo of treatment and at least 3 mo of normalization (or stabilization in the case of residual chronic kidney disease) of kidney function. Earlier cessation (at 3 mo) may be considered in patients (especially children) with pathogenic variants in membrane cofactor protein if there has been rapid remission and recovery of renal function. Patients on dialysis or eculizumab should be maintained for at least 4 to 6 mo before discontinuation. In this setting, assessment of fibrotic changes in kidney biopsy may be helpful. In transplant patients, especially patients who have lost previous allografts, discontinuation is not recommended. Adapted from Goodship *et al*[12].

immunosuppression. A statin has also been added. Further precautions include: lowering BP as much as tolerable and minimizing the cold ischemic time. For the next 16-21 mo, no recurrence or rejection events have been reported[118]. The following conclusion has been addressed: successful kidney transplantation in recurrent aHUS patients can be achieved with an EZ-free regimen through: (1) decreasing cold ischemic time; (2) minimizing the risk of rejection; and (3) preserving endothelial integrity[118].

Renal transplantation in TMA

Timing of transplant: six months after commencing, dialysis should elapse before proceeding in transplant, as renal recovery can be observed several months after initiation of EZ therapy[137,138]. Two prerequisites should be fulfilled before commencing renal transplantation: (1) resolution of the extrarenal manifestations of TMA; and (2) recovery of TMA hematological parameters. The magnitude of recurrence risk may be used to evaluate the recipient's need for complement blockade (Table 9)[1].

CONCLUSION

The role of complement cascade in the evolution of kidney diseases either in the native kidney or post-transplant is well recognized. The prognosis of aHUS and, in some cases, C3G is greatly improved after commencing complement blockade. These agents are not only curative, but also successful in preventing post-transplant disease recurrence. Owing to the inherited nature of most of these diseases, the maintenance of this therapy is recommended despite cost burden. Consequently, the need for regimens allowing safe withdrawal of these agents is urgently required. However, newer therapies (*e.g.*, new monoclonal

antibodies, recombinant proteins, and small interfering RNA (siRNA) agents) hold promise for the near future[139,140].

REFERENCES

1 **Salvadori M**, Bertoni E. Complement related kidney diseases: Recurrence after transplantation. *World J Transplant* 2016; **6**: 632-645 [PMID: 28058212 DOI: 10.5500/wjt.v6.i4.632]

2 **Nester CM**, Barbour T, de Cordoba SR, Dragon-Durey MA, Fremeaux-Bacchi V, Goodship TH, Kavanagh D, Noris M, Pickering M, Sanchez-Corral P, Skerka C, Zipfel P, Smith RJ. Atypical aHUS: State of the art. *Mol Immunol* 2015; **67**: 31-42 [PMID: 25843230 DOI: 10.1016/j.molimm.2015.03.246]

3 **Zipfel PF**, Skerka C, Chen Q, Wiech T, Goodship T, Johnson S, Fremeaux-Bacchi V, Nester C, de Córdoba SR, Noris M, Pickering M, Smith R. The role of complement in C3 glomerulopathy. *Mol Immunol* 2015; **67**: 21-30 [PMID: 25929733 DOI: 10.1016/j.molimm.2015.03.012]

4 **Sethi S**, Fervenza F C. Understanding MPGN in the Native and Transplanted Kidney. Kidney Week 2017. Available from: URL: http://www.kidneynews.org/kidney-news/special-sections/glomerular-disease/understanding-mpgn-in-the-native-and-transplanted-kidney

5 **Sethi S**, Fervenza FC, Zhang Y, Zand L, Vrana JA, Nasr SH, Theis JD, Dogan A, Smith RJ. C3 glomerulonephritis: clinicopathological findings, complement abnormalities, glomerular proteomic profile, treatment, and follow-up. *Kidney Int* 2012; **82**: 465-473 [PMID: 22673887 DOI: 10.1038/ki.2012.212]

6 **Braun MC**, Stablein DM, Hamiwka LA, Bell L, Bartosh SM, Strife CF. Recurrence of membranoproliferative glomerulonephritis type II in renal allografts: The North American Pediatric Renal Transplant Cooperative Study experience. *J Am Soc Nephrol* 2005; **16**: 2225-2233 [PMID: 15888559 DOI: 10.1681/ASN.2005020175]

7 **Zand L**, Lorenz EC, Cosio FG, Fervenza FC, Nasr SH, Gandhi MJ, Smith RJ, Sethi S. Clinical findings, pathology, and outcomes of C3GN after kidney transplantation. *J Am Soc Nephrol* 2014; **25**: 1110-1117 [PMID: 24357668 DOI: 10.1681/ASN.2013070715]

8 **Hou J**, Markowitz GS, Bomback AS, Appel GB, Herlitz LC, Barry Stokes M, D'Agati VD. Toward a working definition of C3 glomerulopathy by immunofluorescence. *Kidney Int* 2014; **85**: 450-456 [PMID: 24067430 DOI: 10.1038/ki.2013.340]

9 **Larsen CP**, Ambuzs JM, Bonsib SM, Boils CL, Cossey LN, Messias NC, Silva FG, Wang YH, Gokden N, Walker PD. Membranous-like glomerulopathy with masked IgG kappa deposits. *Kidney Int* 2014; **86**: 154-161 [PMID: 24429395 DOI: 10.1038/ki.2013.548]

10 **Larsen CP**, Messias NC, Walker PD, Fidler ME, Cornell LD, Hernandez LH, Alexander MP, Sethi S, Nasr SH. Membranoproliferative glomerulonephritis with masked monotypic immunoglobulin deposits. *Kidney Int* 2015; **88**: 867-873 [PMID: 26154922 DOI: 10.1038/ki.2015.195]

11 **Barbour TD**, Pickering MC, Terence Cook H. Dense deposit disease and C3 glomerulopathy. *Semin Nephrol* 2013; **33**: 493-507 [PMID: 24161036 DOI: 10.1016/j.semnephrol.2013.08.002]

12 **Goodship TH**, Cook HT, Fakhouri F, Fervenza FC, Frémeaux-Bacchi V, Kavanagh D, Nester CM, Noris M, Pickering MC, Rodríguez de Córdoba S, Roumenina LT, Sethi S, Smith RJ; Conference Participants. Atypical hemolytic uremic syndrome and C3 glomerulopathy: conclusions from a "Kidney Disease: Improving Global Outcomes" (KDIGO) Controversies Conference. *Kidney Int* 2017; **91**: 539-551 [PMID: 27989322 DOI: 10.1016/j.kint.2016.10.005]

13 **Karthikeyan V**, Parasuraman R, Shah V, Vera E, Venkat KK. Outcome of plasma exchange therapy in thrombotic microangiopathy after renal transplantation. *Am J Transplant* 2003; **3**: 1289-1294 [PMID: 14510703 DOI: 10.1046/j.1600-6143.2003.00222.x]

14 **Nadasdy T**. Thrombotic microangiopathy in renal allografts: the diagnostic challenge. *Curr Opin Organ Transplant* 2014; **19**: 283-292 [PMID: 24811438 DOI: 10.1097/MOT.0000000000000074]

15 **Bouatou Y**, Bacchi VF, Villard J, Moll S, Martin PY, Hadaya K. Atypical Hemolytic Uremic Syndrome Recurrence after Renal Transplantation: C3-Glomerulonephritis as an Initial Presentation. *Transplant Direct* 2015; **1**: e9 [PMID: 27500215 DOI: 10.1097/TXD.0000000000000518]

16 **Matar D**, Naqvi F, Racusen LC, Carter-Monroe N, Montgomery RA, Alachkar N. Atypical hemolytic uremic syndrome recurrence after kidney transplantation. *Transplantation* 2014; **98**: 1205-1212 [PMID: 24933457 DOI: 10.1097/TP.0000000000000200]

17 **Recalde S**, Tortajada A, Subias M, Anter J, Blasco M, Maranta R, Coco R, Pinto S, Noris M, García-Layana A, Rodríguez de Córdoba S. Molecular Basis of Factor H R1210C Association with Ocular and Renal Diseases. *J Am Soc Nephrol* 2016; **27**: 1305-1311 [PMID: 26376859 DOI: 10.1681/ASN.2015050580]

18 **Fritsche LG**, Chen W, Schu M, Yaspan BL, Yu Y, Thorleifsson G, Zack DJ, Arakawa S, Cipriani V, Ripke S, Igo RP Jr, Buitendijk GH, Sim X, Weeks DE, Guymer RH, Merriam JE, Francis PJ, Hannum G, Agarwal A, Armbrecht AM, Audo I, Aung T, Barile GR, Benchaboune M, Bird AC, Bishop PN, Branham KE, Brooks M, Brucker AJ, Cade WH, Cain MS, Campochiaro PA, Chan CC, Cheng CY, Chew EY, Chin KA, Chowers I, Clayton DG, Cojocaru R, Conley YP, Cornes BK, Daly MJ, Dhillon B, Edwards AO, Evangelou E, Fagerness J, Ferreyra HA, Friedman JS, Geirsdottir A, George RJ, Gieger C, Gupta N, Hagstrom SA, Harding SP, Haritoglou C, Heckenlively JR, Holz FG, Hughes G, Ioannidis JP, Ishibashi T, Joseph P, Jun G, Kamatani Y, Katsanis N, N Keilhauer C, Khan JC, Kim IK, Kiyohara Y, Klein BE, Klein R, Kovach JL, Kozak I, Lee CJ, Lee KE, Lichtner P, Lotery AJ, Meitinger T, Mitchell P, Mohand-Saïd S, Moore AT, Morgan DJ, Morrison MA, Myers CE, Naj AC, Nakamura Y, Okada Y, Orlin A, Ortube MC, Othman MI, Pappas C, Park KH, Pauer GJ, Peachey NS, Poch O, Priya RR, Reynolds R, Richardson AJ, Ripp R, Rudolph G, Ryu E, Sahel JA, Schaumberg DA, Scholl HP, Schwartz SG, Scott WK, Shahid H, Sigurdsson H, Silvestri G, Sivakumaran TA, Smith RT, Sobrin L, Souied EH, Stambolian DE, Stefansson H, Sturgill-Short GM, Takahashi A, Tosakulwong N, Truitt BJ, Tsironi EE, Uitterlinden AG, van Duijn CM, Vijaya L, Vingerling JR, Vithana EN, Webster AR, Wichmann HE, Winkler TW, Wong TY, Wright AF, Zelenika D, Zhang M, Zhao L, Zhang K, Klein ML, Hageman GS, Lathrop GM, Stefansson K, Allikmets R, Baird PN, Gorin MB, Wang JJ, Klaver CC, Seddon JM, Pericak-Vance MA, Iyengar SK, Yates JR, Swaroop A, Weber BH, Kubo M, Deangelis MM, Léveillard T, Thorsteinsdottir U, Haines JL, Farrer LA, Heid IM, Abecasis GR; AMD Gene Consortium. Seven new loci associated with age-related macular degeneration. *Nat Genet* 2013; **45**: 433-439, 439e1-439e2 [PMID: 23455636 DOI: 10.1038/ng.2578]

19 **Mathieson PW**, Würzner R, Oliveria DB, Lachmann PJ, Peters DK. Complement-mediated adipocyte lysis by nephritic factor sera. *J Exp Med* 1993; **177**: 1827-1831 [PMID: 8496694 DOI: 10.1084/jem.177.6.1827]

20 **Sethi S**, Fervenza FC, Zhang Y, Nasr SH, Leung N, Vrana J, Cramer C, Nester CM, Smith RJ. Proliferative glomerulonephritis secondary to dysfunction of the alternative pathway of complement. *Clin J Am Soc Nephrol* 2011; **6**: 1009-1017 [PMID: 21415311 DOI: 10.2215/CJN.07110810]

21 **Sethi S**, Fervenza FC. Membranoproliferative glomerulonephritis: pathogenetic heterogeneity and proposal for a new classification. *Semin Nephrol* 2011; **31**: 341-348 [PMID: 21839367 DOI: 10.1016/j.semnephrol.2011.06.005]

22 **Rincón B**, Bernis C, Garcia A, Traver JA. Mesangiocapillary glomerulonephritis associated with hydatid disease. *Nephrol Dial Transplant* 1993; **8**: 783-784 [PMID: 8414165 DOI: 10.1093/ndt/8.8.783]

23 **Goules A**, Masouridi S, Tzioufas AG, Ioannidis JP, Skopouli FN, Moutsopoulos HM. Clinically significant and biopsy-documented renal involvement in primary Sjögren syndrome. *Medicine* (Baltimore) 2000; **79**: 241-249 [PMID: 10941353 DOI: 10.1097/00005792-200007000-00005]

24 **Servais A**, Noël LH, Roumenina LT, Le Quintrec M, Ngo S,

Dragon-Durey MA, Macher MA, Zuber J, Karras A, Provot F, Moulin B, Grünfeld JP, Niaudet P, Lesavre P, Frémeaux-Bacchi V. Acquired and genetic complement abnormalities play a critical role in dense deposit disease and other C3 glomerulopathies. *Kidney Int* 2012; **82**: 454-464 [PMID: 22456601 DOI: 10.1038/ki.2012.63]

25 **Dragon-Durey MA**, Blanc C, Marinozzi MC, van Schaarenburg RA, Trouw LA. Autoantibodies against complement components and functional consequences. *Mol Immunol* 2013; **56**: 213-221 [PMID: 23790637 DOI: 10.1016/j.molimm.2013.05.009]

26 **Smith RJ**, Harris CL, Pickering MC. Dense deposit disease. *Mol Immunol* 2011; **48**: 1604-1610 [PMID: 21601923 DOI: 10.1016/j.molimm.2011.04.005]

27 **Thomas S**, Ranganathan D, Francis L, Madhan K, John GT. Current concepts in C3 glomerulopathy. *Indian J Nephrol* 2014; **24**: 339-348 [PMID: 25484526 DOI: 10.4103/0971-4065.134089]

28 **Chen Q**, Müller D, Rudolph B, Hartmann A, Kuwertz-Bröking E, Wu K, Kirschfink M, Skerka C, Zipfel PF. Combined C3b and factor B autoantibodies and MPGN type II. *N Engl J Med* 2011; **365**: 2340-2342 [PMID: 22168663 DOI: 10.1056/NEJMc1107484]

29 **Goodship TH**, Pappworth IY, Toth T, Denton M, Houlberg K, McCormick F, Warland D, Moore I, Hunze EM, Staniforth SJ, Hayes C, Cavalcante DP, Kavanagh D, Strain L, Herbert AP, Schmidt CQ, Barlow PN, Harris CL, Marchbank KJ. Factor H autoantibodies in membranoproliferative glomerulonephritis. *Mol Immunol* 2012; **52**: 200-206 [PMID: 22721707 DOI: 10.1016/j.molimm.2012.05.009]

30 **Lorcy N**, Rioux-Leclercq N, Lombard ML, Le Pogamp P, Vigneau C. Three kidneys, two diseases, one antibody? *Nephrol Dial Transplant* 2011; **26**: 3811-3813 [PMID: 21813829 DOI: 10.1093/ndt/gfr436]

31 **Dragon-Durey MA**, Frémeaux-Bacchi V, Loirat C, Blouin J, Niaudet P, Deschenes G, Coppo P, Herman Fridman W, Weiss L. Heterozygous and homozygous factor h deficiencies associated with hemolytic uremic syndrome or membranoproliferative glomerulonephritis: report and genetic analysis of 16 cases. *J Am Soc Nephrol* 2004; **15**: 787-795 [PMID: 14978182 DOI: 10.1097/01.ASN.0000115702.28859.A7]

32 **Servais A**, Noël LH, Dragon-Durey MA, Gübler MC, Rémy P, Buob D, Cordonnier C, Makdassi R, Jaber W, Boulanger E, Lesavre P, Frémeaux-Bacchi V. Heterogeneous pattern of renal disease associated with homozygous factor H deficiency. *Hum Pathol* 2011; **42**: 1305-1311 [PMID: 21396679 DOI: 10.1016/j.humpath.2010.11.023]

33 **Skerka C**, Chen Q, Fremeaux-Bacchi V, Roumenina LT. Complement factor H related proteins (CFHRs). *Mol Immunol* 2013; **56**: 170-180 [PMID: 23830046 DOI: 10.1016/j.molimm.2013.06.001]

34 **Besbas N**, Gulhan B, Gucer S, Korkmaz E, Ozaltin F. A novel CFHR5 mutation associated with C3 glomerulonephritis in a Turkish girl. *J Nephrol* 2014; **27**: 457-460 [PMID: 24536001 DOI: 10.1007/s40620-013-0008-1]

35 **Chen Q**, Manzke M, Hartmann A, Büttner M, Amann K, Pauly D, Wiesener M, Skerka C, Zipfel PF. Complement Factor H-Related 5-Hybrid Proteins Anchor Properdin and Activate Complement at Self-Surfaces. *J Am Soc Nephrol* 2016; **27**: 1413-1425 [PMID: 26432903 DOI: 10.1681/ASN.2015020212]

36 **Gale DP**, de Jorge EG, Cook HT, Martinez-Barricarte R, Hadjisavvas A, McLean AG, Pusey CD, Pierides A, Kyriacou K, Athanasiou Y, Voskarides K, Deltas C, Palmer A, Frémeaux-Bacchi V, de Cordoba SR, Maxwell PH, Pickering MC. Identification of a mutation in complement factor H-related protein 5 in patients of Cypriot origin with glomerulonephritis. *Lancet* 2010; **376**: 794-801 [PMID: 20800271 DOI: 10.1016/S0140-6736(10)60670-8]

37 **Zhang Y**, Nester CM, Holanda DG, Marsh HC, Hammond RA, Thomas LJ, Meyer NC, Hunsicker LG, Sethi S, Smith RJ. Soluble CR1 therapy improves complement regulation in C3 glomerulopathy. *J Am Soc Nephrol* 2013; **24**: 1820-1829 [PMID: 23907509 DOI: 10.1681/ASN.2013010045]

38 **Malik TH**, Lavin PJ, Goicoechea de Jorge E, Vernon KA, Rose KL, Patel MP, de Leeuw M, Neary JJ, Conlon PJ, Winn MP, Pickering MC. A hybrid CFHR3-1 gene causes familial C3 glomerulopathy. *J Am Soc Nephrol* 2012; **23**: 1155-1160 [PMID: 22626820 DOI: 10.1681/ASN.2012020166]

39 **Tortajada A**, Garcia SP, Gastoldi S, Fernandez JG, Martin Merinero H, Arjona E, Noris M, Rodriguez de Cordoba S. Prevalent FHR1 mutant protein generated by gene conversion reveals crucial role of factor H polymorhisms in atypical hemolytic uremic syndrome. *Immunobiology* 2016; **221**: 1199 [DOI: 10.1016/j.imbio.2016.06.166]

40 **Thurman JM**. Complement in kidney disease: core curriculum 2015. *Am J Kidney Dis* 2015; **65**: 156-168 [PMID: 25441433 DOI: 10.1053/j.ajkd.2014.06.035]

41 **Goicoechea de Jorge E**, Pickering MC. Atypical hemolytic uremic syndrome: telling the difference between H and Y. *Kidney Int* 2010; **78**: 721-723 [PMID: 20877372 DOI: 10.1038/ki.2010.222]

42 **Noris M**, Remuzzi G. Glomerular Diseases Dependent on Complement Activation, Including Atypical Hemolytic Uremic Syndrome, Membranoproliferative Glomerulonephritis, and C3 Glomerulopathy: Core Curriculum 2015. *Am J Kidney Dis* 2015; **66**: 359-375 [PMID: 26032627 DOI: 10.1053/j.ajkd.2015.03.040]

43 **Cochat P**, Fargue S, Mestrallet G, Jungraithmayr T, Koch-Nogueira P, Ranchin B, Zimmerhackl LB. Disease recurrence in paediatric renal transplantation. *Pediatr Nephrol* 2009; **24**: 2097-2108 [PMID: 19247694 DOI: 10.1007/s00467-009-1137-6]

44 **Medjeral-Thomas NR**, O'Shaughnessy MM, O'Regan JA, Traynor C, Flanagan M, Wong L, Teoh CW, Awan A, Waldron M, Cairns T, O'Kelly P, Dorman AM, Pickering MC, Conlon PJ, Cook HT. C3 glomerulopathy: clinicopathologic features and predictors of outcome. *Clin J Am Soc Nephrol* 2014; **9**: 46-53 [PMID: 24178974 DOI: 10.2215/CJN.04700513]

45 **Angelo JR**, Bell CS, Braun MC. Allograft failure in kidney transplant recipients with membranoproliferative glomerulonephritis. *Am J Kidney Dis* 2011; **57**: 291-299 [PMID: 21215503 DOI: 10.1053/j.ajkd.2010.09.021]

46 **McCaughan JA**, O'Rourke DM, Courtney AE. Recurrent dense deposit disease after renal transplantation: an emerging role for complementary therapies. *Am J Transplant* 2012; **12**: 1046-1051 [PMID: 22233157 DOI: 10.1111/j.1600-6143.2011.03923.x]

47 **Andresdottir MB**, Assmann KJ, Hoitsma AJ, Koene RA, Wetzels JF. Renal transplantation in patients with dense deposit disease: morphological characteristics of recurrent disease and clinical outcome. *Nephrol Dial Transplant* 1999; **14**: 1723-1731 [PMID: 10435883 DOI: 10.1093/ndt/14.7.1723]

48 **Ponticelli C**, Glassock RJ. Posttransplant recurrence of primary glomerulonephritis. *Clin J Am Soc Nephrol* 2010; **5**: 2363-2372 [PMID: 21030574 DOI: 10.2215/CJN.06720810]

49 **Andresdottir MB**, Assmann KJ, Hoitsma AJ, Koene RA, Wetzels JF. Recurrence of type I membranoproliferative glomerulonephritis after renal transplantation: analysis of the incidence, risk factors, and impact on graft survival. *Transplantation* 1997; **63**: 1628-1633 [PMID: 9197358 DOI: 10.1097/00007890-199706150-00016]

50 **Little MA**, Dupont P, Campbell E, Dorman A, Walshe JJ. Severity of primary MPGN, rather than MPGN type, determines renal survival and post-transplantation recurrence risk. *Kidney Int* 2006; **69**: 504-511 [PMID: 16395262 DOI: 10.1038/sj.ki.5000084]

51 **Wong L**, Moran S, Lavin PJ, Dorman AM, Conlon PJ. Kidney transplant outcomes in familial C3 glomerulopathy. *Clin Kidney J* 2016; **9**: 403-407 [PMID: 27274824 DOI: 10.1093/ckj/sfw020]

52 **Zuber J**, Le Quintrec M, Morris H, Frémeaux-Bacchi V, Loirat C, Legendre C. Targeted strategies in the prevention and management of atypical HUS recurrence after kidney transplantation. *Transplant Rev* (Orlando) 2013; **27**: 117-125 [PMID: 23937869 DOI: 10.1016/j.trre.2013.07.003]

53 **Pickering MC**, D'Agati VD, Nester CM, Smith RJ, Haas M, Appel GB, Alpers CE, Bajema IM, Bedrosian C, Braun M, Doyle M, Fakhouri F, Fervenza FC, Fogo AB, Frémeaux-Bacchi V, Gale DP, Goicoechea de Jorge E, Griffin G, Harris CL, Holers VM, Johnson S, Lavin PJ, Medjeral-Thomas N, Paul Morgan B, Nast CC, Noel LH, Peters DK, Rodríguez de Córdoba S, Servais A, Sethi S, Song WC, Tamburini P, Thurman JM, Zavros M, Cook HT. C3 glomerulopathy: consensus report. *Kidney Int* 2013; **84**: 1079-1089 [PMID: 24172683 DOI: 10.1038/ki.2013.377]

54 **Schwimmer J**, Nadasdy TA, Spitalnik PF, Kaplan KL, Zand MS. De novo thrombotic microangiopathy in renal transplant recipients: a comparison of hemolytic uremic syndrome with localized renal thrombotic microangiopathy. *Am J Kidney Dis* 2003; **41**: 471-479 [PMID: 12552512 DOI: 10.1053/ajkd.2003.50058]

55 **Satoskar AA**, Pelletier R, Adams P, Nadasdy GM, Brodsky S, Pesavento T, Henry M, Nadasdy T. De novo thrombotic microangiopathy in renal allograft biopsies-role of antibody-mediated rejection. *Am J Transplant* 2010; **10**: 1804-1811 [PMID: 20659088 DOI: 10.1111/j.1600-6143.2010.03178.x]

56 **Fortin MC**, Raymond MA, Madore F, Fugère JA, Pâquet M, St-Louis G, Hébert MJ. Increased risk of thrombotic microangiopathy in patients receiving a cyclosporin-sirolimus combination. *Am J Transplant* 2004; **4**: 946-952 [PMID: 15147429 DOI: 10.1111/j.1600-6143.2004.00428.x]

57 **Garg N**, Rennke HG, Pavlakis M, Zandi-Nejad K. De novo thrombotic microangiopathy after kidney transplantation. *Transplant Rev* (Orlando) 2018; **32**: 58-68 [PMID: 29157988 DOI: 10.1016/j.trre.2017.10.001]

58 **Java A**, Gaut JP, Brennan DC. De novo membranoproliferative glomerulonephritis III in a renal transplant patient: case report and review of the literature. *Transpl Int* 2012; **25**: e58-e61 [PMID: 22380572 DOI: 10.1111/j.1432-2277.2012.01452.x]

59 **Habbig S**, Mihatsch MJ, Heinen S, Beck B, Emmel M, Skerka C, Kirschfink M, Hoppe B, Zipfel PF, Licht C. C3 deposition glomerulopathy due to a functional factor H defect. *Kidney Int* 2009; **75**: 1230-1234 [PMID: 18633337 DOI: 10.1038/ki.2008.354]

60 **Nester CM**, Smith RJ. Treatment options for C3 glomerulopathy. *Curr Opin Nephrol Hypertens* 2013; **22**: 231-237 [PMID: 23318699 DOI: 10.1097/MNH.0b013e32835da24c]

61 **Rabasco C**, Cavero T, Román E, Rojas-Rivera J, Olea T, Espinosa M, Cabello V, Fernández-Juarez G, González F, Ávila A, Baltar JM, Díaz M, Alegre R, Elías S, Antón M, Frutos MA, Pobes A, Blasco M, Martín F, Bernis C, Macías M, Barroso S, de Lorenzo A, Ariceta G, López-Mendoza M, Rivas B, López-Revuelta K, Campistol JM, Mendizábal S, de Córdoba SR, Praga M; Spanish Group for the Study of Glomerular Diseases (GLOSEN). Effectiveness of mycophenolate mofetil in C3 glomerulonephritis. *Kidney Int* 2015; **88**: 1153-1160 [PMID: 26221755 DOI: 10.1038/ki.2015.227]

62 **Bomback AS**, Smith RJ, Barile GR, Zhang Y, Heher EC, Herlitz L, Stokes MB, Markowitz GS, D'Agati VD, Canetta PA, Radhakrishnan J, Appel GB. Eculizumab for dense deposit disease and C3 glomerulonephritis. *Clin J Am Soc Nephrol* 2012; **7**: 748-756 [PMID: 22403278 DOI: 10.2215/CJN.12901211]

63 **Sanghera P**, Ghanta M, Ozay F, Ariyamuthu VK, Tanriover B. Kidney Diseases Associated With Alternative Complement Pathway Dysregulation and Potential Treatment Options. *Am J Med Sci* 2017; **354**: 533-538 [PMID: 29208248 DOI: 10.1016/j.amjms.2017.03.024]

64 **Gurkan S**, Fyfe B, Weiss L, Xiao X, Zhang Y, Smith RJ. Eculizumab and recurrent C3 glomerulonephritis. *Pediatr Nephrol* 2013; **28**: 1975-1981 [PMID: 23689905 DOI: 10.1007/s00467-013-2503-y]

65 **Herlitz LC**, Bomback AS, Markowitz GS, Stokes MB, Smith RN, Colvin RB, Appel GB, D'Agati VD. Pathology after eculizumab in dense deposit disease and C3 GN. *J Am Soc Nephrol* 2012; **23**: 1229-1237 [PMID: 22677550 DOI: 10.1681/ASN.2011121186]

66 **Garg N**, Zhang Y, Nicholson-Weller A, Khankin EV, Borsa NG, Meyer NC, McDermott S, Stillman IE, Rennke HG, Smith RJ, Pavlakis M. C3 glomerulonephritis secondary to mutations in factors H and I: rapid recurrence in deceased donor kidney transplant effectively treated with eculizumab. *Nephrol Dial Transplant* 2018 [PMID: 29370420 DOI: 10.1093/ndt/gfx369]

67 **Sethi S**, Nester CM, Smith RJ. Membranoproliferative glomerulonephritis and C3 glomerulopathy: resolving the confusion. *Kidney Int* 2012; **81**: 434-441 [PMID: 22157657 DOI: 10.1038/ki.2011.399]

68 **Rose KL**, Paixao-Cavalcante D, Fish J, Manderson AP, Malik TH, Bygrave AE, Lin T, Sacks SH, Walport MJ, Cook HT, Botto M, Pickering MC. Factor I is required for the development of membranoproliferative glomerulonephritis in factor H-deficient mice. *J Clin Invest* 2008; **118**: 608-618 [PMID: 18202746 DOI: 10.1172/JCI32525]

69 **Risitano AM**, Ricklin D, Huang Y, Reis ES, Chen H, Ricci P, Lin Z, Pascariello C, Raia M, Sica M, Del Vecchio L, Pane F, Lupu F, Notaro R, Resuello RR, DeAngelis RA, Lambris JD. Peptide inhibitors of C3 activation as a novel strategy of complement inhibition for the treatment of paroxysmal nocturnal hemoglobinuria. *Blood* 2014; **123**: 2094-2101 [PMID: 24497537 DOI: 10.1182/blood-2013-11-536573]

70 **Reis ES**, DeAngelis RA, Chen H, Resuello RR, Ricklin D, Lambris JD. Therapeutic C3 inhibitor Cp40 abrogates complement activation induced by modern hemodialysis filters. *Immunobiology* 2015; **220**: 476-482 [PMID: 25468722 DOI: 10.1016/j.imbio.2014.10.026]

71 **Paixão-Cavalcante D**, Torreira E, Lindorfer MA, Rodriguez de Cordoba S, Morgan BP, Taylor RP, Llorca O, Harris CL. A humanized antibody that regulates the alternative pathway convertase: potential for therapy of renal disease associated with nephritic factors. *J Immunol* 2014; **192**: 4844-4851 [PMID: 24729617 DOI: 10.4049/jimmunol.1303131]

72 **Subías M**, Tortajada A, Gastoldi S, Galbusera M, López-Perrote A, Lopez Lde J, González-Fernández FA, Villegas-Martínez A, Dominguez M, Llorca O, Noris M, Morgan BP, Rodríguez de Córdoba S. A novel antibody against human factor B that blocks formation of the C3bB proconvertase and inhibits complement activation in disease models. *J Immunol* 2014; **193**: 5567-5575 [PMID: 25355917 DOI: 10.4049/jimmunol.1402013]

73 **Pauly D**, Nagel BM, Reinders J, Killian T, Wulf M, Ackermann S, Ehrenstein B, Zipfel PF, Skerka C, Weber BH. A novel antibody against human properdin inhibits the alternative complement system and specifically detects properdin from blood samples. *PLoS One* 2014; **9**: e96371 [PMID: 24797388 DOI: 10.1371/journal.pone.0096371]

74 **Zhang Y**, Shao D, Ricklin D, Hilkin BM, Nester CM, Lambris JD, Smith RJ. Compstatin analog Cp40 inhibits complement dysregulation in vitro in C3 glomerulopathy. *Immunobiology* 2015; **220**: 993-998 [PMID: 25982307 DOI: 10.1016/j.imbio.2015.04.001]

75 **Bonucchi D**, Leonelli M, Damiano F, Granito M, Ghiandai G, De Amicis S, Americo C, Ligabue G, Albertazzi V, Cappelli G. [Post-transplant recurrence of glomerulonephritis: a complex clinical case]. *G Ital Nefrol* 2010; **27** Suppl 52: S82-S84 [PMID: 21132668]

76 **Daina E**, Noris M, Remuzzi G. Eculizumab in a patient with dense-deposit disease. *N Engl J Med* 2012; **366**: 1161-1163 [PMID: 22435382 DOI: 10.1056/NEJMc1112273]

77 **Garnier AS**, Augusto JF, Pellier I, Subra JF, Sayegh J. Successful long-term outcome of kidney transplantation in a patient with X-linked thrombocytopenia: 9-year follow-up. *Transplantation* 2014; **98**: e57-e58 [PMID: 25221901 DOI: 10.1097/TP.0000000000000338]

78 **Inman M**, Prater G, Fatima H, Wallace E. Eculizumab-induced reversal of dialysis-dependent kidney failure from C3 glomerulonephritis. *Clin Kidney J* 2015; **8**: 445-448 [PMID: 26251714 DOI: 10.1093/ckj/sfv044]

79 **Kerns E**, Rozansky D, Troxell ML. Evolution of immunoglobulin deposition in C3-dominant membranoproliferative glomerulopathy. *Pediatr Nephrol* 2013; **28**: 2227-2231 [PMID: 23892798 DOI: 10.1007/s00467-013-2565-x]

80 **Le Quintrec M**, Lionet A, Kandel C, Bourdon F, Gnemmi V, Colombat M, Goujon JM, Frémeaux-Bacchi V, Fakhouri F. Eculizumab for treatment of rapidly progressive C3 glomerulopathy. *Am J Kidney Dis* 2015; **65**: 484-489 [PMID: 25530108 DOI: 10.1053/j.ajkd.2014.09.025]

81 **Oosterveld MJ**, Garrelfs MR, Hoppe B, Florquin S, Roelofs JJ, van den Heuvel LP, Amann K, Davin JC, Bouts AH, Schriemer PJ, Groothoff JW. Eculizumab in Pediatric Dense Deposit Disease. *Clin J Am Soc Nephrol* 2015; **10**: 1773-1782 [PMID: 26316621 DOI: 10.2215/CJN.01360215]

82 **Ozkaya O**, Nalcacioglu H, Tekcan D, Genc G, Meydan BC, Ozdemir BH, Baysal MK, Keceligil HT. Eculizumab therapy in a patient with dense-deposit disease associated with partial

lipodystropy. *Pediatr Nephrol* 2014; **29**: 1283-1287 [PMID: 24464478 DOI: 10.1007/s00467-013-2748-5]
83 **Radhakrishnan S**, Lunn A, Kirschfink M, Thorner P, Hebert D, Langlois V, Pluthero F, Licht C. Eculizumab and refractory membranoproliferative glomerulonephritis. *N Engl J Med* 2012; **366**: 1165-1166 [PMID: 22435384 DOI: 10.1056/NEJMc1106619]
84 **Rousset-Rouvière C**, Cailliez M, Garaix F, Bruno D, Laurent D, Tsimaratos M. Rituximab fails where eculizumab restores renal function in C3nef-related DDD. *Pediatr Nephrol* 2014; **29**: 1107-1111 [PMID: 24408225 DOI: 10.1007/s00467-013-2711-5]
85 **Sánchez-Moreno A**, De la Cerda F, Cabrera R, Fijo J, López-Trascasa M, Bedoya R, Rodríguez de Córdoba S, Ybot-González P. Eculizumab in dense-deposit disease after renal transplantation. *Pediatr Nephrol* 2014; **29**: 2055-2059 [PMID: 24908321 DOI: 10.1007/s00467-014-2839-y]
86 **Vivarelli M**, Pasini A, Emma F. Eculizumab for the treatment of dense-deposit disease. *N Engl J Med* 2012; **366**: 1163-1165 [PMID: 22435383 DOI: 10.1056/NEJMc1111953]
87 **Lu DF**, Moon M, Lanning LD, McCarthy AM, Smith RJ. Clinical features and outcomes of 98 children and adults with dense deposit disease. *Pediatr Nephrol* 2012; **27**: 773-781 [PMID: 22105967 DOI: 10.1007/s00467-011-2059-7]
88 **Czubkowski P**, Pawłowska J, Jankowska I, Teisseyre M, Kamińska D, Markiewicz M, Ryżko J. Successful sirolimus rescue in tacrolimus-induced thrombotic microangiopathy after living-related liver transplantation. *Pediatr Transplant* 2012; **16**: E261-E264 [PMID: 22066835 DOI: 10.1111/j.1399-3046.2011.01601.x]
89 **Franco A**, Hernandez D, Capdevilla L, Errasti P, Gonzalez M, Ruiz JC, Sanchez J; HUS-Sirolimus Spanish Study Group. De novo hemolytic-uremic syndrome/thrombotic microangiopathy in renal transplant patients receiving calcineurin inhibitors: role of sirolimus. *Transplant Proc* 2003; **35**: 1764-1766 [PMID: 12962787 DOI: 10.1016/S0041-1345(03)00614-6]
90 **Epperla N**, Hemauer K, Hamadani M, Friedman KD, Kreuziger LB. Impact of treatment and outcomes for patients with posttransplant drug-associated thrombotic microangiopathy. *Transfusion* 2017; **57**: 2775-2781 [PMID: 28836275 DOI: 10.1111/trf.14263]
91 **McLeod BC**. Therapeutic apheresis: use of human serum albumin, fresh frozen plasma and cryosupernatant plasma in therapeutic plasma exchange. *Best Pract Res Clin Haematol* 2006; **19**: 157-167 [PMID: 16377548 DOI: 10.1016/j.beha.2005.01.004]
92 **Bell WR**, Braine HG, Ness PM, Kickler TS. Improved survival in thrombotic thrombocytopenic purpura-hemolytic uremic syndrome. Clinical experience in 108 patients. *N Engl J Med* 1991; **325**: 398-403 [PMID: 2062331 DOI: 10.1056/NEJM199108083250605]
93 **Le Quintrec M**, Lionet A, Kamar N, Karras A, Barbier S, Buchler M, Fakhouri F, Provost F, Fridman WH, Thervet E, Legendre C, Zuber J, Frémeaux-Bacchi V. Complement mutation-associated de novo thrombotic microangiopathy following kidney transplantation. *Am J Transplant* 2008; **8**: 1694-1701 [PMID: 18557729 DOI: 10.1111/j.1600-6143.2008.02297.x]
94 **Djamali A**, Kaufman DB, Ellis TM, Zhong W, Matas A, Samaniego M. Diagnosis and management of antibody-mediated rejection: current status and novel approaches. *Am J Transplant* 2014; **14**: 255-271 [PMID: 24401076 DOI: 10.1111/ajt.12589]
95 **Masson P**, Henderson L, Chapman JR, Craig JC, Webster AC. Belatacept for kidney transplant recipients. *Cochrane Database Syst Rev* 2014; **(11)**: CD010699 [PMID: 25416857 DOI: 10.1002/14651858.CD010699.pub2]
96 **Legendre CM**, Licht C, Muus P, Greenbaum LA, Babu S, Bedrosian C, Bingham C, Cohen DJ, Delmas Y, Douglas K, Eitner F, Feldkamp T, Fouque D, Furman RR, Gaber O, Herthelius M, Hourmant M, Karpman D, Lebranchu Y, Mariat C, Menne J, Moulin B, Nürnberger J, Ogawa M, Remuzzi G, Richard T, Sberro-Soussan R, Severino B, Sheerin NS, Trivelli A, Zimmerhackl LB, Goodship T, Loirat C. Terminal complement inhibitor eculizumab in atypical hemolytic-uremic syndrome. *N Engl J Med* 2013; **368**: 2169-2181 [PMID: 23738544 DOI: 10.1056/NEJMoa1208981]
97 **Chua JS**, Baelde HJ, Zandbergen M, Wilhelmus S, van Es LA, de Fijter JW, Bruijn JA, Bajema IM, Cohen D. Complement Factor C4d Is a Common Denominator in Thrombotic Microangiopathy. *J Am Soc Nephrol* 2015; **26**: 2239-2247 [PMID: 25573909 DOI: 10.1681/ASN.2014050429]
98 **Shochet L**, Kanellis J, Simpson I, Ta J, Mulley W. De novo thrombotic microangiopathy following simultaneous pancreas and kidney transplantation managed with eculizumab. *Nephrology* (Carlton) 2017; **22** Suppl 1: 23-27 [PMID: 28176480 DOI: 10.1111/nep.12936]
99 **Dedhia P**, Govil A, Mogilishetty G, Alloway RR, Woodle ES, Abu Jawdeh BG. Eculizumab and Belatacept for De Novo Atypical Hemolytic Uremic Syndrome Associated With CFHR3-CFHR1 Deletion in a Kidney Transplant Recipient: A Case Report. *Transplant Proc* 2017; **49**: 188-192 [PMID: 28104134 DOI: 10.1016/j.transproceed.2016.11.008]
100 **Ikeda T**, Okumi M, Unagami K, Kanzawa T, Sawada A, Kawanishi K, Omoto K, Ishida H, Tanabe K. Two cases of kidney transplantation-associated thrombotic microangiopathy successfully treated with eculizumab. *Nephrology* (Carlton) 2016; **21** Suppl 1: 35-40 [PMID: 26970541 DOI: 10.1111/nep.12768]
101 **Safa K**, Logan MS, Batal I, Gabardi S, Rennke HG, Abdi R. Eculizumab for drug-induced de novo posttransplantation thrombotic microangiopathy: A case report. *Clin Nephrol* 2015; **83**: 125-129 [PMID: 24495904 DOI: 10.5414/CN108163]
102 **Loirat C**, Babu S, Furman R, Sheerin N, Cohen D, Gaber O. Eculizumab Efficacy and Safety in Patients With Atypical Hemolytic Uremic Syndrome (aHUS) Resistant to Plasma Exchange/Infusion. Poster presented at the 16th Congress of European Hematology Association (EHA); 2011 June 9-12; London, UK: Abstract 0979
103 **Loirat C**, Muus P, Legendre C, Douglas K, Hourmant M, Delmas Y. A Phase II Study of Eculizumab in Patients with Atypical Hemolytic Uremic Syndrome Receiving Chronic Plasma Exchange/Infusion. Poster presented at the 16th Congress of European Hematology Association (EHA); 2011 June 9-12; London, UK: Abstract 0979
104 **Stegall MD**, Diwan T, Raghavaiah S, Cornell LD, Burns J, Dean PG, Cosio FG, Gandhi MJ, Kremers W, Gloor JM. Terminal complement inhibition decreases antibody-mediated rejection in sensitized renal transplant recipients. *Am J Transplant* 2011; **11**: 2405-2413 [PMID: 21942930 DOI: 10.1111/j.1600-6143.2011.03757.x]
105 **González-Roncero F**, Suñer M, Bernal G, Cabello V, Toro M, Pereira P, Angel Gentil M. Eculizumab treatment of acute antibody-mediated rejection in renal transplantation: case reports. *Transplant Proc* 2012; **44**: 2690-2694 [PMID: 23146495 DOI: 10.1016/j.transproceed.2012.09.038]
106 **Chehade H**, Rotman S, Matter M, Girardin E, Aubert V, Pascual M. Eculizumab to treat antibody-mediated rejection in a 7-year-old kidney transplant recipient. *Pediatrics* 2015; **135**: e551-e555 [PMID: 25624380 DOI: 10.1542/peds.2014-2275]
107 **Locke JE**, Magro CM, Singer AL, Segev DL, Haas M, Hillel AT, King KE, Kraus E, Lees LM, Melancon JK, Stewart ZA, Warren DS, Zachary AA, Montgomery RA. The use of antibody to complement protein C5 for salvage treatment of severe antibody-mediated rejection. *Am J Transplant* 2009; **9**: 231-235 [PMID: 18976298 DOI: 10.1111/j.1600-6143.2008.02451.x]
108 **Lonze BE**, Dagher NN, Simpkins CE, Locke JE, Singer AL, Segev DL, Zachary AA, Montgomery RA. Eculizumab, bortezomib and kidney paired donation facilitate transplantation of a highly sensitized patient without vascular access. *Am J Transplant* 2010; **10**: 2154-2160 [PMID: 20636451 DOI: 10.1111/j.1600-6143.2010.03191.x]
109 **Stewart ZA**, Collins TE, Schlueter AJ, Raife TI, Holanda DG, Nair R, Reed AI, Thomas CP. Case report: Eculizumab rescue of severe accelerated antibody-mediated rejection after ABO-incompatible kidney transplant. *Transplant Proc* 2012; **44**: 3033-3036 [PMID: 23195021 DOI: 10.1016/j.transproceed.2012.03.053]
110 **Tran D**, Boucher A, Collette S, Payette A, Royal V, Senécal L. Eculizumab for the Treatment of Severe Antibody-Mediated Rejection: A Case Report and Review of the Literature. *Case Rep Transplant* 2016; **2016**: 9874261 [PMID: 27478676 DOI: 10.1155/2016/9874261]
111 **Orandi BJ**, Zachary AA, Dagher NN, Bagnasco SM, Garonzik-

Wang JM, Van Arendonk KJ, Gupta N, Lonze BE, Alachkar N, Kraus ES, Desai NM, Locke JE, Racusen LC, Segev DL, Montgomery RA. Eculizumab and splenectomy as salvage therapy for severe antibody-mediated rejection after HLA-incompatible kidney transplantation. *Transplantation* 2014; **98**: 857-863 [PMID: 25121475 DOI: 10.1097/TP.0000000000000298]

112 **Loupy A**, Viglietti D, Mengel M. Complement inhibition in HLA-incompatible kidney transplants: persisting antibody-mediated injury despite marked decrease of clinical ABMR. *Am J Transplant* 2015; **15**: 1139-1140 [PMID: 25731892 DOI: 10.1111/ajt.13172]

113 **Vernon KA**, Gale DP, de Jorge EG, McLean AG, Galliford J, Pierides A, Maxwell PH, Taube D, Pickering MC, Cook HT. Recurrence of complement factor H-related protein 5 nephropathy in a renal transplant. *Am J Transplant* 2011; **11**: 152-155 [PMID: 21114651 DOI: 10.1111/j.1600-6143.2010.03333.x]

114 **Loirat C**, Fremeaux-Bacchi V. Hemolytic uremic syndrome recurrence after renal transplantation. *Pediatr Transplant* 2008; **12**: 619-629 [PMID: 18482212 DOI: 10.1111/j.1399-3046.2008.00910.x]

115 **Zuber J**, Le Quintrec M, Sberro-Soussan R, Loirat C, Frémeaux-Bacchi V, Legendre C. New insights into postrenal transplant hemolytic uremic syndrome. *Nat Rev Nephrol* 2011; **7**: 23-35 [PMID: 21102542 DOI: 10.1038/nrneph.2010.155]

116 **Noris M**, Remuzzi G. Overview of complement activation and regulation. *Semin Nephrol* 2013; **33**: 479-492 [PMID: 24161035 DOI: 10.1016/j.semnephrol.2013.08.001]

117 **Zuber J**, Le Quintrec M, Krid S, Bertoye C, Gueutin V, Lahoche A, Heyne N, Ardissino G, Chatelet V, Noël LH, Hourmant M, Niaudet P, Frémeaux-Bacchi V, Rondeau E, Legendre C, Loirat C; French Study Group for Atypical HUS. Eculizumab for atypical hemolytic uremic syndrome recurrence in renal transplantation. *Am J Transplant* 2012; **12**: 3337-3354 [PMID: 22958221 DOI: 10.1111/j.1600-6143.2012.04252.x]

118 **Verhave JC**, Westra D, van Hamersvelt HW, van Helden M, van de Kar NC, Wetzels JF. Living kidney transplantation in adult patients with atypical haemolytic uraemic syndrome. *Neth J Med* 2013; **71**: 342-347 [PMID: 24038559]

119 **Goicoechea de Jorge E**, Caesar JJ, Malik TH, Patel M, Colledge M, Johnson S, Hakobyan S, Morgan BP, Harris CL, Pickering MC, Lea SM. Dimerization of complement factor H-related proteins modulates complement activation in vivo. *Proc Natl Acad Sci USA* 2013; **110**: 4685-4690 [PMID: 23487775 DOI: 10.1073/pnas.1219260110]

120 **Kwon T**, Dragon-Durey MA, Macher MA, Baudouin V, Maisin A, Peuchmaur M, Fremeaux-Bacchi V, Loirat C. Successful pre-transplant management of a patient with anti-factor H autoantibodies-associated haemolytic uraemic syndrome. *Nephrol Dial Transplant* 2008; **23**: 2088-2090 [PMID: 18326881 DOI: 10.1093/ndt/gfn063]

121 **Waters AM**, Pappworth I, Marchbank K, Bockenhauer D, Tullus K, Pickering MC, Strain L, Sebire N, Shroff R, Marks SD, Goodship TH, Rees L. Successful renal transplantation in factor H autoantibody associated HUS with CFHR1 and 3 deficiency and CFH variant G2850T. *Am J Transplant* 2010; **10**: 168-172 [PMID: 19951285 DOI: 10.1111/j.1600-6143.2009.02870.x]

122 **Zimmerhackl LB**, Hofer J, Cortina G, Mark W, Würzner R, Jungraithmayr TC, Khursigara G, Kliche KO, Radauer W. Prophylactic eculizumab after renal transplantation in atypical hemolytic-uremic syndrome. *N Engl J Med* 2010; **362**: 1746-1748 [PMID: 20445192 DOI: 10.1056/NEJMc1001060]

123 **Román-Ortiz E**, Mendizabal Oteiza S, Pinto S, López-Trascasa M, Sánchez-Corral P, Rodríguez de Cordoba S. Eculizumab long-term therapy for pediatric renal transplant in aHUS with CFH/CFHR1 hybrid gene. *Pediatr Nephrol* 2014; **29**: 149-153 [PMID: 23982707 DOI: 10.1007/s00467-013-2591-8]

124 **Nester C**, Stewart Z, Myers D, Jetton J, Nair R, Reed A, Thomas C, Smith R, Brophy P. Pre-emptive eculizumab and plasmapheresis for renal transplant in atypical hemolytic uremic syndrome. *Clin J Am Soc Nephrol* 2011; **6**: 1488-1494 [PMID: 21617085 DOI: 10.2215/CJN.10181110]

125 **Krid S**, Roumenina LT, Beury D, Charbit M, Boyer O, Frémeaux-Bacchi V, Niaudet P. Renal transplantation under prophylactic eculizumab in atypical hemolytic uremic syndrome with CFH/CFHR1 hybrid protein. *Am J Transplant* 2012; **12**: 1938-1944 [PMID: 22494769 DOI: 10.1111/j.1600-6143.2012.04051.x]

126 **Pérez-Caballero D**, González-Rubio C, Gallardo ME, Vera M, López-Trascasa M, Rodríguez de Córdoba S, Sánchez-Corral P. Clustering of missense mutations in the C-terminal region of factor H in atypical hemolytic uremic syndrome. *Am J Hum Genet* 2001; **68**: 478-484 [PMID: 11170895 DOI: 10.1086/318201]

127 **Licht C**, Heinen S, Józsi M, Löschmann I, Saunders RE, Perkins SJ, Waldherr R, Skerka C, Kirschfink M, Hoppe B, Zipfel PF. Deletion of Lys224 in regulatory domain 4 of Factor H reveals a novel pathomechanism for dense deposit disease (MPGN II). *Kidney Int* 2006; **70**: 42-50 [PMID: 16612335 DOI: 10.1038/sj.ki.5000269]

128 **West CD**, Witte DP, McAdams AJ. Composition of nephritic factor-generated glomerular deposits in membranoproliferative glomerulonephritis type 2. *Am J Kidney Dis* 2001; **37**: 1120-1130 [PMID: 11382679 DOI: 10.1053/ajkd.2001.24511]

129 **Dragon-Durey MA**, Sethi SK, Bagga A, Blanc C, Blouin J, Ranchin B, André JL, Takagi N, Cheong HI, Hari P, Le Quintrec M, Niaudet P, Loirat C, Fridman WH, Frémeaux-Bacchi V. Clinical features of anti-factor H autoantibody-associated hemolytic uremic syndrome. *J Am Soc Nephrol* 2010; **21**: 2180-2187 [PMID: 21051740 DOI: 10.1681/ASN.2010030315]

130 **Khandelwal P**, Gupta A, Sinha A, Saini S, Hari P, Dragon Durey MA, Bagga A. Effect of plasma exchange and immunosuppressive medications on antibody titers and outcome in anti-complement factor H antibody-associated hemolytic uremic syndrome. *Pediatr Nephrol* 2015; **30**: 451-457 [PMID: 25217328 DOI: 10.1007/s00467-014-2948-7]

131 **Noris M**, Galbusera M, Gastoldi S, Macor P, Banterla F, Bresin E, Tripodo C, Bettoni S, Donadelli R, Valoti E, Tedesco F, Amore A, Coppo R, Ruggenenti P, Gotti E, Remuzzi G. Dynamics of complement activation in aHUS and how to monitor eculizumab therapy. *Blood* 2014; **124**: 1715-1726 [PMID: 25037630 DOI: 10.1182/blood-2014-02-558296]

132 **Tanimoto T**, Oshima Y, Kami M. Eculizumab in atypical hemolytic-uremic syndrome. *N Engl J Med* 2013; **369**: 1378-1379 [PMID: 24088109 DOI: 10.1056/NEJMc1308826]

133 **Zuber J**, Fakhouri F, Roumenina LT, Loirat C, Frémeaux-Bacchi V; French Study Group for aHUS/C3G. Use of eculizumab for atypical haemolytic uraemic syndrome and C3 glomerulopathies. *Nat Rev Nephrol* 2012; **8**: 643-657 [PMID: 23026949 DOI: 10.1038/nrneph.2012.214]

134 **Sheerin NS**, Kavanagh D, Goodship TH, Johnson S. A national specialized service in England for atypical haemolytic uraemic syndrome-the first year's experience. *QJM* 2016; **109**: 27-33 [PMID: 25899302 DOI: 10.1093/qjmed/hcv082]

135 **Wetzels JF**, van de Kar NC. Discontinuation of eculizumab maintenance treatment for atypical hemolytic uremic syndrome. *Am J Kidney Dis* 2015; **65**: 342 [PMID: 25616634 DOI: 10.1053/j.ajkd.2014.04.039]

136 **Ardissino G**, Possenti I, Tel F, Testa S, Salardi S, Ladisa V. Discontinuation of eculizumab treatment in atypical hemolytic uremic syndrome: an update. *Am J Kidney Dis* 2015; **66**: 172-173 [PMID: 26111906 DOI: 10.1053/j.ajkd.2015.04.010]

137 **Povey H**, Vundru R, Junglee N, Jibani M. Renal recovery with eculizumab in atypical hemolytic uremic syndrome following prolonged dialysis. *Clin Nephrol* 2014; **82**: 326-331 [PMID: 23557793 DOI: 10.5414/CN107958]

138 **Licht C**, Greenbaum LA, Muus P, Babu S, Bedrosian CL, Cohen DJ, Delmas Y, Douglas K, Furman RR, Gaber OA, Goodship T, Herthelius M, Hourmant M, Legendre CM, Remuzzi G, Sheerin N, Trivelli A, Loirat C. Efficacy and safety of eculizumab in atypical hemolytic uremic syndrome from 2-year extensions of phase 2 studies. *Kidney Int* 2015; **87**: 1061-1073 [PMID: 25651368 DOI: 10.1038/ki.2014.423]

139 **Thurman JM**, Le Quintrec M. Targeting the complement cascade:

novel treatments coming down the pike. *Kidney Int* 2016; **90**: 746-752 [PMID: 27325183 DOI: 10.1016/j.kint.2016.04.018]
140 **Le KN**, Gibiansky L, van Lookeren Campagne M, Good J, Davancaze T, Loyet KM, Morimoto A, Strauss EC, Jin JY. Population Pharmacokinetics and Pharmacodynamics of Lampalizumab Administered Intravitreally to Patients With Geographic Atrophy. *CPT Pharmacometrics Syst Pharmacol* 2015; **4**: 595-604 [PMID: 26535160 DOI: 10.1002/psp4.12031]

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SYSTEMATIC REVIEW

Impact of machine perfusion of the liver on post-transplant biliary complications: A systematic review

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Abstract

AIM

To review the clinical impact of machine perfusion (MP) of the liver on biliary complications post-transplantation, particularly ischaemic-type biliary lesions (ITBL).

METHODS

This systematic review was performed in accordance with the Preferred Reporting Systematic Reviews and Meta-Analysis (PRISMA) protocol. The following databases were searched: PubMed, MEDLINE and Scopus. The keyword "liver transplantation" was used in combination with the free term "machine perfusion". Clinical studies reporting results of transplantation of donor human livers following *ex situ* or *in situ* MP were analysed. Details relating to donor characteristics, recipients, technique of MP performed and post-operative biliary complications (ITBL, bile leak and anastomotic strictures) were critically analysed.

RESULTS

Fifteen articles were considered to fit the criteria for this review. *Ex situ* normothermic MP was used in 6

studies, *ex situ* hypothermic MP in 5 studies and the other 4 studies investigated *in situ* normothermic regional perfusion (NRP) and controlled oxygenated rewarming. MP techniques which have *per se* the potential to alleviate ischaemia-reperfusion injury: Such as hypothermic MP and NRP, have also reported lower rates of ITBL. Other biliary complications, such as biliary leak and anastomotic biliary strictures, are reported with similar incidences with all MP techniques. There is currently less clinical evidence available to support normothermic MP as a mitigator of biliary complications following liver transplantation. On the other hand, restoration of organ to full metabolism during normothermic MP allows assessment of hepatobiliary function before transplantation, although universally accepted criteria have yet to be validated.

CONCLUSION

MP of the liver has the potential to have a positive impact on post-transplant biliary complications, specifically ITBL, and expand extended criteria donor livers utilisation.

Key words: Liver transplantation; *Ex situ* machine perfusion of the liver; Donation after circulatory death; Non-anastomotic intra-hepatic stricture; Ischemic-type biliary lesions; Extended criteria donors

Core tip: Post-transplant biliary complications are one of the main culprits responsible for the high patient morbidity following extended criteria donor liver transplantation. In its most severe form, ischaemic-type biliary lesions, can lead to graft failure and re-transplantation. Machine perfusion (MP) of the liver is a promising approach in reconditioning high-risk organs. Clinical studies have, so far, focussed on the impact of MP on hepatocellular function recovery and assessment. In this review we present the clinical evidence of the effect of MP on post-transplant biliary complications and discuss how, in the future, this approach can reduce these complications further.

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INTRODUCTION

Post-transplant biliary complications: The current scenario

Post-transplant biliary complications often require laborious and costly interventions, placing a heavy burden on health resources and adversely affecting patient outcomes[1,2]. The incidence of these complications is increasing as a result of the growing utilisation of extended criteria donor (ECD) organs, mainly from donation after circulatory death (DCD). Biliary complications such as biliary leak and anastomotic strictures are primarily related to surgical technicalities and are usually successfully managed with endoscopic procedures[3]. The most severe form of post-transplant biliary complication is non-anastomotic intrahepatic strictures (NAS). NAS is characterised by the occurrence of diffuse intra-hepatic strictures in the biliary tree and it was initially associated with hepatic artery thrombosis[4]. The ischaemic donor biliary tree was found to develop necrosis with fibrotic strictures, dilatations and potentially biliary casts[4]. Thereafter it was demonstrated that similar lesions occurred in the presence of a patent hepatic artery without evidence of recurrence of biliary disease. This entity was subsequently classified as ischaemic-type biliary lesion (ITBL)[5].

The reported incidence of ITBL is approximately 10%-30% for controlled DCD and 1%-3% for donation after brain death (DBD) organs[6-10]. Patients generally present with elevated liver function tests suggesting cholestasis (bilirubin, alkaline phosphatase and gamma-glutamyltransferase) within a few months of transplantation and may be asymptomatic initially. Initial work-up includes exclusion of hepatic artery thrombosis and anastomotic biliary strictures. Imaging investigations consist of non-invasive magnetic resonance cholangiopancreatography (MRCP) and computed tomographic cholangiography, or direct cholangiographic methods, such as endoscopic retrograde cholangiopancreatography and percutaneous transhepatic cholangiography. Due to the high reliability of current non-invasive imaging techniques in diagnosing biliary strictures, invasive procedures are currently reserved for scenarios where an intervention is planned, such as stricture dilatation, stenting or stone extraction[11,12]. With ITBL, imaging confirms the presence of fibrotic strictures, in most cases located around the bifurcation of the common bile duct leading to dilatation of the intra-hepatic biliary system[1,8]. Figure 1 illustrates these typical imaging features of ITBL following liver transplantation. The obstructive strictures cause cholestasis with formation of sludge and casts that predispose to cholangitis, frequently requiring surgical or endoscopic intervention. Despite these measures, approximately 50% of patients with ITBL require re-transplantation or die[13].

Although the pathogenesis of ITBL is still not fully understood a growing body of evidence suggest that it is partially associated with ischaemia-reperfusion injury (IRI)[14,15]. Noack *et al*[16] in a well-designed *in-vitro* study using rat-derived bile duct cells showed that they were more resistant to anoxia than hepatocytes, however during reoxygenation they produced higher amounts of reactive oxygen species (ROS). This was associated with increased rates of bile duct cell death when compared to hepatocytes[16]. It has been shown that mitochondrial ischaemic induced injury leads to ROS production during reperfusion which in turn causes

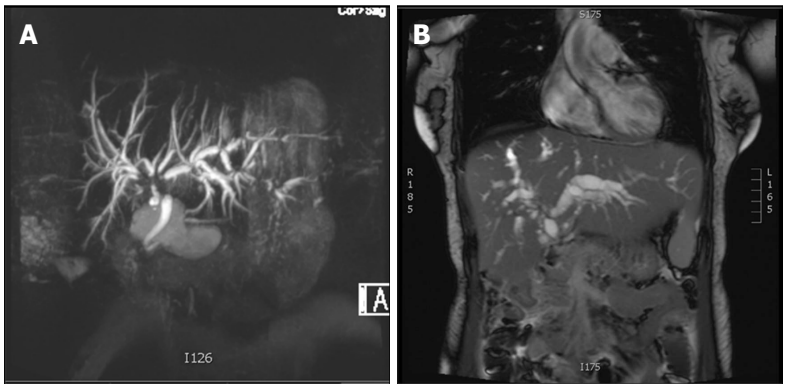

Figure 1 Magnetic resonance cholangiopancreatography images of ischemic-type biliary lesions following liver transplantation. The images show two recipients of livers from donation after circulatory death donors that developed ischemic-type biliary lesions within 60 d following transplantation. Hepatic artery thrombosis and anastomotic biliary strictures were ruled out. A: A typical lesion is seen affecting the bifurcation of the common hepatic bile duct with moderate dilatation of the intrahepatic biliary tree; B: The image shows strictures at the bifurcation of the common hepatic bile duct, diffuse intra-hepatic strictures and a severe dilatation of the intrahepatic biliary tree.

oxidative injury and activation of the inflammatory cascade[17,18]. Conversely, clinical series have reported severe injury to the biliary epithelium just after cold static storage[19,20]. Garcia-Valdecasas *et al*[15] using a porcine transplantation model suggested a direct relationship between prolonged ischaemic times and cell injury. Indeed, other clinical series have confirmed the association of longer cold ischaemic time (CIT) and higher rates of ITBL[21-24]. A similar relationship has been observed with warm ischemic time in DCD liver transplantation[15,25]. A large clinical series of donor bile duct biopsies before liver transplantation showed similar injury to the biliary epithelium after static cold storage (SCS), and that it was exacerbated after reperfusion; however, this did not correlate with the development of ITBL[26]. Nevertheless, the authors reported a strong association between ITBL and damage to the peribiliary vascular plexus and peribiliary glands. As progenitor biliary cells are known to reside in the peribiliary glands, the former finding suggests an association between ITBL and an attenuated regenerative capacity of the biliary epithelium[26,27]. Ischaemic injury is likely to play a major role in ITBL pathogenesis, although other factors have also been shown to be implicated. Immunological mediated injury to the biliary epithelium has been associated with ITBL[28]. It may be the result of direct immunological damage to the biliary epithelium *via* a rejection reaction[29]; or, indirect, secondary to the development of arteriopathy[29,30]. This cross reactivity is described in scenarios of cytomegalovirus infection[30], ABO incompatibility[31] and transplantation for primary sclerosing cholangitis[1]. Bile salt toxicity has also been investigated as a potential cause for ITBL by having a direct detergent effect on phospholipid cellular membranes of the biliary epithelium[28]. Flushing of the biliary tree during organ procurement is necessary in order to remove all bile salts that could damage cholangiocytes[5,28]. Furthermore, an imbalance in the post-transplant bile composition, with a higher bile salt/phospholipid ratio, due to inefficient ATP-dependent biliary transporters has been suggested as a predictive factor for ITBL[32]. While detail of the pathogenesis of ITBL is beyond the scope of this review, information on the implicated mechanisms can be found in a number of published reviews[9,28].

Machine perfusion of donor livers

The utilisation of DCD livers is increasing. In 2017, in the United Kingdom, they constituted 28% of the livers transplanted[33]. Furthermore, the rising prevalence of donor obesity (body mass index greater than 30 kg/m^2) and an ageing population continue to compound the risks to those livers[33]. These high-risk ECD organs are associated not only with a higher risk of graft dysfunction post-transplantation but also increased rates of ITBL[34]. Despite these disadvantages, their utilisation is required to tackle the ever-growing discrepancy between organ donor supply and demand. Machine perfusion (MP) of the liver is being developed as a means of assessment and reconditioning of ECD donors, potentially allowing for safer transplantation of these high-risk livers[34,35]. Different techniques of MP have been developed; it can be performed *in situ* during organ procurement or *ex situ* after the procedure. With regards to livers, the only technique of *in situ* MP described so far is normothermic regional perfusion (NRP)[8]. *Ex situ* MP protocols vary in terms of oxygenation (active or pre-charged oxygenation), perfusate temperature (hypothermic, subnormothermic, gradual rewarming and normothermic), timing of perfusion (preservation or end-ischemic) and via of organ perfusion (portal vein alone or dual portal vein and hepatic artery perfusion)[34,36].

Hypothermic machine perfusion (HMP) has been performed around 10 ℃ in most studies[37,38]. At this temperature liver metabolism is reduced; and, passive oxygen delivery by diffusion in an oxygen carrier-free perfusate is enough to support the organ[39]. The first published clinical series employed pre-charged oxygen delivery to the organs[37], technique that was later followed by active oxygenation of the perfusate[40].

Hypothermic oxygenated MP can be performed *via* portal vein alone (HOPE) or *via* portal vein and hepatic artery (dual hypothermic oxygenated perfusion - D-HOPE)[41-43]. Both techniques have shown the capacity of improve mitochondrial oxidative function prior to rewarming, resulting in increased adenosine triphosphate (ATP) synthesis and a reduction in ROS production, oxidative tissue injury and activation of the inflammatory cascade[42,43].

Normothermic machine perfusion (NMP) maintains the organ at physiological temperatures (37 ℃) and therefore restores full metabolic activity. This enables the possibility of functional or viability assessment prior to transplantation, a major advantage of NMP when compared to other perfusion techniques[44,45]. It also opens up a window of opportunity for *ex situ* therapeutic interventions[34]. Furthermore, previous studies have reported on the safety of extended normothermic perfusion of organs, which may facilitate transportation and logistical management of busy transplant units[46]. However, potential drawbacks of NMP are that it requires obligatorily the inclusion of an oxygen carrier in the perfusate, and NMP inevitably induces reperfusion injury to some extent.

Subnormothermic machine perfusion (SMP) has been performed at around 20 ℃ in most studies. It encompasses purely SMP and the controlled oxygenated rewarming (COR) from 10 ℃ to 20 ℃[47,48]. The increase in temperature from HMP to SMP is suggested to be enough to increase liver metabolism to an extent that it would allow assessment of organ function without inducing the detrimental changes associated with organ reperfusion at normothermic temperatures[48]. Evidence for the clinical benefits is available for COR perfusions, it was associated with lower markers of hepatocellular injury after transplantation and enhanced graft function through the avoidance of subtle changes in organ temperature[47].

For DCD livers, there are encouraging reports of *in situ* oxygenated NRP. It has been successfully applied to controlled DCD donors (withdrawal of life support in patients with irreversible clinical conditions) and uncontrolled DCD (witnessed cardiac arrest without response to resuscitative measures)[8,49,50]. NRP limits ischaemia and prevents depletion of energy stores prior to SCS and this is suggested to be essential for uncontrolled DCD donors and beneficial for controlled DCD[8].

More recently, combinations of MP techniques have been shown to merge the advantages of individual protocols, enhancing the rescue of liver function what may potentially improve graft function after transplantation[51,52]. Despite differences between techniques, MP has the potential to limit ischaemic injury to the organ, thus offering a safer preservation environment and an opportunity for organ reconditioning which could mitigate IRI.

As discussed herein, the current evidence shows that cholangiocytes are more vulnerable to IRI than hepatocytes and that the pathogenesis for biliary injury goes beyond IRI. Therefore, investigation of the impact of MP on biliary function specifically, and not only on hepatocellular function, is fundamental. The aim of this review was to investigate the current clinical evidence available regarding the effect of MP on post-transplant biliary complications, focusing on ITBL.

MATERIALS AND METHODS

This systematic review was performed in accordance with the Preferred Reporting Systematic Reviews and Meta-Analysis (PRISMA) protocol[53].

The following databases were searched for the development of this review: PubMed, MEDLINE and Scopus. The keyword "liver transplantation" was used in combination with the free term "machine perfusion". The literature review was performed until June 20, 2018 and there were no limits on the date for inclusion of publications. The literature search strategy used for one database is presented in the Supplementary Table S1.

The screening and selection of articles were independently performed by two authors (Yuri L Boteon and Amanda PCS Boteon). There was no disagreement in study selection between authors. Manuscript titles that were not related to the main scope of the review were excluded. Full abstracts were then read and excluded if found not to be relevant to the review. Finally, full papers were assessed for eligibility and included in this review. The flow diagram for the literature selection process is shown in Figure 2.

Inclusion criteria were: (1) clinical studies reporting results of transplantation of donor human livers following *ex situ* or *in situ* MP; and (2) articles written in English and published. Exclusion criteria were: (1) absence of transplantation following MP; (2) exclusively animal models; (3) single case report; (4) review articles; and (5) articles not written in English.

Details relating to donor characteristics [type, age, donor risk index (DRI), warm ischaemic time (WIT), CIT], recipients [age, model for end-stage liver disease (MELD)], perfusion (type of perfusion, oxygenation, timings) and post-operative biliary complications (ITBL, leak and anastomotic strictures) were retrieved from each manuscript and critically analysed. Studies were assessed in terms of study design, methods and outcomes. No review protocol was registered before this review was started. No simplifications or assumptions were made, and any identified risk of bias is discussed throughout the review.

RESULTS

Fifteen articles were considered to fit the criteria for this review. A diagrammatic summary of the screening process is provided in Figure 2.

MP and ischemic type biliary lesions (ITBL)

Eight out of fifteen clinical studies utilised an end-

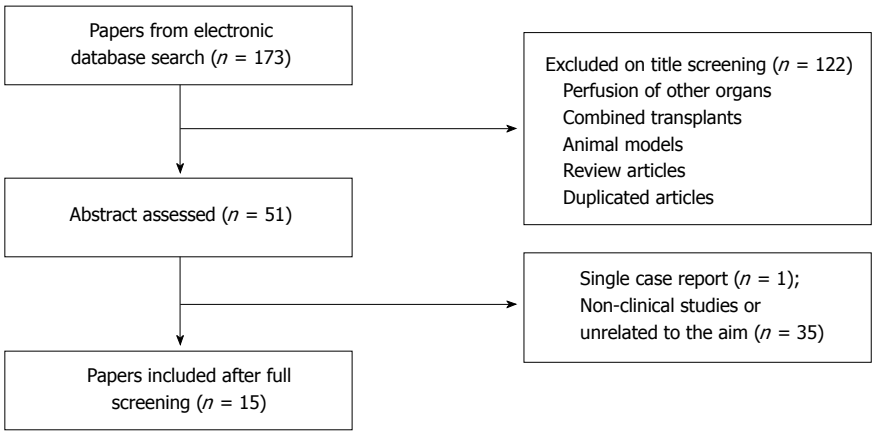

Figure 2 Study flow diagram for systematic review of the literature on the impact of machine perfusion of the liver and post-transplant biliary complications. Following literature search duplicate articles were excluded and the titles screened. The selected abstracts were then read and non-clinical studies or reports unrelated to the aim of the review were excluded.

ischemic model of MP (MP commenced after a variable period of SCS), 4 studies utilised preservation MP (MP from organ procurement up to transplantation) and 3 employed NRP. NMP was used in 6 studies, HMP in 5 studies and the other 4 studies investigated NRP and COR. HMP with active perfusate oxygenation (HOPE and D-HOPE) studies were seen to be currently focused on DCD organs and HMP with pre-charged oxygenation on DBD organs. NMP studies used both donor types, however preservation studies explored a higher proportion of DBD compared to DCD organs. The contrary was seen for end-ischemic NMP.

Donor and recipients characteristics, of the cases included in individual studies, are presented in Table 1. It also reports the rates of ITBL. Table 2 describes the incidence of bile leak and anastomotic biliary stricture within the different studies. Studies characteristics were described therein, as it was their design.

NMP and post-transplant biliary complications

The largest clinical trial involving NMP as a preservation strategy was recently published by Nasralla *et al*[46]. Following procurement, transplantable livers were randomised and allocated to the intervention group that had NMP up to the point of transplantation or a control group that had conventional SCS. From the 121 livers perfused, 87 were from DBD donors and 34 from DCD donors. Results did not shown differences in bile duct complications between groups, with one patient in each arm developing ITBL within the first year, both requiring re-transplantation. On MRCP, the rates of NAS were similar between groups for DBD (NMP 7.4% *vs* SCS 5.4%; $P = 0.678$) and DCD (NMP 11.1% *vs* SCS 26.3%; $P = 0.180$). The incidence of anastomotic strictures was also similar for DBD or DCD organs (NMP 40.7% *vs* SCS 41.8%; $P = 0.909$; and, NMP 48.1% *vs* SCS 57.9%; $P = 0.515$, respectively)[46].

Other clinical studies investigating NMP using a preservation approach[54-56] involved smaller patient numbers, the majority of which were from DBD donors, and did not specifically report the incidence of ITBL (Table 1). Ravikumar *et al*[55] published the first phase 1 clinical trial demonstrating the safety and feasibility of NMP in a preservation approach, as an alternative to SCS. In all, 20 donor livers (16 DBD and 4 DCD) were transplanted following NMP. The 30-day graft survival was similar to static cold stored livers and the median peak aspartate aminotransferase within the first 7 post-operative days was lower. In terms of biliary complications, the authors reported the occurrence of 4 cases of anastomotic biliary strictures in the NMP group[55].

The two studies of NMP after a period of SCS (end-ischaemic model) involved organs that were deemed too high risk for transplantation[57,58]. These studies predominantly used DCD livers and applied predefined viability criteria prior to transplantation. Mergental *et al*[57] did not observe any biliary complications at 7 mo of follow up post-transplantation. Watson *et al*[58] reported the occurrence of 4 cases of ITBL in 16 DCD liver transplants, of which 3 needed re-transplantation. The authors of the latter study concluded that that NMP per se does not prevent ITBL but may provide biomarkers to identify livers that are high risk, such as maximum bile pH > 7.5 and bile glucose ≤ 3 mmol/L or ≥ 10 mmol less than perfusate glucose[58].

HMP and post-transplant biliary complications

The first clinical study using HMP prior to transplantation was performed by Guarrera *et al*[37] Twenty DBD livers were perfused after a period of SCS in a non-actively oxygenated model of HMP. ITBL rate was reported as 5%, half of the incidence of the control matched cohort that was subjected to SCS. Additionally, there was one case of bile leak and 1 report of anastomotic biliary stricture[37]. The same approach was repeated later in a study of DBD livers declined by the United Network for Organ Sharing region for transplantation[59]. The authors found a significant decrease in the rate of biliary stricture in comparison with SCS (10% *vs* 33%, $P = 0.031$). One report of bile leak was noted in the HMP group and 3 in

Table 1 Comparison between donor, recipient, perfusion characteristics and the reported rates of ischemic-type biliary lesions

Ref.	Yr	Perfusion type	Timing MP	*n*	Donor age	Donor risk index	Recipient age	Recipient MELD	DBD (*n*)	DCD (*n*)	DBD ITBL (%)	DCD ITBL (%)	CIT (min)	Func. WIT (min)	Re-Tx (*n*)
Ex situ normothermic machine perfusion															
Nasralla *et al*[46]	2018	NMP	Preserv	121	56 (16-84)	1.7[3]	55	13 (6-35)	87	34	7.4	11.1	126	21	3
Selzner *et al*[54]	2016	NMP	Preserv	10	48 (17-75)	1.9	57	21 (8-40)	8	2	0	0	103	NA	0
Bral *et al*[56]	2017	NMP	Preserv	9	56 (14-71)	1.6 (0.9-2.7)	53 (28-67)	13 (9-32)	6	3	0	0	167 (95-293)	22	0
Ravikumar *et al*[55]	2016	NMP	Preserv	20	58 (21-85)	NA	NA	12 (7-27)	16	4	0	0	NA	21	0
Watson *et al*[58]	2018	NMP	End-Isc	22	57	2.3	NA	NA	6	16	0	25	386	12	3
Mergental *et al*[57]	2016	NMP	End-Isc	5	49 (29-54)	2.3	56 (47-66)	8 (8-13)	1	4	0	0	422	28	0
Ex situ hypothermic non-oxygenated machine perfusion															
Guarrera *et al*[59]	2015	HMP	End-Isc	31	57 (± 18)[1]	1.9 (± 0.5)[1]	57 (± 8.0)[1]	19 (± 5.9)[1]	31	0	9.7	NA	558	NA	0
Guarrera *et al*[37]	2010	HMP	End-Isc	20	39 (± 2.5)[1]	NA	55 (± 6.2)[1]	17 (± 7.4)[1]	20	0	5	NA	306	26	0
Ex situ hypothermic oxygenated machine perfusion															
van Rijn *et al*[43]	2017	DHOPE	End-Isc	10	53 (47-57)	1.9 (1.5-2.2)	57 (54-62)	16 (15-22)	0	10	NA	10	331	15	0
Dutkowski *et al*[38]	2015	HOPE	End-Isc	25	54 (36-63)	NA	60 (57-64)	13 (9-15)	0	25	NA	0	188 (141-264)	31 (26-36)	0
Dutkowski *et al*[40]	2014	HOPE	End-Isc	8	54 (NA)	2.2 (NA)	60 (NA)	12 (NA)	0	8	NA	0	141 (NA)	31 (22-41)	0
In situ normothermic regional perfusion															
De Carlis *et al*[60]2	2017	NRP	NRP	7	48[1]	NA	54[1]	10.6[1]	0	7	NA	0	414[1]	33	0
Oniscu *et al*[49]	2014	NRP	NRP	11	46 (16-74)	NA	68 (43-74)	NA	0	11	NA	0	389 (169-450)	26 (13-48)	0
Minambres *et al*[50]	2017	NRP	NRP	11	58 (50-67)	NA	55 (± 13)[1]	NA	0	11	NA	0	266 (± 82.7)[1]	12 (11-16)	0
Controlled oxygenated rewarming															
Hoyer *et al*[47]	2016	COR	End-Isc	6	58 (51-71)	1.9 (1.5-2.5)	52 (43-65)	18 (11-23)	6	0	0	NA	508 (369-870)	NA	0

[1]Data presented as median or median (± SD), if available. Otherwise, all data presented as median (Interquartile range); [2]Combined hypothermic oxygenated machine perfusion after normothermic regional perfusion. Six uncontrolled DCD were included in this study; [3]Eurotransplant DRI. MP: Machine perfusion; MELD: Model for end stage liver disease; DBD: Donation after brain death; DCD: Donation after circulatory death; ITBL: Ischemic-type biliary lesions; CIT: Cold ischemic time; Func; WIT: Functional warm ischemic time; Re-Tx: Re-transplantation; NA: Not applicable or not available; Preserv: Preservation; End-Isc: End ischemic; NMP: Normothermic machine perfusion; HMP: Hypothermic machine perfusion; DHOPE: Dual vessel hypothermic oxygenated machine perfusion; HOPE: Hypothermic oxygenated machine perfusion; NRP: Normothermic regional perfusion; COR: Controlled oxygenated rewarming.

SCS respectively (Table 2).

Following these initial studies, the Zurich group developed the concept of HOPE, with active oxygenation of the perfusate, and applied this MP strategy to DCD donors[38,40]. Their first clinical trial was published in 2015, reporting the results of transplantation of 25 DCD livers[38]. The authors reported no cases of ITBL at one year follow-up of patients who received perfused DCD livers, whereas control livers subjected to SCS developed a significantly higher rate of ITBL (0/25 *vs* 11/50, $P = 0.013$). The same benefit of HOPE was not seen for extra-hepatic biliary complications, as the reported rates of leaks and anastomotic strictures were similar (HOPE 5/25 *vs* Control 12/50)[38].

The Groningen group published the first clinical series using D-HOPE in 2017[43]. Ten DCD livers were transplanted following two hours of D-HOPE, one patient in the perfusion group developed ITBL compared to 7 out of 20 in the control group. The case in the D-HOPE group was described as NAS in segments II and III of the liver and was managed with endoscopic stenting. Three control livers which developed ITBL required re-transplantation. The rate of anastomotic biliary strictures was comparable between groups (D-HOPE 2 *vs* Control 3, $P = 1.000$) as was the reported rate of biliary cast formation (D-HOPE 3 *vs* Control 3, $P = 0.372$)[43].

Normothermic reginal perfusion and post-transplant biliary complications

The first series reporting the results for transplantation of livers following NRP was published in 2014 by Oniscu *et al*[49] The authors reported the results of transplantation of

Table 2 Prevalence of bile leak and anastomotic biliary strictures between clinical studies using different techniques of machine perfusion of donor livers

Ref.	Yr	Study design	Perfusion type	Timing machine perfusion	*n*	DBD (*n*)	DCD (*n*)	Bile leak (*n*)	Anastomotic stricture (*n*)
Ex situ normothermic machine perfusion									
Nasralla *et al*[46]	2018	RCT	NMP	Preservation	121	87	34	0	0
Selzner *et al*[54]	2016	PS	NMP	Preservation	10	8	2	0	0
Bral *et al*[56]	2017	PS	NMP	Preservation	9	6	3	0	0
Ravikumar *et al*[55]	2016	PS	NMP	Preservation	20	16	4	0	4 (DBD)
Watson *et al*[58]	2018	DS	NMP	End-Ischaemic	22	6	16	0	0
Mergental *et al*[57]	2016	DS	NMP	End-Ischaemic	5	1	4	0	0
Ex situ hypothermic non-oxygenated machine perfusion									
Guarrera *et al*[59]	2015	PS	HMP	End-Ischaemic	31	31	0	1	0
Guarrera *et al*[37]	2010	NCS	HMP	End-Ischaemic	20	20	0	1	1
Ex situ hypothermic oxygenated machine perfusion									
van Rijn *et al*[43]	2017	PS	DHOPE	End-Ischaemic	10	0	10	0	2
Dutkowski *et al*[38]	2015	PS	HOPE	End-Ischaemic	25	0	25	5 (in total)	
Dutkowski *et al*[40]	2014	PS	HOPE	End-Ischaemic	8	0	8	1	1
In situ normothermic regional perfusion									
De Carlis *et al*[60]1	2017	DS	NRP	NRP	7	0	7*	0	1
Oniscu *et al*[49]	2014	DS	NRP	NRP	11	0	11	1	1
Minambres *et al*[50]	2017	DS	NRP	NRP	11	0	11	NA	NA
Controlled Oxygenated Rewarming									
Hoyer *et al*[47]	2016	PS	COR	End-Ischaemic	6	6	0	NA	NA

[1]Combined hypothermic oxygenated machine perfusion after normothermic regional perfusion. Six uncontrolled DCD were included in this study. RCT: Randomised controlled trial; PS: Single-arm non-randomised pilot study; DS: Descriptive study; NCS: Non-randomised cohort studies; DBD: Donation after brain death; DCD: Donation after circulatory death; NA: Not applicable or not available; NMP: Normothermic machine perfusion; HMP: Hypothermic machine perfusion; DHOPE: Dual vessel hypothermic oxygenated machine perfusion; HOPE: Hypothermic oxygenated machine perfusion; NRP: Normothermic regional perfusion; COR: Controlled oxygenated rewarming.

11 controlled DCD livers, with a minimum follow-up of 3 mo, with no clinical or radiological evidence of ITBL. One patient developed an anastomotic stricture, treated endoscopically by cholangio-pancreatography (exact intervention performed is not described), and one patient had a bile leak[49]. Minambres *et al*[50] 2017, studying controlled DCD transplantation after NRP, reported no cases of ITBL after 1-year follow-up. De Carlis *et al*[60] 2017 performed NRP on 1 controlled DCD liver and 6 uncontrolled DCD. On arrival at the transplant centre, the livers were subjected to D-HOPE until transplantation. No cases of ITBL were observed and one patient had an anastomotic biliary stricture 45 d after transplantation, which was successfully treated with endoscopic stenting[60]. In terms of SMP, Hoyer *et al*[47] reported transplantation of 6 DBD livers following COR perfusion. No biliary complications were reported within a follow-up period of six months.

DISCUSSION

Post-transplant biliary complications are associated with high rates of morbidity and re-transplantation and are a major obstacle to the wider clinical utilisation of ECD livers. There is a growing body of evidence suggesting that MP can offer safer organ preservation when compared to SCS, and also offer an opportunity for organ assessment and/or reconditioning prior to transplantation[38,43,46,49,58]. In this review we have assessed the available literature investigating the impact of MP on post-transplant biliary complications, with special reference to ITBL. MP techniques which have per se the potential to alleviate IRI, such as HMP and NRP, have also reported lower rates of ITBL. Other biliary complications, such as biliary leak and anastomotic biliary strictures, are reported with similar incidences with all MP techniques.

Liver IRI is thought to be a major driver of biliary injury and, therefore, it is associated with complications following transplantation. More specifically, during ischemia, without oxygen as a terminal acceptor of electrons in the electron transport chain, succinate accumulates and acts as a store for electrons. Succinate oxidation during the early stage of reperfusion, blocks mitochondrial complex II of the electron transport chain resulting in a reverse flow of electrons towards mitochondrial complex I leading to accentuated leakage of electrons, and generation of ROS[61]. Various experimental findings using the HOPE technique have shown that oxygen at hypothermic temperatures is able to promote mitochondrial metabolism of succinate prior to reperfusion[36,42,62]. By re-establishing adequate mitochondrial oxidative function, HOPE is able to recover ATP stores, since during hypothermia mitochondria have lower energy requirements due to a minimum activation of the organ metabolism. Therefore, mechanistically, HOPE can in theory prevent the reverse flow of electrons during reperfusion, ROS generation and activation of the inflammatory cascade[36]. These factors may mitigate IRI, which would be beneficial not only

for hepatocellular function but also for the prevention of further biliary injury.

Extensive research focussing on the effect of oxygenated HMP on post-transplant biliary complications has been performed by the Groningen group. In a recent publication exploring the effects of D-HOPE on bile duct biopsies from a previous published series of cases, they showed less injury to deep and periluminal peribiliary glands after reperfusion during transplantation in the perfused group in comparison with SCS control livers[43,63]. Peribiliary glands have been described as stores for biliary progenitor cells, therefore injury to them would potentially decrease the regenerative capacity of the biliary system[64,65]. The authors acknowledge that definitive evidence to support this would require a clinical randomized trial that has since been initiated at their centre[63].

There is currently less clinical evidence available to support NMP as a mitigator of biliary complications following liver transplantation[58]. Preservation NMP shortens the ischaemic injury and offers a more physiological environment for the organ before transplantation. Nevertheless, as previously discussed, the injury to biliary cells would not be restricted to an ischaemic mechanism but may also be worsened during reperfusion. This observation could imply that NMP is of limited benefit in terms of biliary complications, since biliary injury may worsen during organ reperfusion on the machine and is not prevented or mitigated beforehand. NMP restores the full metabolism of the organ, resulting inevitably in the production and circulation of ROS and potential activation of the inflammatory response leading to tissue injury[66]. On the other hand, restoration of organ to full metabolism allows assessment of hepatobiliary function before transplantation, although universally accepted criteria have yet to be validated[35]. Watson *et al*[58] suggested bile pH and glucose content as markers of bile duct injury and associated those with the development of ITBL, however the authors recognise that NMP was not able to prevent biliary damage.

Promisingly, *in situ* NRP has shown excellent biliary outcomes after transplantation of DCD livers[49,50,60]. NRP may potentially prevent ischaemic injury and deterioration of ATP stores during organ procurement. Additionally, NRP allows assessment of the liver metabolism even before SCS[8]. Despite these points, there is no mechanistic evidence available to demonstrate any alleviation in IRI after reperfusion. It is also difficult to rule out the possibility that this beneficial effect was as a result of a potential selection bias when recruiting organs for transplantation during the procedure.

The present body of work has several limitations. First and foremost, donor livers and recipient characteristics as well as MP technique protocols exhibit a high degree of variability between studies. So far, there has been no standardisation in terms of methodology and reporting of results. Furthermore, some studies neglect to report important data variables, such as DRI, recipient age, recipient MELD and CIT. All these features are presented in Table 1 to allow an unbiased assessment of the retrieved information by the readers. Additionally, few clinical studies from each MP technique are available and most of them are originated from small pilot studies, which limit definitive interpretation of the data. MRCP was performed in some of the studies at different post-operative periods, but the significance of findings without clinical correlation is not clear. In addition, they have focussed mainly on evaluation of hepatocellular function rather than biliary function and injury. Despite the subject of this review being a relevant topic with important clinical implications, the direct effects of MP on biliary tree integrity are still relatively under-researched. More clinical randomized trials will be reported in the field in the next few years.

Higher rates of ITBL following transplantation of ECD livers, mainly DCD, place a major restraint on the wider use of these marginal livers. Each technique of MP offers different advantages and they all have the potential to tackle this problem. A feasibility study has shown that a combination of HOPE and NMP increased the rescue of metabolic parameters of high-risk ECD organs[52]. This approach may derive benefits from the individual methods, thus optimising gains also in terms of biliary function. Pharmacological interventions during NMP may potentially alleviate IRI, positively affecting biliary cells[67], and may have a direct effect on post-transplant biliary complications. Supplementation of the perfusate with substances that may induce proliferation and maturation of progenitor cells from peribiliary glands may be a feasible option to be considered[9]. We hypothesize that therapies promoting increase in secretion of phospholipids and cholesterol in the bile would equilibrate the phospholipids/bile salts balance mitigating further injury to the biliary tree. Although promising, these are options that still need to be explored in future studies. A diagrammatic summary of the current and future impact of MP on ITBL is presented in Figure 3.

The high incidence of post-transplant biliary complications, specifically ITBL, is a major constraint to wider utilisation of ECD livers. MP is currently considered a promising tool to increase ECD utilisation. However, the focus of most of the studies up to date has been the effect of MP on hepatocellular function. In this review we explored the clinical evidence currently available for the impact of MP on post-transplant biliary complications. From those studies that have looked at the effects of MP on biliary integrity, oxygenated HMP and NRP studies have been shown to exhibit better postoperative biliary outcomes in comparison with NMP and non-oxygenated HMP. However, larger clinical studies and randomised clinical trials powered for the occurrence of biliary complications as a primary endpoint are needed to confirm this data.

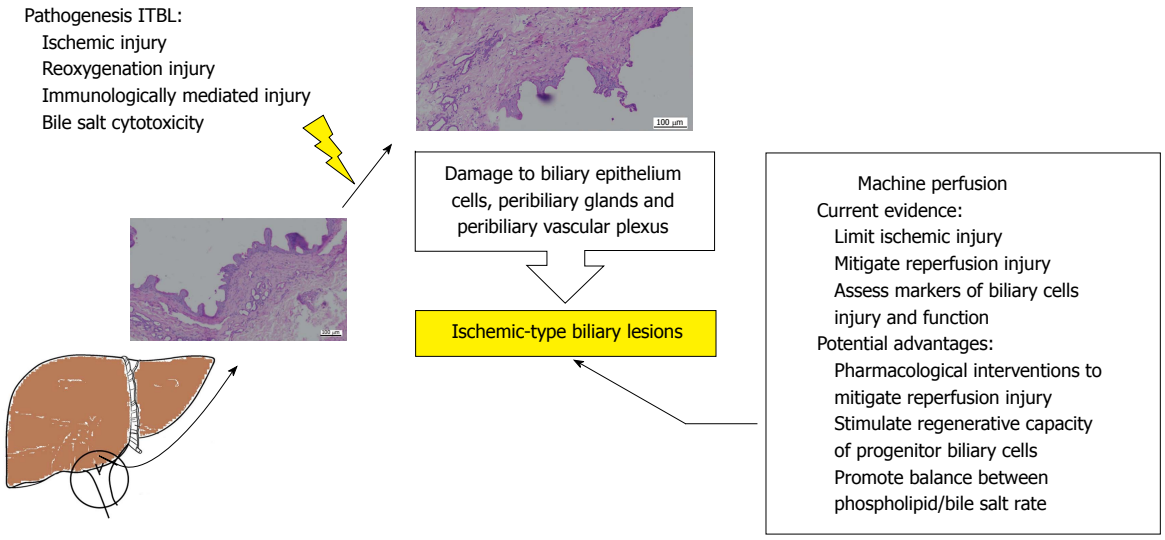

Figure 3 Diagrammatic summary of the current evidence for the impact of machine perfusion of the liver on post-transplant ischemic-type biliary lesions and future perspectives. The current evidence suggests that ischaemic-type biliary lesions (ITBL) have a multifactorial pathogenesis. These diverse factors lead to injury to the biliary epithelium, peribiliary glands and peribiliary vascular plexus. Currently, there is evidence for the potential benefits of machine perfusion on post-transplant ITBL. The figure summarises those and possible future interventions that could enhance increase these benefits further.

ARTICLE HIGHLIGHTS

Research background

The ever-growing discrepancy between donor organ availability and patients on the transplant waiting list has led to increased acceptance of extended criteria donors (ECD). However, ECD liver transplantation, mainly donation after circulatory death, is associated with poor patient and graft outcome. A major factor is the increased risk of biliary complications, in particular ischaemic type biliary lesions (ITBL). Machine perfusion (MP) of the liveris a promising tool to recondition ECD organs prior to transplantation. Therefore investigation of the impact of MP on post-transplant biliary complications is a highly relevant topic.

Research motivation

Understanding the current evidence available for the effect of MP on post-transplant biliary complications, in particular ITBL, may guide further studies in this field.

Research objectives

Revise the current clinical evidence available regarding the effect of MP on post-transplant biliary complications, focusing on ITBL.

Research methods

A systematic review was carried out with literature searches in PubMed, MEDLINE and Scopus databases. The keyword "liver transplantation" was used in combination with the free term "machine perfusion". Only clinical studies reporting results of transplantation of donor human livers following ex situ or in situ MP were included.

Research results

MP techniques which have demonstrated the potential to mitigate ischaemia reperfusion injury, such as *ex situ* oxygenated hypothermic MP and *in situ* normothermic regional perfusion, have also reported lower rates of ITBL. Other biliary complications, such as biliary leak and anastomotic biliary strictures, are reported with similar incidences with all MP techniques. Clinical studies have focused on evaluation of hepatocellular function rather than biliary function and injury so far. The direct effects of MP on biliary tree integrity are still relatively under-researched and further studies are needed.

Research conclusions

Post-transplant biliary complications are a major obstacle to the wider utilisation of ECD livers. MP has the potential to have a positive impact on this issue, specifically ITBL, and expand ECD livers utilisation. Mechanistically, mitigation of ischaemia-reperfusion injury appears to be the key mechanism involved.

Research perspectives

Supplementation of the perfusion fluid during *ex situ* MP with drugs can stimulate protective/regenerative mechanisms of the biliary tree. Pharmacological strategies may potentially modulate progenitor cells proliferation and equilibrate the phospholipid/bile salts balance in the bile.

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REFERENCES

1 **Buis CI**, Verdonk RC, Van der Jagt EJ, van der Hilst CS, Slooff MJ, Haagsma EB, Porte RJ. Nonanastomotic biliary strictures after liver

transplantation, part 1: Radiological features and risk factors for early vs. late presentation. *Liver Transpl* 2007; **13**: 708-718 [PMID: 17457932 DOI: 10.1002/lt.21166]
2 **Wojcicki M**, Milkiewicz P, Silva M. Biliary tract complications after liver transplantation: a review. *Dig Surg* 2008; **25**: 245-257 [PMID: 18628624 DOI: 10.1159/000144653]
3 **DeOliveira ML**, Jassem W, Valente R, Khorsandi SE, Santori G, Prachalias A, Srinivasan P, Rela M, Heaton N. Biliary complications after liver transplantation using grafts from donors after cardiac death: results from a matched control study in a single large volume center. *Ann Surg* 2011; **254**: 716-722; discussion 722-723 [PMID: 22042467 DOI: 10.1097/SLA.0b013e318235c572]
4 **Zajko AB**, Campbell WL, Logsdon GA, Bron KM, Tzakis A, Esquivel CO, Starzl TE. Cholangiographic findings in hepatic artery occlusion after liver transplantation. *AJR Am J Roentgenol* 1987; **149**: 485-489 [PMID: 3303874 DOI: 10.2214/ajr.149.3.485]
5 **Sanchez-Urdazpal L**, Gores GJ, Ward EM, Maus TP, Wahlstrom HE, Moore SB, Wiesner RH, Krom RA. Ischemic-type biliary complications after orthotopic liver transplantation. *Hepatology* 1992; **16**: 49-53 [PMID: 1618482 DOI: 10.1002/hep.1840160110]
6 **Chan EY**, Olson LC, Kisthard JA, Perkins JD, Bakthavatsalam R, Halldorson JB, Reyes JD, Larson AM, Levy AE. Ischemic cholangiopathy following liver transplantation from donation after cardiac death donors. *Liver Transpl* 2008; **14**: 604-610 [PMID: 18433032 DOI: 10.1002/lt.21361]
7 **Laing RW**, Scalera I, Isaac J, Mergental H, Mirza DF, Hodson J, Wilkin RJ, Perera MT, Muiesan P. Liver Transplantation Using Grafts From Donors After Circulatory Death: A Propensity Score-Matched Study From a Single Center. *Am J Transplant* 2016; **16**: 1795-1804 [PMID: 26725645 DOI: 10.1111/ajt.13699]
8 **Hessheimer AJ**, Cárdenas A, García-Valdecasas JC, Fondevila C. Can we prevent ischemic-type biliary lesions in donation after circulatory determination of death liver transplantation? *Liver Transpl* 2016; **22**: 1025-1033 [PMID: 27082839 DOI: 10.1002/lt.24460]
9 **de Vries Y**, von Meijenfeldt FA, Porte RJ. Post-transplant cholangiopathy: Classification, pathogenesis, and preventive strategies. *Biochim Biophys Acta* 2018; **1864**: 1507-1515 [PMID: 28645651 DOI: 10.1016/j.bbadis.2017.06.013]
10 **Jay CL**, Lyuksemburg V, Ladner DP, Wang E, Caicedo JC, Holl JL, Abecassis MM, Skaro AI. Ischemic cholangiopathy after controlled donation after cardiac death liver transplantation: a meta-analysis. *Ann Surg* 2011; **253**: 259-264 [PMID: 21245668 DOI: 10.1097/SLA.0b013e318204e658]
11 **Akbar A**, Tran QT, Nair SP, Parikh S, Bilal M, Ismail M, Vanatta JM, Eason JD, Satapathy SK. Role of MRCP in Diagnosing Biliary Anastomotic Strictures After Liver Transplantation: A Single Tertiary Care Center Experience. *Transplant Direct* 2018; **4**: e347 [PMID: 29796418 DOI: 10.1097/TXD.0000000000000789]
12 **Jorgensen JE**, Waljee AK, Volk ML, Sonnenday CJ, Elta GH, Al-Hawary MM, Singal AG, Taylor JR, Elmunzer BJ. Is MRCP equivalent to ERCP for diagnosing biliary obstruction in orthotopic liver transplant recipients? A meta-analysis. *Gastrointest Endosc* 2011; **73**: 955-962 [PMID: 21316670 DOI: 10.1016/j.gie.2010.12.014]
13 **Buis CI**, Hoekstra H, Verdonk RC, Porte RJ. Causes and consequences of ischemic-type biliary lesions after liver transplantation. *J Hepatobiliary Pancreat Surg* 2006; **13**: 517-524 [PMID: 17139425 DOI: 10.1007/s00534-005-1080-2]
14 **Cutrin JC**, Cantino D, Biasi F, Chiarpotto E, Salizzoni M, Andorno E, Massano G, Lanfranco G, Rizzetto M, Boveris A, Poli G. Reperfusion damage to the bile canaliculi in transplanted human liver. *Hepatology* 1996; **24**: 1053-1057 [PMID: 8903374 DOI: 10.1002/hep.510240512]
15 **García-Valdecasas JC**, Tabet J, Valero R, Deulofeu R, Taurá P, Rull R, Capdevila L, Cifuentes A, González FX, Net M, Beltran J, López-Boado MA, Palacin J, García F, Visa J. Evaluation of ischemic injury during liver procurement from non-heart-beating donors. *Eur Surg Res* 1999; **31**: 447-456 [PMID: 10861340 DOI: 10.1159/000008724]
16 **Noack K**, Bronk SF, Kato A, Gores GJ. The greater vulnerability of bile duct cells to reoxygenation injury than to anoxia. Implications for the pathogenesis of biliary strictures after liver transplantation. *Transplantation* 1993; **56**: 495-500 [PMID: 8212138]
17 **Jaeschke H**, Farhood A. Neutrophil and Kupffer cell-induced oxidant stress and ischemia-reperfusion injury in rat liver. *Am J Physiol* 1991; **260**: G355-G362 [PMID: 2003603 DOI: 10.1152/ajpgi.1991.260.3.G355]
18 **Jaeschke H**, Woolbright BL. Current strategies to minimize hepatic ischemia-reperfusion injury by targeting reactive oxygen species. *Transplant Rev (Orlando)* 2012; **26**: 103-114 [PMID: 22459037 DOI: 10.1016/j.trre.2011.10.006]
19 **Brunner SM**, Junger H, Ruemmele P, Schnitzbauer AA, Doenecke A, Kirchner GI, Farkas SA, Loss M, Scherer MN, Schlitt HJ, Fichtner-Feigl S. Bile duct damage after cold storage of deceased donor livers predicts biliary complications after liver transplantation. *J Hepatol* 2013; **58**: 1133-1139 [PMID: 23321317 DOI: 10.1016/j.jhep.2012.12.022]
20 **Hansen T**, Hollemann D, Pitton MB, Heise M, Hoppe-Lotichius M, Schuchmann M, Kirkpatrick CJ, Otto G. Histological examination and evaluation of donor bile ducts received during orthotopic liver transplantation--a morphological clue to ischemic-type biliary lesion? *Virchows Arch* 2012; **461**: 41-48 [PMID: 22588496 DOI: 10.1007/s00428-012-1245-8]
21 **Guichelaar MM**, Benson JT, Malinchoc M, Krom RA, Wiesner RH, Charlton MR. Risk factors for and clinical course of non-anastomotic biliary strictures after liver transplantation. *Am J Transplant* 2003; **3**: 885-890 [PMID: 12814481 DOI: 10.1034/j.1600-6143.2003.00165.x]
22 **Fisher A**, Miller CH. Ischemic-type biliary strictures in liver allografts: the Achilles heel revisited? *Hepatology* 1995; **21**: 589-591 [PMID: 7843733 DOI: 10.1002/hep.1840210245]
23 **Theilmann L**, Küppers B, Kadmon M, Roeren T, Notheisen H, Stiehl A, Otto G. Biliary tract strictures after orthotopic liver transplantation: diagnosis and management. *Endoscopy* 1994; **26**: 517-522 [PMID: 7828563 DOI: 10.1055/s-2007-1009026]
24 **Heidenhain C**, Pratschke J, Puhl G, Neumann U, Pascher A, Veltzke-Schlieker W, Neuhaus P. Incidence of and risk factors for ischemic-type biliary lesions following orthotopic liver transplantation. *Transpl Int* 2010; **23**: 14-22 [PMID: 19691661 DOI: 10.1111/j.1432-2277.2009.00947.x]
25 **Taner CB**, Bulatao IG, Perry DK, Sibulesky L, Willingham DL, Kramer DJ, Nguyen JH. Asystole to cross-clamp period predicts development of biliary complications in liver transplantation using donation after cardiac death donors. *Transpl Int* 2012; **25**: 838-846 [PMID: 22703372 DOI: 10.1111/j.1432-2277.2012.01508.x]
26 **op den Dries S**, Westerkamp AC, Karimian N, Gouw AS, Bruinsma BG, Markmann JF, Lisman T, Yeh H, Uygun K, Martins PN, Porte RJ. Injury to peribiliary glands and vascular plexus before liver transplantation predicts formation of non-anastomotic biliary strictures. *J Hepatol* 2014; **60**: 1172-1179 [PMID: 24560661 DOI: 10.1016/j.jhep.2014.02.010]
27 **Sutton ME**, op den Dries S, Koster MH, Lisman T, Gouw AS, Porte RJ. Regeneration of human extrahepatic biliary epithelium: the peribiliary glands as progenitor cell compartment. *Liver Int* 2012; **32**: 554-559 [PMID: 22171992 DOI: 10.1111/j.1478-3231.2011.02721.x]
28 **Op den Dries S**, Sutton ME, Lisman T, Porte RJ. Protection of bile ducts in liver transplantation: looking beyond ischemia. *Transplantation* 2011; **92**: 373-379 [PMID: 21629175 DOI: 10.1097/TP.0b013e318223a384]
29 **Oguma S**, Belle S, Starzl TE, Demetris AJ. A histometric analysis of chronically rejected human liver allografts: insights into the mechanisms of bile duct loss: direct immunologic and ischemic factors. *Hepatology* 1989; **9**: 204-209 [PMID: 2643544 DOI: 10.1002/hep.1840090207]
30 **Martelius T**, Krogerus L, Höckerstedt K, Bruggeman C, Lautenschlager I. Cytomegalovirus infection is associated with increased inflammation and severe bile duct damage in rat liver allografts. *Hepatology* 1998; **27**: 996-1002 [PMID: 9537439 DOI: 10.1002/hep.510270415]
31 **Wu J**, Ye S, Xu X, Xie H, Zhou L, Zheng S. Recipient outcomes after ABO-incompatible liver transplantation: a systematic review and meta-analysis. *PLoS One* 2011; **6**: e16521 [PMID: 21283553

DOI: 10.1371/journal.pone.0016521]
32 **Buis CI**, Geuken E, Visser DS, Kuipers F, Haagsma EB, Verkade HJ, Porte RJ. Altered bile composition after liver transplantation is associated with the development of nonanastomotic biliary strictures. *J Hepatol* 2009; **50**: 69-79 [PMID: 19012987 DOI: 10.1016/j.jhep.2008.07.032]
33 **NHS Blood and Transplant**. Interim report on liver transplantation. REPORT FOR 2016/2017 (1 October 2015 - 30 September 2016). Published April 2017. Available from: URL: https://nhsbtdbe.blob.core.windows.net/umbraco-assets-corp/5947/nhsbt-interim-liver-report-1617.pdf
34 **Boteon YL**, Afford SC, Mergental H. Pushing the Limits: Machine Preservation of the Liver as a Tool to Recondition High-Risk Grafts. *Curr Transplant Rep* 2018; **5**: 113-120 [PMID: 29774176 DOI: 10.1007/s40472-018-0188-7]
35 **Watson CJE**, Jochmans I. From "Gut Feeling" to Objectivity: Machine Preservation of the Liver as a Tool to Assess Organ Viability. *Curr Transplant Rep* 2018; **5**: 72-81 [PMID: 29564205 DOI: 10.1007/s40472-018-0178-9]
36 **Schlegel A**, Muller X, Dutkowski P. Hypothermic Machine Preservation of the Liver: State of the Art. *Curr Transplant Rep* 2018; **5**: 93-102 [PMID: 29564206 DOI: 10.1007/s40472-018-0183-z]
37 **Guarrera JV**, Henry SD, Samstein B, Odeh-Ramadan R, Kinkhabwala M, Goldstein MJ, Ratner LE, Renz JF, Lee HT, Brown RS Jr, Emond JC. Hypothermic machine preservation in human liver transplantation: the first clinical series. *Am J Transplant* 2010; **10**: 372-381 [PMID: 19958323 DOI: 10.1111/j.1600-6143.2009.02932.x]
38 **Dutkowski P**, Polak WG, Muiesan P, Schlegel A, Verhoeven CJ, Scalera I, DeOliveira ML, Kron P, Clavien PA. First Comparison of Hypothermic Oxygenated PErfusion Versus Static Cold Storage of Human Donation After Cardiac Death Liver Transplants: An International-matched Case Analysis. *Ann Surg* 2015; **262**: 764-70; discussion 770-1 [PMID: 26583664 DOI: 10.1097/SLA.0000000000001473]
39 **Dutkowski P**, de Rougemont O, Clavien PA. Machine perfusion for 'marginal' liver grafts. *Am J Transplant* 2008; **8**: 917-924 [PMID: 18416733 DOI: 10.1111/j.1600-6143.2008.02165.x]
40 **Dutkowski P**, Schlegel A, de Oliveira M, Müllhaupt B, Neff F, Clavien PA. HOPE for human liver grafts obtained from donors after cardiac death. *J Hepatol* 2014; **60**: 765-772 [PMID: 24295869 DOI: 10.1016/j.jhep.2013.11.023]
41 **Schlegel A**, Kron P, De Oliveira ML, Clavien PA, Dutkowski P. Is single portal vein approach sufficient for hypothermic machine perfusion of DCD liver grafts? *J Hepatol* 2016; **64**: 239-241 [PMID: 26432684 DOI: 10.1016/j.jhep.2015.09.015]
42 **Schlegel A**, Kron P, Graf R, Clavien PA, Dutkowski P. Hypothermic Oxygenated Perfusion (HOPE) downregulates the immune response in a rat model of liver transplantation. *Ann Surg* 2014; **260**: 931-7; discussion 937-8 [PMID: 25243553 DOI: 10.1097/SLA.0000000000000941]
43 **van Rijn R**, Karimian N, Matton APM, Burlage LC, Westerkamp AC, van den Berg AP, de Kleine RHJ, de Boer MT, Lisman T, Porte RJ. Dual hypothermic oxygenated machine perfusion in liver transplants donated after circulatory death. *Br J Surg* 2017; **104**: 907-917 [PMID: 28394402 DOI: 10.1002/bjs.10515]
44 **Laing RW**, Mergental H, Yap C, Kirkham A, Whilku M, Barton D, Curbishley S, Boteon YL, Neil DA, Hübscher SG, Perera MTPR, Muiesan P, Isaac J, Roberts KJ, Cilliers H, Afford SC, Mirza DF. Viability testing and transplantation of marginal livers (VITTAL) using normothermic machine perfusion: study protocol for an open-label, non-randomised, prospective, single-arm trial. *BMJ Open* 2017; **7**: e017733 [PMID: 29183928 DOI: 10.1136/bmjopen-2017-017733]
45 **op den Dries S**, Karimian N, Sutton ME, Westerkamp AC, Nijsten MW, Gouw AS, Wiersema-Buist J, Lisman T, Leuvenink HG, Porte RJ. Ex vivo normothermic machine perfusion and viability testing of discarded human donor livers. *Am J Transplant* 2013; **13**: 1327-1335 [PMID: 23463950 DOI: 10.1111/ajt.12187]
46 **Nasralla D**, Coussios CC, Mergental H, Akhtar MZ, Butler AJ, Ceresa CDL, Chiocchia V, Dutton SJ, García-Valdecasas JC, Heaton N, Imber C, Jassem W, Jochmans I, Karani J, Knight SR, Kocabayoglu P, Malagò M, Mirza D, Morris PJ, Pallan A, Paul A, Pavel M, Perera MTPR, Pirenne J, Ravikumar R, Russell L, Upponi S, Watson CJE, Weissenbacher A, Ploeg RJ, Friend PJ; Consortium for Organ Preservation in Europe. A randomized trial of normothermic preservation in liver transplantation. *Nature* 2018; **557**: 50-56 [PMID: 29670285 DOI: 10.1038/s41586-018-0047-9]
47 **Hoyer DP**, Mathé Z, Gallinat A, Canbay AC, Treckmann JW, Rauen U, Paul A, Minor T. Controlled Oxygenated Rewarming of Cold Stored Livers Prior to Transplantation: First Clinical Application of a New Concept. *Transplantation* 2016; **100**: 147-152 [PMID: 26479280 DOI: 10.1097/TP.0000000000000915]
48 **Bruinsma BG**, Yeh H, Ozer S, Martins PN, Farmer A, Wu W, Saeidi N, Op den Dries S, Berendsen TA, Smith RN, Markmann JF, Porte RJ, Yarmush ML, Uygun K, Izamis ML. Subnormothermic machine perfusion for ex vivo preservation and recovery of the human liver for transplantation. *Am J Transplant* 2014; **14**: 1400-1409 [PMID: 24758155 DOI: 10.1111/ajt.12727]
49 **Oniscu GC**, Randle LV, Muiesan P, Butler AJ, Currie IS, Perera MT, Forsythe JL, Watson CJ. In situ normothermic regional perfusion for controlled donation after circulatory death--the United Kingdom experience. *Am J Transplant* 2014; **14**: 2846-2854 [PMID: 25283987 DOI: 10.1111/ajt.12927]
50 **Miñambres E**, Suberviola B, Dominguez-Gil B, Rodrigo E, Ruiz-San Millan JC, Rodríguez-San Juan JC, Ballesteros MA. Improving the Outcomes of Organs Obtained From Controlled Donation After Circulatory Death Donors Using Abdominal Normothermic Regional Perfusion. *Am J Transplant* 2017; **17**: 2165-2172 [PMID: 28141909 DOI: 10.1111/ajt.14214]
51 **De Carlis L**, De Carlis R, Lauterio A, Di Sandro S, Ferla F, Zanierato M. Sequential Use of Normothermic Regional Perfusion and Hypothermic Machine Perfusion in Donation After Cardiac Death Liver Transplantation With Extended Warm Ischemia Time. *Transplantation* 2016; **100**: e101-e102 [PMID: 27495774 DOI: 10.1097/TP.0000000000001419]
52 **Boteon YL**, Laing RW, Schlegel A, Wallace L, Smith A, Attard J, Bhogal RH, Neil DA, Hübscher S, Perera MTP, Mirza DF, Afford SC, Mergental H. Combined Hypothermic and Normothermic Machine Perfusion Improves Functional Recovery of Extended Criteria Donor Livers. *Liver Transpl* 2018 [PMID: 30058119 DOI: 10.1002/lt.25315]
53 **Moher D**, Liberati A, Tetzlaff J, Altman DG; PRISMA Group. Preferred reporting items for systematic reviews and meta-analyses: the PRISMA statement. *J Clin Epidemiol* 2009; **62**: 1006-1012 [PMID: 19631508 DOI: 10.1016/j.jclinepi.2009.06.005]
54 **Selzner M**, Goldaracena N, Echeverri J, Kaths JM, Linares I, Selzner N, Serrick C, Marquez M, Sapisochin G, Renner EL, Bhat M, McGilvray ID, Lilly L, Greig PD, Tsien C, Cattral MS, Ghanekar A, Grant DR. Normothermic ex vivo liver perfusion using steen solution as perfusate for human liver transplantation: First North American results. *Liver Transpl* 2016; **22**: 1501-1508 [PMID: 27339754 DOI: 10.1002/lt.24499]
55 **Ravikumar R**, Jassem W, Mergental H, Heaton N, Mirza D, Perera MT, Quaglia A, Holroyd D, Vogel T, Coussios CC, Friend PJ. Liver Transplantation After Ex Vivo Normothermic Machine Preservation: A Phase 1 (First-in-Man) Clinical Trial. *Am J Transplant* 2016; **16**: 1779-1787 [PMID: 26752191 DOI: 10.1111/ajt.13708]
56 **Bral M**, Gala-Lopez B, Bigam D, Kneteman N, Malcolm A, Livingstone S, Andres A, Emamaullee J, Russell L, Coussios C, West LJ, Friend PJ, Shapiro AM. Preliminary Single-Center Canadian Experience of Human Normothermic Ex Vivo Liver Perfusion: Results of a Clinical Trial. *Am J Transplant* 2017; **17**: 1071-1080 [PMID: 27639262 DOI: 10.1111/ajt.14049]
57 **Mergental H**, Perera MT, Laing RW, Muiesan P, Isaac JR, Smith A, Stephenson BT, Cilliers H, Neil DA, Hübscher SG, Afford SC, Mirza DF. Transplantation of Declined Liver Allografts Following Normothermic Ex-Situ Evaluation. *Am J Transplant* 2016; **16**: 3235-3245 [PMID: 27192971 DOI: 10.1111/ajt.13875]
58 **Watson CJE**, Kosmoliaptsis V, Pley C, Randle L, Fear C, Crick K, Gimson AE, Allison M, Upponi S, Brais R, Jochmans I, Butler AJ.

Observations on the ex situ perfusion of livers for transplantation. *Am J Transplant* 2018; **18**: 2005-2020 [PMID: 29419931 DOI: 10.1111/ajt.14687]

59 **Guarrera JV**, Henry SD, Samstein B, Reznik E, Musat C, Lukose TI, Ratner LE, Brown RS Jr, Kato T, Emond JC. Hypothermic machine preservation facilitates successful transplantation of "orphan" extended criteria donor livers. *Am J Transplant* 2015; **15**: 161-169 [PMID: 25521639 DOI: 10.1111/ajt.12958]

60 **De Carlis R**, Di Sandro S, Lauterio A, Ferla F, Dell'Acqua A, Zanierato M, De Carlis L. Successful donation after cardiac death liver transplants with prolonged warm ischemia time using normothermic regional perfusion. *Liver Transpl* 2017; **23**: 166-173 [PMID: 27783454 DOI: 10.1002/lt.24666]

61 **Chouchani ET**, Pell VR, Gaude E, Aksentijević D, Sundier SY, Robb EL, Logan A, Nadtochiy SM, Ord ENJ, Smith AC, Eyassu F, Shirley R, Hu CH, Dare AJ, James AM, Rogatti S, Hartley RC, Eaton S, Costa ASH, Brookes PS, Davidson SM, Duchen MR, Saeb-Parsy K, Shattock MJ, Robinson AJ, Work LM, Frezza C, Krieg T, Murphy MP. Ischaemic accumulation of succinate controls reperfusion injury through mitochondrial ROS. *Nature* 2014; **515**: 431-435 [PMID: 25383517 DOI: 10.1038/nature13909]

62 **Schlegel A**, Kron P, Dutkowski P. Hypothermic machine perfusion in liver transplantation. *Curr Opin Organ Transplant* 2016; **21**: 308-314 [PMID: 26918882 DOI: 10.1097/MOT.0000000000000303]

63 **van Rijn R**, van Leeuwen OB, Matton APM, Burlage LC, Wiersema-Buist J, van den Heuvel MC, de Kleine RHJ, de Boer MT, Gouw ASH, Porte RJ. Hypothermic oxygenated machine perfusion reduces bile duct reperfusion injury after transplantation of donation after circulatory death livers. *Liver Transpl* 2018; **24**: 655-664 [PMID: 29369470 DOI: 10.1002/lt.25023]

64 **DiPaola F**, Shivakumar P, Pfister J, Walters S, Sabla G, Bezerra JA. Identification of intramural epithelial networks linked to peribiliary glands that express progenitor cell markers and proliferate after injury in mice. *Hepatology* 2013; **58**: 1486-1496 [PMID: 23703727 DOI: 10.1002/hep.26485]

65 **Irie T**, Asahina K, Shimizu-Saito K, Teramoto K, Arii S, Teraoka H. Hepatic progenitor cells in the mouse extrahepatic bile duct after a bile duct ligation. *Stem Cells Dev* 2007; **16**: 979-987 [PMID: 18004941 DOI: 10.1089/scd.2007.0037]

66 **Selten J**, Schlegel A, de Jonge J, Dutkowski P. Hypo- and normothermic perfusion of the liver: Which way to go? *Best Pract Res Clin Gastroenterol* 2017; **31**: 171-179 [PMID: 28624105 DOI: 10.1016/j.bpg.2017.04.001]

67 **Boteon YL**, Boteon APCS, Attard J, Mergental H, Mirza DF, Bhogal RH, Afford SC. Ex situ machine perfusion as a tool to recondition steatotic donor livers: Troublesome features of fatty livers and the role of defatting therapies. A systematic review. *Am J Transplant* 2018; **18**: 2384-2399 [PMID: 29947472 DOI: 10.1111/ajt.14992]

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CASE REPORT

Treatment of transplant renal artery pseudoaneurysm using expandable hydrogel coils: A case report and review of literature

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Author contributions: Marie Y and Shrestha B managed the patient and prepared the manuscript; Kumar A, Hinchliffe S and Curran S were involved in managing the patient and contributed to manuscript preparation; Brown P and Turner D performed radiological investigations, managed the patient and contributed to manuscript preparation.

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Abstract

Transplant renal artery (TRA) pseudoaneurysm can result in bleeding, infection, graft dysfunction and graft loss. We report the management of a renal transplant recipient who presented five months after renal transplantation with deterioration of renal function, who was found to have TRA pseudoaneurysm and TRA stenosis. Both were treated radiologically by using expandable hydrogel coils (EHC) in combination with stenting. Improvement in clinical, biochemical and radiological parameters were observed after the intervention. To our knowledge, this is the first report in the transplant literature on the use of EHC for the treatment of a TRA pseudoaneurysm.

Key words: Pseudoaneurysm; Transplant renal artery; Expandable hydrogel coils; Outcomes

Core tip: Transplant renal artery (TRA) pseudoaneurysm is an uncommon complication after renal transplantation, which can cause transplant dysfunction, bleeding, infection

and graft loss. Expandable hydrogel coils should be considered in the treatment of TRA pseudoaneurysm as they have been effective in our patient.

Marie Y, Kumar A, Hinchliffe S, Curran S, Brown P, Turner D, Shrestha B. Treatment of transplant renal artery pseudoaneurysm using expandable hydrogel coils: A case report and review of literature. *World J Transplant* 2018; 8(6): 232-236 Available from: URL: http://www.wjgnet.com/2220-3230/full/v8/i6/232.htm DOI: http://dx.doi.org/10.5500/wjt.v8.i6.232

INTRODUCTION

Transplant renal artery (TRA) pseudoaneurysm can result in bleeding, infection, graft dysfunction, graft loss, lower limb ischaemia, limb loss and mortality[1]. The treatment of TRA pseudoaneurysm remains challenging. The expandable hydrogel coil (EHC) embolization system is a relatively new type of device that has been described to successfully treat intracranial and peripheral pseudoaneurysms. These are helical platinum coils coated with exapandable hydrogel polymer. The hydrogel coating undergoes full expansion within 20 min attending a size between 4-5 times the size of the coils on coming in contact with blood. The stasis of the blood causes organization of thrombus, which fills the aneurysm causing complete its occlusion[1]. To our knowledge, there is no published data in the transplant literature on the application of EHC in the treatment of TRA pseudoaneurysm. We describe the successful management of a case of TRA pseudoaneurysm using EHC and review the pertinent literature.

CASE REPORT

A 38-year-old male patient received a renal transplant (RT) of a kidney from a donation after circulatory death donor. The right donor kidney had a single real artery on an aortic patch and the short renal vein which was elongated by using a segment of inferior vena cava. The kidney was implanted in the right iliac fossa by anastomosing the renal artery to the external iliac artery in an end-to-side fashion using continuous 5/0 prolene sutures (Ethicon Inc., United Kingdom) and renal vein to the external iliac vein in the similar fashion. An extravesical ureteroneocystostomy was performed as describe by Lich-Gregoir. The vascular anastomosis time was 45 min while the total cold ischaemic time was 15 h and 38 min. The patient received basiliximab (Sandoz, United Kingdom) and methyl prednisolone as induction therapy and tacrolimus, mycophenolate mofetil and prednisolone as maintenance immunosuppression.

The transplant had delayed graft function and required haemodialysis during the first week until renal function started to improve. The initial ultrasound scan of the transplant kidney showed a well perfused graft with no evidence of hydronephrosis or any collection and the resistive indices (RI) were within normal limits. The renal function was stable with a serum creatinine of 136 μmol/L and an estimated glomerular filtration rate (eGFR) of 51 mL/min per 1.73 m^2 at 3 mo post-transplantation.

At five months post-transplantation, on routine outpatient review, deterioration in kidney renal function with a rise in serum creatinine to 633 μmol/L (eGFR 13 mL/min per 1.73 m^2) was observed. A duplex ultrasound scan showed a well-perfused kidney with no evidence of hydronephrosis. An ultrasound-guided biopsy of the kidney, which was treated with three pulses of intravenous methyl prednisolone, showed features of acute cellular rejection. However, there was no improvement in renal function. A repeat duplex ultrasound scan showed damped flow signals on the intra-renal blood vessels with reduced RI ranging between 0.4 and 0.45. There were associated high velocities at the transplant artery origin which were suspicious of TRA stenosis. A computerized tomography (CT) scan was done which showed a 20 mm × 25 mm pseudoaneurysm arising from the aortic patch and the TRA origin lying adjacent to the pseudoaneurysm was tightly narrowed (Figures 1 and 2). After discussion in the departmental multidisciplinary team meeting and with patient's informed consent, he underwent radiological intervention as described below.

Under ultrasound guidance, the right common femoral artery was punctured, and a 5Fr sheath was inserted. 7500 unit of heparin was administered intravenously. The dimensions of the TRA were confirmed and were found to be like those of CT scan findings. The TRA was catheterized using a size 4Fr Berenstein® catheter and Terumo® wire (Terumo Medical Corporation, United States), subsequently exchanged for a 0.014 Thruway wire (Boston Scientific Inc., Ireland) and was left in situ as a "safety wire". The left common femoral artery was punctured and a 6Fr destination sheath was placed over the aortic bifurcation. Through the ipsilateral 6Fr sheath, a 10 mm percutaneous transluminal angioplasty (PTA) balloon was placed opposite the aneurysm neck. From the 6Fr sheath, a 4Fr Cobra (Cook Medical, United States) and 2/7 Progreat® microcatheter (Terumo Medical Corporation, United States) were used to gain access to the aneurysm sac. Within the right external iliac artery, the PTA balloon was inflated to reduce the risk of coil prolapse and migration and the aneurysm was embolized using two Azur® 20 mm Framing coils (Terumo Medical Corporation, United States), and packed with Azur® Hydrogel Coils. The pseudoaneurysm was filled with coils and hydrogel leading to its complete occlusion. Subsequently, the stenosed transplant RA was stented using a 6 mm × 20 mm Hippocampus stent (Medtronic, United Kingdom), which restored the patency of the stenosed renal artery and normal blood flow (Figure 3).

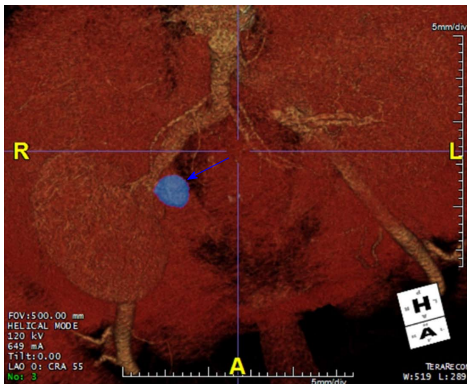

Figure 1 Computerized tomography angiogram showing a 20 mm x 25 mm pseudoaneurysm arising from the aortic patch (blue arrow).

Figure 2 Angiogram showing transplant renal artery stenosis (blue arrow) due to compression caused by the pseudoaneurysm. Guide wire is present within the right common and external iliac artery.

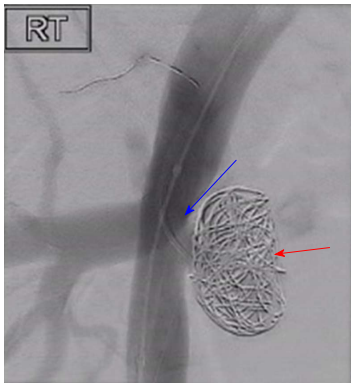

Figure 3 Successful coiling of the pseudoaneurysm (red arrow) and stenting of the transplant renal artery stenosis (blue arrow).

Following embolization of the pseudoaneurysm and stenting of the TRA, improvement in renal function occurred leading to a fall in the serum creatinine level to 159 μmol/L (eGFR 47 mL/min per 1.73 m^2). A follow-up CT angiogram one month after the intervention showed a patent TRA with successful coiling of the pseudoaneurysm and satisfactory position of the stent with no evidence of TRA stenosis. The patient continues to be followed up in the routine RT clinic and has a serum creatinine of 150 μmol/L (eGFR 49 mL/min per 1.73 m^2).

DISCUSSION

Renal Transplantation remains the treatment of choice in end stage renal disease patients. Vascular complications after RT include TRA stenosis, TRA thrombosis, transplant renal vein thrombosis, arteriovenous fistula and TRA pseudoaneurysm[2]. TRA pseudoaneurysm is an uncommon complication and can be classified anatomically as intrarenal or extrarenal based on the involvement of either TRA or iliac artery, respectively. Aetiologically, TRA pseudoaneurysms can be of infective or non- infective origin. Infective pseudoaneurysms are more common and can be of fungal (mycotic) and non-fungal (non-mycotic) origin. Amongst the infective pseudoaneurysms, Candida albicans and Aspergillus species have been reported to be the predominant microorganisms, while Pseudomonas species were the leading cause of non-mycotic infective pseudoaneurysms[2-8]. Non-infective TRA pseudoaneurysms can result from injury to the arterial wall, faulty suture techniques[9-11] or following a biopsy[12,13].

TRA pseudoaneurysms can be asymptomatic[14] or can present with RT dysfunction, fever, pain (mainly at the site of the transplant) or a combination of these presentations. Graft loss is a recognized complication of TRA pseudoaneurysms and sometimes bleeding from the ruptured pseudoaneurysm can lead to hemorrhagic shock or death of the patient[8,15]. Lumbar plexopathy has been reported in a previous literature because of pressure effect of the pseudoaneurysm[16], while malignant hypertension is a rare presentation[17].

The choice of the modality of treatment of TRA pseudoaneurysms depends on several factors including the aetiology, haemodynamic stability of the patient, presentation, anatomy, graft function and the radiological features of the pseudoaneurysm[18-22]. Aneurysms larger than 25 mm, progressive enlargement, deterioration of renal function or presentation with symptoms are the main indications for repair[18]. Treatment modalities include minimally invasive techniques using mainly exclusion stents to the external iliac artery, but this may sacrifice the graft[14]. Ultrasound-guided percutaneous thrombin injection in combination with a covered stent has been reported as a successful way of treating TRA pseudoaneurysm with preservation of renal function[18,19].

Traditional aneurysm coiling in general can be associated with complications such as migration, non-target embolisation, inadequate filling, compaction and the technical difficulty in placing the coils leading to added risk to organs, patients and increase in cost[23].

Expandable hydrogel technology coils have been described to treat intracranial and peripheral pseudo-aneurysms successfully for number of years. The main

advantage of using EHC is related to their superior mechanical occlusion properties resulting in fewer coils deployed and a lower recurrence rate. They are also compatible with imaging modalities[24-26].

In our case, successful radiological and clinical outcomes were achieved with return of serum creatinine to baseline within 48 h of intervention without any complication related to the RT or lower limb. We have employed EHC system to treat TRA pseudoaneurysm, achieved excellent volumetric filling and targeted embolisation and subsequently deployed a stent leading to restoration of transplant renal function to its normality. It offers a new non- invasive technique to treat TRA pseudoaneurysms with preservation of renal grafts; therefore, it should be considered as a first line treatment modality in this clinical situation.

ARTICLE HIGHLIGHTS

Case characteristics

A 38-year-old male, who had received a deceased donor renal transplant presented with deterioration of renal function five months post-transplantation.

Clinical diagnosis

On examination, there were clinical features pointing to definitive diagnosis.

Differential diagnosis

Differential diagnosis included obstructive uropathy, acute rejection, infections, drug nephrotoxicity and transplant renal artery stenosis.

Laboratory diagnosis

The serum creatinine was significantly elevated.

Imaging diagnosis

The Duplex ultrasound scan showed reduced resistive index with high velocity flow in the renal artery suggestive of transplant renal artery stenosis. A computerized tomography angiogram showed a 20 mm × 25 mm pseudoaneurysm at the anastomosis site and stenosis of the transplant renal artery adjacent to the pseudoaneurysm.

Treatment

Endovascular embolisation of the pseudoaneurysm using expandable hydrogel coils (EHC) followed by deployment of stent lead to resolution of the pseudoaneurysm and transplant renal artery stenosis and restoration of renal function to normality.

Related reports

Follow-up of the patient was satisfactory with no adverse events related to the procedure.

Experiences and lessons

This is the first reported case of treatment of a transplant renal artery pseudoaneurysm with wide neck with the use of EHC leading to successful outcomes.

REFERENCES

1 **Ding YH**, Dai D, Lewis DA, Cloft HJ, Kallmes DF. Angiographic and histologic analysis of experimental aneurysms embolized with platinum coils, Matrix, and HydroCoil. *AJNR Am J Neuroradiol* 2005; **26**: 1757-1763 [PMID: 16091526]

2 **Aktas S**, Boyvat F, Sevmis S, Moray G, Karakayali H, Haberal M. Analysis of vascular complications after renal transplantation. *Transplant Proc* 2011; **43**: 557-561 [PMID: 21440760 DOI: 10.1016/j.transproceed.2011.01.007]

3 **Kountidou CS**, Stier K, Niehues SM, Lingnau A, Schostak M, Fuller TF, Lützenberg R. Successful repair of post-transplant mycotic aneurysm of iliac artery with renal graft preservation: a case report. *Urology* 2012; **80**: 1151-1153 [PMID: 22999448 DOI: 10.1016/j.urology.2012.07.048]

4 **Minz M**, Sharma A, Kumar S, Singh S, Shivaprakash MR, Bag S. Use of autogenous internal iliac artery for bridging the external iliac artery after excision of Aspergillus mycotic aneurysm in renal transplant recipients. *J Vasc Surg* 2011; **53**: 802-804 [PMID: 21215589 DOI: 10.1016/j.jvs.2010.10.102]

5 **Smeds MR**, Ofstein R, Peterson GJ, Peterson BG, Jacobs DL. Endovascular repair of a para-anastomotic pseudoaneurysm after renal autotransplantation: an alternative to open reconstruction. *Ann Vasc Surg* 2013; **27**: 110.e5-110.e8 [PMID: 23079504 DOI: 10.1016/j.avsg.2012.06.003]

6 **Chandak P**, Kessaris N, Uwechue RU, Abboudi H, Hossain M, Harris F, Jones K, Fronek J. Successful excision of a suspected mycotic transplant renal artery patch aneurysm with renal allograft autotransplantation. *Transplantation* 2014; **97**: e25-e26 [PMID: 24492430 DOI: 10.1097/01.TP.0000438628.75848.9a]

7 **Orlando G**, Di Cocco P, Gravante G, D'Angelo M, Famulari A, Pisani F. Fatal hemorrhage in two renal graft recipients with multi-drug resistant Pseudomonas aeruginosa infection. *Transpl Infect Dis* 2009; **11**: 442-447 [PMID: 19508700 DOI: 10.1111/j.1399-3062.2009.00412.x]

8 **Burkey SH**, Vazquez MA, Valentine RJ. De novo renal artery aneurysm presenting 6 years after transplantation: a complication of recurrent arterial stenosis? *J Vasc Surg* 2000; **32**: 388-391 [PMID: 10918000 DOI: 10.1067/mva.2000.106943]

9 **Nguan CY**, Luke PP. Renal artery pseudoaneurysm of infectious etiology: a life-threatening complication after renal transplantation. *Urology* 2006; **68**: 668-669 [PMID: 16979702 DOI: 10.1016/j.urology.2006.04.017]

10 **Rivera M**, Villacorta J, Jiménez-Alvaro S, Quereda C. Asymptomatic large extracapsular renal pseudoaneurysm following kidney transplant biopsy. *Am J Kidney Dis* 2011; **57**: 175-178 [PMID: 21184923 DOI: 10.1053/j.ajkd.2010.07.020]

11 **Duwe KM**, Newhouse JH, Fayter J, Stern L, Budorick NE. Conservative management of an extrarenal pseudoaneurysm after percutaneous needle biopsy of a renal allograft. *J Ultrasound Med* 2000; **19**: 281-283 [PMID: 10759353 DOI: 10.7863/jum.2000.19.4.281]

12 **Antonopoulos IM**, Yamacake KG, Tiseo BC, Carnevale FC, Z E Jr, Nahas WC. Renal pseudoaneurysm after core-needle biopsy of renal allograft successfully managed with superselective embolization. *Int Braz J Urol* 2016; **42**: 165-167 [PMID: 27136484 DOI: 10.1590/S1677-5538.IBJU.2014.0315]

13 **Patrono D**, Verhelst R, Buemi A, Darius T, Godefroid N, Mourad M. Presentation and management of mycotic pseudoaneurysm after kidney transplantation. *Transpl Infect Dis* 2015; **17**: 129-136 [PMID: 25620391 DOI: 10.1111/tid.12346]

14 **Orlic P**, Vukas D, Drescik I, Ivancic A, Blecic G, Budiselic B, Velcic G, Maricic A, Oguic R, Mozetic V, Valencic M, Sotosek S, Vukas D Jr. Vascular complications after 725 kidney transplantations during 3 decades. *Transplant Proc* 2003; **35**: 1381-1384 [PMID: 12826165 DOI: 10.1016/S0041-1345(03)00506-2]

15 **Luzzio CC**, Waclawik AJ, Gallagher CL, Knechtle SJ. Iliac artery pseudoaneurysm following renal transplantation presenting as lumbosacral plexopathy. *Transplantation* 1999; **67**: 1077-1078 [PMID: 10221499 DOI: 10.1097/00007890-199904150-00026]

16 **Madhav D**, Kumar P, Mohan C; Vijay, Mahesh U; Anusha; Suneetha; Suryaprakash. Candida-associated pseudo-aneurysm of the transplant renal artery presenting as malignant hypertension and managed successfully without nephrectomy. *Saudi J Kidney Dis Transpl* 2015; **26**: 1000-1005 [PMID: 26354578 DOI: 10.4103/1319-2442.164591]

17 **Bracale UM**, Santangelo M, Carbone F, Del Guercio L, Maurea

S, Porcellini M, Bracale G. Anastomotic pseudoaneurysm complicating renal transplantation: treatment options. *Eur J Vasc Endovasc Surg* 2010; **39**: 565-568 [PMID: 20122855 DOI: 10.1016/j.ejvs.2009.12.010]

18 **Asztalos L**, Olvasztó S, Fedor R, Szabó L, Balázs G, Lukács G. Renal artery aneurysm at the anastomosis after kidney transplantation. *Transplant Proc* 2006; **38**: 2915-2918 [PMID: 17112863 DOI: 10.1016/j.transproceed.2006.08.115]

19 **Zavos G**, Pappas P, Kakisis JD, Leonardou P, Manoli E, Bokos J, Kostakis A. Endovascular repair as first-choice treatment of iliac pseudoaneurysms following renal transplantation. *Transplant Proc* 2005; **37**: 4300-4302 [PMID: 16387102 DOI: 10.1016/j.transproceed.2005.11.034]

20 **Kubal C**, Cacciola R, Riley P, Ready A. Internal iliac artery pseudoaneurysm following renal transplant biopsy successfully treated with endovascular stenting and thrombosis: a case report. *Transplant Proc* 2007; **39**: 1676-1678 [PMID: 17580217 DOI: 10.1016/j.transproceed.2007.03.018]

21 **Dimitroulis D**, Bokos J, Zavos G, Nikiteas N, Karidis NP, Katsaronis P, Kostakis A. Vascular complications in renal transplantation: a single-center experience in 1367 renal transplantations and review of the literature. *Transplant Proc* 2009; **41**: 1609-1614 [PMID: 19545690 DOI: 10.1016/j.transproceed.2009.02.077]

22 **Sharron JA**, Esterl RM, Washburn WK, Abrahamian GA. Surgical treatment of an extrarenal pseudoaneurysm after kidney transplantation. *Vasc Endovascular Surg* 2009; **43**: 317-321 [PMID: 19223384 DOI: 10.1177/1538574409331697]

23 **Sluzewski M**, van Rooij WJ, Slob MJ, Bescós JO, Slump CH, Wijnalda D. Relation between aneurysm volume, packing, and compaction in 145 cerebral aneurysms treated with coils. *Radiology* 2004; **231**: 653-658 [PMID: 15118115 DOI: 10.1148/radiol.2313030460]

24 **Kallmes DF**, Fujiwara NH. New expandable hydrogel-platinum coil hybrid device for aneurysm embolization. *AJNR Am J Neuroradiol* 2002; **23**: 1580-1588 [PMID: 12372752]

25 **Deshaies EM**, Adamo MA, Boulos AS. A prospective single-center analysis of the safety and efficacy of the hydrocoil embolization system for the treatment of intracranial aneurysms. *J Neurosurg* 2007; **106**: 226-233 [PMID: 17410704 DOI: 10.3171/jns.2007.106.2.226]

26 **White PM**, Lewis SC, Nahser H, Sellar RJ, Goddard T, Gholkar A; HELPS Trial Collaboration. HydroCoil Endovascular Aneurysm Occlusion and Packing Study (HELPS trial): procedural safety and operator-assessed efficacy results. *AJNR Am J Neuroradiol* 2008; **29**: 217-223 [PMID: 18184832 DOI: 10.3174/ajnr.A0936]

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