World Journal of *Gastroenterology*

World J Gastroenterol 2022 January 7; 28(1): 1-175





Published by Baishideng Publishing Group Inc

WJG

World Journal of Gastroenterology

Contents

Weekly Volume 28 Number 1 January 7, 2022

FRONTIER

1 Advances in traction methods for endoscopic submucosal dissection: What is the best traction method and traction direction?

Nagata M

23 Gastric pentadecapeptide BPC 157 in cytoprotection to resolve major vessel occlusion disturbances, ischemia-reperfusion injury following Pringle maneuver, and Budd-Chiari syndrome

Sikiric P, Skrtic A, Gojkovic S, Krezic I, Zizek H, Lovric E, Sikiric S, Knezevic M, Strbe S, Milavic M, Kokot A, Boban Blagaic A, Seiwerth S

REVIEW

- 47 Transfusion-transmitted hepatitis E: What we know so far? Cheung CKM, Wong SH, Law AWH, Law MF
- 76 Viral hepatitis in 2021: The challenges remaining and how we should tackle them Dunn R, Wetten A, McPherson S, Donnelly MC

MINIREVIEWS

96 Risk of hepatocellular carcinoma after hepatitis C virus cure Luna-Cuadros MA, Chen HW, Hanif H, Ali MJ, Khan MM, Lau DTY

108 Artificial intelligence in the diagnosis and management of colorectal cancer liver metastases Rompianesi G, Pegoraro F, Ceresa CD, Montalti R, Troisi RI

ORIGINAL ARTICLE

Basic Study

Focal adhesion kinase-related non-kinase ameliorates liver fibrosis by inhibiting aerobic glycolysis via the 123 FAK/Ras/c-myc/ENO1 pathway

Huang T, Li YQX, Zhou MY, Hu RH, Zou GL, Li JC, Feng S, Liu YM, Xin CQ, Zhao XK

Retrospective Cohort Study

140 Dynamics of cytokines predicts risk of hepatocellular carcinoma among chronic hepatitis C patients after viral eradication

Lu MY, Yeh ML, Huang CI, Wang SC, Tsai YS, Tsai PC, Ko YM, Lin CC, Chen KY, Wei YJ, Hsu PY, Hsu CT, Jang TY, Liu TW, Liang PC, Hsieh MY, Lin ZY, Chen SC, Huang CF, Huang JF, Dai CY, Chuang WL, Yu ML

SYSTEMATIC REVIEWS

154 Current guidelines for the management of celiac disease: A systematic review with comparative analysis Raiteri A, Granito A, Giamperoli A, Catenaro T, Negrini G, Tovoli F



Contents

Weekly Volume 28 Number 1 January 7, 2022

ABOUT COVER

Editorial Board Member of World Journal of Gastroenterology, Pilar Codoñer-Franch, MD, PhD, Professor, Chief, Department of Pediatrics, Gastroenterology and Nutrition, University of Valencia, Dr. Peset University Hospital, Avda Gaspar Aguilar 90, Valencia 46017, Spain. pilar.codoner@uv.es

AIMS AND SCOPE

The primary aim of World Journal of Gastroenterology (WJG, World J Gastroenterol) is to provide scholars and readers from various fields of gastroenterology and hepatology with a platform to publish high-quality basic and clinical research articles and communicate their research findings online. WJG mainly publishes articles reporting research results and findings obtained in the field of gastroenterology and hepatology and covering a wide range of topics including gastroenterology, hepatology, gastrointestinal endoscopy, gastrointestinal surgery, gastrointestinal oncology, and pediatric gastroenterology.

INDEXING/ABSTRACTING

The WJG is now indexed in Current Contents®/Clinical Medicine, Science Citation Index Expanded (also known as SciSearch®), Journal Citation Reports®, Index Medicus, MEDLINE, PubMed, PubMed Central, and Scopus. The 2021 edition of Journal Citation Report® cites the 2020 impact factor (IF) for WJG as 5.742; Journal Citation Indicator: 0.79; IF without journal self cites: 5.590; 5-year IF: 5.044; Ranking: 28 among 92 journals in gastroenterology and hepatology; and Quartile category: Q2. The WJG's CiteScore for 2020 is 6.9 and Scopus CiteScore rank 2020: Gastroenterology is 19/136.

RESPONSIBLE EDITORS FOR THIS ISSUE

Production Editor: Ying-Yi Yuan; Production Department Director: Xiang Li; Editorial Office Director: Ze-Mao Gong.

NAME OF JOURNAL	INSTRUCTIONS TO AUTHORS
World Journal of Gastroenterology	https://www.wjgnet.com/bpg/gerinfo/204
ISSN	GUIDELINES FOR ETHICS DOCUMENTS
ISSN 1007-9327 (print) ISSN 2219-2840 (online)	https://www.wjgnet.com/bpg/GerInfo/287
LAUNCH DATE	GUIDELINES FOR NON-NATIVE SPEAKERS OF ENGLISH
October 1, 1995	https://www.wjgnet.com/bpg/gerinfo/240
FREQUENCY	PUBLICATION ETHICS
Weekly	https://www.wjgnet.com/bpg/GerInfo/288
EDITORS-IN-CHIEF	PUBLICATION MISCONDUCT
Andrzej S Tarnawski	https://www.wjgnet.com/bpg/gerinfo/208
EDITORIAL BOARD MEMBERS	ARTICLE PROCESSING CHARGE
http://www.wjgnet.com/1007-9327/editorialboard.htm	https://www.wjgnet.com/bpg/gerinfo/242
PUBLICATION DATE	STEPS FOR SUBMITTING MANUSCRIPTS
January 7, 2022	https://www.wjgnet.com/bpg/GerInfo/239
COPYRIGHT	ONLINE SUBMISSION
© 2022 Baishideng Publishing Group Inc	https://www.f6publishing.com

© 2022 Baishideng Publishing Group Inc. All rights reserved. 7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA E-mail: bpgoffice@wjgnet.com https://www.wjgnet.com



WJG

World Journal of Gastroenterology

Submit a Manuscript: https://www.f6publishing.com

World J Gastroenterol 2022 January 7; 28(1): 1-22

DOI: 10.3748/wjg.v28.i1.1

ISSN 1007-9327 (print) ISSN 2219-2840 (online)

FRONTIER

Advances in traction methods for endoscopic submucosal dissection: What is the best traction method and traction direction?

Mitsuru Nagata

ORCID number: Mitsuru Nagata 0000-0002-5697-5953.

Author contributions: Nagata M has been associated with conception, drafting of the article, and final approval of the article.

Conflict-of-interest statement: No financial relationships with a commercial entity producing health-care related products and/or services relevant to this article

Country/Territory of origin: Japan

Specialty type: Gastroenterology and hepatology

Provenance and peer review: Invited article; Externally peer reviewed.

Peer-review model: Single blind

Peer-review report's scientific quality classification

Grade A (Excellent): A, A Grade B (Very good): B, B Grade C (Good): 0 Grade D (Fair): 0 Grade E (Poor): 0

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative

Mitsuru Nagata, Department of Endoscopy, Shonan Fujisawa Tokushukai Hospital, Kanagawa 251-0041, Japan

Corresponding author: Mitsuru Nagata, MD, Chief Doctor, Department of Endoscopy, Shonan Fujisawa Tokushukai Hospital, 1-5-1 Tsujidokandai, Fujisawa, Kanagawa 251-0041, Japan. mitsuru10jp@yahoo.co.jp

Abstract

Endoscopic submucosal dissection (ESD) has been developed as a treatment for superficial gastrointestinal neoplasms, which can achieve en bloc resection regardless of the lesion size. However, ESD is technically difficult because endoscopists cannot bring their hand into the gastrointestinal tract, unlike surgeons in regular surgery. It is difficult to obtain sufficient tension in the dissection plane and a good field of vision. Therefore, ESD is associated with a long procedure time and a high risk of adverse events in comparison with endoscopic mucosal resection. Traction methods have been developed to provide sufficient tension for the dissection plane and a good field of vision during the ESD procedure. However, traction direction is limited in most traction methods, resulting in insufficient effect in some cases. Although traction direction is considered important, there have been few investigations of its effect. In the first half of this review, important traction methods are discussed, including traction direction. In second half, appropriate traction methods for each organ are considered. Other important considerations for traction method, such as ability to adjust traction strength, interference between traction device and endoscope, and the need for specialized devices are also discussed.

Key Words: Endoscopic submucosal dissection; Traction method; Countertraction; Traction direction; Vertical traction

©The Author(s) 2022. Published by Baishideng Publishing Group Inc. All rights reserved.

Core Tip: Endoscopic submucosal dissection is associated with a long procedure time and adverse events (e.g., perforation) due to technical difficulty-the absence of tension for the dissection plane and poor field of vision. Traction methods allow efficient dissection and a good field of vision. Although many traction methods have



WJG https://www.wjgnet.com

Commons Attribution

NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: htt p://creativecommons.org/License s/by-nc/4.0/

Received: March 18, 2021 Peer-review started: March 18, 2021 First decision: July 3, 2021 Revised: July 13, 2021 Accepted: December 28, 2021 Article in press: December 28, 2021 Published online: January 7, 2022

P-Reviewer: Li XB, Libânio D, Noh CK, Tsou YK S-Editor: Zhang H L-Editor: A P-Editor: Zhang H



been developed, traction direction is limited in most. Each traction method has advantages and disadvantages. It is important to select an appropriate traction method to obtain proper traction direction, depending on lesion location. We discuss the characteristics of different traction methods and their effects depending on traction direction.

Citation: Nagata M. Advances in traction methods for endoscopic submucosal dissection: What is the best traction method and traction direction? World J Gastroenterol 2022; 28(1): 1-22 URL: https://www.wjgnet.com/1007-9327/full/v28/i1/1.htm DOI: https://dx.doi.org/10.3748/wjg.v28.i1.1

INTRODUCTION

Endoscopic submucosal dissection (ESD) allows en bloc resection, regardless of lesion size, where endoscopic mucosal resection (EMR) is considered impossible, enabling accurate pathological assessment and a lower recurrence^[1]. However, ESD is still a challenging therapy due to technical difficulty, which results in long procedure time and high perforation rate^[2]. Surgeons may use the nondominant hand to provide traction for the lesion while they dissect using the dominant hand. By contrast, endoscopists cannot use their nondominant hand to provide traction for the lesion during ESD because they cannot put their hand into the gastrointestinal tract – it is like cutting a steak using only a knife, with no fork. It is important to obtain traction during ESD because this enables two important effects: creating a visual field by turning over the mucosal flap, and facilitating dissection by providing tension for the dissection plane. A basic hood attached to the endoscope can be used to obtain traction. However, this is occasionally insufficient.

The clip-with-line method (Figure 1), which may be the first traction method ever used, was reported in 2002[3,4]. Many other methods have since been developed. Although the clip-with-line method is simple and low cost, its traction direction is limited to the direction in which the line is pulled. In a multicenter prospective randomized controlled trial comparing the conventional and the clip-with-line methods, the clip-with-line method did not demonstrate a reduction in procedure time for gastric ESD^[5] but did for esophageal ESD^[6]. These results suggest the efficacy of the traction method is different depending on traction direction, because traction direction in the clip-with-line method is limited to the direction toward the endoscope in esophageal ESD, while it changes depending on the lesion location in gastric ESD.

Unlike the clip-with-line method, several other traction methods can provide traction in any direction. These include the internal traction method, which uses an spring-and-loop with clip (S-O clip; Zeon Medical, Tokyo, Japan)[7-10] (Figure 2). We reported a single-center prospective randomized controlled trial comparing the conventional and the S-O clip-assisted methods in gastric ESD, which demonstrated that the S-O clip-assisted method reduced the median gastric ESD procedure time (29.1 min vs 52.6 min; P = 0.005)[11]. In this study, a direction vertical to the gastric wall was selected for the S-O clip-assisted method, using its multidirectional traction function.

These outcomes suggest that traction direction is the important factor for tractionassisted ESD. However, little study has been done to explore the influence of traction direction during the procedure. Each traction method has characteristics other than traction direction, and it is necessary to understand the characteristics in order to use the methods effectively. The purpose of this article is to review the characteristic of traction methods. Then follows a discussion of appropriate traction methods for each organ, based on the results of clinical trials.

BASICS OF TRACTION

Definition of traction

In published literature of ESD, the terms traction and countertraction are used interchangeably; the unclear distinction results in a potential for confusion[12]. In this



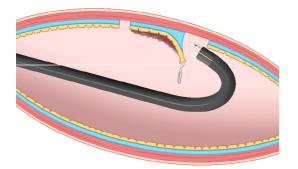


Figure 1 Clip-with-line method. This method provides traction for the lesion by pulling the line. The traction direction is limited to the direction in which the line is pulled.

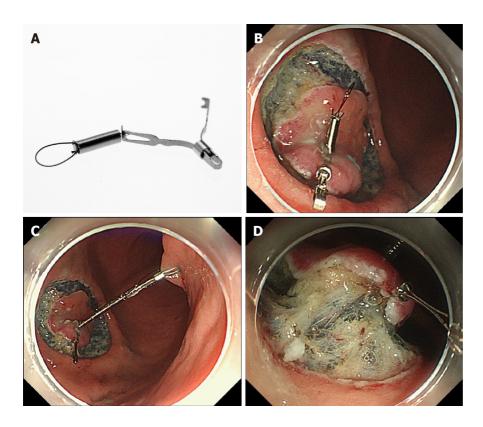


Figure 2 Internal traction method using the spring-and-loop with clip (Zeon Medical, Tokyo, Japan). A: The spring-and-loop with clip (S–O clip) has a 5-mm long spring and a 4-mm long loop at one side of the clip claws; B: The S-O clip is attached to the lesion; C: The regular clip anchors the loop of the S-O clip on the opposite side of the lesion; D: The extension of the spring provides traction on the lesion. Citation: Mitsuru Nagata. Internal traction method using a springand-loop with clip (S–O clip) allows countertraction in gastric endoscopic submucosal dissection. Surg Endosc 2020; 34(8): 3722–3733. Copyright © 2020 Mitsuru Nagata[10].

> article, we do not use the term countertraction. We define traction as force acting on the target lesion.

Classification of traction direction

As a force, traction can be represented by a vector, characterized by size and direction. Traction direction can be divided into the following five categories, according to the relationship with the endoscope tip and the gastrointestinal wall: Proximal, diagonally proximal, vertical, diagonally distal, and distal (Figure 3). Of these five categories, vertical traction may be appropriate in any situation because it provides two important effects: Enabling visualization of the submucosa, by turning over the mucosa; and facilitating submucosal dissection by providing tension to submucosa (Figure 3A).

Proximal traction can provide sufficient tension to the submucosa. However, the mucosal flap falls toward the endoscope (Figure 3B). If the endoscope tip is not parallel to the gastrointestinal wall, it can be difficult to approach the submucosa, even



WJG | https://www.wjgnet.com

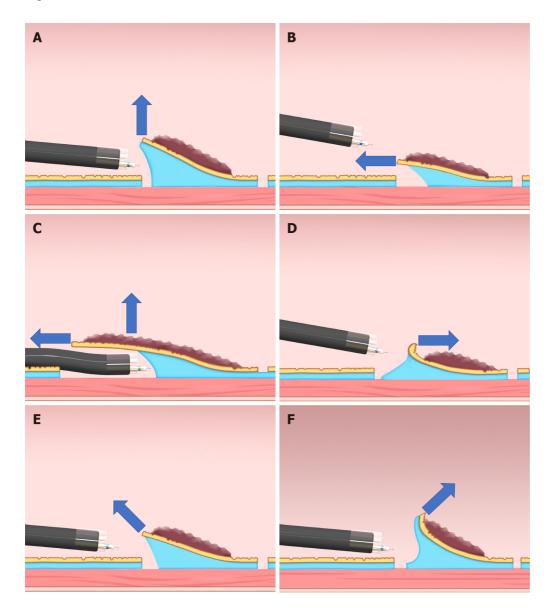


Figure 3 Classification of the traction direction. A: Vertical traction; B: Proximal traction; C: Proximal traction combined with hood traction; D: Distal traction; E: Diagonally proximal traction; F: Diagonally distal traction.

with proximal traction. If the endoscope tip is parallel to the gastrointestinal wall, it is easy to approach the submucosa, even if the mucosal flap falls down. Moreover, once the endoscope tip gets under the mucosal flap, proximal traction is combined with hood traction, resulting in diagonally proximal or vertical traction (Figure 3C). Proximal traction is suitable for situations where the endoscope tip can be placed parallel to the gastrointestinal wall, for example, in esophageal ESD.

Distal traction can cause the submucosal dissection plane to fall distally as submucosal dissection advances, resulting in submucosal thinning and subsequently, cutting the muscle layer or mucosa because of misrecognition of the layer (Figure 3D). Moreover, distal traction may decrease the effectiveness of the tension for the submucosal dissection plane, leading to inefficient dissection. Hence, distal traction may be the least useful approach for submucosal dissection in most cases.

Diagonally proximal traction (Figure 3E) and diagonally distal traction (Figure 3F) can be decomposed into horizontal and vertical vectors. The larger the horizontal component, the closer to the proximal or distal traction. The larger the vertical component, the closer to vertical traction.

MODALITY OF TRACTION

Traction can be roughly classified into hood traction, natural traction, and device-



WJG https://www.wjgnet.com

assisted traction. Natural traction and device-assisted traction can be further subdivided.

Hood traction

A hood attached to the endoscope tip is used in ESD procedures primarily to secure the visual field. The hood also can be used to obtain traction; it can turn the mucosal flap and provide tension for the submucosa after the endoscope tip is inserted under the mucosa. The straight hood has a wide field of view but is sometimes difficult to get under the mucosa. A hood with a tapered tip may be effective in such a situation (Figure 4)[13,14]. However, in a situation where it is difficult to keep the endoscope tip under the mucosa, such as in severe submucosal fibrosis, substantial lesion movement (due to patient respiration), and vertical confrontation with the lesion, the hood alone is not effective.

Natural traction

Natural traction is defined as traction using natural power, such as gravity, mucosal tension, buoyancy, and water pressure. The advantage of natural traction is that it is easy to switch to other methods and there is no need for any special device.

Gravity: When the lesion is gravitationally upward, gravity keeps the mucosal flap open and provides tension for the submucosa (Figure 5). Changing the patient's posture to raise the lesion against gravity is the basic strategy for ESD. However, in esophageal, gastric, and duodenal ESD, the patient's posture is primarily the left lateral decubitus position, which is difficult to change. By contrast, it is easy to change the patient's posture in colorectal ESD (*e.g.*, left lateral, supine, right lateral, and prone positions). However, changing the patient's posture sometimes makes the ESD procedure complicated, for example, through poor maneuverability of the endoscope, a vertical approach to the lesion, and difficulty opening the lumen.

Mucosal tension, pocket creation method, and endoscopic submucosal tunnel dissection: When the mucosa around the lesion is incised circumferentially, the lesion loses tension from the surrounding mucosa and submucosa, making it difficult to get the endoscope tip under the mucosal flap. By leaving a part of the mucosa around the lesion, the remaining mucosa gives tension to the lesion. In conventional ESD, traction at the lesion can be maintained by using mucosal tension, as follows. A C-shaped or inverted C-shaped mucosal incision is made. Next, the submucosa under the lesion is dissected, while the remaining mucosa gives tension at the dissection plane. Finally, a circumferential mucosal incision is made and the remaining submucosa dissected.

The pocket creation method (PCM) and endoscopic submucosal tunnel dissection (ESTD) use the same principle, using mucosal tension for traction[15-19]. In PCM (Figure 6), an initial mucosal incision on the proximal side of the lesion is first performed, to make entry to the submucosa. Then, the submucosa under the lesion is dissected, followed by creation of a submucosal space. Finally, the mucosa and submucosa around the submucosal space is dissected to achieve en bloc resection. In ESTD, a mucosal incision on the distal side of the lesion is performed before creation of a submucosal space is completed, unlike the PCM procedure.

PCM and ESTD procedures have similar advantages, as follows. The endoscope inside the submucosal space provides tension for the dissection plane. The endoscope tip can take a parallel approach to the muscle layer. The submucosal space holds the endoscope, which achieves stabilization of the endoscope. Thus, PCM and ESTD may be particularly suitable for lesions that are located where maneuverability of the endoscope is poor. Moreover, minimal mucosal incision until completion of submucosal dissection may prevent leakage of the injected solution.

Buoyancy and water pressure: We have reported the usefulness of underwater ESD for buoyancy, easier use of water pressure from an endoscope that has a water supply function, clear visual field, and heat sink effect[20,21]. Buoyancy and water pressure can provide a traction function, where buoyancy acts opposite to gravitational pull.

In conventional ESD, when the lesion is located at the gravitationally lower side, the opening of the mucosal flap is obstructed by gravity. Moreover, the lesion is half-way submerged due to gravity, while the boundary between gas and liquid obstructs the visual field (Figure 7A and B). By switching from conventional ESD to underwater ESD, buoyancy aids opening of the mucosal flap and provides tension for the submucosa, while the visual field is unaffected by a gas-liquid boundary (Figure 7C and D). Water pressure from the endoscope (using its water supply function) also assists opening the mucosal flap. Whereas water pressure can temporarily deteriorate

Nagata M. Advances in traction methods for ESD



Figure 4 Small-caliber tip transparent hood (Fujifilm, Tokyo, Japan). A: DH-33GR [small-caliber tip transparent (ST) hood] with 7-mm tip opening diameter and 7-mm tip protruding length; B: DH-28GR (short ST hood) with 8-mm tip opening diameter and 7-mm tip protruding length.

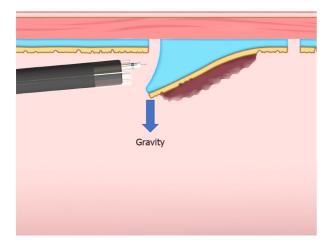


Figure 5 Gravity provides favorable traction when the lesion is located at the upper side of the gravitational force.

the visual field, due to splashing under gas insufflated conditions, it can be used without splashing in underwater conditions, allowing seamless submucosal dissection.

Saline solution is preferable for underwater ESD due to saline solution's higher specific gravity, compared with water, which provides a greater flotation effect.

Device-assisted traction method

A traction method using a device other than a hood can be defined as device-assisted. Device-assisted traction method implies traction in the narrow sense. They are broadly classified into external, internal, and other methods; each of these is further subclassified (Table 1).

External traction methods: External traction can be defined as a method where the traction device acts from outside the body. Representative external traction method includes the clip-with-line method, pulley method, sheath traction method, external forceps method, double scope method, and magnetic anchor method.

Clip-with-line method: The clip-with-line method was reported by Oyama et al[3,4] in 2002 (Figure 1). It is performed as follows. After circumferential mucosal incision, the endoscope is withdrawn. The clip applicator is deployed into the accessory channel of the endoscope. The clip-with-line (a clip with a line tied to its arm) is attached to the clip applicator. The endoscope is inserted, then the clip-with-line is attached to the edge of the lesion. Using this procedure, the line comes out of the body without passing through the accessory channel of the endoscope, while pulling the line provides traction at the lesion.

WJG | https://www.wjgnet.com

Table 1 Classification of device-assisted traction method				
	Traction direction	Control of traction force	Withdrawal of the endoscope	Recommended lesion location
External traction methods				
Clip-with-line method	One	Strengthen	Required	Esophagus, Greater curvature of the upper and middle third of the stomach, Colorectum
Pulley method	Any	Strengthen	Required	unclear because of fewer reports
Sheath traction method				
Clip-and-snare method	Two	Strengthen and weaken	Required	Stomach, Rectum
Endo Trac	Two	Strengthen and weaken	Required	Stomach, Rectum
External forceps method	Two	Strengthen and weaken	Required	Esophagus; Stomach except for cardia, lesser curvature or posterior wall of the upper gastric body; Rectum
Double scope method	Any	Strengthen and weaken	None	Stomach
Magnetic anchor method	Any	Strengthen and weaken	Required	Stomach, Colorectum
Internal traction method				
S-O clip	Any	Strengthen and weaken	None	Stomach, Colorectum
Ring thread	Any	Strengthen and weaken	None	Colorectum
Multiloop	Any	Strengthen and weaken	None	Colorectum
Double clip and rubber band	Any	Strengthen and weaken	None	Colorectum
Others				

The advantages of this method are its simplicity, low cost, and no requirement for a special device. The disadvantage is that traction direction is limited to the direction in which the line is pulled; therefore, submucosal dissection may be difficult, depending on traction direction. Although increasing traction force is possible, by pulling the line, it is difficult to weaken traction force. Moreover, friction between the endoscope and the line in the narrow space generates interference, which sometimes causes strong traction resulting in slip-off of the clip. In fact, the slip-off rate is reported to be 16.4% in esophageal ESD[6] and 13.2% in gastric ESD[5].

Pulley method: The pulley method is a modified clip-with-line method. By anchoring the line to the gastrointestinal wall, the direction of traction can be controlled in any direction (Figure 8). The pulley method can be classified into two types according to the pulley system used: Clip pulley[22] or suture pulley[23,24]. There are only a few reports on the pulley method, and its effectiveness needs to be verified.

Sheath traction method: In the sheath traction method, the line part of the clip-withline method is replaced with a sheath. Since the sheath is harder than the line, it can provide not only pulling force but also pushing force to the lesion, thus allowing two traction directions (Figure 9). Sheath traction method includes the clip-and-snare method[25-29] and the Endo Trac[30,31] (TOP, Tokyo, Japan) (Figure 10A).

The clip-and-snare method requires only a polypectomy snare and a clip; therefore, this method may be performed anywhere. In the conventional clip-and-snare method, the snare is grasped with forceps and delivered to the lesion[25]. However, this procedure is sometimes difficult. The prelooping technique was developed to improve the delivery of the snare[26-29]. The prelooping technique for the clip-and-snare method is performed as follows. After circumferential mucosal incision, the endoscope is withdrawn to preloop the snare on the tip of the endoscope. Then, the endoscope is inserted to the lesion along with the snare sheath. The clip is attached to the edge of Nagata M. Advances in traction methods for ESD

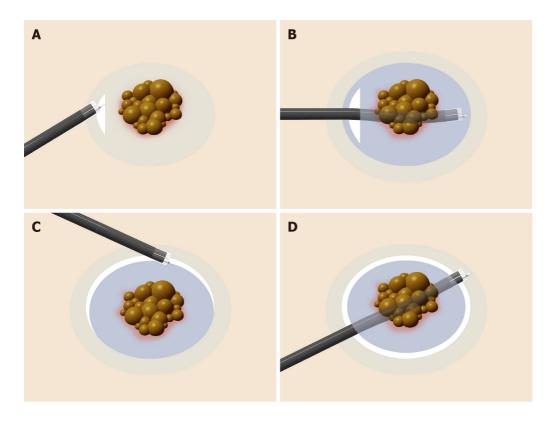


Figure 6 Procedure for the pocket creation method of endoscopic submucosal dissection. A: A minimal initial mucosal incision (approximately 20mm in width) is made for entry into the submucosa; B: Submucosal dissection is performed, followed by creation of a submucosal pocket under the lesion; C: Additional mucosal incision of the gravitational lower side of the submucosal pocket; D: Opening around the submucosal pocket, then, en bloc resection is achieved.

> the lesion. Then, the snare is loosened so that it can slide over the clip applicator toward the clip. Finally, the snare holds the clip, and the clip applicator is withdrawn.

> Endo Trac is a product developed for the sheath traction method. Whereas interference between the sheath and the tip of the endoscope sometimes makes access to the submucosa difficult when the clip-and-snare method is used, the Endo Trac has a structure that can release the sheath from the lesion, to avoid interference between the sheath and the endoscope tip (Figure 10B).

> The clip-and-snare method and the Endo Trac require withdrawal and reinsertion of the endoscope to set the traction system. Therefore, these methods are not suitable for colonic lesions, in which insertion of the endoscope is difficult and time-consuming. Moreover, interference between the sheath and the endoscope, due to friction, is possibly greater than with the clip-with-line method, due to the sheath being thicker than the line. In fact, it has been reported that even with a thin sheath (with a maximum diameter of 1.8 mm), interference with the endoscope can occur to some extent; the operator needs to move the endoscope carefully to avoid detachment of the snare from the clipped lesion[29].

> External forceps method: The external forceps method is performed as follows[32-36]. After circumferential mucosal incision, the endoscope is withdrawn. The endoscope is reinserted with external forceps that are grasped by second forceps inserted through the accessory channel of the endoscope. External forceps grasp the edge of the lesion (Figure 11A) while the second forceps are withdrawn. The external forceps provide traction to two directions by pulling or pushing the lesion (Figure 11B). This method allows changing the traction point, by releasing and re-grasping the lesion.

> However, this method has some disadvantages. It is difficult to deliver the external forceps, depending on lesion location, such as the cardia, lesser curvature or posterior wall of the upper gastric body, duodenum, and colon. Interference between the endoscope and the external forceps may be relatively strong compared with that of the clip-with-line and the sheath traction methods, because the forceps is thicker than the line and the sheath. Great care should be taken regarding potential damage to mucosa grasped by the external forceps, because of the strong traction.

> Double scope method: The double scope method is performed by two experienced endoscopists with main and second endoscopes[37,38]. The second endoscope is inserted alongside the main endoscope. Then, the second endoscope deploys the forceps, through the accessory channel, and grasps the lesion to provide traction. A



WJG | https://www.wjgnet.com

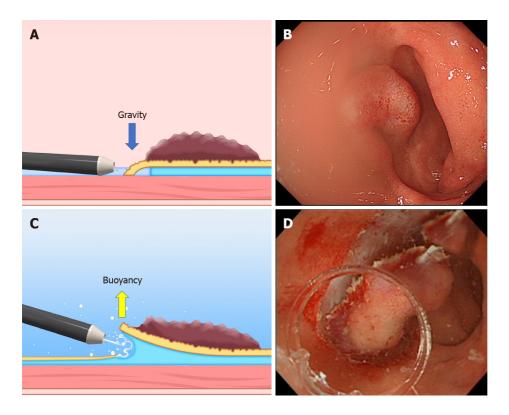


Figure 7 The difference between the conventional endoscopic submucosal dissection and the underwater endoscopic submucosal dissection. A and B: The conventional endoscopic submucosal dissection for the lesion is located at the gravitational lower side. Gravity obstructs the opening of the mucosal flap. Incomplete submersion deteriorates the visual field; C and D: The underwater condition aids the opening of the mucosal flap by buoyancy. Water pressure from the endoscope (using its water supply function) also assists in opening the mucosal flap. Complete submersion improves the visual field. Citation: Mitsuru Nagata. Underwater endoscopic submucosal dissection in saline solution using a bent-type knife for duodenal tumor. VideoGIE 2018; 3(12): 375-377. Copyright © 2018 American Society for Gastrointestinal Endoscopy. Published by Elsevier Inc[21].

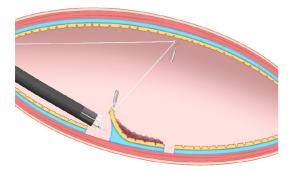


Figure 8 Pulley method. This method is the modified clip-with-line method, which can provide traction in any direction depending on the pulley site.

thin endoscope is recommended for the second endoscope, to avoid interference with the main endoscope.

This method has the great advantage that traction direction can be easily controlled by the second endoscope. Although the indication may be limited, as this method requires two experienced endoscopists and two endoscope systems, it may be a useful option for difficult cases, such as gastric cancers with ulcer scar[39]. This method has been reported to be useful in the treatment of superficial pharyngeal cancers[40] and gastric submucosal tumors[41].

Magnetic anchor method: The magnetic anchor method, as initially reported, used a large external electromagnet to provide traction, by moving an internal magnet attached to the lesion[42,43]. However, it was necessary to miniaturize the external electromagnet in clinical practice. Recently, use of neodymium rare earth magnets has allowed the external electromagnet to be minimized [44,45]; the feasibility of this method in clinical practice has been demonstrated in a prospective trial[46]. Although this method requires a special magnetic device and is not yet widespread, it is a promising method for the future due to the great advantage that it can provide Nagata M. Advances in traction methods for ESD

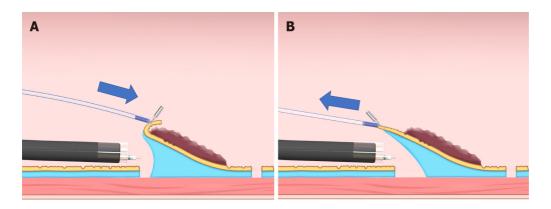


Figure 9 Sheath traction method. This method has two traction directions by pushing or pulling the sheath. A: Pushing the sheath; B: Pulling the sheath.

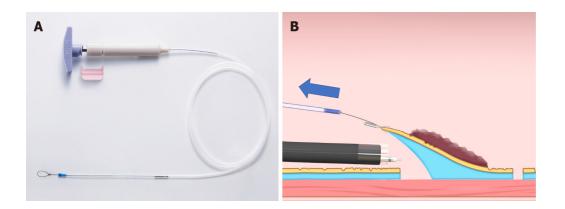


Figure 10 Endo Trac (TOP, Tokyo, Japan). A: This device can be used for the sheath traction method; B: This device has a structure that can release the sheath from the lesion to avoid interference between the sheath and the endoscope tip.

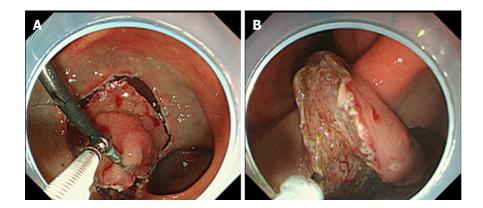


Figure 11 Endoscopic submucosal dissection using external forceps. A: External grasping forceps was anchored at the distal margin of the lesion in the lesser curvature of the antrum under the control of the endoscope and a second grasping forceps; B: With gentle oral traction applied with the external grasping forceps, the submucosal layer was dissected in retroversion from the aboral side. Citation: Imaeda H, Hosoe N, Kashiwagi K, Ohmori T, Yahagi N, Kanai T, Ogata H. Advanced endoscopic submucosal dissection with traction. World J Gastrointest Endosc 2014; 6(7): 286-295. Copyright © 2014 Baishideng Publishing Group Inc. Published by Baishideng Publishing Group Inc[35].

traction in any direction and control traction force. It should be noted that patients with a cardiac pacemaker or implantable cardioverter-defibrillator are not indicated for this method.

Internal traction methods: An internal traction method can be defined as a method in which the traction device acts only inside the gastrointestinal tract. Devices for internal traction include the S-O clip[7,8] (Zeon Medical, Tokyo, Japan) (Figure 2), ring thread [47], multiloop[48], double clip and rubber band[49], and clip band[50]. The principle for generating traction is the same in these devices, as follows. First, the clip with the



Zaishidena® WJG https://www.wjgnet.com

specific mechanism (e.g., spring, thread, and band) for generating the traction is attached to the lesion (Figure 12A). Second, the regular clip anchors the tip of the mechanism to the gastrointestinal wall (Figure 12B). Finally, extension of the mechanism provides traction to the lesion (Figure 12C).

In these devices, the traction direction can be controlled in any direction by the anchor site. Traction force can also be controlled to some extent by inflating or collapsing the lumen. These devices are useful especially for colorectal ESD, as withdrawal of the endoscope is not required. The disadvantage is that a certain distance between the anchor clip and the clip attached to the lesion is required to generate the traction force. Therefore, these devices are usually utilized for gastric ESD [9-11] or colorectal ESD[47-50].

These devices are designed primarily for use with the forward endoscopic position, because there is a possibility that, in the retroflexed position, the endoscope may stretch the traction device, resulting in laceration of the mucosal flap or slip-off or breakage of the traction device. In gastric ESD, the retroflexed endoscopic position is as common as the forward position, due to the large lumen, unlike colorectal ESD. Therefore, we developed a modified method for attaching the S-O clip, to avoid stretching of the spring by the endoscope[9-11] (Figure 13). Although there are several devices for internal traction, the S-O clip may be the most appropriate in gastric ESD, as the S-O clip has a spring with higher elasticity than a thread or band. The elasticity of the spring can be easily adjusted for a large lumen, preventing laceration of the mucosal flap, slip-off, or breakage of the traction device. The S-O clip is sold only in Japan currently. However, it will be sold in future in Asian countries under the brand name "Countertraction CLIP".

The management of the anchor clip after traction is not standardized. Conventionally, the traction mechanism of the device (e.g., thread, band) is cut to detach the resected specimen, while the anchor clip remains on the gastrointestinal wall. In colorectal ESD, the anchor clip may naturally drop by vermiculation. In contrast, vermiculation of the stomach is poor, except in the pars pylorica; there is a possibility of a permanent residual of the anchor clip after gastric ESD[51]. Therefore, we usually detach the anchor clip with forceps. So far, we have not experienced any adverse events from detaching the anchor clip (e.g., perforation, post-ESD bleeding from the anchor site), probably because of the thicker stomach wall, compared with other organ of the gastrointestinal tract[10,11]. The safety of this management method for the anchor clip should be assessed in many gastric cases.

Others

Pocket creation method with traction device: The combination of the PCM and traction device (TD) for internal traction has been reported to facilitate better mucosal flap formation and opening of the submucosal pocket, compared with conventional PCM[52]. A retrospective study demonstrated that the median dissection speed in PCM with TD was significantly greater than in conventional ESD with TD (16.6 mm² $/\min vs \ 12.2 \ mm^2 / min; P = 0.003)[53]$. Additional studies are needed to confirm whether TD has an additional effect in PCM, by comparing PCM alone against PCM with TD.

Clip flap method: The clip flap method has been reported as using a clip attached to the edge of the lesion to substitute for the mucosal flap until it is made (Figure 14)[54-56]. By using the clip flap method together with clip-based traction (e.g., clip-withline), the procedure of getting under the mucosal flap can be facilitated, especially when proximal traction makes the mucosal flap fall down (Figure 3B). Although a randomized controlled trial comparing the conventional method and clip flap method in gastric ESD demonstrated that the clip flap method had no advantage in efficacy and safety[57], this method may be effective when it is used along with other clipbased traction methods.

TRACTION METHODS ASSOCIATED WITH LESION SITE

Esophageal ESD

Representative traction methods that are reported to be effective in esophageal ESD include the clip-with-line method and ESTD. A multicenter randomized controlled trial demonstrated that the median ESD procedure time was significantly shorter with the clip-with-line method (n = 116) than with the conventional method (n = 117) (44.5



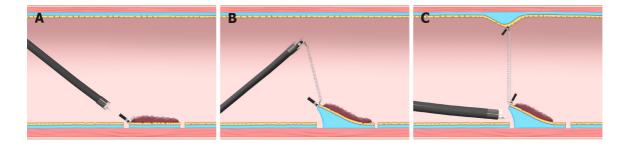


Figure 12 Internal traction method using the spring-and-loop with clip (Zeon Medical, Tokyo, Japan). A: The spring-and-loop with clip (S–O clip) is attached to the lesion; B: The regular clip anchors the loop part of the S–O clip on the gastrointestinal wall; C: The extension of the spring provides traction on the lesion. The traction direction can be controlled by the anchor site. Citation: Nagata M, Fujikawa T, Munakata H. Comparing a conventional and a spring-and-loop with clip traction method of endoscopic submucosal dissection for superficial gastric neoplasms: a randomized controlled trial (with videos). *Gastrointest Endosc* 2021; 93(5): 1097-1109. Copyright © 2021 American Society for Gastrointestinal Endoscopy. Published by Elsevier Inc[11].

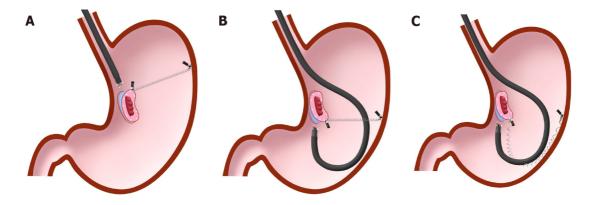


Figure 13 Spring-and-loop with clip-assisted gastric endoscopic submucosal dissection. A: Forward endoscopic position; B: Retroflexed endoscopic position; C: The endoscope has the possibility to stretch the spring, resulting in a loss of spring elasticity in the retroflexed endoscopic position. In this situation, the modified attachment method that we described in detail in the previous papers[9-11] is required to avoid this problem. Citation: Nagata M, Fujikawa T, Munakata H. Comparing a conventional and a spring-and-loop with clip traction method of endoscopic submucosal dissection for superficial gastric neoplasms: a randomized controlled trial (with videos). *Gastrointest Endosc* 2021; 93(5): 1097-1109. Copyright © 2021 American Society for Gastrointestinal Endoscopy. Published by Elsevier Inc[11].

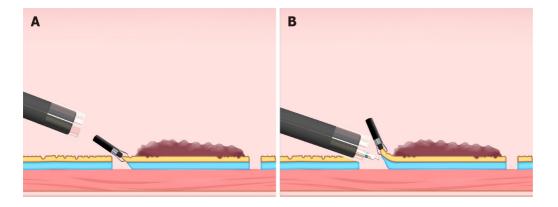


Figure 14 Clip flap method. A: The clip is attached to the edge of the lesion; B: The clip can be used as a substitute for the mucosal flap.

min vs 60.5 min; P < 0.001)[6]. Although traction using the clip-with-line method in esophageal ESD is limited to proximal traction, because the forward endoscopic position is predominantly used, due to the narrow cylindrical esophageal lumen, proximal traction may be effective, because the endoscope tip can approach parallel to the esophageal wall and can easily access the submucosa without vertical traction. After getting under the mucosal flap, hood traction and proximal traction using the clip-with-line method are combined, providing diagonally proximal or vertical traction to the submucosa (Figure 3C). Remarkably, the conventional method was changed to the clip-with-line method in six patients (5.2%) because of technical

difficulties. Moreover, five patients (4.3%) experienced perforation under the conventional method, whereas one patient (0.9%) could not complete the ESD procedure because of perforation. Conversely, no perforations were observed in the clip-with-line method.

In a multicenter randomized controlled trial, the clip slip-off rate was reported as 16.4%[6]. If clip slip-off occurs, there is a possibility that histopathological evaluation for the margin is made difficult due to damage to the specimen. Moreover, clip slip-off requires reattaching the clip-with-line, which is time-consuming. Interference between the endoscope and the line due to friction causes slip-off. Therefore, once the clip-withline is attached to the lesion, unnecessary movement and withdrawal of the endoscope should be avoided.

A propensity score matching analysis^[58] showed that ESTD had a shorter median ESD procedure time (38.0 min vs 48.0 min; P = 0.006) and lower muscle injury rate (28.9% vs 52.6%; P = 0.036) compared with conventional ESD. Furthermore, a metaanalysis including 17 studies[59] showed that ESTD had significantly higher en bloc resection rate, shorter ESD procedure time, and lower muscle injury rate. In ESTD, the endoscope tip is held by the submucosal tunnel, which allows stabilization of the endoscope and a parallel approach to the muscle layer. The endoscope tip inside the submucosal tunnel pushes up the lesion, providing sufficient tension at the dissection plane. These advantages of the ESTD may provide a shorter ESD procedure time and a lower muscle injury rate.

In conclusion, many promising results have been reported for the clip-with-line method and ESTD. At present, it may be better to select either of these two methods. Most studies on traction method for esophageal ESD have been reported from Asia, mainly targeting squamous cell carcinoma. There are not many reports of tractionassisted ESD for Barrett's esophageal adenocarcinoma, located around the esophagogastric junction; future studies should focus on this issue.

Gastric ESD

As the stomach lumen is large, both the forward and retroflexed endoscopic positions are common, unlike in esophageal, duodenal, and colonic ESD. Therefore, it is desirable that a traction method for gastric ESD is easy to utilize in both forward and retroflexed endoscopic positions. The popular traction methods for gastric ESD include clip-with-line, internal traction, sheath traction, and ESTD.

The clip-with-line method may be the first traction method for gastric ESD, and was reported in 2002[3,4]. However, a multicenter randomized controlled trial[5] comparing the conventional ESD (n = 316) and the clip-with-line method (n = 319) failed to show a reduction in the mean procedure time for gastric ESD in the total population (conventional ESD, 60.7 min vs clip-with-line method, 58.1 min; P = 0.45). Since traction by the clip-with-line method in gastric ESD is limited to the cardia, the direction of traction varies depending on the lesion location. In the retroflexed endoscopic position, the traction is likely to be distal, especially for lesions located at the lesser couverture side of the gastric body (Figure 15A). Distal traction may cause the submucosal dissection plane to fall distally, making the procedure difficult and prolonging the procedure time in some cases. In the forward endoscopic position, the traction may be proximal or diagonally proximal (Figure 15B). If the endoscope tip cannot be parallel to the gastric wall, proximal traction may cause the mucosal flap to fall proximally, making it difficult to approach the submucosal layer. In contrast, a subgroup analysis based on lesion location demonstrated that the mean ESD procedure time for lesions located at the greater curvature of the upper and middle third of the stomach was significantly shorter in the clip-with-line method (104.1 min vs 57.2 min; P = 0.01). From an anatomical point of view, these results seem reasonable, because it is difficult for the clip-with-line method to provide vertical traction unless the lesion is located at the greater curvature (Figure 15C). In a subgroup analysis based on operator experience, the mean ESD procedure time was not significantly different between the conventional and clip-with-line methods in an expert group (58.0 min vs 58.0 min; P = 1.00). Conversely, the mean ESD procedure time in a trainee group tended to be better in the clip-with-line method (68.9 min vs 58.3 min; P = 0.13). However, the lesions managed by trainees were primarily easy cases; therefore, a simple comparison may be inaccurate. Nonetheless, the analysis result suggests that the benefit from clip-with-line method differs depending on the operator experience.

S-O clip-assisted ESD is classified as internal traction, and can provide traction in any direction. Since the use of the S-O clip in gastric ESD has a potential for the endoscope in the retroflexed position to stretch the spring, we have developed a modified attachment method for the S-O clip, to avoid interference between the endoscope and spring part of the clip[9,10]. We reported a single-center randomized



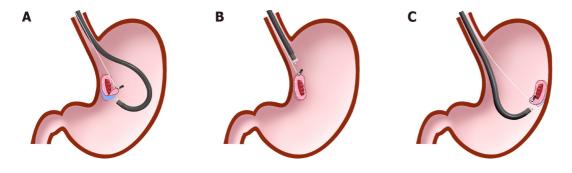


Figure 15 Difference in traction direction depending on the lesion location in the clip-with-line method. A: Distal traction; B: Proximal traction; C: Vertical traction.

controlled trial comparing conventional (n = 40) and S-O clip-assisted ESD (n = 40), which showed that the median ESD procedure time was significantly shorter in S-O clip-assisted ESD than in conventional ESD (29.1 min vs 52.6 min; P = 0.005)[11]. According to the subgroup analysis comparing the ESD procedure time by lesion location, the median ESD procedure time of S-O clip-assisted ESD was significantly shorter than that of conventional ESD in the upper and middle third of the stomach (39.4 min vs 58.3 min; P = 0.005). In the lower third of the stomach, the two methods were not significantly different (22.8 min vs 36.2 min; P = 0.146). Essentially, the ESD procedure performed in the lower third of the stomach is easier than that performed in the upper and middle third of the stomach [60,61]. The difference in the difficulty of the ESD procedure may explain the difference in the subgroup analysis outcomes. Therefore, the S-O clip-assisted ESD is especially recommended for lesions in the upper and middle third of the stomach. Meanwhile, en bloc resection, R0 resection, perforation, and post-ESD bleeding showed no significant difference between the two groups. The S-O clip slip-off rate was only 2.5%, probably because the modified attachment method prevented interference between the endoscope and spring part of the clip. In this trial, vertical traction was selected for the S–O clip-assisted ESD using its multidirectional traction function. Considering this result and the result of subgroup analysis of the above-mentioned multicenter randomized controlled trial of the clip-with-line method, vertical traction may be the optimal traction direction for gastric ESD. Although other internal traction methods, including the pulley, double scope, and magnetic anchor methods, may be able to provide vertical traction, their feasibility in gastric ESD is unclear and needed to be assessed.

The usefulness of the sheath traction method in gastric ESD has been reported. Unlike the clip-with-line method, the sheath traction method allows traction not only in the pulling direction but also in the pushing direction, so distal traction can be avoided in both forward and retroflexed endoscopic positions. A retrospective study comparing conventional ESD (n = 20) and the clip-and-snare method (n = 20) demonstrated that the clip-and-snare method significantly reduced the median ESD procedure time (38.5 min vs 59.5 min; P = 0.023)[29]. En bloc resection was achieved without perforation in all the patients in both groups. A case series of 21 challenging gastric ESD cases treated using the Endo Trac reported that the ability to change the traction direction in both proximal and distal sides was 100% [31]. Although these results are promising, the stress on the operator due to possible interference between the sheath and the endoscope is concerning. Moreover, these studies have the limitations of being retrospective studies with small numbers of cases. Therefore, evaluation in large-scale studies is warranted.

ESTD was reported to be useful not only for esophageal ESD but also for gastric ESD. A retrospective study evaluating 799 consecutive cases of gastric ESD in single institution showed that resection speed using ESTD was greater than with conventional ESD (19.3 mm²/min vs 17.7 mm²/min; P = 0.009)[62]. Perforation was significantly less frequent in ESTD (0.9% vs 6.0%; P = 0.035). However, the creation of a submucosal tunnel in the stomach may be more difficult compared to the esophagus because the stomach lumen is not straight. It has been reported difficult to form a submucosal tunnel for a lesion located at the pylorus ring or the greater curvature side of the fornix[62]. On the other hand, lesions located at the cardia, the lesser curvature of the gastric corpus, and the greater curvature of the antrum are reported to be suitable for ESTD[63].

In summary, the internal traction method using the S-O clip with modified attachment method has the potential to be the most appropriate traction method for

gastric ESD. The clip-with-line method and ESTD may be effective methods in gastric ESD if the lesion location is appropriate for these methods.

Colon and rectal ESD

Colonic ESD is more challenging than esophageal and gastric ESD because the maneuverability of the endoscope is limited, the colorectal lumen is angulated, and the muscle layer is thin and easy to perforate. Withdrawal and reinsertion of the endoscope is time-consuming in colonic ESD, unlike ESD in the upper gastrointestinal tract and rectum. Therefore, for colonic ESD, a traction method that does not require withdrawal and reinsertion of the endoscope is suitable. By contrast, it is easy to utilize most traction methods in rectal ESD.

Internal traction methods are suitable not only for rectal ESD but also colonic ESD because they do not require withdrawal and reinsertion of the endoscope. Recently, several novel devices for internal traction in colorectal ESD have been reported, such as S-O clip[7,8], ring thread[47], multiloop[48], double clip and rubber band[49], and clip band^[50]. These devices have the common advantage of controlling traction direction at anchor site. Among them, the S-O clip is made of a highly elastic spring that can be used flexibly, regardless of the lesion location. A prospective randomized controlled trial comparing conventional (n = 27) and S–O clip-assisted ESD (n = 23) demonstrated that the mean ESD procedure time for S-O clip-assisted ESD was significantly shorter than that for conventional ESD (37.4 min vs 67.1 min; P = 0.03) [64]. No significant differences were observed in en bloc resection, perforation, and post-ESD bleeding. Although the conventional ESD was converted into the S-O clipassisted ESD in eight cases, these cases remained in the conventional ESD group. In most of these conversion cases, the lesions were located in flexural areas where endoscope maneuverability is poor; these areas were the sigmoid colon, hepatic flexure, and splenic flexure. In these areas, reaching under the mucosal flap by the endoscope tip is difficult when only used with hood traction. The S-O clip helps the endoscope tip reach under the mucosal flap, providing proper visualization of the submucosa, despite poor endoscope maneuverability. Traction-assisted ESD using ring thread^[47], multiloop^[65], or double clip and rubber band^[49] also showed promising treatment results in clinical trial, compared with conventional ESD. Further studies should focus on which traction direction is appropriate for colorectal ESD by using multidirectional traction function of internal traction methods.

PCM is another traction method that does not require withdrawal and reinsertion of the endoscope. In this method, the submucosal pocket holds the endoscope, allowing stable endoscope maneuverability. Moreover, the endoscope inside the submucosal pocket pushes up the lesion and provides sufficient tension at the dissection plane. A prospective randomized controlled trial comparing PCM (n = 59) and the conventional method (n = 55), conducted at three Japanese institutions, reported that the rate of ESD completion (defined as completion of colorectal ESD in three hours with en bloc resection using the assigned ESD method without changing to other methods or other devices and without perforation during the procedure) was significantly higher in PCM compared with conventional ESD (93% vs 73%; P = 0.01)[66]. By contrast, the median dissection speed was not significantly different between the two methods (15.9 $mm^2/min vs 17.4 mm^2/min; P = 0.81$). This was unforeseen, as several retrospective studies had reported dissection speed significantly greater in PCM than in the conventional method [67,68]. A novel method that combines the PCM and internal traction has been developed[52,53], and it can possibly accelerate the dissection speed. A metaanalysis including five studies (two randomized controlled trials and three retrospective studies) evaluated the efficacy and safety of PCM in comparison with the conventional method for superficial colorectal neoplasms^[69]. PCM achieved a higher R0 resection rate (93.5% vs 78.1%; OR, 3.4; 95%: 1.3-8.9; I² = 58%), a higher en bloc resection rate (99.8% vs 92.8%; OR, 9.9; 95%CI: 2.7-36.2; I² = 0), a shorter procedure time (min) [mean difference (MD), -11.5; 95%CI: -19.9 to -3.1; I² = 72%], a faster dissection speed (mm²/min) (MD, 3.6; 95%CI: 2.8-4.5; $I^2 = 0$), and a lower overall adverse event rate (4.4% vs 6.6%; OR, 0.6; 95% CI: 0.3–1.0; I² = 0) than the conventional method. However, all the included studies were conducted in Japan, with only two randomized controlled trials. Hence, further study is needed, especially regarding dissection speed.

The conventional clip-with-line method requires withdrawal and reinsertion of the endoscope, which may be troublesome during colonic ESD for lesions located where it is difficult to insert the endoscope. Modified preparation techniques for the clip-withline method have been developed that eliminate withdrawal and reinsertion of the endoscope[70]. A single-center prospective randomized controlled trial comparing the clip-with-line method with modified preparation technique (n = 42) against the

conventional method (n = 42) demonstrated that the median colorectal ESD procedure time was significantly shorter in the modified clip-with-line method than in the conventional method (40 min vs 70 min; P < 0.0001)[71]. No significant differences were noted in en bloc resection, R0 resection, perforation, and post-ESD bleeding. In this study, two experts and two intermediates performed the colorectal ESD procedures. When the intermediates encountered difficult situations, the experts took over the procedure. The intermediates' self-completion rate was significantly higher in the modified clip-with-line method than in the conventional method (100% vs 90%; P =0.04). Although this modified preparation technique is a little tricky, and the clip-withline method is not able to control the traction direction, it has the advantage that does not require any special device. Since colorectal ESD is generally performed in the forward endoscopic position, the clip-with-line method may provide diagonally proximal or proximal traction for colorectal ESD. The clip-with-line method may be effective, as long as the endoscope has a parallel approach to colorectal wall.

The sheath traction, clip-and-snare, and Endo Trac methods also requires withdrawal and reinsertion of the endoscope. However, the sheath traction method was reported to be utilized even for colonic lesions[27,31]. A retrospective study reported that the clip-and-snare method (n = 17) significantly reduced mean colorectal ESD procedure time compared with conventional method (n = 123) (45.6 min vs 70.1 min; P = 0.047)[28]. There were no significant differences in en bloc resection, curative resection and adverse events (perforation and post-ESD bleeding). The sheath traction method has the great advantage that it can control traction direction to some extent by pushing or pulling the sheath, which may facilitate the ESD procedure. Moreover, the clip-and-snare method may be useful in any country, because it does not require any special device. Although reinsertion of the endoscope during the sheath traction method is occasionally troublesome, a balloon overtube may help address the issue, simplifying insertion of the endoscope.

The usefulness of underwater techniques during colorectal ESD has been reported [20,72-75]. Underwater conditions provide buoyancy (classified as natural traction) which can help turn over the mucosal flap of a lesion located lower gravitationally (Figure 7C and D). Although colorectal ESD is generally performed with the patient's posture such that the target lesion is on the upper side of gravity, to open the mucosal flap by gravity, it is difficult to select this posture in some cases due to poor endoscope maneuverability, a vertical approach to the lesion, and difficulty opening the lumen. Water pressure from the endoscope using its water supply function can be used as a traction method at any time. Water pressure can be used even in the conventional method. However, splashing can sometimes obstruct the visual field. In underwater conditions, splashing can be avoidable, which makes it easier to get under the mucosal flap. The underwater condition provides a good field of vision through a zoom effect and the disappearance of halation; this facilitates colorectal ESD in a poor field of vision due to severe submucosal fibrosis or fat tissue (Figure 16). We reported a case series study that demonstrated the feasibility and safety of underwater techniques for colorectal ESD[20]. However, additional studies are needed to evaluate whether underwater techniques improve colorectal ESD procedures compared with conventional methods.

In conclusion, PCM and internal traction methods (e.g., S-O clip, ring thread, multiloop, double clip and rubber band, clip band) are especially recommended for colorectal ESD, based on the results of recent studies.

Duodenal ESD

Duodenal ESD is extremely challenging due to the fragile muscle layer, thin submucosal layer, and poor maneuverability of the endoscope, resulting in a high perforation rate which has been reported as 8.8% to 27.0% [76-78]. However, there are few reports of effective methods for reducing the procedural difficulty of duodenal ESD, because superficial duodenal epithelial tumors indicated for ESD are rare.

PCM has been reported to be useful for the treatment of superficial duodenal epithelial tumors. A retrospective study showed that perforation was significantly less frequent in PCM [7% (2/28)] than in the conventional method [29% (5/17; P = 0.046)] [79]. PCM demonstrated a faster dissection speed (9.4 mm²/min vs 6.5 mm²/min; P =0.09) and a higher en bloc resection rate (100% vs 88%; P = 0.07) than the conventional method, although the differences were not statistically significant. PCM may prevent leakage of the injected solution from the submucosa due to minimal mucosal incision until completion of submucosal dissection. Moreover, the submucosal pocket holds the endoscope, providing stable endoscope maneuverability. In the conventional method for duodenal ESD, the mucosa around the lesion is incised widely before the completion of submucosal dissection, while the injected solution in the submucosa may



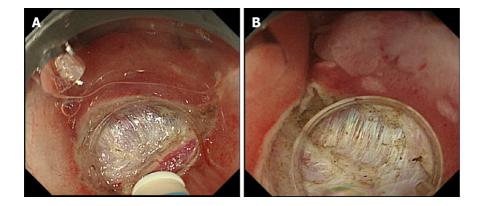


Figure 16 Comparison of aerial and underwater endoscopic view. A: Severe fibrosis and halation makes the boundary between the submucosal layer and muscle layer unclear; B: Submergence enables a detailed observation through a natural zoom effect and causes halation disappearance, thereby clarifying the boundary between the submucosal layer and muscle layer. Citation: Nagata M. Usefulness of underwater endoscopic submucosal dissection in saline solution with a monopolar knife for colorectal tumors (with videos). Gastrointest Endosc 2018; 87(5): 1345-1353. Copyright © 2018 American Society for Gastrointestinal Endoscopy. Published by Elsevier Inc[20].

easily flow out and the endoscope operability deteriorates.

Underwater techniques are another option for the treatment of superficial duodenal epithelial tumors [21,80]. Severe submucosal fibrosis in duodenal submucosa occasionally exists and causes insufficient submucosal elevation, even with a large quantity of injection. Although submucosal fibrosis generally makes it difficult to get under the mucosal flap during the first half of duodenal ESD, water pressure from the endoscope using its water supply function helps turn over the mucosal flap and enable the endoscope to get under the mucosal flap. Since the underwater condition eliminates splashing, unlike under gas supply conditions, the endoscope can get under the mucosal flap seamlessly after generating water pressure. This technique is classified as natural traction, and can be used repeatedly at any time.

The underwater condition has several useful effects based on nature, such as a zoom effect, the disappearance of halation, buoyancy, and a heat sink effect. The zoom effect and the disappearance of halation allow recognition of the proper dissection plane despite severe submucosal fibrosis. Buoyancy can be classified as natural traction and aids the opening of the mucosal flap when the lesion is gravitationally lower (Figure 7C and D). The heat sink effect minimizes thermal damage to the muscle layer from the ESD procedure. Thermal damage may increase the risk of delayed perforation after duodenal ESD, which leads to serious complications[81]; therefore, the underwater techniques may be suitable for duodenal ESD.

The underwater techniques may be able to be combined with other traction methods. In fact, a case report showed the usefulness of the underwater techniques with PCM and internal traction during duodenal ESD[82]. Although underwater techniques have major disadvantages, such as visual field loss due to active bleeding, gel immersion endoscopy, which secures the visual field during bleeding, may help address this issue^[83]. Underwater techniques have the potential to reduce difficulty in ESD procedures through unique effects not found in conventional methods under gas supply condition. Further study should focus on the efficacy of ESD with underwater techniques, especially in the duodenum.

Several traction methods other than PCM and underwater techniques for duodenal ESD have been reported, such as internal traction using the S–O clip[84] and the sheath traction method[31]. However, there are few reports about these methods, and the efficacy of these methods is still unclear.

In summary, PCM and underwater techniques have the potential to facilitate the duodenal ESD procedure, decreasing the risk of perforation.

CONCLUSION

The purpose of traction during ESD is to create a visual field by turning over the mucosal flap and facilitate dissection by providing tension for the dissection plane. In order to achieve these effects, it is important to understand the advantages and disadvantages of each traction method and to use a traction method that is most appropriate as per the situation. The results of previous studies suggest that traction



direction affects the effectiveness of the traction method. Therefore, traction direction should be considered when choosing a traction method. Although there are increasing reports of methods that can control the traction direction, further studies should focus on investigating the optimal traction direction and its influence on the effectiveness of the traction method.

REFERENCES

- Suzuki H, Oda I, Abe S, Sekiguchi M, Mori G, Nonaka S, Yoshinaga S, Saito Y. High rate of 5-year survival among patients with early gastric cancer undergoing curative endoscopic submucosal dissection. Gastric Cancer 2016; 19: 198-205 [PMID: 25616808 DOI: 10.1007/s10120-015-0469-0]
- Rösch T, Sarbia M, Schumacher B, Deinert K, Frimberger E, Toermer T, Stolte M, Neuhaus H. 2 Attempted endoscopic en bloc resection of mucosal and submucosal tumors using insulated-tip knives: a pilot series. Endoscopy 2004; 36: 788-801 [PMID: 15326574 DOI: 10.1055/s-2004-825838]
- Oyama T, Kikuchi Y, Shimaya S, Tomori T, Hotta K, Miyata Y, Yamada S. Endoscopic mucosal resection using a hooking knife (hooking EMR) [in Japanese]. Stomach Intest 2002; 37: 1151-1161 [DOI: 10.11477/mf.1403104523]
- Oyama T. Counter traction makes endoscopic submucosal dissection easier. Clin Endosc 2012; 45: 4 375-378 [PMID: 23251884 DOI: 10.5946/ce.2012.45.4.375]
- Yoshida M, Takizawa K, Suzuki S, Koike Y, Nonaka S, Yamasaki Y, Minagawa T, Sato C, Takeuchi 5 C, Watanabe K, Kanzaki H, Morimoto H, Yano T, Sudo K, Mori K, Gotoda T, Ono H; CONNECT-G Study Group. Conventional versus traction-assisted endoscopic submucosal dissection for gastric neoplasms: a multicenter, randomized controlled trial (with video). Gastrointest Endosc 2018; 87: 1231-1240 [PMID: 29233673 DOI: 10.1016/j.gie.2017.11.031]
- 6 Yoshida M, Takizawa K, Nonaka S, Shichijo S, Suzuki S, Sato C, Komori H, Minagawa T, Oda I, Uedo N, Hirasawa K, Matsumoto K, Sumiyoshi T, Mori K, Gotoda T, Ono H; CONNECT-E Study Group. Conventional versus traction-assisted endoscopic submucosal dissection for large esophageal cancers: a multicenter, randomized controlled trial (with video). Gastrointest Endosc 2020; 91: 55-65.e2 [PMID: 31445039 DOI: 10.1016/j.gie.2019.08.014]
- Sakamoto N, Osada T, Shibuya T, Beppu K, Matsumoto K, Shimada Y, Konno A, Kurosawa A, 7 Nagahara A, Ohkusa T, Ogihara T, Watanabe S. The facilitation of a new traction device (S-O clip) assisting endoscopic submucosal dissection for superficial colorectal neoplasms. Endoscopy 2008; 40 Suppl 2: E94-E95 [PMID: 19085712 DOI: 10.1055/s-2007-995603]
- 8 Sakamoto N, Osada T, Shibuya T, Beppu K, Matsumoto K, Mori H, Kawabe M, Nagahara A, Otaka M, Ogihara T, Watanabe S. Endoscopic submucosal dissection of large colorectal tumors by using a novel spring-action S-O clip for traction (with video). Gastrointest Endosc 2009; 69: 1370-1374 [PMID: 19403131 DOI: 10.1016/j.gie.2008.12.245]
- Nagata M. Modified attachment method using an S-O clip for gastric endoscopic submucosal dissection. VideoGIE 2019; 4: 151-153 [PMID: 31032463 DOI: 10.1016/j.vgie.2018.12.015]
- Nagata M. Internal traction method using a spring-and-loop with clip (S-O clip) allows 10 countertraction in gastric endoscopic submucosal dissection. Surg Endosc 2020; 34: 3722-3733 [PMID: 32350668 DOI: 10.1007/s00464-020-07590-9]
- 11 Nagata M, Fujikawa T, Munakata H. Comparing a conventional and a spring-and-loop with clip traction method of endoscopic submucosal dissection for superficial gastric neoplasms: a randomized controlled trial (with videos). Gastrointest Endosc 2021; 93: 1097-1109 [PMID: 33058886 DOI: 10.1016/j.gie.2020.09.049
- 12 Uppal DS, Wang AY, Traction-assisted endoscopic submucosal dissection in the esophagus: Should we all be flossing? Gastrointest Endosc 2020; 91: 66-69 [PMID: 31865997 DOI: 10.1016/j.gie.2019.09.038]
- 13 Yamamoto H, Kawata H, Sunada K, Sasaki A, Nakazawa K, Miyata T, Sekine Y, Yano T, Satoh K, Ido K, Sugano K. Successful en-bloc resection of large superficial tumors in the stomach and colon using sodium hyaluronate and small-caliber-tip transparent hood. Endoscopy 2003; 35: 690-694 [PMID: 12929067 DOI: 10.1055/s-2003-41516]
- 14 Hayashi Y, Nomura T, Lee RF, Miura Y, Shinozaki S, Sunada K, Yamamoto H. Introducing the next evolution of the small-caliber-tip transparent hood: enhancing the pocket-creation method by building on previous successes. Endoscopy 2020; 52: E297-E299 [PMID: 32066191 DOI: 10.1055/a-1093-0621]
- 15 Arantes V, Albuquerque W, Freitas Dias CA, Demas Alvares Cabral MM, Yamamoto H. Standardized endoscopic submucosal tunnel dissection for management of early esophageal tumors (with video). Gastrointest Endosc 2013; 78: 946-952 [PMID: 23810327 DOI: 10.1016/j.gie.2013.05.031]
- Linghu E, Feng X, Wang X, Meng J, Du H, Wang H. Endoscopic submucosal tunnel dissection for 16 large esophageal neoplastic lesions. Endoscopy 2013; 45: 60-62 [PMID: 23254407 DOI: 10.1055/s-0032-1325965
- Choi HS, Chun HJ, Seo MH, Kim ES, Keum B, Seo YS, Jeen YT, Lee HS, Um SH, Kim CD, Ryu 17 HS. Endoscopic submucosal tunnel dissection salvage technique for ulcerative early gastric cancer. World J Gastroenterol 2014; 20: 9210-9214 [PMID: 25083097 DOI: 10.3748/wjg.v20.i27.9210]



- Hayashi Y, Sunada K, Takahashi H, Shinhata H, Lefor AT, Tanaka A, Yamamoto H. Pocket-creation 18 method of endoscopic submucosal dissection to achieve en bloc resection of giant colorectal subpedunculated neoplastic lesions. Endoscopy 2014; 46 Suppl 1: E421-E422 [PMID: 25314173] DOI: 10.1055/s-0034-1377438]
- Miura Y, Hayashi Y, Lefor AK, Osawa H, Yamamoto H. The pocket-creation method of ESD for 19 gastric neoplasms. Gastrointest Endosc 2016; 83: 457-458 [PMID: 26358325 DOI: 10.1016/j.gie.2015.08.068]
- 20 Nagata M. Usefulness of underwater endoscopic submucosal dissection in saline solution with a monopolar knife for colorectal tumors (with videos). Gastrointest Endosc 2018; 87: 1345-1353 [PMID: 29242059 DOI: 10.1016/j.gie.2017.11.032]
- 21 Nagata M. Underwater endoscopic submucosal dissection in saline solution using a bent-type knife for duodenal tumor. VideoGIE 2018; 3: 375-377 [PMID: 30505999 DOI: 10.1016/j.vgie.2018.09.015]
- 22 Li CH, Chen PJ, Chu HC, Huang TY, Shih YL, Chang WK, Hsieh TY. Endoscopic submucosal dissection with the pulley method for early-stage gastric cancer (with video). Gastrointest Endosc 2011; 73: 163-167 [PMID: 21030018 DOI: 10.1016/j.gie.2010.08.041]
- Aihara H, Kumar N, Ryou M, Abidi W, Ryan MB, Thompson CC. Facilitating endoscopic 23 submucosal dissection: the suture-pulley method significantly improves procedure time and minimizes technical difficulty compared with conventional technique: an ex vivo study (with video). Gastrointest Endosc 2014: 80: 495-502 [PMID: 24679655 DOI: 10.1016/j.gie.2014.01.050]
- Ge PS, Thompson CC, Jirapinyo P, Aihara H. Suture pulley countertraction method reduces 24 procedure time and technical demand of endoscopic submucosal dissection among novice endoscopists learning endoscopic submucosal dissection: a prospective randomized ex vivo study. Gastrointest Endosc 2019; 89: 177-184 [PMID: 30148993 DOI: 10.1016/j.gie.2018.08.032]
- 25 Yasuda M, Naito Y, Kokura S, Yoshida N, Yoshikawa T. Newly-developed ESD (CSL-ESD) for early gastric cancer using convenient and low-cost lifting method (lifting method using clips and snares) for lesions is clinically useful. Gastrointest Endosc 2012; 75: AB244 [DOI: 10.1016/j.gie.2012.04.207]
- Yoshida N, Doyama H, Ota R, Tsuji K. The clip-and-snare method with a pre-looping technique 26 during gastric endoscopic submucosal dissection. Endoscopy 2014; 46 Suppl 1 UCTN: E611-E612 [PMID: 25502265 DOI: 10.1055/s-0034-1390752]
- 27 Ota R, Doyama H, Tsuji K, Yamada S. Deep colonic endoscopic submucosal dissection using a modified clip and snare method incorporating a pre-looping technique. BMJ Case Rep 2015; 2015 [PMID: 25616654 DOI: 10.1136/bcr-2014-207918]
- Yamada S, Doyama H, Ota R, Takeda Y, Tsuji K, Tsuji S, Yoshida N. Impact of the clip and snare 28 method using the prelooping technique for colorectal endoscopic submucosal dissection. Endoscopy 2016; 48: 281-285 [PMID: 26517845 DOI: 10.1055/s-0034-1393241]
- 29 Yoshida N, Doyama H, Ota R, Takeda Y, Nakanishi H, Tominaga K, Tsuji S, Takemura K. Effectiveness of clip-and-snare method using pre-looping technique for gastric endoscopic submucosal dissection. World J Gastrointest Endosc 2016; 8: 451-457 [PMID: 27358671 DOI: 10.4253/wjge.v8.i12.451]
- Tanaka S, Toyonaga T, Kaku H, Sakaguchi H, Baba S, Takao T, Kodama Y. A novel traction device 30 (EndoTrac) for use during endoscopic submucosal dissection. Endoscopy 2019; 51: E90-E91 [PMID: 30731488 DOI: 10.1055/a-0830-4556]
- 31 Kaku H, Toyonaga T, Tanaka S, Takihara H, Baba S, Tsubouchi E, Ikeda Y, Orita H, Nakamoto M, Horikawa Y, Chiba H, Ban H, Furumoto Y, Morita R, Kodama Y. Endoscopic Submucosal Dissection Using EndoTrac, a Novel Traction Device. Digestion 2021; 102: 714-721 [PMID: 33352560 DOI: 10.1159/000511731]
- Imaeda H, Iwao Y, Ogata H, Ichikawa H, Mori M, Hosoe N, Masaoka T, Nakashita M, Suzuki H, 32 Inoue N, Aiura K, Nagata H, Kumai K, Hibi T. A new technique for endoscopic submucosal dissection for early gastric cancer using an external grasping forceps. Endoscopy 2006; 38: 1007-1010 [PMID: 16673308 DOI: 10.1055/s-2006-925264]
- 33 Imaeda H, Hosoe N, Ida Y, Kashiwagi K, Morohoshi Y, Suganuma K, Nagakubo S, Komatsu K, Suzuki H, Saito Y, Aiura K, Ogata H, Iwao Y, Kumai K, Kitagawa Y, Hibi T. Novel technique of endoscopic submucosal dissection using an external grasping forceps for superficial gastric neoplasia. Dig Endosc 2009; 21: 122-127 [PMID: 19691787 DOI: 10.1111/j.1443-1661.2009.00842.x]
- Imaeda H, Hosoe N, Ida Y, Nakamizo H, Kashiwagi K, Kanai T, Iwao Y, Hibi T, Ogata H. Novel 34 technique of endoscopic submucosal dissection by using external forceps for early rectal cancer (with videos). Gastrointest Endosc 2012; 75: 1253-1257 [PMID: 22624814 DOI: 10.1016/j.gie.2012.02.018]
- 35 Imaeda H, Hosoe N, Kashiwagi K, Ohmori T, Yahagi N, Kanai T, Ogata H. Advanced endoscopic submucosal dissection with traction. World J Gastrointest Endosc 2014; 6: 286-295 [PMID: 25031787 DOI: 10.4253/wjge.v6.i7.286]
- Wang F, Leng X, Gao Y, Zhao K, Sun Y, Bian H, Liu H, Liu P. Endoscopic submucosal dissection of 36 distal intestinal tumors using grasping forceps for traction. Tech Coloproctol 2019; 23: 1079-1083 [PMID: 31659559 DOI: 10.1007/s10151-019-02102-x]
- 37 Uraoka T, Kato J, Ishikawa S, Harada K, Kuriyama M, Takemoto K, Kawahara Y, Saito Y, Okada H. Thin endoscope-assisted endoscopic submucosal dissection for large colorectal tumors (with videos). Gastrointest Endosc 2007; 66: 836-839 [PMID: 17905031 DOI: 10.1016/j.gie.2007.04.028]
- Uraoka T, Ishikawa S, Kato J, Higashi R, Suzuki H, Kaji E, Kuriyama M, Saito S, Akita M, Hori K,



Harada K, Ishiyama S, Shiode J, Kawahara Y, Yamamoto K. Advantages of using thin endoscopeassisted endoscopic submucosal dissection technique for large colorectal tumors. Dig Endosc 2010; 22: 186-191 [PMID: 20642607 DOI: 10.1111/j.1443-1661.2010.00992.x]

- 39 Higuchi K, Tanabe S, Azuma M, Sasaki T, Katada C, Ishido K, Naruke A, Mikami T, Koizumi W. Double-endoscope endoscopic submucosal dissection for the treatment of early gastric cancer accompanied by an ulcer scar (with video). Gastrointest Endosc 2013; 78: 266-273 [PMID: 23472995 DOI: 10.1016/j.gie.2013.01.010]
- 40 Yoshio T, Tsuchida T, Ishiyama A, Omae M, Hirasawa T, Yamamoto Y, Fujisaki J, Sato Y, Sasaki T, Kawabata K, Igarashi M. Efficacy of double-scope endoscopic submucosal dissection and long-term outcomes of endoscopic resection for superficial pharyngeal cancer. Dig Endosc 2017; 29: 152-159 [PMID: 27525634 DOI: 10.1111/den.12712]
- 41 Fujii L, Onkendi EO, Bingener-Casey J, Levy MJ, Gostout CJ. Dual-scope endoscopic deep dissection of proximal gastric tumors (with video). Gastrointest Endosc 2013; 78: 365-369 [PMID: 23394839 DOI: 10.1016/j.gie.2012.12.010]
- Kobayashi T, Gotohda T, Tamakawa K, Ueda H, Kakizoe T. Magnetic anchor for more effective 42 endoscopic mucosal resection. Jpn J Clin Oncol 2004; 34: 118-123 [PMID: 15078906 DOI: 10.1093/jjco/hyh025]
- Gotoda T, Oda I, Tamakawa K, Ueda H, Kobayashi T, Kakizoe T. Prospective clinical trial of 43 magnetic-anchor-guided endoscopic submucosal dissection for large early gastric cancer (with videos). Gastrointest Endosc 2009; 69: 10-15 [PMID: 18599053 DOI: 10.1016/j.gie.2008.03.1127]
- Aihara H, Ryou M, Kumar N, Ryan MB, Thompson CC. A novel magnetic countertraction device for endoscopic submucosal dissection significantly reduces procedure time and minimizes technical difficulty. Endoscopy 2014; 46: 422-425 [PMID: 24573770 DOI: 10.1055/s-0034-1364940]
- 45 Matsuzaki I, Miyahara R, Hirooka Y, Funasaka K, Furukawa K, Ohno E, Nakamura M, Kawashima H, Maeda O, Watanabe O, Ando T, Kobayashi M, Goto H. Simplified magnetic anchor-guided endoscopic submucosal dissection in dogs (with videos). Gastrointest Endosc 2014; 80: 712-716 [PMID: 25085334 DOI: 10.1016/j.gie.2014.05.334]
- Matsuzaki I, Hattori M, Hirose K, Esaki M, Yoshikawa M, Yokoi T, Kobayashi M, Miyahara R, 46 Hirooka Y, Goto H. Magnetic anchor-guided endoscopic submucosal dissection for gastric lesions (with video). Gastrointest Endosc 2018; 87: 1576-1580 [PMID: 29352971 DOI: 10.1016/j.gie.2018.01.015]
- 47 Mori H, Kobara H, Nishiyama N, Fujihara S, Matsunaga T, Masaki T. Novel effective and repeatedly available ring-thread counter traction for safer colorectal endoscopic submucosal dissection. Surg Endosc 2017; 31: 3040-3047 [PMID: 27858210 DOI: 10.1007/s00464-016-5326-7]
- Sudo G, Tanuma T, Suzuki Y, Nakase H. Multiloop method for traction during colorectal endoscopic 48 submucosal dissection. VideoGIE 2019; 4: 11-13 [PMID: 30623150 DOI: 10.1016/j.vgie.2018.10.002]
- 49 Jacques J, Charissoux A, Bordillon P, Legros R, Rivory J, Hervieu V, Albouys J, Guyot A, Ponchon T, Sautereau D, Kerever S, Pioche M. High proficiency of colonic endoscopic submucosal dissection in Europe thanks to countertraction strategy using a double clip and rubber band. Endosc Int Open 2019; 7: E1166-E1174 [PMID: 31475236 DOI: 10.1055/a-0965-8531]
- Ge PS, Aihara H. A novel clip-band traction device to facilitate colorectal endoscopic submucosal 50 dissection and defect closure. VideoGIE 2020; 5: 180-186 [PMID: 32426563 DOI: 10.1016/j.vgie.2020.01.012
- 51 Kim SH, Lee JK, Lim YJ, Kim JH. The risk factors for prolonged hemostatic clip retention after endoscopic submucosal dissection for gastric neoplasm. Surg Endosc 2021 [PMID: 33629182 DOI: 10.1007/s00464-021-08379-0]
- 52 Ide D, Saito S, Chino A, Ohya TR. Submucosal pocket creation using a traction device in colorectal endoscopic submucosal dissection. Ann Gastroenterol 2018; 31: 380 [PMID: 29720866 DOI: 10.20524/aog.2018.0258
- Ide D, Ohya TR, Saito S, Mitsuyoshi Y, Hatamori H, Ikenoyama Y, Suzuki K, Ishioka M, Yakabi S, 53 Yasue C, Chino A, Igarashi M, Saruta M, Fujisaki J. Clinical utility of the pocket-creation method with a traction device for colorectal endoscopic submucosal dissection. Surg Endosc 2021; 35: 2110-2118 [PMID: 32382886 DOI: 10.1007/s00464-020-07614-4]
- Yamamoto K, Hayashi S, Nakabori T, Shibuya M, Ichiba M, Inada M. Endoscopic submucosal 54 dissection using endoclips to assist in mucosal flap formation (novel technique: "clip flap method"). Endoscopy 2012; 44 Suppl 2: E334-E335 [PMID: 23012008 DOI: 10.1055/s-0032-1309860]
- Yamamoto K, Hayashi S, Nishida T, Saiki H, Naito M, Michida T, Ito T. Effective use of the "clip-55 flap" method for the endoscopic submucosal dissection of a difficult-to-approach superficial gastric tumor. Endoscopy 2015; 47 Suppl 1: E318-E319 [PMID: 26115394 DOI: 10.1055/s-0034-1392319]
- Yamamoto K, Hayashi S, Saiki H, Indo N, Nakabori T, Yamamoto M, Shibuya M, Nishida T, Ichiba M, Inada M. Endoscopic submucosal dissection for large superficial colorectal tumors using the "clipflap method". Endoscopy 2015; 47: 262-265 [PMID: 25412089 DOI: 10.1055/s-0034-1390739]
- Ban H, Sugimoto M, Otsuka T, Murata M, Nakata T, Hasegawa H, Inatomi O, Bamba S, Andoh A. 57 Usefulness of the clip-flap method of endoscopic submucosal dissection: A randomized controlled trial. World J Gastroenterol 2018; 24: 4077-4085 [PMID: 30254412 DOI: 10.3748/wjg.v24.i35.4077]
- 58 Huang R, Cai H, Zhao X, Lu X, Liu M, Lv W, Liu Z, Wu K, Han Y. Efficacy and safety of endoscopic submucosal tunnel dissection for superficial esophageal squamous cell carcinoma: a propensity score matching analysis. Gastrointest Endosc 2017; 86: 831-838 [PMID: 28286094 DOI:



10.1016/j.gie.2017.03.001]

- 59 Zhang T, Zhang H, Zhong F, Wang X. Efficacy of endoscopic submucosal tunnel dissection versus endoscopic submucosal dissection for superficial esophageal neoplastic lesions: a systematic review and meta-analysis. Surg Endosc 2021; 35: 52-62 [PMID: 32856152 DOI: 10.1007/s00464-020-07925-6]
- 60 Imagawa A, Okada H, Kawahara Y, Takenaka R, Kato J, Kawamoto H, Fujiki S, Takata R, Yoshino T, Shiratori Y. Endoscopic submucosal dissection for early gastric cancer: results and degrees of technical difficulty as well as success. Endoscopy 2006; 38: 987-990 [PMID: 17058162 DOI: 10.1055/s-2006-944716
- Kim SJ, Choi CW, Nam HS, Kang DH, Kim HW, Park SB, Ryu DG. Factors associated with 61 conversion to snare resection during gastric endoscopic submucosal dissection. Surg Endosc 2020; 34: 1585-1591 [PMID: 31209610 DOI: 10.1007/s00464-019-06918-4]
- 62 Ojima T, Takifuji K, Nakamura M, Nakamori M, Hayata K, Kitadani J, Yamaue H. Endoscopic submucosal tunnel dissection versus conventional endoscopic submucosal dissection for early gastric cancers: outcomes of 799 consecutive cases in a single institution. Surg Endosc 2020; 34: 5625-5631 [PMID: 32748265 DOI: 10.1007/s00464-020-07849-1]
- Feng X, Linghu E, Chai N, Lu Z, Wang X, Tang P, Meng J, Du H, Wang H. Endoscopic Submucosal 63 Tunnel Dissection for Large Gastric Neoplastic Lesions: A Case-Matched Controlled Study. Gastroenterol Res Pract 2018; 2018: 1419369 [PMID: 29692806 DOI: 10.1155/2018/1419369]
- Ritsuno H, Sakamoto N, Osada T, Goto SP, Murakami T, Ueyama H, Mori H, Matsumoto K, Beppu 64 K, Shibuya T, Nagahara A, Ogihara T, Watanabe S. Prospective clinical trial of traction deviceassisted endoscopic submucosal dissection of large superficial colorectal tumors using the S-O clip. Surg Endosc 2014; 28: 3143-3149 [PMID: 24879138 DOI: 10.1007/s00464-014-3572-0]
- 65 Suzuki Y, Tanuma T, Nojima M, Sudo G, Murakami Y, Ishii T, Akahonai M, Kobayashi Y, Hamamoto H, Aoki H, Harada T, Katanuma A, Nakase H. Comparison of dissection speed during colorectal ESD between the novel Multiloop (M-loop) traction method and ESD methods without traction. Endosc Int Open 2020; 8: E840-E847 [PMID: 32617388 DOI: 10.1055/a-1161-8596]
- Yamashina T, Nemoto D, Hayashi Y, Fukuda H, Okada M, Takezawa T, Aizawa M, Sakamoto H, 66 Miura Y, Sunada K, Lefor AK, Togashi K, Yamamoto H. Prospective randomized trial comparing the pocket-creation method and conventional method of colorectal endoscopic submucosal dissection. Gastrointest Endosc 2020; 92: 368-379 [PMID: 32119937 DOI: 10.1016/j.gie.2020.02.034]
- Sakamoto H, Hayashi Y, Miura Y, Shinozaki S, Takahashi H, Fukuda H, Okada M, Ino Y, Takezawa 67 T. Sunada K. Lefor AK, Yamamoto H. Pocket-creation method facilitates endoscopic submucosal dissection of colorectal laterally spreading tumors, non-granular type. Endosc Int Open 2017; 5: E123-E129 [PMID: 28337483 DOI: 10.1055/s-0042-122778]
- Takezawa T, Hayashi Y, Shinozaki S, Sagara Y, Okada M, Kobayashi Y, Sakamoto H, Miura Y, 68 Sunada K, Lefor AK, Yamamoto H. The pocket-creation method facilitates colonic endoscopic submucosal dissection (with video). Gastrointest Endosc 2019; 89: 1045-1053 [PMID: 30716306 DOI: 10.1016/j.gie.2019.01.022]
- Pei Q, Qiao H, Zhang M, Wang G, Feng H, Pan J, Shi Y. Pocket-creation method versus conventional 69 method of endoscopic submucosal dissection for superficial colorectal neoplasms: a meta-analysis. Gastrointest Endosc 2021; 93: 1038-1046.e4 [PMID: 33484729 DOI: 10.1016/j.gie.2021.01.007]
- 70 Yamasaki Y, Takeuchi Y, Hanaoka N, Higashino K, Uedo N, Ishihara R, Iishi H. A novel traction method using an endoclip attached to a nylon string during colonic endoscopic submucosal dissection. Endoscopy 2015; 47 Suppl 1: E238-E239 [PMID: 26069981 DOI: 10.1055/s-0034-1391868]
- Yamasaki Y, Takeuchi Y, Uedo N, Kanesaka T, Kato M, Hamada K, Tonai Y, Matsuura N, Akasaka 71 T, Hanaoka N, Higashino K, Ishihara R, Okada H, Iishi H. Efficacy of traction-assisted colorectal endoscopic submucosal dissection using a clip-and-thread technique: A prospective randomized study. Dig Endosc 2018; 30: 467-476 [PMID: 29424030 DOI: 10.1111/den.13036]
- Despott EJ, Hirayama Y, Lazaridis N, Koukias N, Telese A, Hayashi Y, Miura Y, Yamamoto H, 72 Murino A. Saline immersion therapeutic endoscopy facilitated pocket-creation method for endoscopic submucosal dissection (with video). Gastrointest Endosc 2019; 89: 652-653 [PMID: 30784510 DOI: 10.1016/j.gie.2018.10.005]
- Mavrogenis G, Mavrogenis I, Anastasiadis S, Bazerbachi F. Underwater endoscopic submucosal 73 dissection in saline solution with rubber-band countertraction for a cecal polyp extending into a diverticulum. Ann Gastroenterol 2019; 32: 527 [PMID: 31474803 DOI: 10.20524/aog.2019.0396]
- 74 Ramos-Zabala F, García-Mayor M, Domínguez-Pino A, Alzina-Pérez A, Moreno-Almazán L. Combination of immersion in saline solution, pocket-creation method, water-jet hydrodissection, and hybrid knife "probe mode" simplifies endoscopic submucosal dissection in giant rectal polyp. VideoGIE 2019; 4: 478-480 [PMID: 31709336 DOI: 10.1016/j.vgie.2019.05.009]
- 75 Caruso A, Sgamato C, Bertani H, Pigò F, Mangiafico S, Grande G, Conigliaro RL. Underwater endoscopic submucosal dissection of an obstructing giant colonic lipoma. Endoscopy 2020; 52: E90-E91 [PMID: 31561257 DOI: 10.1055/a-1011-3617]
- Hoteya S, Furuhata T, Takahito T, Fukuma Y, Suzuki Y, Kikuchi D, Mitani T, Matsui A, Yamashita 76 S, Nomura K, Kuribayashi Y, Iizuka T, Kaise M. Endoscopic Submucosal Dissection and Endoscopic Mucosal Resection for Non-Ampullary Superficial Duodenal Tumor. Digestion 2017; 95: 36-42 [PMID: 28052275 DOI: 10.1159/000452363]
- Matsuda Y, Sakamoto K, Kataoka N, Yamaguchi T, Tomita M, Makimoto S. Perforation associated 77 with endoscopic submucosal dissection for duodenal neoplasm without a papillary portion. World J



Gastrointest Surg 2017; 9: 161-166 [PMID: 28824748 DOI: 10.4240/wjgs.v9.i7.161]

- Yahagi N, Kato M, Ochiai Y, Maehata T, Sasaki M, Kiguchi Y, Akimoto T, Nakayama A, Fujimoto 78 A, Goto O, Uraoka T. Outcomes of endoscopic resection for superficial duodenal epithelial neoplasia. Gastrointest Endosc 2018; 88: 676-682 [PMID: 29753040 DOI: 10.1016/j.gie.2018.05.002]
- Miura Y, Shinozaki S, Hayashi Y, Sakamoto H, Lefor AK, Yamamoto H. Duodenal endoscopic 79 submucosal dissection is feasible using the pocket-creation method. Endoscopy 2017; 49: 8-14 [PMID: 27875854 DOI: 10.1055/s-0042-116315]
- Yahagi N, Nishizawa T, Sasaki M, Ochiai Y, Uraoka T. Water pressure method for duodenal 80 endoscopic submucosal dissection. Endoscopy 2017; 49: E227-E228 [PMID: 28759932 DOI: 10.1055/s-0043-113556]
- 81 Inoue T, Uedo N, Yamashina T, Yamamoto S, Hanaoka N, Takeuchi Y, Higashino K, Ishihara R, Iishi H, Tatsuta M, Takahashi H, Eguchi H, Ohigashi H. Delayed perforation: a hazardous complication of endoscopic resection for non-ampullary duodenal neoplasm. Dig Endosc 2014; 26: 220-227 [PMID: 23621427 DOI: 10.1111/den.12104]
- Kono M, Nagami Y, Kitagawa D, Manabe T, Ominami M, Fukunaga S, Fujiwara Y. Underwater 82 endoscopic submucosal dissection for a duodenal neuroendocrine tumor using pocket creation and ring-shaped thread countertraction methods. Endoscopy 2021; 53: E110-E111 [PMID: 32659806 DOI: 10.1055/a-1198-4153]
- Yano T, Nemoto D, Ono K, Miyata Y, Numao N, Iwashita C, Nagayama M, Takahashi H, Lefor AK, 83 Yamamoto H. Gel immersion endoscopy: a novel method to secure the visual field during endoscopy in bleeding patients (with videos). Gastrointest Endosc 2016; 83: 809-811 [PMID: 26463338 DOI: 10.1016/j.gie.2015.09.048]
- Hashimoto R, Hirasawa D. Duodenal endoscopic submucosal dissection with traction method using 84 the S-O clip. Dig Endosc 2017; 29: 635 [PMID: 28295649 DOI: 10.1111/den.12860]



WŨ

World Journal of Gastroenterology

Submit a Manuscript: https://www.f6publishing.com

World J Gastroenterol 2022 January 7; 28(1): 23-46

DOI: 10.3748/wjg.v28.i1.23

ISSN 1007-9327 (print) ISSN 2219-2840 (online)

FRONTIER

Cytoprotective gastric pentadecapeptide BPC 157 resolves major vessel occlusion disturbances, ischemia-reperfusion injury following Pringle maneuver, and Budd-Chiari syndrome

Predrag Sikiric, Anita Skrtic, Slaven Gojkovic, Ivan Krezic, Helena Zizek, Eva Lovric, Suncana Sikiric, Mario Knezevic, Sanja Strbe, Marija Milavic, Antonio Kokot, Alenka Boban Blagaic, Sven Seiwerth

ORCID number: Predrag Sikiric 0000-0002-7952-2252; Anita Skrtic 0000-0002-9275-7283; Slaven Gojkovic 0000-0003-4020-326X; Ivan Krezic 0000-0001-7994-5645; Helena Zizek 0000-0001-9863-4164; Eva Lovric 0000-0003-3346-7005; Suncana Sikiric 0000-0001-9337-9248; Mario Knezevic 0000-0001-6448-7274; Sanja Strbe 0000-0001-7199-4530; Marija Milavic 0000-0003-3404-0644; Antonio Kokot 0000-0002-4836-7536; Alenka Boban Blagaic 0000-0003-1266-3402; Sven Seiwerth 0000-0002-5894-419X.

Author contributions: Knezevic M, Kokot A, and Skrtic A designed the research study; Zizek H, Gojkovic S, and Krezic I performed the research; Milavic M, Sikiric S, and Lovric E analyzed the data; Strbe S contributed new reagents and analytic tools; Sikiric P, Seiwerth S, and Boban AB wrote the manuscript; all authors have read and approved the final manuscript.

Conflict-of-interest statement: The authors state that they have no conflicts of interest to disclose.

Supported by University of Zagreb, Zagreb, Croatia, No. BM 099.

Country/Territory of origin: Croatia

Specialty type: Medicine, research

Predrag Sikiric, Slaven Gojkovic, Ivan Krezic, Helena Zizek, Mario Knezevic, Sanja Strbe, Alenka Boban Blagaic, Department of Pharmacology, School of Medicine, University of Zagreb, Zagreb 10000, Croatia

Anita Skrtic, Eva Lovric, Suncana Sikiric, Marija Milavic, Sven Seiwerth, Department of Pathology, School of Medicine, University of Zagreb, Zagreb 10000, Croatia

Antonio Kokot, Department of Anatomy and Neuroscience, Faculty of Medicine Osijek, J.J.Strossmayer University of Osijek, Osijek 31000, Croatia

Corresponding author: Predrag Sikiric, MD, PhD, Full Professor, Department of Pharmacology, School of Medicine, University of Zagreb, Salata 11, Zagreb 10000, Croatia. sikiric@mef.hr

Abstract

The stable gastric pentadecapeptide BPC 157 counteracts various venous occlusion-induced syndromes. Summarized are all these arguments, in the Robert's cytoprotection concept, to substantiate the resolution of different major vessel occlusion disturbances, in particular ischemia-reperfusion injury following the Pringle maneuver and Budd-Chiari syndrome, which was obtained by BPC 157 therapy. Conceptually, there is a new point, namely, endothelium maintenance to epithelium maintenance (the recruitment of collateral blood vessels to compensate for vessel occlusion and reestablish blood flow or bypass the occluded or ruptured vessel). In this paper, we summarize the evidence of the native cytoprotective gastric pentadecapeptide BPC 157, which is stable in the human gastric juice, is a membrane stabilizer and counteracts gut-leaky syndrome. As a particular target, it is distinctive from the standard peptide growth factors, involving particular molecular pathways and controlling VEGF and NO pathways. In the early 1990s, BPC 157 appeared as a late outbreak of the Robert's and Szabo's cytoprotection-organoprotection concept, like the previous theoretical/practical breakthrough in the 1980s and the brain-gut axis and gutbrain axis. As the time went on, with its reported effects, it is likely most useful theory practical implementation and justification. Meantime, several reviews suggest that BPC 157, which does not have a lethal dose, has profound cytoprotective activity, used to be demonstrated in ulcerative colitis and multiple sclerosis trials. Likely, it may bring the theory to practical application, starting with the



WJG | https://www.wjgnet.com

and experimental

Provenance and peer review:

Invited article; Externally peer reviewed.

Peer-review report's scientific quality classification

Grade A (Excellent): 0 Grade B (Very good): B Grade C (Good): 0 Grade D (Fair): 0 Grade E (Poor): 0

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: htt ps://creativecommons.org/Licens es/by-nc/4.0/

Received: March 21, 2021 Peer-review started: March 21, 2021 First decision: April 29, 2021 Revised: May 14, 2021 Accepted: December 21, 2021 Article in press: December 21, 2021 Published online: January 7, 2022

P-Reviewer: Vukojević J S-Editor: Ma YI L-Editor: Wang TQ P-Editor: Ma YJ



initial argument, no degradation in human gastric juice for more than 24 h, and thereby, the therapeutic effectiveness (including via a therapeutic per-oral regimen) and pleiotropic beneficial effects.

Key Words: Gastric pentadecapeptide BPC 157; Cytoprotection; Major vessel occlusion disturbances; Pringle maneuver; Budd-Chiari syndrome; Therapy

©The Author(s) 2022. Published by Baishideng Publishing Group Inc. All rights reserved.

Core Tip: Summarizing the essential epithelium and endothelium protection interplay described in Robert's and Szabo's cytoprotection concept, and the role of the stable pentadecapeptide BPC 157 as a likely mediator, we suggest that BPC 157 may be a useful cytoprotective therapy. The hope is that it could finally bring into practice the huge theoretical importance of all aspects of the cytoprotection concept. Conceptually, there is a new point to discuss, namely, endothelium maintenance to epithelium maintenance (recruitment of collateral blood vessels to compensate for vessel occlusion and reestablish blood flow or bypass the occluded or ruptured vessel). BPC 157 counteracts various venous occlusion-induced syndromes, as well as inferior caval vein syndrome, ischemia-reperfusion injury following Pringle maneuver, and Budd-Chiari syndrome in rats. Activation of the alternative collateral pathways to bypass occlusion and reestablish alternative blood flow, results in the counteraction of the consequent syndromes. The severe venous occlusion-induced disturbances, the high portal and caval hypertension, aortal hypotension, arterial and venous thrombosis, both peripherally and centrally, and various organ lesions (i.e., gastrointestinal, liver, kidney, heart, and brain) were all attenuated and/or eliminated. Furthermore, this particular beneficial effect may be competing with the Virchow's triad that can be a common presentation [i.e., duodenal venous congestion lesions, perforated cecum, ischemic/reperfusion colitis, bile duct ligation-induced liver cirrhosis and portal hypertension, temporary portal triad occlusion (ischemia-reperfusion injury following the Pringle maneuver), and suprahepatic occlusion of the inferior caval vein (Budd-Chiari syndrome)]. The resolution of these various venous occlusion-induced syndromes emphasizes the evidence that as the native cytoprotective gastric peptide and a stable gastric pentadecapeptide membrane stabilizer, BPC 157, which is stable in the human gastric juice and counteracts gut-leaky syndrome, is a particular target and easily distinguished from standard peptide growth factors, involving particular molecular pathways, and specifically controlling the VEGF and NO pathways, in particular the prostaglandin pathway.

Citation: Sikiric P, Skrtic A, Gojkovic S, Krezic I, Zizek H, Lovric E, Sikiric S, Knezevic M, Strbe S, Milavic M, Kokot A, Blagaic AB, Seiwerth S. Cytoprotective gastric pentadecapeptide BPC 157 resolves major vessel occlusion disturbances, ischemia-reperfusion injury following Pringle maneuver, and Budd-Chiari syndrome. World J Gastroenterol 2022; 28(1): 23-46 URL: https://www.wjgnet.com/1007-9327/full/v28/i1/23.htm DOI: https://dx.doi.org/10.3748/wjg.v28.i1.23

INTRODUCTION

The current review aims to evaluate whether the stable gastric pentadecapeptide BPC 157, which has consistent efficacy in the co-, pre-, and post-treatment regimens, with a rapid onset of the therapeutic effect, as well as the parenteral and per-oral effectiveness, may bring the Robert's cytoprotection theory into practical application[1].

As previously stated^[1], all of the studies to date that have tested the stable gastric pentadecapeptide BPC 157 as a treatment have demonstrated extremely positive healing effects for various injury types in numerous organ systems. Its practical significance as a prototypic cytoprotective agent and an important mediator of Robert's cytoprotection[1], and its contribution to resolving Selye's stress response[1], and brain-gut and gut-brain axis activity have been reported^[2]. Additional particular points are its wide interactions with the nitric oxide (NO) system[1] and prostaglandin

system and counteraction of the toxicity of non-steroidal-anti-inflammatory drugs[1]. Its therapeutic effects on fistula healing[1] and damaged skin, muscles, tendons, ligaments, and bone comparable to those in the gastrointestinal tract[3,4] and wound healing, in particular[3], are also reviewed. The counteraction of tumor-induced muscle cachexia and the signaling process implicated in cancer cachexia^[5] and leaky gut, and its membrane stabilizer and free radical scavenger activity [6] are highlighted. The final focus is on the particular effect of BPC 157 on blood vessels and vessel recruitment[1,7]. In addition, BPC 157, due to its profound cytoprotective activity, which has been demonstrated in ulcerative colitis and applied to multiple sclerosis trials, may be used, since it does not have a lethal dose (LD1)[1,6]. In one of the most recent studies[3], BPC 157 was found to be distributed in the gastrointestinal mucosa, lung, bronchial epithelium, epidermal layer of the skin, and kidney glomeruli by in situ hybridization and immunostaining. These data suggest that BPC 157 may have additional regulatory roles in the function of the lungs, kidneys, and skin in humans, in addition to being isolated from gastric juice and primarily acting in the gastrointestinal system[3]. BPC 157 has also been reviewed in several other articles[8].

In the present review, we discuss the cytoprotective activity of the gastric pentadecapeptide BPC 157[1] to resolve major vessel occlusion disturbances, ischemiareperfusion injury following the Pringle maneuver, and Budd-Chiari syndrome[9-11], providing evidence that it may bring the cytoprotection theory to practical application. On the other hand, as mentioned above, the stable gastric pentadecapeptide BPC 157 perfectly matched with the original Robert's cytoprotective requirements for the stomach, or even extended it[1]. These requirements are the protection of the epithelium ("epithelial pathway") and endothelium ("endothelial pathway"), and the maintenance of gastrointestinal mucosal integrity to obtain a large beneficial effect inside and outside the gastrointestinal tract. Typically, human gastric juice rapidly destroyed standard growth factors within 15 min[12,13]. In contrast, BPC 157, with its essential gastric juice origin and stability in human gastric juice for more than 24 h [12], was matched in the local level (stomach, the permanent maintenance of the mucosal integrity, and thereby the entire gastrointestinal tract)[1]. Therefore, BPC 157 has particular therapeutic effectiveness, including via a therapeutic per-oral regimen, and pleiotropic beneficial effects. This local stomach and gastrointestinal tract protection was further extended to the general level (protection of other organs) (cytoprotection to organoprotection)[1]. As previously mentioned, BPC 157 could follow both the "epithelial" and "endothelial" pathways in Robert's cytoprotection[1,7, 12,14].

According to Andre Robert[14] in 1975, the first indication for cytoprotection was the evidence that certain prostaglandins (PGF2 and PGFB, which could not affect gastric acid secretion) protected the gastric mucosa against indomethacin, in a gastric acid-independent manner, via a mechanism other than the inhibition of gastric acid secretion[14-16]. Therefore, the term cytoprotection pioneered by Robert[14-16] was introduced in 1979 against the noxious effect of both intragastric alcohol application and the use of nonsteroidal anti-inflammatory drugs (NSAIDs). According to Robert's concept, the rapid onset of gastric cytoprotection would be the most remarkable aspect [14-16]. Prostaglandins reduce the development of gastric necrotic lesions when given orally at an appropriate dose as late as 1 min before the administration of absolute ethanol. Knowing this "before but not after effectiveness" as a limitation, Robert appreciated the curing of the already existing lesions as the further possibility[14-16]. The full explanation was provided in a few subsequent reviews[14-16] and the full argument was later substantiated[14-16]. In our view of the new cytoprotection principle, the essential evidence of Robert (epithelial protection) is the remarkable ability of endogenous and exogenous cytoprotective agents (i.e., prostaglandins) to prevent rapidly acute gastric hemorrhagic lesions induced by diverse noxious stimuli such as ethanol, bile acids, hyperosmolar solutions, and NSAIDs such as aspirin or indomethacin. According to the claims of Robert [i.e., cytoprotection preventing mucosal necrosis caused by noxious agents due to the direct damage of cells or a local deficiency of cytoprotective mediators (i.e., prostaglandins)], the cytoprotection concept also goes beyond peptic ulcer therapy [14-16]. Moreover, in Robert's view [14], the demonstration of adaptive cytoprotection suggests that cytoprotection by prostaglandins may be a physiological phenomenon^[14]. One milliliter of 20% ethanol (as small irritant) given orally (note, in Robert's publication, "orally" implies administration via a tube into the stomach, or rather intragastric application, see Chapter 2.1.3. Epithelial pathway to adaptive cytoprotection) to fasted rats 15 min prior to giving absolute ethanol (regarded as a strong irritant) prevented the gastric mucosal necrosis caused by the latter[17]. For the effectiveness of cytoprotective agents, the concept holds the protection of stomach cells (cytoprotection) and other epithelia (organoprotection)



against direct injury to cells induced by various noxious agents [14-16]. In addition, see the notation for the cytoprotection-organoprotection for other agents[18,19].

In the early 1980s, the concept obtained an additional key, the concomitant protection of the stomach endothelium[20,21] or Szabo's vascular injury, as an early pathogenic factor in the development of ethanol-induced gastric hemorrhagic erosions. Demonstration of the vascular injury was seen within 1 min, as was the estimated effect of the agents [20,21]. This vascular point in ethanol-induced gastric lesions was fully elaborated in a series of subsequent reports^[22]. Since then, the rapid recovery of damaged endothelium may be considered a shared effect of the cytoprotective agents [23].

Thus, the cytoprotection theory holds that cytoprotective agents should exert direct epithelial and endothelial cell protection inside and outside of the gastrointestinal tract [14-16], via the "epithelial pathway" and "endothelial pathway". There is an essential evolvement in the stomach protection: Endothelium protection to epithelium protection[14-16]. Each of these pathways originates as a result of the increase in cytoprotective activity, together manifested as an increased therapeutic effect in both the prophylactic process (important for maintaining undisturbed organ function) and the therapeutic process (important for the possible reversal of the damaged tissue to a normal structure, and the interruption of damaging events). Unfortunately, such practical realization of the highly conceptualized theory is lacking. The anticipated huge range of organ lesions that should be counteracted and protection against nonspecific lesions^[1], as well as the rapid onset of action implemented in the agents' efficacy, as a resolving outcome, remains obscure.

On the other hand, in addition to the proposed role of BPC 157[1,7], within Robert's concept of cytoprotection, different points of view and different highlights can be clearly seen. Certainly, such a multitude illustrates the essential value of the potential application of the concept. The emphasis was on NO, carbon monoxide and hydrogen sulfide[24-26], sulfhydryls (SH)[20,27,28] in parallel with prostaglandins, as well as histamine^[23], prostaglandins^[17], EP1 and IP receptors^[29,30], the healing action of antacid[31], sucralfate[31,32], heat shock protein 70 (HSP70)[33], and the reninangiotensin system and active angiotensin metabolites[34]. Further illustrative emphases include opioids[35], alpha-2 adrenoreceptors[36], glucocorticoids[20,37-39], thyrotropin-releasing hormone (TRH)[40-43], capsaicin[44-46], dopamine[19,47-51], somatostatin[18,52,53], epidermal growth factor (EGF)[53-55], bombesin[56], ghrelin [17,34,57], cholecystokinin (CCK) and leptin[58,59], melatonin[60], neurotensin[61], fibroblast growth factor (FGF)[62,63], agmatine[64], amino acids[65], secondgeneration histamine H(2)-receptor antagonists[66], hemeoxygenase-1[67,68], and the molecular basis of alcohol-related gastric and colon cancer (acetaldehyde)[69].

Finally, a historical cytoprotection review, along with many original details, is given by Mozsik[70].

The term cytoprotection was commonly coined in other organ studies, i.e., the heart and brain[71,72], kidney[73], liver[74], eye[75], skin and wounds[76], bone[77], and skeletal muscle^[78].

Unfortunately, the multitude of agents supposed to be involved did not resolve the conceptual problems that were initially shown with the prime agents, providing the limited therapeutic potential of prostaglandins in stomach lesions (i.e., prostaglandins might only prevent rather than cure any already established stomach lesions)[14-16]. Likewise, there was an even more limited therapeutic potential in the healing of other organs (prostaglandins were only effective in a few organ lesions)[79-81]. The switching to other cytoprotective agents (i.e., sulfhydryls[19,21], somatostatin[18], EGF [53], TRH[41,67,82], opioids[83], dopamine[50,51,61], and CCK) led to similar incomplete results in both stomach and other organ lesions (for review see[1,7]). Consequently, considering the application and efficacy of standard agents, a considerable gap remains between the theoretical potential and practical realization[1,7]. Considering the supplemental endothelium protection, after initial demonstration in the stomach, no endothelial protection outside the stomach was investigated at the time[22]. Of note, BPC 157 appears to resolve both of these issues, *i.e.*, the "epithelium pathway" and "endothelium pathway" in cytoprotection[1,7], and may both prevent lesion development and cure any established lesions.

Likewise, to illustrate the failed realization of the concept with standard antiulcer agents, in addition to only prophylactic effectiveness in stomach lesions and a few other organs in which effectiveness was shown, the theoretical/practical problem is that standard cytoprotective agents also demonstrated the opposite outcome[42,84]. The intriguing point is the sulfhydryl prototype, cysteamine, and sulfhydryl conceptual involvement[19,21]. Cysteamine is highly protective in alcohol-induced stomach lesions[19,21], but, in contrast, cysteamine application provided the most



valuable standard model for the induction of duodenal[85,86] and colon[87] lesions. Also, we emphasized [1] that Robert's concept [14-17] largely applied the antecedent Selye's stress concept[88,89] which essentially contributed[90] to the introduction of corticosteroid therapy[91]. Of note, both concepts act against various nonspecific agents that would induce non-specific lesions[1]. Both concepts also hold organoprotection (Selve's concept of homeostasis that should be reestablished by the stress response^[88] vs Robert's direct stomach cell protection that should be generalized by the application of cytoprotective agents [14-16]), and adaptation [Selye's small stress that protects against severe stress^[89] vs Robert's small irritants that protect against strong irritants (adaptive cytoprotection[17])]. However, the essential first mediator of Selye's stress concept[88,89], which would integrate the adaptive bodily stress response and reestablish organoprotective bodily homeostasis, remained undiscovered, and appeared to be a major weakness of the concept that would preclude its practical realization[92,93].

For the classic concepts of Robert and Selye[14-16,88], the adverse effects of the prototype agents (*i.e.*, mediators) appeared to be an additional pitfall. Obviously, protection against direct injury to the cell in Robert's cytoprotection concept[14-16] certainly precludes any adverse effects, which are quite common with the application of prostaglandin analogues[65]. Likewise, the reestablishing of homeostasis (Selye's stress response defined "as such")[88] does not include the adverse effects that have been commonly known for the application of corticosteroids since early times [94]. As BPC 157 appears to be very safe and LD1 was not achieved, with no side effects reported in clinical trials, the possible switching of beneficial effects to negative ones (over-shutting phenomenon) appears to be highly unlikely^[1].

However, whatever the pitfalls may be, these two concepts[14-16,88] provided a firm theoretical frame for the development of novel agents and therapies. It should be practically realized and demonstrated, in addition to the local (stomach) beneficial effect, by the agents' pleiotropic beneficial effects[1,7]. If properly followed, it may fulfill the first conceptual beneficial point (starting with Robert's cytoprotection[14-16], local protection and therapy of the stomach and gastrointestinal tract achieved) by the next extended beneficial point (the protection of other organs [epithelia] and the achievement of therapy), and bring them together to a reality that can no longer be disputed.

Thus, in the early 1990s, pentadecapeptide BPC 157[1,7] appeared as a late outbreak of the cytoprotection-organoprotection concept of Robert and Szabo[14-16,18,19], for epithelial and endothelial protection, like the previous theoretical/practical breakthrough in the 1980s[14-16,18,19] and the brain-gut axis and gut-brain axis[3]. As time went on, with its reported effects, BPC 157 could be most useful in the practical implementation and justification of the theory[1]. All arguments were given to bring the long-standing theory into practice, starting with the initial argument of the lack of degradation in human gastric juice for more than 24 h[12], and thereby the therapeutic effectiveness (including via a therapeutic per-oral regimen) and pleiotropic beneficial effect[1,7].

BPC 157 IN CYTOPROTECTION

Overall, and in particular for the role and cytoprotective effectiveness of BPC 157, it is safe to speculate that the efficacy and activity limitation of this agent, and thereby its practical application, would be determined by the foundation of the standing concepts, and vice versa. Briefly, the agent "runs" within the concept frame, and vice versa. Ideally, agent and concept can match completely (as may be seen with the achieved extent of the obtained beneficial effects; cytoprotection can be manifested as a huge range of beneficial effects, both inside and outside the gastrointestinal tract). General pitfalls may be the number of mentioned cytoprotective agents that have previously failed to match the required cytoprotection concept. In general, this means more problems with the use of new agents, and more problems for the concept to maintain its validity and less possibility (enthusiasm and belief) to be once applied. Alternatively, if there were no known agents which fulfilled the requirements of the standing concepts, the agent's efficacy and activity would determine the opposition to the "law" of the standing concepts and form a new relevant concept.

Illustratively, regarding the sympathetic system function, Alhquist's receptor concept[95] (i.e., six catecholamines, and their different order of potency depending on the tissue involved, to anticipate the presentation of the particular alpha and beta receptors) discharged the long-standing "law" of physiology, Cannon's concept of two



WJG | https://www.wjgnet.com

mediator substances (sympathin E and sympathin I)[96]. Although there was an overlap of several years, Alhquist's receptor concept accuracy[95] envisaged the development of specific blocking agents in the subsequent years[97], and the consequent regular use of beta blockers in a large range of indications[98]. However, similar general acceptance and applicability did not arrive for the cytoprotection concept, nor was there any proof[99,100], when years later, as alternative gastric acid-nondependent, Robert's cytoprotection theory challenged the peptic ulcer therapy[14-16]. The lack of a practical solution and the absence of any commonly applicable cytoprotective therapy[99,100] mean that the "law" "no acid-no ulcer" and the superiority of H2-blockers were not discharged until the present time[101].

In the early 1990s, BPC 157 was introduced as a pentadecapeptide with cytoprotective effects[1,18,19], many years after the breakthrough of the original concepts of Robert[14-16] and Selye[88]. The surveillance of these two major concepts[14-16,88] and their development and achievements lacking full realization and adequate practical application[92,93,99,100] considered the introduction of BPC 157 to be too late a challenge. Seeing from the achieved perspective of all agents tested as standard cytoprotective agents, it was safe to speculate that a novel agent would hardly achieve a wider range of pleiotropic beneficial effects and drug characteristics that remained elusive for years.

However, conceptually, there is a new point, namely, endothelium maintenance to epithelium maintenance is upgraded to endothelium maintenance to epithelium maintenance (collateral blood vessels to compensate for vessel occlusion and reestablish blood flow or bypass the occluded or ruptured vessel^{y[1,7]}. The recruitment of collateral blood vessels would compensate for vessel occlusion and reestablish blood flow[1,7,9-11]. BPC 157 counteracted various venous occlusion-induced syndromes[9-11], inferior caval vein syndrome[9], ischemia-reperfusion injury following Pringle maneuver[10], and Budd-Chiari syndrome[11] in rats. This beneficial effect was also shown for other syndromes, i.e., duodenal venous congestion lesions, perforated cecum, ischemic/reperfusion colitis, and bile duct ligation induced liver cirrhosis and portal hypertension[1,7]. The resolution of these various venous occlusion-induced syndromes[1,7,9-11] emphasized the practical evidence. The stable gastric pentadecapeptide BPC 157, as a membrane stabilizer[5], likely acts as the native cytoprotective gastric peptide[1,3,7], which is resistant and stable in human gastric juice[12], and counteracts gut-leaky syndrome[6]. As a particular target, it is distinct from the standard peptide growth factors[3], involving particular molecular pathways [102-105], particularly controlling VEGF and NO pathways[1,106,107], and the prostaglandin pathway[1].

Epithelial pathway in stomach and gastrointestinal tract healing for cytoprotection against direct cell injury produced by direct contact with noxious agents

BPC 157 consistently counteracted the gastric lesions induced by 96% alcohol^[1]. Of note, epithelial protection, as direct cytoprotection against direct cell injury produced by direct contact with noxious agents (*i.e.*, alcohol)[14-16], appears to be essential to resolve the follow-up of Robert's stomach cytoprotection ("epithelial pathway")[14-16]. As with Robert's alcohol intragastric application, this was a more advantageous therapeutic effect, overriding previous common limitations shared by standard cytoprotective agents (*i.e.*, prophylactic effect that may only counteract lesion development, but is unable to cure already existing lesions upgraded to the equal therapeutic ability[1]). BPC 157 demonstrated very consistent efficacy in alcoholinduced gastric lesions for co-, pre-, and post-treatment regimens, with a rapid onset of therapeutic effect, thereby providing consistent evidence for undistributed pertinent and specific effects, such as protection and healing, and the likely positive effects of an unusually high range[1]. This essential stomach point is confirmed and appreciated by others[108]. The BPC 157 equipotent (co-, pre-, and post-treatment regimens, per-oral and parenteral) beneficial effect is particular. There are constant interactions with the NO system and capsaicin-sensitive somatosensory neurons, since it consistently appears in naive rats as well as in those challenged with NOS blockade (NOS blocker L-NAME), NOS substrate L-arginine (NOS over-activity), NO system immobilization (concomitant application of L-NAME and L-arginine), capsaicin (as newborn or adult), or small exogenous or endogenous irritants[1,7]. A comparable beneficial effect was also achieved in vitro (denervated (isolated) gastric mucosal cells)[109,110].

Further supporting evidence included a strong reduction of the Monastral blue staining in ethanol-treated rats and, thereby, endothelium maintenance[1,7] and comparable beneficial effect in the stress gastric ulcer model[1] and cysteamine-duodenal ulcer model[1]. The same high efficacy was observed for both intragastric

Zaishideng® WJG | https://www.wjgnet.com

and intraperitoneal regimens[1,7]. The evidence that BPC 157 fully counteracted NSAID-induced gastric and intestinal lesions is consistent with the prostaglandin requirement of Robert's model, and the beneficial effect of BPC 157 in the entire gastrointestinal tract[1]. Also, in addition to cysteamine- or ischemia/reperfusioninduced colitis[1,7], BPC 157 counteracted trinitrobenzene sulfonic acid (TNBS)[111] or iodoacetamide[112,113]-induced ulcerative colitis. Of note, the beneficial effect of BPC 157 is long-lasting, and may also counteract ulcer recidivation (*i.e.*, cysteamine ulcerative colitis)[1,7]. Also important for the issue of cytoprotection is the evidence that BPC 157 may counteract stomach ulcer and induce ulcer regression (i.e., clopidogrel-induced)[114], as recently demonstrated in another prototype model of direct injury, Okabe's direct acetic acid application into stomach-induced gastric lesions[115,116], which is also commonly used in cytoprotection studies[1].

Provided that the essential point for lesions in Robert's cytoprotection model would be the injury made by direct contact (damage) to the cells[14-16], the perforation lesion instantly made by surgery is thereby a prototype[1]. The healing of perforated injury by the application of BPC 157 is an important conceptual point[1]. Further consequent evidence includes the healing of skin wounds and other wounds[1,3]. Importantly, proper wound healing includes the achievement of all four major events (vascular constriction, loose platelet plug, fibrin mesh to ensure stability of platelet plug, and dissolution of the clot) that occur in a set order following the loss of vascular integrity [3]. As a result, an agent applied in wound healing, such as stable gastric pentadecapeptide BPC 157, which is shown to be effective in wound healing, should also be effective in bleeding disorders[3].

Together, these consistent beneficial effects clearly indicated a full potential, in addition to the achievement of local protection and therapy of the stomach and gastrointestinal tract[1], toward Robert's point (other organ (epithelia) protection and therapy achieved)[14-16]. Of note, as pointed out, these studies indicated the use of the stress gastric ulcer models as a "cytoprotective" model (i.e., not related to gastric acid secretion)[39]. The significance of the stress gastric ulcer models is fairly described in several reviews[117-121]. Likewise, the connection with the prostaglandin system (and thereby, Robert's cytoprotection) is fully substantiated[31]. For BPC 157, the use of the prolonged restraint stress procedure[1] was important, provided that the use of the restraint stress methodology by gradually modulating/increasing the level of the stress[39,120] (e.g., usual cold + 3 h[39,120] vs 48 h restraint stress[1]) fully highlighted its efficacy[1]. Thereby, we could consistently suggest the effectiveness of BPC 157 over the application of standard H2-blockers or dopamine agonists[1].

Likewise, providing protection against the possible negative influence of gastric acid (hyper)secretion, in addition to the counteraction of Shay stomach ulcers induced by pylorus ligation[122] (but no influence on gastric acid secretion[123]) by BPC 157, there is some antagonism of the cysteamine-induced duodenal ulcer[1]. Since the Szabo's study[86], cysteamine-duodenal ulcers are commonly related to gastric acid hypersecretion[124-127]. However, we should consider stress lesions[85] as cytoprotection before Robert's cytoprotection[14-16], and thereby, Selye's "stress view" [85]. The introduction of cysteamine duodenal ulcers in rats will overcome the problems arising from multiple gastric erosions as the most characteristic rat gastrointestinal manifestations of exposure to stress, and would closely mimic human "stress ulcers", which are frequently localized in the duodenum[85]. Selye and Szabo considered the duodenal ulcer potency of various agents, and also emphasized "some relation to nonspecific stress" since cysteamine was the most potent agent of the other agents assessed (acetanilide, allylchloride, acetaminophen, 4,4-diaminodiphenylmethane, proprionitrile, and 3,4-toluendiamine) which were capable of inducing such lesions [85]. Yet, at that time, no mention was made on any influence of dopamine or gastric acid secretion[85]. With such particular "stress" notation to the duodenal lesions[85], initiation goes along with the emergence of the histamine, and the H2 receptor blocker-mediated resolution of peptic ulcers[128]. The subsequent cysteamine report by Szabo in the Lancet revealed the dopamine and gastric acid hypersecretion background, meaning that it became a seminal dopamine paper [86].

Also, this beneficial effect in cysteamine-induced duodenal ulcers[1] combined BPC 157 application with the dopamine system. Szabo provided cysteamine as a dopamine antagonist and its close similarity with the parkinsongenic neurotoxin 1-methyl-4phenyl-1,2,3,6-tetrahydropyridine (MPTP) (that also induced duodenal ulcers)[129, 130] in support of the theory holding schizophrenia, Parkinson's disease, and ulcer disease as dopamine system failures, and dopamine antagonists (ulcerogenic potential)/dopamine agonists (therapy) in peptic ulcer therapy[131]. Lately, the interaction of BPC 157 with the dopamine system was reviewed[2]; BPC 157 counteracted the effect of neuroleptics (haloperidol), MPTP, and reserpine (i.e.,



akinesia, catalepsy, hypothermia, and gastric lesions). Also, BPC 157 counteracted the models resembling positive-like symptoms of schizophrenia[55], and haloperidolinduced catalepsy and gastric ulcers^[2]. This effect, as a close interaction with dopamine system functioning, was able to determine an active gut-brain axis or braingut axis functioning[2]. It should be noted that BPC 157 also counteracted various encephalopathies and behavioral disturbances, and may therefore represent essential brain-gut and gut-brain axis activities^[2]. As an extension of the therapeutic effect, BPC 157 also counteracted the typical and atypical neuroleptic-induced arrhythmias, QTcinterval prolongation[1]. The counteraction of the prolonged QT interval appeared as part of the large therapeutic effect of BPC 157 on the heart disturbances noted in the prevention and reversal of doxorubicin-induced chronic heart failure[1], and the counteraction of various arrhythmias[1], including those induced by venous occlusion [9-11].

Subsequently, again with the 96% alcohol-induced gastric lesion[1], the cytoprotective effect of BPC 157 was closely related to the NO system that should have an essential role in the maintenance of gastrointestinal mucosa integrity, and, more importantly, in endothelial functioning[1,8]. BPC 157 induced NO release from homogenate supernatants of the gastric mucosa from the rat stomach, which is particularly resistant to the NOS blocker N(G)-nitro-L-arginine methylester (L-NAME), and may counteract the NOS substrate L-arginine-induced NO over-release^[1]. This particular interaction may be seen in various models and species with the ability of BPC 157 to counteract the adverse effects of L-NAME and L-arginine application[1].

Also, an essential point to remember in the cytoprotective effect of agents is capsaicin-sensitive afferent neurons[45,46], which regulate vascular function in many somatic and visceral tissues, including the regulation of local blood flow in the gastrointestinal tract. Thereby, the important point is that the beneficial effect of BPC 157 in gastric lesions induced by ethanol, restraint stress, or indomethacin was combined with the maintained as well as restored capsaicin-sensitive afferent neurons [1]. Quite recently, this cytoprotective notation was confirmed with the evidence that BPC 157 acts via inhibition of the release of enteric serotonin, an increase in the rat and human survival rate of cultured enteric neurons, and the proliferation of cultured enteric glial cells (EGCs)[132]. It was suggested that the inhibition of the release of enteric serotonin may be related to the release of serotonin noted in several brain areas (*i.e.*, nigrostriatum) after the administration of BPC 157[2,132].

Together, these findings clearly indicate a complex involvement of BPC 157 in the practical realization of cytoprotection as a non-gastric acid dependent phenomenon and "direct cell injury to cell - direct cell protection" principle. Furthermore, unlike its ulcerogenic effect ascribed to gastric acid hypersecretion[86], we showed that the application of cysteamine after gastrectomy induced duodenal ulcers in gastrectomized rats, and BPC 157, as well as all standard anti-ulcer agents, may clearly antagonize these cysteamine-induced ulcers in gastrectomized rats[1]. Interestingly, sialoadenectomy abolished the beneficial effect of standard antiulcer agents on cysteamine-induced duodenal ulcers, while BPC 157 was also effective in sialectomized rats[1]. As mentioned above, further evidence showed cysteamine enemainduced ulcerative colitis^[1]. Thus, these findings may be used as a full argument that cysteamine-induced ulcer appears as originally suggested (stress ulcer, non-gastric acid-dependent)[5,8] while cytoprotection, as the non-gastric acid-dependent phenomenon and "direct cell injury to cell - direct cell protection" principle, is continuously operating[1].

Epithelial pathway for innate cytoprotection both inside and outside the

gastrointestinal tract

The wider range of BPC 157 therapy[1,7] follows the definition of the innate cytoprotective potential in additional circumstances (*i.e.*, other epithelial healing), which has to combine the healing of different tissues, and is thus a pleiotropic beneficial effect[14-16]. Provided that cytoprotection represents a huge range of beneficial effects as the prototype model[1], there was a consistent demonstration of the strong therapeutic effect of BPC 157[1]. As emphasized, it not only occurred in the entire gastrointestinal tract[1,2], but also in various liver lesions, acute pancreatitis, and heart, lung, and kidney disturbances^[1]. The consistent beneficial effects that include a considerable number of models may clearly verify the large range of therapeutic effects[1]. For instance, there are therapeutic effects in the liver lesion network against prolonged restraint stress, bile duct and hepatic artery ligation, CCl4 application, chronic alcohol drinking, NSAID over-dose application, insulin over-dose, and bile duct ligationinduced cirrhosis[1]. In particular, the beneficial effects occur against ischemiareperfusion injury following Pringle maneuver[10], and Budd-Chiari syndrome[11] in



rats. Acute pancreatitis models were represented by bile duct ligation or lower esophageal and pyloric sphincter dysfunction[1]. As already mentioned, heart disturbance counteraction[1] was based on doxorubicin-induced chronic heart failure [57], and the counteraction of various, quite distinctive arrhythmias. This may be clearly seen providing the wide range of noxious events tested (*i.e.*, digitalis, hyperkalemia, bupivacaine, and lidocaine)[1] and venous occlusion procedures applied[9-11]. Likewise, the lung lesion counteraction is based on edema of the interstitium, and substantial dilatation and congestion of the capillaries in the alveolar septum in the lung of rats with venous occlusion syndromes[10,11]. If not corrected, the lung congestion appears as a common outcome (*i.e.*, time-dependent and time-independent features that can be acute respiratory distress syndrome exudative phase features); acute lung injury is a primary component of multiple organ dysfunction syndromes triggered by intestinal ischemia-reperfusion, which results in high mortality and acute lung injury[133,134].

Likewise, as a general follow-up of Robert's cytoprotection, the BPC 157 wound healing studies appear to be well founded [1,3]. As well as gastrointestinal ulcers, consistent evidence includes various skin wounds. In addition to the incisional wound and deep burns and fistula wounds[1,3], there were also diabetic ulcers[102,135] and alkali wounds[105]. These beneficial effects also include the healing of muscle (i.e., the healing of the transected, crushed and denervated muscle), tendons (transected Achilles' tendon and Achilles' tendon detached from the calcaneus), ligaments (transected medial collateral ligament), and bone (alveolar bone loss and radial pseudoarthrosis)[3]. The delivery of BPC 157 was through local (*i.e.*, cream application) and systemic (*i.e.*, intraperitoneally, or intragastrically, or per-orally in drinking water) methods[1,3]. The therapeutic effects of BPC 157 on tendon and muscle healing was also investigated[3,103,104,107]. Moreover, there is a strong practical distinction from the standard angiogenic factors[3]. As pointed out, bFGF, EGF, and VEGF gastrointestinal tract studies demonstrated improved healing[3]. However, most of their corresponding studies on tendon, muscle, and bone injuries provide evidence of their increased presentation along with various procedures used to produce beneficial effects, compared to fewer studies in vitro[3]. In vivo healing evidence of these standard angiogenic growth factors was limited, commonly to local application. Evidently, providing the use of different carriers with corresponding peptides, there is an obvious attribution problem due to different combinations of peptide + carrier complex. Thereby, for the standard growth factors and use of different carriers, there is inadequate evidence due to diverse healing evidence with diverse carriers and delivery systems^[3]. Contrary to this, BPC 157, using the same regimens as gastrointestinal healing studies (always given alone, without carrier), improves tendon, ligament and bone healing, accurately implementing its own angiogenic effect in healing[3]. Important for the particular effect on angiogenesis (particular in consideration of the corneal avascularity as "angiogenic privilege", no formation of corneal neovascularization which is essential for corneal wound healing)[136], later studies also included corneal wound healing and maintained corneal transparency (rescued total debridement of the corneal epithelium and perforating corneal incisions)[3]. The evidence that BPC 157 eye drops successfully close perforating corneal incisions in live rats is consistent with the cytoprotection/endothelial/mucosal protection model[3]. Regardless of its complex function in the corneal endothelium, endothelial maintenance by BPC 157 is also implicated in the healing of corneal ulcers in live rats [3]. Since this model is sensible, we suggest that BPC 157 should have tissue-specific healing effects[3]. Thus, we can envisage a particular healing potential in cytoprotection terms. From the method viewpoint [1,3], all of these lesions are within the scope of Robert's direct cell injury produced by direct contact[14-16].

In addition, there is quite indicative evidence about the simultaneous healing of different tissues. There is healing of various anastomoses (vessel, nerve, and gastrointestinal tract) and of various fistulas (surgically induced by defects and anastomosis creation), both external and internal[1]. Together, these findings showed that this additional extent (*i.e.*, the healing of other epithelia) may be combined in the simultaneous healing of different tissues, such as the simultaneous healing of fistula defects and the closing of fistulas[1]. A particular point is that these rat fistulas are severe, considering the significant size of the defect relative to the small size of the corresponding rat tissue[1]. Illustratively, rectovaginal fistulas in rats, with a 5 mm defect *vs* a 2.4 cm vaginal length, result in long-lasting defects and spontaneous patency of the fistula, leading to fecal matter leaking through the vagina; this actually mimics severe fistulas that may not spontaneously heal, thereby clearly emphasizing the beneficial effect of BPC 157[1].

Further, for BPC 157, in an additional cytoprotective extent (i.e., other epithelia healing), epithelium protection is based on the extended relevance of the intragastric alcohol or NSAIDs on the stomach lesions commonly used in Robert's cytoprotection studies[1,14-16]. Namely, Robert's first epithelium protection, or the direct cell protection against cell injury produced by direct contact with the noxious agents, used intragastric alcohol or NSAIDs to induce stomach lesions[14-16]. Consequently, further evidence toward an additional extent (*i.e.*, other epithelia healing) follows other adverse effects of alcohol and NSAIDs and their consistent counteraction [1,7].

In addition to the 96% alcohol intragastric application-induced gastric lesions, BPC 157 largely counteracted chronic alcohol drinking-induced stomach lesions, liver failure, and portal hypertension, providing evidence that it may act as an alcohol antagonist[1]. Likewise, BPC 157 promptly counteracted acute alcohol (4 g/kg intraperitoneally) intoxication (i.e., quickly produced and sustained anesthesia, hypothermia, increased ethanol blood values, 25% fatality, 90-min assessment period) given before or after ethanol[1]. In addition, BPC 157 counteracted chronic (withdrawal) alcohol intoxication, and was suggested as an alcohol antagonist[1], peripherally and centrally (of note, BPC 157 may attenuate the effect of thiopental anesthesia)[2].

Confronted with the over-dose application of various NSAIDs^[1], similar beneficial effects occurred against various gastrointestinal lesions, and liver and encephalopathies; the worst damaged areas showed the most evident therapeutic effect[1]. Prolonged bleeding, consequent thrombocytopenia, and thrombocyte malfunctioning were also attenuated and/or counteracted[1,3,7]. Therefore, it seems that BPC 157 may particularly affect the functioning of the prostaglandins system[1,3,7] (interestingly, unlike NSAIDs and corticosteroids, BPC 157 strongly prevented adjuvant arthritis development and reversed the already formed adjuvant arthritis in rats[1]). The final clue may be that BPC 157 counteracted indomethacin-induced leaky gut syndrome[6]. It acts via increasing tight junction protein ZO-1 expression, and transepithelial resistance, inhibiting the mRNA of inflammatory mediators (iNOS, IL-6, IFNy, and $TNF-\alpha$), and increasing the expression of HSP 70 and 90, and antioxidant proteins, such as HO-1,NQO-1, glutathione reductase, glutathione peroxidase 2, and GST-pi[6]. Considering the importance of the leaky gut as an essential mechanism responsible for various severe systemic diseases, this may fully substantiate the significance of BPC 157 in the realization of that additional cytoprotective extent (*i.e.*, other epithelia healing)[6]. Also, BPC 157 counteracted other encephalopathies induced by various noxious events (insulin over-dose, cuprizone, multiple sclerosis mimicking neurotoxin, magnesium over-dose, brain trauma, spinal cord compression, and stroke)[1,2].

Epithelial pathway for adaptive cytoprotection

We also demonstrated that BPC 157 may regulate cytoprotection adaptation processes (adaptive cytoprotection)[1], functioning of the endogenous adaptive processes essential for permanent mucosal maintenance, and afford defensive reactions that start after any injurious event [1]. This follows Robert's connotation about the cytoprotection as a physiologic process^[14-16] based on the adaptive cytoprotection evidence of Robert's small irritant to the stomach that precedes and protects against any subsequent major injurious event (*i.e.*, Robert's strong irritant to the stomach)[17]. Evidently, cytoprotective agents should have a more extensive action, participate in Robert's first epithelium protection, exhibit direct cell protection against cell injury produced by direct contact with noxious agents, and also participate in adaptive cytoprotection, in the next defensive reaction, and afford its final beneficial effects (*i.e.*, permanently attenuated lesion consequences)[1]. Thus, whatever the small irritant may be, whether exogenous (mild alcohol) or endogenous (*i.e.*, accumulated gastric juice, gastric acid, *i.e.*, made by gastrojejunal anastomosis), BPC 157 administration strongly contributed to the final attenuation of stomach lesions^[1]. Thus, BPC 157 strongly contributes to and improves the presentation of adaptive cytoprotection processes[1]. Specifically, BPC 157 would improve adaptation processes in the damaged intestine, through a prostaglandin-related process, as it may be strongly aggravated by the application of NSAIDs^[1]. In rats with short bowel surgery, the BPC 157 therapy, per-oral (in drinking water) and parenteral, causes constant weight gain (even more than preoperative values), with all three wall layers accordingly increased (i.e., villus height, crypt depth, and muscle thickness [inner (circular) muscular layer] also increased), but no difference in jejunal and ileal diameters, and increased anastomosis strength. These beneficial effects of BPC 157 (i.e., the weight gain in the BPC 157 rats with short bowel, all three wall layers accordingly increased) appear to be particular[1]. Namely, standard growth factors [even using a special application route (e.g., subcutaneous pump)][137,138] at best may induce a decrease in weight loss[139-



142], with an increase in one layer, but not in the other. There is also some caution about the use of peptidergic agents, and adaptation processes, particularly on a longterm basis^[140]. There is some growth of several tumor cell lines (EGF)^[143,144], and hyperplastic lesions in the colon (subjects treated with GLP-2[145]). In contrast to adequately controlled adaptive processes, supportive evidence for BPC 157 (i.e., BPC 157 administration showed no toxic effect and was limit test negative, with LD1 not achieved, and no side-effects in trials[1,3]) shows that it inhibits the growth of several tumor cell lines and counteracts the tumor-promoting effect of vascular endothelial growth factor (VEGF)[1,4]. In mice with C26 colon adenocarcinoma, BPC 157 counteracted tumor-cachexia and markedly prolonged survival[5]. BPC 157 afforded significant mitigating action against cancer cachexia-induced muscle degeneration, inflammation, and catabolism. BPC 157 significantly corrected deranged muscle proliferation as well as myogenesis, counteracted an increase in proinflammatory cytokines such as IL-6 and TNF- α looking at muscle metabolism relevant to cancer cachexia, as well as any changes in the expression of FoxO3a, p-AKT, p-mTOR, and P-GSK-36[5]. Also important for its likely control of the adaptation processes, and prostaglandinssystem function, in the short bowel rats, BPC 157 may counteract gastrointestinal lesions and the concomitant liver and brain lesions, and the additional aggravation that would otherwise appear with the application of diclofenac[1].

Also, it is possible that BPC 157 would afford an adaptive cytoprotective reaction regardless of the site of its initiation in the gastrointestinal tract[1]. Supporting evidence was also provided showing that adaptation cytoprotection accordingly occurs in the complete gastrointestinal tract, lasting for a considerable time, depending on the part that is initially targeted by the small irritant, stomach, duodenum, or colon, enabling the other parts to be more resistant to any subsequent strong irritant challenge[1]. Considering the eating and drinking habits, the adaptive cytoprotection in the gastrointestinal tract starts in the upper parts, in the stomach and duodenum, and may beneficially affect other parts (and thereby, adaptive cytoprotection occurs between stomach \rightarrow stomach; stomach \rightarrow duodenum, stomach \rightarrow colon; duodenum \rightarrow duodenum; duodenum \rightarrow stomach, duodenum \rightarrow colon)[1]. The colon seems to be distinctive and passive, as it could not initiate an adaptive cytoprotection response[1]. We used combinations of specific agents for initial small lesion and final more severe lesion [1 mL/rat of 25% or 96% ethanol intragastrically (stomach); cysteamine 40 mg/kg or 400 mg/kg subcutaneously (duodenum); cysteamine 40 mg/kg or 400 mg/kg intrarectally (colon)[1]. All of these ulcerogens were known to be inhibited by BPC 157[1].

Finally, with normal eating and drinking, Robert's adaptive cytoprotection (i.e., Robert's small irritant to the stomach and Robert's strong irritant to the stomach) showed another essential point. We used the tongue as the initial target[1]. Within the very short time needed to swallow, the stomach is immediately affected, and the lesions are considerably less than those obtained with the direct instillation of alcohol into the stomach[1]. The application of BPC 157 considerably afforded this spontaneous healing effect, and additionally mitigated tongue, esophageal, gastric, and duodenal lesions, and reversed lower esophageal and pyloric sphincter impairment, through an action which seems to be NO system dependent[1]. Actually, it means that Robert's cytoprotection and adaptive cytoprotection following the direct application of noxious agents into the stomach completely avoid the regular defensive response that would occur with the tongue (and not the stomach) as the initial target.

On the other hand, this emphasizes the original significance of Robert's application of alcohol directly into the stomach, and thereby cytoprotection, as the direct cell protection against direct cell injury produced by direct contact with the noxious agents. Robert's regimen (alcohol applied in the stomach directly, by tube) regularly skips the existing defensive system (*i.e.*, starting with the tongue). Consequently, spontaneous rapid healing mechanisms remain skipped and not activated. Thereby, the essential ability of the cytoprotective agents would depend more on their own healing capacity, and their ability to act rapidly to induce healing.

Thus, such a huge range of healing effects, as noted with the applications of BPC 157, should be a prerequisite for realizing the endothelium pathway (blood vessel recruitment and activation towards defect or bypassing vessel occlusion)[1,7].

Endothelium pathway (endothelium maintenance to epithelium maintenance)

We already emphasized [1,7] the original cytoprotection studies [14-16,20,21] from the 1980s, which demonstrated significant stomach endothelium lesions, and verified the consequent change in stomach injuries via the endothelium pathway (endothelium maintenance \rightarrow epithelium maintenance)[14-16,20,21]. Since that time, the cytoprotective endothelium pathway remained to be fully elaborated for therapeutic



purposes, both as a stomach therapy and as a more general therapy. With this high therapy requirement, we summarize the additional evidence. These highlights include the Virchow triad situation, endothelium injury, thrombus, and stasis, preceding the current demonstration that the administration of BPC 157 may finally induce the rapid recruitment of existing blood vessels and activate particular collateral pathways when confronted with vessel occlusion[1,7,9-11]. That pathway activation would accordingly compensate for the occlusion of vessels, and mitigate the consequent noxious chain of events[1,7,9-11]. We would analyze the possible cause in the indicated cytoprotection terms leading to an extension of the endothelium pathway to blood vessel recruitment and activation towards defect or bypassing vessel occlusion[1,7,9-11].

After presenting the initial cytoprotection concept (*i.e.*, epithelial pathway)[14-16], the endothelial pathway appeared as a further clarification of the development of stomach lesions in the concept of cytoprotection [14-16,20,21]. As a common point [47] appeared the evidence that with alcohol intragastric instillation, these vascular changes are early events, even before the appearance of gross hemorrhagic lesions, occurring within seconds or minutes during the development of moderate or severe gastric mucosal injury with interstitial hemorrhage and the necrosis of glandular epithelial cells[14-16,20,21]. Additionally, there is early stasis of mucosal blood flow and thrombi formation (within 30 s), often in regions without deep necrotic lesions. Even more, there was the rapid and complete cessation of blood flow to areas of mucosal damage consequent to ethanol administration[146]. Thus, although this was not initially claimed for the beneficial effect of prostaglandin and cysteamine application[14-16], we can envisage the particular Virchow triad presentation[1,7]. Unlike the initial claim for generalization of the epithelial stomach protection to other epithelial protection (cytoprotection \rightarrow organoprotection)[14-16,19], at that time, these studies[14-16,20,21] made no attempt to generalize the findings seen for endothelium recovery in the stomach.

A strong reduction of Monastral blue staining and maintenance of the endothelium integrity after alcohol intragastric application was considered to be essential for the healing effect of BPC 157[1]. An interesting insight appeared after absolute alcohol instillation in the fully distended rat stomach, and gastric, esophageal, and duodenal lesions. Throughout the next 3 min, left gastric artery blood vessels clearly disappeared at the serosal site, indicative of the loss of vessel integrity and function. In contrast, constant vessel presentation could predict the beneficial effect of the applied agent. After pentadecapeptide BPC 157 instillation into the stomach, the vessel presentation remained constant, and lesions of the stomach, esophagus, and duodenum were inhibited[1]. Standards (atropine, ranitidine, and omeprazole) could only slightly improve the vessel presentation compared to control values, and only had a partial effect on the lesions[1]. Furthermore, for BPC 157, this maintenance of the endothelium integrity initially revealed a strong inhering angiogenic effect, which was more potent than those noted for standard antiulcer agents[1,7]. This appeared as a follow-up of "direct" cellular pharmacological treatment for ulcer with growth factors, notably bFGF and PDGF, that should result in the superior quality of ulcer healing by optimal angiogenesis, and thereby dense granulation tissue, as well as the complete reepithelization and restoration with minimal inflammation[32]. Moreover, with BPC 157, it appears that angiogenesis was closely related to its wound healing and promotion, as well as healing in other tissues (*i.e.*, muscle, tendon, and ligament, known to be hypovascular tissues)[3]. In particular, in both muscle (transected or crushed) and transected tendon healing, we noted an increase in early angiogenesis (and the increased expression of VEGF, Factor VIII, and CD34), while late angiogenesis decreased (and the expression of VEGF, Factor VIII, and CD34 was decreased)[3]. The therapeutic potential (*i.e.*, acceleration of the blood flow recovery and vessel number in rats with hind limb ischemia) of pro-angiogenic BPC 157 is associated with VEGFR2 activation and up-regulation[107]. It also immediately triggered the internalization of VEGFR2 and subsequently activated the phosphorylation of VEGFR2 and Akt, and the eNOS signaling pathway without the need for other known ligands or shear stress [107].

On the other hand, as the reduction of Monastral blue staining and maintenance of the endothelium integrity after alcohol intragastric application is an immediate effect of BPC 157, we should consider the pleiotropic beneficial effects of BPC 157 in the entire gastrointestinal tract[1,7]. This should provide evidence that it effectively combines its particular mediator role (as an original anti-ulcer peptide which is stable in human gastric juice for longer than 24 h [12] and thereby, in Robert's stomach cytoprotection, protection against direct cell injury made by direct contact of various noxious agents and required endothelium protection and maintenance of the endothelium function [1,7]. This has to be an immediate and rapid effect [1,7]. Thereby,



BPC 157, as a cytoprotective agent in the entire gastrointestinal tract, may both prevent and reverse the Virchow triad situation, and have an additional modulatory role[1,7].

As the first evidence of the implementation of the endothelium maintenance originally noted in stomach cytoprotection studies[1], BPC 157 prevents and reverses thrombosis formation after abdominal aorta anastomosis, or major vein occlusion [1,7, 9-11]. Furthermore, BPC 157 may attenuate the prolonged bleeding that appeared after different injuries (i.e., tail or leg amputation, organ perforation, and prolonged occlusion of the inferior caval vein) or anticoagulants, such as heparin or warfarin, and aspirin and the NOS substrate L-arginine application [1,7,9]. Also, it was shown that BPC 157 maintains thrombocyte function, without interfering with coagulation pathways[1,7]. Furthermore, there is evidence that BPC 157 counteracted stroke, given in reperfusion, after clamping of the common carotid arteries [i.e., both early and delayed neural hippocampal damage, achieving full functional recovery (Morris water maze test, inclined beam-walking test, and lateral push test)][147]. Together, this may be a particular modulatory effect or NO-system-related effect[1]. BPC 157 may counteract both the NOS blocker L-NAME's pro-thrombotic effect and the NOS substrate L-arginine's anti-thrombotic effect in the same way that it counteracted both L-NAME-induced hypertension and L-arginine-induced hypotension, and could induce the NO release on its own, which is quite resistant to L-NAME application[1]. Finally, in addition to the VEGFR2-Akt-eNOS signaling pathway being activated without the need for other known ligands or shear stress[107], there is a direct effect on vasomotor tone (i.e., specific activation of Src-Caveolin-1-endothelial nitric oxide synthase (eNOS) pathway)[106]. Also, it should be recalled that four major events (vascular constriction, loose platelet plug, fibrin mesh to insure stability of platelet plug, and dissolution of the clot) are implicated in the wound healing process and occur in a set order following the loss of vascular integrity[3]. Consequently, it may be not surprising that an agent implemented in wound healing, such as stable gastric pentadecapeptide BPC 157, should be effective in this particular way also in bleeding disorders[1,3], due to its innate cytoprotective effect, and the fact that it has been shown to be an effective therapy in wound healing[3].

Finally, in consideration of the previous original findings in cytoprotection endothelium studies (complete cessation of blood flow to areas of mucosal damage and rapid cloth formation consequent to ethanol administration[146]), and resolving of the presented Virchow triad circumstances, we suggested that the beneficial effect of cytoprotective agents should be related to the resolution of this noxious chain of events[1,7]. Thus, conclusive evidence involves confrontation with permanent major vessel occlusion, and therapeutic evidence that BPC 157 administration quickly recruits vessels to rapidly activate the collateral pathway which would adequately compensate for vessel occlusion and reestablish blood flow[1,7,9-11]. There, the alleviated peripheral vascular occlusion disturbances rapidly activated alternative bypassing pathways[1,7,9-11], appears to be an additional follow-up of its essential endothelium protection[1], which was long ago implemented as an essential class activity of cytoprotective agents^[13]; however, in this way, this has so far only been implemented by the application and beneficial effects of the stable gastric pentadecapeptide BPC 157[1,7,9-11].

Rapid activation of a bypassing loop from the existing vessels

With BPC 157 therapy, when confronted with the occluded vessel in rats with distinctive vascular occlusion disturbances, we first reported the rapid activation of a bypassing loop recruited from the existing vessels (*i.e.*, intestinal arcade vessel network, or the left ovarian vein)[1,7,9-11]. The evidence [1,7,9-11] included the infrarenal occlusion of the inferior caval vein, left colic artery and vein occlusion ischemic/reperfusion ulcerative colitis, superior anterior pancreaticoduodenal veininduced duodenal venous congestion lesions, bile duct ligation-induced liver cirrhosis and portal hypertension, temporary occlusion of the portal triad (Pringle maneuver)induced ischemia-reperfusion injury[10], and suprahepatic occlusion of the inferior caval vein-induced Budd-Chiari syndrome[11]. This occurred in rats with a ligated part of the left colic artery and vein, ischemic/reperfusion colitis, or an infrarenal ligation of the inferior vena cava[1,7,9-11]. Evidently, the BPC 157 application-induced activation of the collateral pathways (the left ovarian vein and other veins in rats with infrarenal occlusion of the inferior caval vein) may rapidly resolve any systemic disturbances (i.e., caval hypertension, aortal hypotension, heart dysfunction, thrombosis, and consequent thrombocytopenia, and induced bleeding prolongation in rats with infrarenal occlusion of the inferior caval vein)[1,7,9-11]. Likewise, there is also the local injury counteraction (attenuated/counteracted ischemia/reperfusion injury) in a rat study of the ischemic/reperfusion colitis[1,7]. As emphasized[1], with

part of the left colic vein and artery excluded by two ligations, along with BPC 157 application, blood vessels propagated toward the injury obstruction, bypassing it, interconnecting collaterals between arcades, and reestablishing the inside-outside point. In reperfusion, the application of BPC 157 after the initiation of full reperfusion with both ligations removed resulted in increased vessel presentation and arcade interconnections. With application of BPC 157 in ischemia as well as in reperfusion, the mucosal folds were recovered, and the pale areas were small and markedly reduced in size^[1]. In the ischemia and even more so in the reperfusion, oxidative stress was counteracted, and the otherwise increased MDA (as a result of the lysis of endothelial cells^[148,149] and NO levels in colon tissue were found to be normal in rats that received BPC 157 bath treatment^[1]. This occurs as before in both ischemic and reperfusion conditions in the various tissues (i.e., the colon, duodenum, cecum, liver, and veins) and plasma[1,7,9-11]. Thus, the action of BPC 157 as a free radical scavenger (noted also in the other tissues, *i.e.*, gastrointestinal sphincters, stomach, duodenum, bowel adhesions, bladder, and brain [1,7,9-11]) may considerably contribute to its pleiotropic beneficial effects and maintain endothelial function. Notably, BPC 157 contains four carboxylic groups that could be active in scavenger process, and if they are reactivated (by, e.g., glutathione or enzymes), the overall antioxidant activity could be very high [1].

Thus, relieving Virchow's triad situation is the particular activation of collateral pathways corresponding to the damaging occlusion [*i.e.*, mentioned passing through arcade vessels (occlusion of the left colic artery and vein) or the left ovarian vein (infrarenal occlusion of the inferior caval vein)][1,7,9-11]. As pointed out[1,7,9-11], the superior anterior pancreaticoduodenal vein-inferior anterior pancreaticoduodenal vein-superior mesenteric vein appears to counteract duodenal congestion lesions [1,7,9-11]. A porto-caval shunt appears with the portal vein-superior mesenteric vein-inferior mesenteric vein-rectal vein-left iliac vein-inferior caval vein pathway to counteract portal hypertension in rats with bile duct occlusion or ischemia-reperfusion injury following Pringle maneuver[1,7,9-11]. The inferior caval vein - azygos vein - left superior caval vein pathway appears to counteract Budd-Chiari syndrome in rats[1,7, 9-11].

Of note, an adequate compensation regularly occurred. As pointed out in our venous occlusion studies [10,11], there is consistent evidence in rats with bile duct ligation. Preventing the development of portal hypertension, and the rapid reversal of the already established portal hypertension, are both among its additional beneficial effects [150]. We noted that BPC 157 therapy markedly abated jaundice, ascites, and nodular, steatotic livers with large dilatation of the main bile duct, increased liver and/or cyst weight, and decreased body weight[150]. Furthermore, the piecemeal necrosis, focal lytic necrosis, apoptosis, and focal inflammation, disturbed cell proliferation (Ki-67-staining), cytoskeletal structure in the hepatic stellate cell (α -SMA staining), and collagen presentation (Mallory staining) were all counteracted, providing evidence that BPC 157 may affect both liver fibrosis and portal hypertension [150]. Thus, this may be the principle seen in venous occlusion studies[10,11].

As previously reviewed[1], in rats with a perforated cecum, BPC 157 application rapidly reversed the regular noxious course, with the rapid disappearance of blood vessels at the cecum serosa (emptied/disappeared), thereby producing a large immediate defect, with bleeding, the leakage of fluid, increased oxidative stress, and disturbed NO-levels in cecal tissue. With BPC 157, there is immediate blood vessel recruitment and activation ("running") towards the site of injury[150], as was described in the "bypassing" of vessel occlusion via alternative pathways[9-11], which can likely cure rats and reestablish blood flow. Also, a small-vessel network appeared around the perforated defect with BPC 157 bath administration; cecal defect enlargement reversed to defect contraction (i.e., each defect breaks blood flow) may be a result of the reestablishment of blood flow as well as the shortened bleeding time from the perforated cecum[1]. Less bleeding corresponds to the beneficial effects in rats with amputation, anticoagulant or aspirin application, or vein obstruction; direct defect closing corresponds to the closing of various fistula defects, which were also surgically created in corresponding tissues[1,7,9] (i.e., all by Robert's direct injury to the cell by direct contact).

Along with these findings[1,7,9-11] is the beneficial effect of BPC 157 in rats with a damaged peritoneum. Endothelium maintenance → epithelium maintenance = blood vessel recruitment and activation ("running") towards the site of injury, also described as "bypassing" the occlusion via alternative ways [1,7,9-11], was seen with BPC 157 administration after parietal peritoneum excision with an underlying superficial layer of muscle tissue in rats to counteract failed vasculature, and finally to counteract the increased formation of adhesions. Rapid abundant vascular vessels in and close to the

WJG | https://www.wjgnet.com

defect mean that BPC 157 could interfere with the motion of the coagulation cascade once the peritoneum is damaged [1,7,9-11]. When two damaged peritoneal surfaces come into contact with each other, BPC 157 is likely to interfere the temporary role of fibrin in healing without adhesions that must be degraded by the fibrinolytic system for the restoration of normal tissue structure and function, as it reversed the healing that would result in fusion to form a connection, *e.g.*, an adhesion[151,152].

Finally, with the BPC 157 therapy in the Pringle maneuver in rats[10], severe preportal hypertension, temporary portal triad obstruction, ischemia, and short and prolonged reperfusion, we resolved the regular lack of adequate portocaval shunting as the most detrimental feature that should be counteracted [10]. With the stable gastric pentadecapeptide BPC 157, we noted the resolution of damage, either following occlusion or following the re-opening of the hepatic artery, portal vein, and bile duct. Therefore, in the portal triad obstruction syndrome in rats, in the rapidly activated manner, portal vein-superior mesenteric vein-inferior mesenteric vein-rectal vein-left iliac vein-inferior caval vein pathway would appear as specific activation of the collateral circulation, as the bypassing loop that can rapidly circumvent occlusions and decompress portal triad obstruction in rats upon BPC 157 administration[10]. That solution in rats with ischemia and reperfusion following the Pringle maneuver goes along with the resolution of oxidative stress, hemodynamic disturbances, severe portal and caval hypertension, aortic hypotension, rapid cloth formation in the portal vein, superior mesenteric vein, lienal vein, inferior caval vein, and hepatic artery, ascites, peaked P waves, tachycardia, increased serum values, and gross intestine, liver, lung, spleen, and heart lesions[19]. In particular, it goes along with the application of agents during reperfusion. Furthermore, the pentadecapeptide BPC 157 resolved the suprahepatic occlusion of the inferior caval vein in a Budd-Chiari syndrome model in rats[11]. Budd-Chiari syndrome was perceived as originally suggested[153,154], a hepatic venous outflow obstruction and its manifestation, regardless of cause, but this was mostly attributed to thrombosis, which can be located anywhere from the small hepatic veins to the entrance of the inferior vena cava into the right atrium[153,154]. Thereby, bypassing the occlusion in the rat Budd-Chiari syndrome along with pharmacotherapy treatment should be essential. BPC 157 therapy results in the rapidly activated azygos/hemiazygos vein bypassing pathway, upgrading an inadequate rescuing inferior-superior vena cava shunt to an adequate one, as well as a portocaval shunt[11]. Consequently, the caval and portal hypertension and aortal hypotension presented by Budd-Chiari syndrome rats were largely eliminated by BPC 157 therapy [11]. Largely attenuated consequent disturbances (rapid clot formation in the portal vein, superior mesenteric vein, splenic vein, inferior vena cava, hepatic artery, and coronary artery, as well as peaked P waves, significant ST elevation, tachycardia, gross organ lesions, and liver and spleen weight increases) together support this contention [11].

Thus, BPC 157 application may counteract a life-threatening syndrome[9-11]. Characterized by the multiple mutual cause-consequence relationships in vascular occlusion-induced syndrome presentation in rats, the generalized thrombosis and stasis, vascular failure, and heart dysfunction, lung congestion appears to be a common outcome (i.e., time-dependent and time-independent features that the exudative phase features of acute respiratory distress syndrome)[9-11]. Acute lung injury is a primary component of multiple organ dysfunction syndrome triggered by intestinal ischemia-reperfusion. The results may be high mortality and acute lung injury[155,156], followed by liver failure (substantial congestion of central vein as well as branches of portal veins in portal triads), kidney congestion, prominent portal and caval hypertension, aortal hypotension, and consequential gastrointestinal hemorrhagic lesions[9-11]. Therefore, the previously mentioned beneficial effects, in elaborating the cytoprotective "epithelial pathway" (i.e., counteracted various heart or liver lesions), including the combined and simultaneous healing of different tissues[1], may also be essential. In particular, the compensatory efficacy of new functional equilibrium ("endothelium pathway") with the activated specific functioning collateral pathways[13-15] is also ascertained with an important notification for the general pathology of the portal hypertension [1,7]. Namely, BPC 157 counteracted all portal hypertension presentations whatever the cause, post-hepatic, hepatic, and pre-hepatic [1,9-11].

In addition, as in venous-occlusion syndromes[9-11], BPC 157 also counteracted various lung lesions[1,7].

Finally, with holistic concepts, any criticisms about the cytoprotection concept, such as "cytoprotection", "as everything and nothing", and "cytoprotection which is not mechanism", and thereby, criticisms about peptides and cytoprotection, could not be avoided. The general point that animal studies per se may be cautious regarding their results and the relative paucity of BPC 157 clinical data was also reported[1]. On the other hand, it should be noted that BPC 157 was proven to be efficacious in ulcerative colitis, both in clinical settings[157,158] and in experimental ischemic/reperfusion ulcerative colitis studies in rats and other ulcerative colitis models[1]. A particular point is the very safe profile (LD1 could be not achieved)[1,3,7], a point that was recently confirmed in a large study by Xu et al [159]. In this context, the role of the animal model is indispensable, and the practical evidence is even more important. Besides the majority of studies with BPC 157 conducted on rodents that were given an injection of the supplement, there have also been a considerable number of studies, particularly in gastrointestinal research, with intragastric application or peroral application in drinking water (regularly used in fistulas studies[1,3]), that are correspondingly effective. There are also studies in other species, *i.e.*, birds and insects (given in the food), which favor a more general effect of BPC 157 application[1,7]. Lastly, the suitability of the models used for the topic of cytoprotection, in particular, since Robert's original description of the cytoprotection in rats[14-16], evidently resolves the practical/theoretical consideration of the cause-consequence issue. Thus, the suited models and lesion counteraction clearly indicate the beneficial effects. The deciding result exemplified the resolved endothelium pathway (blood vessel recruitment and activation towards defect or bypassing vessel occlusion)[1], but the particular background still needs to be further elaborated. Note, the consistently used range of BPC 157 application (µg-ng) may also suggest a physiological role, in accordance with in situ hybridization and immunostaining for BPC 157 in the gastrointestinal mucosa, lung bronchial epithelium, epidermal layer of the skin, and kidney glomeruli[3]. Thereby, illustrative examples for further research may be the evidence that BPC 157 exhibited a specific effect on the Egr, Nos, Srf, Vegfr, Akt1, Plcy, and Kras pathways in infrarenal occlusion-induced inferior caval vein syndrome in rats. This appears in a timely manner, to be increased, decreased, or unchanged, depending on whether the vessel was blinded (the right ovarian vein and inferior caval vein) or open and served as an alternative operating pathway (the left ovarian vein)[9]. Also, to support the beneficial effect of BPC 157 on brain lesions, given in reperfusion in stroke rats[147], BPC 157 therapy counteracted both early and delayed neural hippocampal damage, showing that achieving full functional recovery can restore recognition memory deficits along with a therapeutic effect [147]. mRNA expression studies at 1 h and 24 h, provided strongly elevated (Egr1, Akt1, Kras, Src, Foxo, Srf, Vegfr2, Nos3, and Nos1) and decreased (Nos2 and Nfkb) gene expression (Mapk1 not changed). This may be how BPC 157 acts[147].

In conclusion, Robert's cytoprotection concept[14-16] was initially of intense interest, but lately received the claim that the concept's foundation ("gastric cytoprotection") is still relevant[23]. Anyway, the essential rebuilding was lacking. Now, the concept has been reexamined for many major reasons (Figure 1): (1) The gastric pentadecapeptide BPC 157, thought to be an essential cytoprotective mediator that is native to and stable in human gastric juice, was noted to have a pleiotropic beneficial effect[1,3]; (2) With the administration of BPC 157, in prophylactic as well as in therapeutic regimens, there is evidence of the innate Robert's cell protection in the stomach epithelium against direct injury (which may be induced by various noxious agents) using either method of application, which provides the ability to realize the protection of other epithelia as well[1-3]; (3) BPC 157 effectively combines its particular mediator role (as an original anti-ulcer peptide that is stable in human gastric juice for longer than 24 h); therefore, in Robert's stomach cytoprotection, it has a protective effect against direct cell injury made by the direct contact of various noxious agents, requiring endothelial protection and the maintenance of endothelial function. This has to be an immediate and rapid effect^[1]; (4) As first evidence of the implementation of the endothelial maintenance originally noted in stomach cytoprotection studies, BPC 157 prevents and reverses thrombosis after abdominal aorta anastomosis, or major vein occlusion[1,7,9-11]. Furthermore, BPC 157 may attenuate the prolonged bleeding that appeared after different injuries or anticoagulant, heparin or warfarin, and aspirin application[1,7,9-11]. Also, BPC 157 maintains thrombocyte function, without interfering with coagulation pathways [1,7,9-11]; and (5) The vessel recruitment activated collateral pathways to bypass vessel occlusion as a new conceptual point in the cytoprotection concept, and cytoprotective agent activity [1,7,9-11]. BPC 157 counteracted various venous occlusion-induced syndromes, inferior caval vein syndrome[9], ischemia/reperfusion injury following Pringle maneuver[10], and Budd-Chiari syndrome^[11] in rats. Activation of the alternative collateral pathways to bypass occlusion, and reestablishing alternative blood flow result in counteraction of the consequent syndromes [1,7,9-11]. Due to the severe venous occlusion-induced disturbances, the high portal and caval hypertension and aortal hypotension, arterial



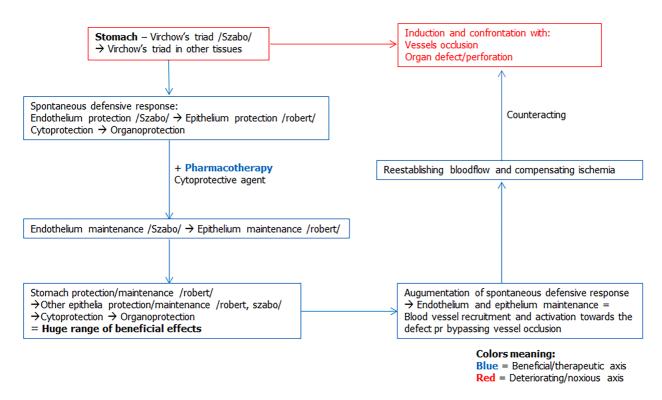


Figure 1 Summarizing the essential epithelium and endothelium protection interplay known in Robert's and Szabo's cytoprotection concept, and the role of the stable pentadecapeptide BPC 157 as a likely mediator, we suggest that BPC 157 may be a useful cytoprotective therapy. Hopefully, it may finally realize in the practice the huge theoretical importance of all aspects of the cytoprotection concept. Conceptually, there is a new point, namely, endothelium maintenance to epithelium maintenance (the recruitment of collateral blood vessels to compensate for vessel occlusion and reestablish blood flow or bypass the occluded or ruptured vessel). BPC 157 counteracts various venous occlusion-induced syndromes, inferior caval vein syndrome, ischemia-reperfusion injury following the Pringle maneuver, and Budd-Chiari syndrome in rats. Activation of the alternative collateral pathways to bypass occlusion, and reestablishing alternative blood flow, result in the counteraction of the full consequent perilous syndromes.

> and venous thrombosis, both peripherally and centrally, and various organs lesions (*i.e.*, gastrointestinal, liver, kidney, heart, and brain) were all attenuated and/or eliminated[1,7,9-11]. Furthermore, this particular beneficial effect may be competing with the Virchow's triad situation that is commonly presented [i.e., duodenal venous congestion lesions, perforated cecum, ischemic/reperfusion colitis, bile duct ligationinduced liver cirrhosis and portal hypertension, portal triad temporary occlusion (ischemia-reperfusion injury following the Pringle maneuver), and suprahepatic occlusion of the inferior caval vein (Budd-Chiari-syndrome)][1,7,9-11,150].

CONCLUSION

BPC 157 may be a useful cytoprotective therapy, which may finally result in the huge theoretical to practical importance of all aspects of the cytoprotection concept[1,7,9-11].

REFERENCES

- Sikiric P. Hahm KB. Blagaic AB. Tyrdeic A. Pavlov KH. Petrovic A. Kokot A. Goikovic S. Krezic I, Drmic D, Rucman R, Seiwerth S. Stable gastric pentadecapeptide BPC 157, Robert's stomach cytoprotection/adaptive cytoprotection/organoprotection, and Selye's stress coping response: Progress, achievements, and the future. Gut Liver 2020; 14: 153-167 [PMID: 31158953 DOI: 10.5009/gnl18490]
- Sikiric P, Seiwerth S, Rucman R, Kolenc D, Vuletic LB, Drmic D, Grgic T, Strbe S, Zukanovic G, Crvenkovic D, Madzarac G, Rukavina I, Sucic M, Baric M, Starcevic N, Krstonijevic Z, Bencic ML, Filipcic I, Rokotov DS, Vlainic J. Brain-gut axis and pentadecapeptide BPC 157: Theoretical and practical implications. Curr Neuropharmacol 2016; 14: 857-865 [PMID: 27138887 DOI: 10.2174/1570159x13666160502153022]
- 3 Seiwerth S, Milavic M, Vukojevic J, Gojkovic S, Krezic I, Vuletic LB, Pavlov KH, Petrovic A, Sikiric S, Vranes H, Prtoric A, Zizek H, Durasin T, Dobric I, Staresinic M, Strbe S, Knezevic M, Sola M, Kokot A, Sever M, Lovric E, Skrtic A, Blagaic AB, Sikiric P. Stable gastric



pentadecapeptide BPC 157 and wound healing. Front Pharmacol 2021; 12: 627533 [PMID: 34267654 DOI: 10.3389/fphar.2021.627533]

- 4 Gwyer D, Wragg NM, Wilson SL. Gastric pentadecapeptide body protection compound BPC 157 and its role in accelerating musculoskeletal soft tissue healing. Cell Tissue Res 2019; 377: 153-159 [PMID: 30915550 DOI: 10.1007/s00441-019-03016-8]
- 5 Kang EA, Han YM, An JM, Park YJ, Sikiric P, Kim DH, Kwon KA, Kim YJ, Yang D, Tchah H, Hahm KB. BPC157 as potential agent rescuing from cancer cachexia. Curr Pharm Des 2018; 24: 1947-1956 [PMID: 29898649 DOI: 10.2174/1381612824666180614082950]
- Park JM, Lee HJ, Sikiric P, Hahm KB. BPC 157 rescued NSAID-cytotoxicity via stabilizing 6 intestinal permeability and enhancing cytoprotection. Curr Pharm Des 2020; 26: 2971-2981 [PMID: 32445447 DOI: 10.2174/1381612826666200523180301]
- Sikiric P, Rucman R, Turkovic B, Sever M, Klicek R, Radic B, Drmic D, Stupnisek M, Misic M, Vuletic LB, Pavlov KH, Barisic I, Kokot A, Peklic M, Strbe S, Blagaic AB, Tvrdeic A, Rokotov DS, Vrcic H, Staresinic M, Seiwerth S. Novel cytoprotective mediator, stable gastric pentadecapeptide BPC 157. Vascular recruitment and gastrointestinal tract healing. Curr Pharm Des 2018; 24: 1990-2001 [PMID: 29879879 DOI: 10.2174/1381612824666180608101119]
- Wood JD. The first nobel prize for integrated systems physiology: Ivan Petrovich Pavlov, 1904. Physiology (Bethesda) 2004; 19: 326-330 [PMID: 15546849 DOI: 10.1152/physiol.00034.2004]
- 9 Vukojević J, Siroglavić M, Kašnik K, Kralj T, Stanćić D, Kokot A, Kolarić D, Drmić D, Sever AZ, Barišić I, Šuran J, Bojić D, Patrlj MH, Sjekavica I, Pavlov KH, Vidović T, Vlainić J, Stupnišek M, Seiwerth S, Sikirić P. Rat inferior caval vein (ICV) ligature and particular new insights with the stable gastric pentadecapeptide BPC 157. Vascul Pharmacol 2018; 106: 54-66 [PMID: 29510201 DOI: 10.1016/j.vph.2018.02.010]
- Kolovrat M, Gojkovic S, Krezic I, Malekinusic D, Vrdoljak B, Kasnik Kovac K, Kralj T, Drmic D, 10 Barisic I, Horvat Pavlov K, Petrovic A, Duzel A, Knezevic M, Mirkovic I, Kokot A, Boban Blagaic A, Seiwerth S, Sikiric P. Pentadecapeptide BPC 157 resolves Pringle maneuver in rats, both ischemia and reperfusion. World J Hepatol 2020; 12: 184-206 [PMID: 32547687 DOI: 10.4254/wjh.v12.i5.184]
- 11 Gojkovic S, Krezic I, Vrdoljak B, Malekinusic D, Barisic I, Petrovic A, Horvat Pavlov K, Kolovrat M, Duzel A, Knezevic M, Kasnik Kovac K, Drmic D, Batelja Vuletic L, Kokot A, Boban Blagaic A, Seiwerth S, Sikiric P. Pentadecapeptide BPC 157 resolves suprahepatic occlusion of the inferior caval vein, Budd-Chiari syndrome model in rats. World J Gastrointest Pathophysiol 2020; 11: 1-19 [PMID: 32226643 DOI: 10.4291/wjgp.v11.i1.1]
- 12 Veljaca M, Chan K, Guglietta A. Digestion of h-EGF, h-TGF alpha, and BPC-15 in human gastric juice. Pharmacol Res 1995; 31: 70 [DOI: 10.1016/1043-6618(95)86539-X]
- 13 Szabo S, Khomenko T, Gombos Z, Deng XM, Jadus MR, Yoshida M. Review article: transcription factors and growth factors in ulcer healing. Aliment Pharmacol Ther 2000; 14 Suppl 1: 33-43 [PMID: 10807401 DOI: 10.1046/j.1365-2036.2000.014s1033.x]
- 14 Robert A. Current history of cytoprotection. Prostaglandins 1981; 21 Suppl: 89-96 [PMID: 7029653 DOI: 10.1016/0090-6980(81)90123-4]
- 15 Robert A. Prostaglandins: effects on the gastrointestinal tract. Clin Physiol Biochem 1984; 2: 61-69 [PMID: 6386280]
- 16 Robert A. Cytoprotection and prostaglandins. Klin Wochenschr 1986; 64 Suppl 7: 40-43 [PMID: 3560780]
- 17 Robert A, Nezamis JE, Lancaster C, Davis JP, Field SO, Hanchar AJ. Mild irritants prevent gastric necrosis through "adaptive cytoprotection" mediated by prostaglandins. Am J Physiol 1983; 245: G113-G121 [PMID: 6869543 DOI: 10.1152/ajpgi.1983.245.1.G113]
- 18 Szabo S, Usadel KH. Cytoprotection - organoprotection by somatostatin: gastric and hepatic lesions. Experientia 1982; 38: 254-256 [PMID: 6120852 DOI: 10.1007/BF01945097]
- Szabo S. Experimental basis for a role for sulfhydryls and dopamine in ulcerogenesis: a primer for 19 cytoprotection-organoprotection. Klin Wochenschr 1986; 64 Suppl 7: 116-122 [PMID: 3560772]
- 20 Szabo S, Trier JS. Pathogenesis of acute gastric mucosal injury: sulfhydrils as a protector, adrenal cortex as modulator, and vascular endothelium as a target. In: Allen A, Flemstrom G, Garner A, Silen W, Turnberg A, eds. Mechanism of mucosal protection in the upper gastrointestinal tract. New York Raven Press, 1984: 287-293
- 21 Szabó S. Role of sulfhydryls and early vascular lesions in gastric mucosal injury. Acta Physiol Hung 1984; 64: 203-214 [PMID: 6532115]
- 22 Tarnawski A, Stachura J, Hollander D, Sarfeh IJ, Bogdal J. Cellular aspects of alcohol-induced injury and prostaglandin protection of the human gastric mucosa. Focus on the mucosal microvessels. J Clin Gastroenterol 1988; 10 Suppl 1: S35-S45 [PMID: 3183341 DOI: 10.1097/00004836-198812001-00008]
- 23 Szabo S. "Gastric cytoprotection" is still relevant. J Gastroenterol Hepatol 2014; 29 Suppl 4: 124-132 [PMID: 25521744 DOI: 10.1111/jgh.12735]
- 24 Wallace JL, Ianaro A, de Nucci G. Gaseous Mediators in Gastrointestinal Mucosal Defense and Injury. Dig Dis Sci 2017; 62: 2223-2230 [PMID: 28733867 DOI: 10.1007/s10620-017-4681-0]
- Vandiver M, Snyder SH. Hydrogen sulfide: a gasotransmitter of clinical relevance. J Mol Med 25 (Berl) 2012; 90: 255-263 [PMID: 22314625 DOI: 10.1007/s00109-012-0873-4]
- 26 Brzozowski T, Konturek PC, Pajdo R, Ptak-Belowska A, Kwiecien S, Pawlik M, Drozdowicz D, Sliwowski Z, Brzozowski B, Konturek SJ, Pawlik WW. Physiological mediators in nonsteroidal



anti-inflammatory drugs (NSAIDs)-induced impairment of gastric mucosal defense and adaptation. Focus on nitric oxide and lipoxins. J Physiol Pharmacol 2008; 59 Suppl 2: 89-102 [PMID: 18812631]

- 27 Szabo S, Trier JS, Frankel PW. Sulfhydryl compounds may mediate gastric cytoprotection. Science 1981; 214: 200-202 [PMID: 7280691 DOI: 10.1126/science.7280691]
- Konturek SJ, Brzozowski T, Piastucki I, Radecki T, Szabo S. Gastric cytoprotection by agents 28 altering gastric mucosal sulfhydryl compounds: role of endogenous prostaglandins. Adv Prostaglandin Thromboxane Leukot Res 1983; 12: 411-416 [PMID: 6221624]
- 29 Takeuchi K, Kato S, Amagase K. Prostaglandin EP receptors involved in modulating gastrointestinal mucosal integrity. J Pharmacol Sci 2010; 114: 248-261 [PMID: 21041985 DOI: 10.1254/jphs.10r06cr
- 30 Takeuchi K. Gastric cytoprotection by prostaglandin E2 and prostacyclin: relationship to EP1 and IP receptors. J Physiol Pharmacol 2014; 65: 3-14 [PMID: 24622825]
- 31 Szabo S, Pihan G. Mechanisms of gastric cytoprotection. J Clin Gastroenterol 1987; 9 Suppl 1: 8-13 [PMID: 3302011 DOI: 10.1097/00004836-198709011-00003]
- 32 Szabo S, Vattay P, Scarbrough E, Folkman J. Role of vascular factors, including angiogenesis, in the mechanisms of action of sucralfate. Am J Med 1991; 91: 158S-160S [PMID: 1715670 DOI: 10.1016/0002-9343(91)90469-e]
- deFoneska A, Kaunitz JD. Gastroduodenal mucosal defense. Curr Opin Gastroenterol 2010; 26: 33 604-610 [PMID: 20948371 DOI: 10.1097/MOG.0b013e32833f1222]
- 34 Brzozowski T, Ptak-Belowska A, Kwiecien S, Krzysiek-Maczka G, Strzalka M, Drozdowicz D, Pajdo R, Olszanecki R, Korbut R, Konturek SJ, Pawlik WW. Novel concept in the mechanism of injury and protection of gastric mucosa: role of renin-angiotensin system and active metabolites of angiotensin. Curr Med Chem 2012; 19: 55-62 [PMID: 22300076 DOI: 10.2174/092986712803413953
- Ray A, Gulati K, Puri S, Sen P. Role of kappa opioid receptors during stress responsiveness in rats. 35 Indian J Exp Biol 1993; 31: 116-119 [PMID: 8388852]
- Rónai AZ, Gyires K, Barna I, Müllner K, Palkovits M. Neonatal monosodium glutamate treatment 36 abolishes both delta opioid receptor-induced and alpha-2 adrenoceptor-mediated gastroprotection in the lower brainstem in rats. J Physiol Paris 2001; 95: 215-220 [PMID: 11595440 DOI: 10.1016/s0928-4257(01)00028-6
- Filaretova LP. Contribution of glucocorticoid hormones to gastroprotection. Usp Fiziol Nauk 2014; 37 45: 44-56 [PMID: 25702452]
- 38 Filaretova LP, Podvigina TT, Bobryshev PY, Bagaeva TR, Tanaka A, Takeuchi K. Hypothalamicpituitary-adrenocortical axis: the hidden gold in gastric mucosal homeostasis. Inflammopharmacology 2006; 14: 207-213 [PMID: 17093902 DOI: 10.1007/s10787-006-1544-2]
- Hernandez DE, Adcock JW, Nemeroff CB, Prange AJ Jr. The role of the adrenal gland in 39 cytoprotection against stress-induced gastric ulcers in rats. J Neurosci Res 1984; 11: 193-201 [PMID: 6708137 DOI: 10.1002/jnr.490110209]
- Kaneko H, Taché Y, Kusugami K. Importance of medullary thyrotropin-releasing hormone in brain-40 gut circuits regulating gastric integrity: preclinical studies. J Gastroenterol 2002; 37 Suppl 14: 128-132 [PMID: 12572880 DOI: 10.1007/BF03326431]
- 41 Taché Y, Yoneda M. Central action of TRH to induce vagally mediated gastric cytoprotection and ulcer formation in rats. J Clin Gastroenterol 1993; 17 Suppl 1: S58-S63 [PMID: 8283016 DOI: 10.1097/00004836-199312001-00013]
- 42 Hernandez DE, Arredondo ME, Xue BG. Imipramine prevents gastric lesions induced by centrally administered thyrotropin-releasing hormone (TRH) in rats. Neurosci Lett 1990; 111: 339-343 [PMID: 2159606 DOI: 10.1016/0304-3940(90)90285-h]
- 43 Király A, Sütó G, Guth PH, Taché Y. Mechanisms mediating gastric hyperemic and acid responses to central TRH analog at a cytoprotective dose. Am J Physiol 1997; 273: G31-G38 [PMID: 9252506 DOI: 10.1152/ajpgi.1997.273.1.G31]
- 44 Mózsik G. Capsaicin as new orally applicable gastroprotective and therapeutic drug alone or in combination with nonsteroidal anti-inflammatory drugs in healthy human subjects and in patients. Prog Drug Res 2014; 68: 209-258 [PMID: 24941671 DOI: 10.1007/978-3-0348-0828-6 9]
- 45 Holzer P. Peptidergic sensory neurons in the control of vascular functions: mechanisms and significance in the cutaneous and splanchnic vascular beds. Rev Physiol Biochem Pharmacol 1992; 121: 49-146 [PMID: 1485073 DOI: 10.1007/BFb0033194]
- 46 Holzer P. Capsaicin: cellular targets, mechanisms of action, and selectivity for thin sensory neurons. Pharmacol Rev 1991; 43: 143-201 [PMID: 1852779]
- Glavin GB, Szabo S. Experimental gastric mucosal injury: laboratory models reveal mechanisms of 47 pathogenesis and new therapeutic strategies. FASEB J 1992; 6: 825-831 [PMID: 1740232 DOI: 10.1096/fasebj.6.3.1740232]
- Szabo S, Vattay P. Experimental gastric and duodenal ulcers. Advances in pathogenesis. 48 Gastroenterol Clin North Am 1990; 19: 67-85 [PMID: 2184131]
- 49 Henke PG. Recent studies of the central nucleus of the amygdala and stress ulcers. Neurosci Biobehav Rev 1988; 12: 143-150 [PMID: 2902539 DOI: 10.1016/s0149-7634(88)80006-x]
- Sikirić P, Rotkvić I, Mise S, Krizanac S, Gjuris V, Jukić J, Suchanek E, Petek M, Udovicić I, 50 Kalogjera L. The influence of dopamine agonists and antagonists on indomethacin lesions in stomach and small intestine in rats. Eur J Pharmacol 1988; 158: 61-67 [PMID: 2906010 DOI:



10.1016/0014-2999(88)90253-11

- 51 Hernandez DE. Involvement of dopamine receptors in experimental ulceration. Int J Tissue React 1987; 9: 407-411 [PMID: 3667110]
- 52 Diel F, Szabo S. Dose-dependent effects of linear and cyclic somatostatin on ethanol-induced gastric erosions: the role of mast cells and increased vascular permeability in the rat. Regul Pept 1986; 13: 235-243 [PMID: 2871590 DOI: 10.1016/0167-0115(86)90042-x]
- 53 Konturek SJ. Role of growth factors in gastroduodenal protection and healing of peptic ulcers. Gastroenterol Clin North Am 1990; 19: 41-65 [PMID: 1970337]
- 54 Brzozowski T, Konturek SJ, Sliwowski Z, Pajdo R, Drozdowicz D, Stachura J. Role of betaadrenoceptors in gastric mucosal integrity and gastroprotection induced by epidermal growth factor. Digestion 1997; 58: 319-331 [PMID: 9324159 DOI: 10.1159/000201462]
- 55 Brzozowski T. Gastro-protection in vivo and in vitro. Patol Pol 1992; 43: 1-9 [PMID: 1296166]
- 56 West SD, Mercer DW. Bombesin-induced gastroprotection. Ann Surg 2005; 241: 227-231 [PMID: 15650631 DOI: 10.1097/01.sla.0000151790.14274.5d]
- Peeters TL. Ghrelin and the gut. Endocr Dev 2013; 25: 41-48 [PMID: 23652390 DOI: 57 10.1159/0003460511
- Lewin MJ, Bado A. Gastric leptin. Microsc Res Tech 2001; 53: 372-376 [PMID: 11376498 DOI: 58 10.1002/jemt.1105]
- Brzozowski T, Konturek PC, Konturek SJ, Pajdo R, Drozdowicz D, Kwiecień S, Hahn EG. 59 Acceleration of ulcer healing by cholecystokinin (CCK): role of CCK-A receptors, somatostatin, nitric oxide and sensory nerves. Regul Pept 1999; 82: 19-33 [PMID: 10458643 DOI: 10.1016/s0167-0115(99)00029-4
- 60 Brzozowski T, Konturek PC, Konturek SJ, Pajdo R, Bielanski W, Brzozowska I, Stachura J, Hahn EG. The role of melatonin and L-tryptophan in prevention of acute gastric lesions induced by stress, ethanol, ischemia, and aspirin. J Pineal Res 1997; 23: 79-89 [PMID: 9392446 DOI: 10.1111/j.1600-079x.1997.tb00339.x]
- Hernandez DE, Stanley DA, Melvin JA, Prange AJ Jr. Role of brain neurotransmitters on 61 neurotensin-induced gastric cytoprotection. Pharmacol Biochem Behav 1985; 22: 509-513 [PMID: 2859609 DOI: 10.1016/0091-3057(85)90266-7]
- 62 Florkiewicz RZ, Ahluwalia A, Sandor Z, Szabo S, Tarnawski AS. Gastric mucosal injury activates bFGF gene expression and triggers preferential translation of high molecular weight bFGF isoforms through CUG-initiated, non-canonical codons. Biochem Biophys Res Commun 2011; 409: 494-499 [PMID: 21600881 DOI: 10.1016/j.bbrc.2011.05.033]
- 63 Folkman J, Szabo S, Stovroff M, McNeil P, Li W, Shing Y. Duodenal ulcer. Discovery of a new mechanism and development of angiogenic therapy that accelerates healing. Ann Surg 1991; 214: 414-425; discussion 426 [PMID: 1719945 DOI: 10.1097/00000658-199110000-00006]
- 64 Zádori ZS, Tóth VE, Fehér Á, Philipp K, Németh J, Gyires K. Evidence for the gastric cytoprotective effect of centrally injected agmatine. Brain Res Bull 2014; 108: 51-59 [PMID: 25171957 DOI: 10.1016/j.brainresbull.2014.07.008]
- Takeuchi K, Nagahama K. Animal model of acid-reflux esophagitis: pathogenic roles of 65 acid/pepsin, prostaglandins, and amino acids. Biomed Res Int 2014; 2014: 532594 [PMID: 24672789 DOI: 10.1155/2014/532594]
- 66 Ichikawa T, Hotta K, Ishihara K. Second-generation histamine H(2)-receptor antagonists with gastric mucosal defensive properties. Mini Rev Med Chem 2009; 9: 581-589 [PMID: 19456288 DOI: 10.2174/138955709788167646
- 67 Chang M, Xue J, Sharma V, Habtezion A. Protective role of hemeoxygenase-1 in gastrointestinal diseases. Cell Mol Life Sci 2015; 72: 1161-1173 [PMID: 25428780 DOI: 10.1007/s00018-014-1790-1]
- 68 Ryter SW. Therapeutic potential of heme oxygenase-1 and carbon monoxide in acute organ Injury, critical illness, and inflammatory disorders. Antioxidants (Basel) 2020; 9 [PMID: 33228260 DOI: 10.3390/antiox9111153]
- 69 Na HK, Lee JY. Molecular basis of alcohol-related gastric and colon cancer. Int J Mol Sci 2017; 18 [PMID: 28538665 DOI: 10.3390/ijms18061116]
- 70 Mózsik G. Gastric cytoprotection 30 years after its discovery by André Robert: a personal perspective. Inflammopharmacology 2010; 18: 209-221 [PMID: 20596896 DOI: 10.1007/s10787-010-0045-5
- Jabůrek M, Průchová P, Holendová B, Galkin A, Ježek P. Antioxidant synergy of mitochondrial 71 phospholipase PNPLA8/iPLA2y with fatty acid-conducting SLC25 gene family transporters. Antioxidants (Basel) 2021; 10 [PMID: 33926059 DOI: 10.3390/antiox10050678]
- 72 Li Y, Wu W, Liu W, Zhou M. Roles and mechanisms of renalase in cardiovascular disease: A promising therapeutic target. Biomed Pharmacother 2020; 131: 110712 [PMID: 32916539 DOI: 10.1016/j.biopha.2020.110712
- 73 Detsika MG, Lianos EA. Regulation of complement activation by heme oxygenase-1 (HO-1) in kidney injury. Antioxidants (Basel) 2021; 10 [PMID: 33418934 DOI: 10.3390/antiox10010060]
- Dimitrova-Shumkovska J, Krstanoski L, Veenman L. Potential beneficial actions of fucoidan in 74 brain and liver injury, disease, and intoxication-potential implication of sirtuins. Mar Drugs 2020; 18 [PMID: 32380741 DOI: 10.3390/md18050242]
- 75 Jabbehdari S, Handa JT. Oxidative stress as a therapeutic target for the prevention and treatment of early age-related macular degeneration. Surv Ophthalmol 2021; 66: 423-440 [PMID: 32961209 DOI:



10.1016/j.survophthal.2020.09.002]

- 76 auf dem Keller U, Kümin A, Braun S, Werner S. Reactive oxygen species and their detoxification in healing skin wounds. J Investig Dermatol Symp Proc 2006; 11: 106-111 [PMID: 17069017 DOI: 10.1038/sj.jidsymp.5650001]
- 77 Chen XD, Tan JL, Feng Y, Huang LJ, Zhang M, Cheng B. Autophagy in fate determination of mesenchymal stem cells and bone remodeling. World J Stem Cells 2020; 12: 776-786 [PMID: 32952858 DOI: 10.4252/wjsc.v12.i8.776]
- Szewczyk A, Bednarczyk P, Jędraszko J, Kampa RP, Koprowski P, Krajewska M, Kucman S, 78 Kulawiak B, Laskowski M, Rotko D, Sęk A, Walewska A, Żochowska M, Wrzosek A. Mitochondrial potassium channels - an overview. Postepy Biochem 2018; 64: 196-212 [PMID: 30656905 DOI: 10.18388/pb.2018 132]
- Robert A, Bundy GL, Field SO, Nezamis JE, Davis JP, Hanchar AJ, Lancaster C, Ruwart MJ. 79 Prevention of cecitis in hamsters by certain prostaglandins. Prostaglandins 1985; 29: 961-980 [PMID: 3898232 DOI: 10.1016/0090-6980(85)90221-7]
- Elliott G, Whited BA, Purmalis A, Davis JP, Field SO, Lancaster C, Robert A. Effect of 16,16-80 dimethyl PGE2 on renal papillary necrosis and gastrointestinal ulcerations (gastric, duodenal, intestinal) produced in rats by mefenamic acid. Life Sci 1986; 39: 423-432 [PMID: 3736334 DOI: 10.1016/0024-3205(86)90522-9
- 81 Robert A, Lum JT, Lancaster C, Olafsson AS, Kolbasa KP, Nezamis JE. Prevention by prostaglandins of caerulein-induced pancreatitis in rats. Lab Invest 1989; 60: 677-691 [PMID: 24698591
- 82 Sato Y, Yoneda M, Nakamura K, Makino I, Terano A. Protective effect of central thyrotropinreleasing hormone on carbon tetrachloride-induced acute hepatocellular necrosis in rats. J Hepatol 2003; **39**: 47-54 [PMID: 12821043 DOI: 10.1016/s0168-8278(03)00146-6]
- 83 Gyires K. Neuroinflammatory reactions in experimental gastric ulcer: target for mucosal protection. Inflammopharmacology 1997; 5: 383-395 [PMID: 17657616 DOI: 10.1007/s10787-997-0034-5]
- 84 Gyires K. Are all "cytoprotective" drugs gastroprotective? Acta Physiol Hung 1992; 80: 247-255 [PMID: 1345194]
- 85 Selye H, Szabo S. Experimental model for production of perforating duodenal ulcers by cysteamine in the rat. Nature 1973; 244: 458-459 [PMID: 4582506 DOI: 10.1038/244458a0]
- Szabo S. Dopamine disorder in duodenal ulceration. Lancet 1979; 2: 880-882 [PMID: 90970 DOI: 86 10.1016/s0140-6736(79)92690-4
- 87 Klicek R, Kolenc D, Suran J, Drmic D, Brcic L, Aralica G, Sever M, Holjevac J, Radic B, Turudic T, Kokot A, Patrlj L, Rucman R, Seiwerth S, Sikiric P. Stable gastric pentadecapeptide BPC 157 heals cysteamine-colitis and colon-colon-anastomosis and counteracts cuprizone brain injuries and motor disability. J Physiol Pharmacol 2013; 64: 597-612 [PMID: 24304574 DOI: 10.1002/cphy.c120035]
- Selye H. A syndrome produced by diverse nocuous agents. Nature 1936; 138: 32 88
- Masson G, Selye H. Réaction générale d'adaptation: Ses indications pratiques. Can J Comp Med 89 1938; 2: 282-285 [PMID: 17647461]
- Selye H. Rheumatic diseases as diseases of adaptation. Br Med J 1950; 1: 1362-1364 [PMID: 90 15420482 DOI: 10.1136/bmj.1.4666.1362]
- Ward LE, Polley HF, Slocumb CH, Hench PS. Cortisone in treatment of rheumatoid arthritis. J Am 91 Med Assoc 1953; 152: 119-126 [PMID: 13034543 DOI: 10.1001/jama.1953.03690020011003]
- 92 Mason JW. A historical view of the stress field. J Human Stress 1975; 1: 6-12 contd [PMID: 798012 DOI: 10.1080/0097840X.1975.9940399]
- 93 Mason JW. A historical view of the stress field. J Human Stress 1975; 1: 22-36 concl [PMID: 798013 DOI: 10.1080/0097840X.1975.9940405]
- Selye H. Production of nephrosclerosis by overdosage with desoxycorticosterone acetate. Can Med 94 Assoc J 1942; 47: 515-519 [PMID: 20322632]
- 95 Ahlquist RP. A study of the adrenotropic receptors. Am J Physiol 1948; 153: 586-600 [PMID: 18882199 DOI: 10.1152/ajplegacy.1948.153.3.586]
- Cannon WB. The adrenal medulla. Bull N Y Acad Med 1940; 16: 3-13 [PMID: 19312138] 96
- Black JW, Stephenson JS. Pharmacology of a new adrenergic beta-receptor-blocking compound 97 (Nethalide). Lancet 1962; 2: 311-314 [PMID: 13869657 DOI: 10.1016/s0140-6736(62)90103-4]
- 98 Fumagalli C, Maurizi N, Marchionni N, Fornasari D. β-blockers: Their new life from hypertension to cancer and migraine. Pharmacol Res 2020; 151: 104587 [PMID: 31809852 DOI: 10.1016/j.phrs.2019.104587]
- 99 Szabo S, Bynum TE. Alternatives to the acid-oriented approach to ulcer disease: does 'cytoprotection' exist in man? Scand J Gastroenterol 1988; 23: 1-6 [PMID: 3278362 DOI: 10.3109/00365528809093839
- Szabó S. Critical and timely review of the concept of gastric cytoprotection. Acta Physiol Hung 100 1989; 73: 115-127 [PMID: 2688357]
- Herszényi L, Bakucz T, Barabás L, Tulassay Z. Pharmacological approach to gastric acid 101 suppression: Past, present, and future. Dig Dis 2020; 38: 104-111 [PMID: 31846972 DOI: 10.1159/000505204]
- 102 Tkalcević VI, Cuzić S, Brajsa K, Mildner B, Bokulić A, Situm K, Perović D, Glojnarić I, Parnham MJ. Enhancement by PL 14736 of granulation and collagen organization in healing wounds and the potential role of egr-1 expression. Eur J Pharmacol 2007; 570: 212-221 [PMID: 17628536 DOI:



10.1016/i.eiphar.2007.05.072]

- 103 Chang CH, Tsai WC, Hsu YH, Pang JH. Pentadecapeptide BPC 157 enhances the growth hormone receptor expression in tendon fibroblasts. Molecules 2014; 19: 19066-19077 [PMID: 25415472 DOI: 10.3390/molecules191119066
- Chang CH, Tsai WC, Lin MS, Hsu YH, Pang JH. The promoting effect of pentadecapeptide BPC 104 157 on tendon healing involves tendon outgrowth, cell survival, and cell migration. J Appl Physiol (1985) 2011; 110: 774-780 [PMID: 21030672 DOI: 10.1152/japplphysiol.00945.2010]
- Huang T, Zhang K, Sun L, Xue X, Zhang C, Shu Z, Mu N, Gu J, Zhang W, Wang Y, Zhang Y. 105 Body protective compound-157 enhances alkali-burn wound healing in vivo and promotes proliferation, migration, and angiogenesis in vitro. Drug Des Devel Ther 2015; 9: 2485-2499 [PMID: 25995620 DOI: 10.2147/DDDT.S82030]
- Hsieh MJ, Lee CH, Chueh HY, Chang GJ, Huang HY, Lin Y, Pang JS. Modulatory effects of BPC 106 157 on vasomotor tone and the activation of Src-Caveolin-1-endothelial nitric oxide synthase pathway. Sci Rep 2020; 10: 17078 [PMID: 33051481 DOI: 10.1038/s41598-020-74022-y]
- 107 Hsieh MJ, Liu HT, Wang CN, Huang HY, Lin Y, Ko YS, Wang JS, Chang VH, Pang JS. Therapeutic potential of pro-angiogenic BPC157 is associated with VEGFR2 activation and upregulation. J Mol Med (Berl) 2017; 95: 323-333 [PMID: 27847966 DOI: 10.1007/s00109-016-1488-y
- 108 Sandor Z, Vince A, Szabo S. The protective effect of a recently isolated peptide PL-10 in acute and chronic gastric injury. FASEB J 1996; 10: 171
- 109 Bódis B, Karádi O, Nagy L, Dohoczky C, Kolega M, Mózsik G. Direct cellular effects of some mediators, hormones and growth factor-like agents on denervated (isolated) rat gastric mucosal cells. J Physiol Paris 1997; 91: 183-187 [PMID: 9403792 DOI: 10.1016/s0928-4257(97)89482-x]
- 110 Bódis B, Karádi O, Németh P, Dohoczky C, Kolega M, Mózsik G. Evidence for direct cellular protective effect of PL-10 substances (synthesized parts of body protection compound, BPC) and their specificity to gastric mucosal cells. Life Sci 1997; 61: PL 243-PL 248 [PMID: 9353174 DOI: 10.1016/s0024-3205(97)00744-3
- 111 Veljaca M, Lesch CA, Pllana R, Sanchez B, Chan K, Guglietta A. BPC-15 reduces trinitrobenzene sulfonic acid-induced colonic damage in rats. J Pharmacol Exp Ther 1995; 272: 417-422 [PMID: 7815358]
- 112 Khomenko T, Szabo S, Deng XM, Sandor Z, Gombos Z, Yoshida M. Cell proliferation, transcription factor Egr-1 and growth factors in experimental ulcerative colitis after treatment with PL 14736: In vitro and in vivo studies. Gastroenterology 2003; 124: 493 [DOI: 10.1016/S0016-5085(03)82495-2]
- 113 Sandor ZS, Vincze A, Jadus MR, Dohoczky C, Erceg D, Szabo S. The protective effect of newly isolated peptide PL-10 in the iodoacetamide colitis model in rats. Gastroenterology 1997; 112: 400
- Wu H, Wei M, Li N, Lu Q, Shrestha SM, Tan J, Zhang Z, Wu G, Shi R. Clopidogrel-induced gastric 114 injury in rats is attenuated by stable gastric pentadecapeptide BPC 157. Drug Des Devel Ther 2020; 14: 5599-5610 [PMID: 33376304 DOI: 10.2147/DDDT.S284163]
- 115 Okabe S, Amagase K. An overview of acetic acid ulcer models--the history and state of the art of peptic ulcer research. Biol Pharm Bull 2005; 28: 1321-1341 [PMID: 16079471 DOI: 10.1248/bpb.28.1321]
- 116 Okabe S, Roth JL, Pfeiffer CJ. A method for experimental, penetrating gastric and duodenal ulcers in rats. Observations on normal healing. Am J Dig Dis 1971; 16: 277-284 [PMID: 5554507 DOI: 10.1007/BF02235252
- 117 Glavin GB, Paré WP, Sandbak T, Bakke HK, Murison R. Restraint stress in biomedical research: an update. Neurosci Biobehav Rev 1994; 18: 223-249 [PMID: 8058215 DOI: 10.1016/0149-7634(94)90027-21
- 118 Glavin GB, Murison R, Overmier JB, Pare WP, Bakke HK, Henke PG, Hernandez DE. The neurobiology of stress ulcers. Brain Res Brain Res Rev 1991; 16: 301-343 [PMID: 1790434 DOI: 10.1016/0165-0173(91)90012-w]
- 119 Paré WP, Glavin GB. Restraint stress in biomedical research: a review. Neurosci Biobehav Rev 1986; 10: 339-370 [PMID: 3095718 DOI: 10.1016/0149-7634(86)90017-5]
- 120 Overmier JB, Murison R, Milde AM. Sensitization and conditioning as contributors to gastrointestinal vulnerability. Auton Neurosci 2006; 125: 22-27 [PMID: 16476574 DOI: 10.1016/j.autneu.2006.01.011]
- 121 Zhao DQ, Xue H, Sun HJ. Nervous mechanisms of restraint water-immersion stress-induced gastric mucosal lesion. World J Gastroenterol 2020; 26: 2533-2549 [PMID: 32523309 DOI: 10.3748/wjg.v26.i20.2533
- Xue XC, Wu YJ, Gao MT, Li WG, Zhao N, Wang ZL, Bao CJ, Yan Z, Zhang YQ. Protective effects 122 of pentadecapeptide BPC 157 on gastric ulcer in rats. World J Gastroenterol 2004; 10: 1032-1036 [PMID: 15052688 DOI: 10.3748/wjg.v10.i7.1032]
- 123 Erceg D, Simicevic VN, Kolega M, Dohoczky C. Some aspects of the effects of PL-10.1.AK-15 on the gastrointestinal tract. J Physiol Paris 1997; 91: 179-181 [PMID: 9403791 DOI: 10.1016/s0928-4257(97)89481-8
- 124 Szabo S, Pihan G. Development and significance of cysteamine and propionitrile models of duodenal ulcer. Chronobiol Int 1987; 4: 31-42 [PMID: 3315259 DOI: 10.1080/07420528709078506
- 125 Szabo S, Pihan G, Gallagher GT, Brown A. Role of local secretory and motility changes in the pathogenesis of experimental duodenal ulcer. Scand J Gastroenterol Suppl 1984; 92: 106-111



[PMID: 6588493]

- Gallagher G, Brown A, Szabo S. Effect of dopamine-related drugs on duodenal ulcer induced by 126 cysteamine or propionitrile: prevention and aggravation may not be mediated by gastrointestinal secretory changes in the rat. J Pharmacol Exp Ther 1987; 240: 883-889 [PMID: 3559980 DOI: 10.1016/0160-5402(87)90040-4]
- Szabo S, Haith LR Jr, Reynolds ES. Pathogenesis of duodenal ulceration produced by cysteamine or 127 propionitrile: influence of vagotomy, sympathectomy, histamine depletion, H-2 receptor antagonists and hormones. Dig Dis Sci 1979; 24: 471-477 [PMID: 37058 DOI: 10.1007/BF01299831]
- 128 Somerville KW, Langman MJ. Newer antisecretory agents for peptic ulcer. Drugs 1983; 25: 315-330 [PMID: 6133734 DOI: 10.2165/00003495-198325030-00003]
- 129 Szabo S, Cho CH. From cysteamine to MPTP: structure-activity studies with duodenal ulcerogens. Toxicol Pathol 1988; 16: 205-212 [PMID: 3055230 DOI: 10.1177/019262338801600213]
- 130 Mangla JC, Pihan G, Brown HA, Rattan S, Szabo S. Effect of duodenal ulcerogens cysteamine, mepirizole, and MPTP on duodenal myoelectric activity in rats. Dig Dis Sci 1989; 34: 537-542 [PMID: 2784758 DOI: 10.1007/BF01536329]
- 131 Szabo S, Neumeyer JL. Dopamine agonists and antagonists in duodenal ulcer disease. In: ACS Symposium Series, eds. Kaiser C, Kebabian W. American Chemical Society Publications. Washington, 1983: 175-199
- 132 Wang XY, Qu M, Duan R, Shi D, Jin L, Gao J, Wood JD, Li J, Wang GD. Cytoprotective mechanism of the novel gastric peptide BPC157 in gastrointestinal tract and cultured enteric neurons and glial cells. Neurosci Bull 2019; 35: 167-170 [PMID: 30116973 DOI: 10.1007/s12264-018-0269-8]
- 133 Breithaupt-Faloppa AC, Fantozzi ET, de Assis Ramos MM, Vitoretti LB, Couto GK, Lino-dos-Santos-Franco A, Rossoni LV, Oliveira-Filho RM, Vargaftig BB, Tavares-de-Lima W. Protective effect of estradiol on acute lung inflammation induced by an intestinal ischemic insult is dependent on nitric oxide. Shock 2013; 40: 203-209 [PMID: 23846411 DOI: 10.1097/SHK.0b013e3182a01e24]
- 134 Koike K, Moore FA, Moore EE, Poggetti RS, Tuder RM, Banerjee A. Endotoxin after gut ischemia/reperfusion causes irreversible lung injury. J Surg Res 1992; 52: 656-662 [PMID: 1326681 DOI: 10.1016/0022-4804(92)90145-p]
- Seveljević-Jaran D, Cuzić S, Dominis-Kramarić M, Glojnarić I, Ivetić V, Radosević S, Parnham 135 MJ. Accelerated healing of excisional skin wounds by PL 14736 in alloxan-hyperglycemic rats. Skin Pharmacol Physiol 2006; 19: 266-274 [PMID: 16785777 DOI: 10.1159/000093982]
- 136 Tshionyi M, Shay E, Lunde E, Lin A, Han KY, Jain S, Chang JH, Azar DT. Hemangiogenesis and lymphangiogenesis in corneal pathology. Cornea 2012; 31: 74-80 [PMID: 22030600 DOI: 10.1097/ICO.0b013e31821dd986
- Kato S, Pinto M, Carvajal A, Espinoza N, Monsó C, Bravo L, Villalon M, Cuello M, Quest AF, 137 Suenaga A, Brosens JJ, Owen GI. Tissue factor is regulated by epidermal growth factor in normal and malignant human endometrial epithelial cells. Thromb Haemost 2005; 94: 444-453 [PMID: 16113838 DOI: 10.1160/TH05-01-0066]
- 138 Sigalet DL, Martin GR. Hormonal therapy for short bowel syndrome. J Pediatr Surg 2000; 35: 360-363; discussion 364 [PMID: 10693697 DOI: 10.1016/s0022-3468(00)90041-1]
- 139 Fiore NF, Ledniczky G, Liu Q, Orazi A, Du X, Williams DA, Grosfeld JL. Comparison of interleukin-11 and epidermal growth factor on residual small intestine after massive small bowel resection. J Pediatr Surg 1998; 33: 24-29 [PMID: 9473093 DOI: 10.1016/s0022-3468(98)90354-2]
- 140 Pereira PM, Bines JE. New growth factor therapies aimed at improving intestinal adaptation in short bowel syndrome. J Gastroenterol Hepatol 2006; 21: 932-940 [PMID: 16724975 DOI: 10.1111/j.1440-1746.2006.04351.x]
- Petersen TI, Kissmeyer-Nielsen P, Flyvbjerg A, Laurberg S, Christensen H. Effect of insulin-like 141 growth factor I (IGF-I) administration on the healing of colonic anastomoses in rats. Int J Colorectal Dis 1996; 11: 19-24 [PMID: 8919336 DOI: 10.1007/BF00418850]
- 142 Seyer-Hansen M, Andreassen TT, Oxlund H. Strength of colonic anastomoses and skin incisional wounds in old rats - influence by diabetes and growth hormone. Growth Horm IGF Res 1999; 9: 254-261 [PMID: 10512691 DOI: 10.1054/ghir.1999.0116]
- 143 Kato Y, Yu D, Schwartz MZ. Enhancement of intestinal adaptation by hepatocyte growth factor. J Pediatr Surg 1998; 33: 235-239 [PMID: 9498393 DOI: 10.1016/s0022-3468(98)90438-9]
- Festuccia C, Angelucci A, Gravina GL, Biordi L, Millimaggi D, Muzi P, Vicentini C, Bologna M. 144 Epidermal growth factor modulates prostate cancer cell invasiveness regulating urokinase-type plasminogen activator activity. EGF-receptor inhibition may prevent tumor cell dissemination. Thromb Haemost 2005; 93: 964-975 [PMID: 15886816 DOI: 10.1160/TH04-09-0637]
- 145 Drucker DJ. Gut adaptation and the glucagon-like peptides. Gut 2002; 50: 428-435 [PMID: 11839727 DOI: 10.1136/gut.50.3.428]
- 146 Guth PH. Gastric blood flow in ethanol injury and prostaglandin cytoprotection. Scand J Gastroenterol Suppl 1986; 125: 86-91 [PMID: 3469743 DOI: 10.3109/00365528609093822]
- 147 Vukojević J, Vrdoljak B, Malekinušić D, Siroglavić M, Milavić M, Kolenc D, Boban Blagaić A, Batelja L, Drmić D, Seiverth S, Sikirić P. The effect of pentadecapeptide BPC 157 on hippocampal ischemia/reperfusion injuries in rats. Brain Behav 2020; 10: e01726 [PMID: 32558293 DOI: 10.1002/brb3.1726]
- Schiller HJ, Reilly PM, Bulkley GB. Tissue perfusion in critical illnesses. Antioxidant therapy. Crit 148 Care Med 1993; 21: S92-102 [PMID: 8428505 DOI: 10.1097/00003246-199302001-00016]



- 149 Rangan U, Bulkley GB. Prospects for treatment of free radical-mediated tissue injury. Br Med Bull 1993; 49: 700-718 [PMID: 8221033 DOI: 10.1093/oxfordjournals.bmb.a072641]
- 150 Sever AZ, Sever M, Vidovic T, Lojo N, Kolenc D, Vuletic LB, Drmic D, Kokot A, Zoricic I, Coric M, Vlainic J, Poljak L, Seiwerth S, Sikiric P. Stable gastric pentadecapeptide BPC 157 in the therapy of the rats with bile duct ligation. Eur J Pharmacol 2019; 847: 130-142 [PMID: 30690000 DOI: 10.1016/j.ejphar.2019.01.030]
- 151 Davey AK, Maher PJ. Surgical adhesions: a timely update, a great challenge for the future. J Minim Invasive Gynecol 2007; 14: 15-22 [PMID: 17218224 DOI: 10.1016/j.jmig.2006.07.013]
- 152 Collen D. On the regulation and control of fibrinolysis. Edward Kowalski Memorial Lecture. Thromb Haemost 1980; 43: 77-89 [PMID: 6450468]
- 153 Ludwig J, Hashimoto E, McGill DB, van Heerden JA. Classification of hepatic venous outflow obstruction: ambiguous terminology of the Budd-Chiari syndrome. Mayo Clin Proc 1990; 65: 51-55 [PMID: 2296212 DOI: 10.1016/s0025-6196(12)62109-0]
- 154 Darwish Murad S, Dom VA, Ritman EL, de Groen PC, Beigley PE, Abraham SC, Zondervan PE, Janssen HL. Early changes of the portal tract on microcomputed tomography images in a newlydeveloped rat model for Budd-Chiari syndrome. J Gastroenterol Hepatol 2008; 23: 1561-1566 [PMID: 19120847 DOI: 10.1111/j.1440-1746.2008.05403.x]
- 155 Bona E, Hagberg H, Løberg EM, Bågenholm R, Thoresen M. Protective effects of moderate hypothermia after neonatal hypoxia-ischemia: short- and long-term outcome. Pediatr Res 1998; 43: 738-745 [PMID: 9621982 DOI: 10.1203/00006450-199806000-00005]
- Murao Y, Loomis W, Wolf P, Hoyt DB, Junger WG. Effect of dose of hypertonic saline on its 156 potential to prevent lung tissue damage in a mouse model of hemorrhagic shock. Shock 2003; 20: 29-34 [PMID: 12813365 DOI: 10.1097/01.shk.0000071060.78689.f1]
- 157 Veljaca M, Pavic-Sladoljev D, Mildner B, Brajsa K, Krnic Z, Bubenik M, Stipanicic S, Tabak-Slosic M, Brnic L, Khan Z, Krznaric Z, Bischoff A, Scroeder A, van Dongen WD, van Schaik F. Safety, tolerability and pharmacokinetics of PL 14736, a novel agent for treatment of ulcerative colitis, in healthy male volunteers. Gut 2003; 51: A309
- 158 Ruenzi M, Stolte M, Veljaca M, Oreskovic K, Peterson J; Ulcerative Colitis Study Group. A multicenter, randomized, double blind, placebo controlled phase II study of PL 14736 enema in the treatment of mild-to-moderate ulcerative colitis. Gastroenterology 2005; 128: 584
- 159 Xu C, Sun L, Ren F, Huang P, Tian Z, Cui J, Zhang W, Wang S, Zhang K, He L, Zhang C, Hao Q, Zhang Y, Li M, Li W. Preclinical safety evaluation of body protective compound-157, a potential drug for treating various wounds. Regul Toxicol Pharmacol 2020; 114: 104665 [PMID: 32334036 DOI: 10.1016/j.yrtph.2020.104665]



WJG

World Journal of Gastroenterology

Submit a Manuscript: https://www.f6publishing.com

World J Gastroenterol 2022 January 7; 28(1): 47-75

DOI: 10.3748/wjg.v28.i1.47

ISSN 1007-9327 (print) ISSN 2219-2840 (online)

REVIEW

Transfusion-transmitted hepatitis E: What we know so far?

Carmen Ka Man Cheung, Sunny Hei Wong, Alvin Wing Hin Law, Man Fai Law

ORCID number: Carmen Ka Man Cheung 0000-0001-9386-506X; Sunny Hei Wong 0000-0002-3354-9310; Alvin Wing Hin Law 0000-0001-8376-4193; Man Fai Law 0000-0003-2462-6625..

Author contributions: Cheung CKM contributed to acquisition, analysis and interpretation of data/references; drafted and approved the manuscript; Wong SH contributed to analysis, interpretation of data/references; revised critically and approved the manuscript; Law AWH contributed to analysis of data/references and approved the manuscript; Law MF contributed to acquisition, analysis and interpretation of data/references, and drafted and approved the manuscript.

Conflict-of-interest statement: The authors declare no conflict of interest

Country/Territory of origin: China

Specialty type: Gastroenterology and Hepatology

Provenance and peer review: Invited article; Externally peer reviewed.

Peer-review model: Single blind

Peer-review report's scientific quality classification Grade A (Excellent): A

Carmen Ka Man Cheung, Man Fai Law, Medicine and Therapeutics, Prince of Wales Hospital, Hong Kong 852, China

Sunny Hei Wong, Institute of Digestive Disease and Department of Medicine and Therapeutics, the Chinese University of Hong Kong, Hong Kong 852, China

Sunny Hei Wong, Lee Kong Chian School of Medicine, Nanyang Technological University, Singapore 639798, Singapore

Alvin Wing Hin Law, West Island School, Hong Kong 852, China

Corresponding author: Man Fai Law, MRCP, Doctor, Medicine and Therapeutics, Prince of Wales Hospital, 30-32 Ngai Shing Street, Shatin, Hong Kong 852, China. mflaw99@yahoo.com.hk

Abstract

Hepatitis E virus (HEV) is a major cause of viral hepatitis globally. There is growing concern about transfusion-transmitted HEV (TT-HEV) as an emerging global health problem. HEV can potentially result in chronic infection in immunocompromised patients, leading to a higher risk of liver cirrhosis and even death. Between 0.0013% and 0.281% of asymptomatic blood donors around the world have HEV viremia, and 0.27% to 60.5% have anti-HEV immunoglobulin G. HEV is infectious even at very low blood concentrations of the virus. Immunosuppressed patients who develop persistent hepatitis E infection should have their immunosuppressant regimen reduced; ribavirin may be considered as treatment. Pegylated interferon can be considered in those who are refractory or intolerant to ribavirin. Sofosbuvir, a nucleotide analog, showed modest antiviral activity in some clinical studies but sustained viral response was not achieved. Therefore, rescue treatment remains an unmet need. The need for HEV screening of all blood donations remains controversial. Universal screening has been adopted in some countries after consideration of risk and resource availability. Various pathogen reduction methods have also been proposed to reduce the risk of TT-HEV. Future studies are needed to define the incidence of transmission through transfusion, their clinical features, outcomes and prognosis.

Key Words: Hepatitis E virus; Acute and chronic hepatitis; Immunosuppression; Blood transfusion; Transplantation

©The Author(s) 2022. Published by Baishideng Publishing Group Inc. All rights reserved.



WJG https://www.wjgnet.com

Grade B (Very good): 0 Grade C (Good): C Grade D (Fair): 0 Grade E (Poor): 0

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: htt ps://creativecommons.org/Licens es/by-nc/4.0/

Received: April 10, 2021 Peer-review started: April 10, 2021 First decision: June 24, 2021 Revised: July 16, 2021 Accepted: December 22, 2021 Article in press: December 22, 2021 Published online: January 7, 2022

P-Reviewer: Anand AC, Kulkarni AV S-Editor: Chang KL L-Editor: Filipodia P-Editor: Chang KL



Core Tip: Transfusion-transmitted hepatitis E virus (HEV) is an emerging global health concern. In immunocompromised patients, chronic HEV infection increases the risk of liver cirrhosis. The prevalence of viremia and anti-HEV immunoglobulin G in asymptomatic blood donors varies widely between countries but even low concentrations of HEV in blood components are infectious, and in most countries blood donations are not routinely screened for HEV. Treatment of persistent infection includes modification of the immunosuppressant regimen followed by ribavirin. The need for screening of HEV in all blood donations remains controversial. Strategies to reduce de novo HEV infection should also be emphasized.

Citation: Cheung CKM, Wong SH, Law AWH, Law MF. Transfusion-transmitted hepatitis E: What we know so far? World J Gastroenterol 2022; 28(1): 47-75 URL: https://www.wjgnet.com/1007-9327/full/v28/i1/47.htm DOI: https://dx.doi.org/10.3748/wjg.v28.i1.47

INTRODUCTION

Hepatitis E virus (HEV) was first discovered as an epidemic of non-A, non-B hepatitis in the 1980s[1], and has since become one of the major global causes of viral hepatitis. The World Health Organization estimated that HEV caused approximately 44000 deaths in 2015, and accounted for 3.3% of global deaths related to viral hepatitis[2]. A recent meta-analysis concluded that approximately 939 million of the global population have ever experienced HEV infection, and 15 to 110 million individuals have recent or ongoing infection[3]. The infection is generally self-limiting; however, it poses a threat to some vulnerable patients resulting in a significant burden of inpatient admissions, chronic infection, organ failure, and death[4]. The mortality rate can be greater than 20% in patients with chronic liver disease, cirrhosis, or pregnancy [4,5]. With a high HEV serological prevalence among the global population, the safety of blood products has become a public health concern. Herein, we review existing evidence on transfusion-transmitted HEV (TT-HEV), and the implications for screening of blood donations.

VIROLOGY

HEV is a positive-sense, single-stranded RNA icosahedral virus belonging to the genus Orthohepevirus within the Hepeviridae family[6]. Orthohepevirus A has eight distinct genotypes, of which HEV-1, -2, -3 and -4 infect humans[7]. HEV genotype C1, belonging to the species Orthohepevirus C, circulates in rats and can cause cross-species infection and sporadic zoonotic transmission to humans^[8].

HEV exists in urine or feces as non-enveloped virions encased by a capsid. It circulates in blood in a membrane-associated, quasi-enveloped form (eHEV) which is considered to be less contagious[9]. The entry mechanisms for HEV are not well characterized, but once the genomic RNA is uncoated and delivered to the cytosol, the replication cycle is initiated[10]. The viral release that initiates subsequent infection requires multivesicular bodies through endosomal sorting complexes required for transport[11].

EPIDEMIOLOGY

The prevalence rates of HEV antibody are higher in developing countries than in developed countries^[12]. The highest anti-HEV immunoglobulin G (IgG) seropositivity rate has been reported in Africa with a mean of 21.76%, followed by Asia (15.80%), Europe (9.31%), North America (8.05%), South America (7.28%), and Oceania (5.99%). In addition, the reported anti-HEV immunoglobulin M (IgM) seroprevalence rate was 3.09%, 1.86%, 0.79%, 0.22% and 2.43% in Africa, Asia, Europe, North America, and South America, respectively[3].



Among the four major genotypes that can infect humans, HEV-1 and -2 are mostly found in developing countries including Asia, Africa, Latin America, and Mexico. Infection is mainly transmitted via fecally contaminated water, but occasionally also by person-to-person and vertical transmission[13]. Hepatitis E occurs as outbreaks as well as sporadic cases of acute hepatitis, with the preponderance of cases among adolescents and young adults. When stratified by age, the estimated incidence of HEV-1 and -2 infection is roughly between 0.5% and 1.0% for ages 0 to 15 years, with rates increasing to between 1.0% and 1.4% for ages 15 years to 20 years, then falling rapidly to a lower rate of 0.2% and below in individuals older than 30 years [14].

HEV-3 accounts for most of the autochthonous infection in developed countries while HEV-4 is mainly found in Asia and sporadically in Europe[15,16]. The reported seroprevalence of HEV-3 ranged from 0.6% to 52.5% in Europe, 6% in United States, 3 to 16% in United Kingdom and up to 52% in some regions of France[17]. HEV-3 and HEV-4 are zoonotic viruses which are frequently transmitted via food, close contact with animals, or transfusion of viremic blood units[18].

CLINICAL FEATURES AND EXTRAHEPATIC MANIFESTATIONS

The incubation period following exposure to HEV ranges from 2 to 6 wks. HEV infection commonly takes a clinically silent, asymptomatic course with around 5% to 30% of infected individuals developing acute hepatitis[19]. Symptoms of acute hepatitis include fever, malaise, anorexia, vomiting, followed by jaundice, tea-colored urine, and hepatomegaly^[20]. It is then followed by a convalescent phase with gradual recovery within a few weeks in immunocompetent patients^[21]. Acute liver failure is rare and occurs more frequently in middle-aged/elderly patients[22]. Fulminant hepatitis with fatal outcome is uncommon, but has been observed in pregnant women or in patients with pre-existing liver disease. The development of fulminant hepatitis appears to be related to host-specific factors rather than virus genotype, variants, or specific substitutions[23]. HEV superinfection may trigger liver decompensation in patients with chronic liver disease or cirrhosis, resulting in acute-on-chronic liver failure, which is associated with significant short-term mortality [24,25]. Further research is needed to clarify the clinical features, course of illness, and prognosis of patients with decompensated cirrhosis who develop HEV infection.

HEV-3 and HEV-4 can persist in immunocompromised patients resulting in chronic infection, defined as viral replication lasting for more than 3 to 6 mo^[26]. It has been well described in patients after solid organ or stem cell transplant, hematology patients receiving chemotherapy, or HIV-infected patients[27-32]. The prevalence of anti-HEV IgG was about 11.6% and viral RNA was 2% in solid organ transplant recipients^[33]. In solid organ transplant recipients who were positive for HEV RNA, more than 60% developed chronic hepatitis[33].

The natural history of chronic hepatitis E infection is not well understood[34]. In liver transplant recipients infected by HEV, histological analyses of liver biopsy revealed atypical morphology that is distinct from those in immunocompetent patients during early phases of infection[35]. Proliferation of, and cytokine production by, CD4+ and CD8+ T-cells were impaired in patients with persistent HEV viremia[36]. Chronic hepatitis E leads to liver fibrosis and cirrhosis. Cases of HEV-related hepatocellular carcinoma have been reported[37].

Although HEV predominantly infects hepatocytes, it may also affect other organs and present as extrahepatic manifestations. The mechanisms by which HEV can induce extrahepatic manifestations are not fully understood, but hypotheses include direct cytopathic tissue damage by extrahepatic replication, or immunological processes induced by an overwhelming host immune response[38]. Details of extrahepatic manifestations are shown in Table 1[39-44].

PREVALENCE IN BLOOD DONORS

Viremia

The prevalence of HEV RNA in blood donors varies around the world. (Table 2)[45-78]. Most countries have a low prevalence of HEV viremia, ranging from 0.0013% to 0.086%. A relatively higher rate of viremia was reported in Germany (0.12%) and China (0.281%)[49,70]. A meta-analysis of 10 studies from China showed a pooled prevalence of HEV RNA of 0.1% [79]. The actual prevalence might have been underes-



Table 1 Extrahepatic manifestation	ons associated with hepatitis E virus infection						
System	Extrahepatic manifestations						
Neurological	Guillain-Barré syndrome (GBS)						
	Neuralgic amyotrophy						
	Neuropathy						
	Bell's palsy						
	Encephalitis						
	Transverse myelitis						
	Myositis						
	Myasthenia gravis						
	Pseudotumor cerebri						
	Seizure						
Renal	Decrease glomerular filtration rate						
	Glomerulonephritis						
	Nephrotic syndrome						
	Mixed cryoglobulinemia						
Hematological	Thrombocytopenia						
	Hemolytic anemia						
	Aplastic anemia						
	Hemophagocytic syndrome						
	Monoclonal gammopathy of uncertain significance (MGUS)						
Others	Thyroiditis						
	Pancreatitis						
	Myocarditis						
	Polyarthritis						

timated as some studies included in the meta-analysis conducted RNA detection only in those donors who were positive for anti-HEV IgM or antigen[79].

The prevalence of HEV-3 and -4 is affected by dietary habits[80]. Consumption of raw pork tartare and undercooked pork liver may represent a relevant risk factor for HEV infection in Germany^[49]. Regular consumption of pork meat and shellfish were also reported in the viremic donors in China^[70].

Since 70% of infections with HEV-3 and -4 are asymptomatic[81], it can be difficult to identify infected blood donors, as viremia occurs primarily during the pre-icteric phase[82]. Katiyar et al[72] described anti-HEV IgG positivity in 60.5% of the tested donors in India and yet none of them were positive for HEV RNA. In India, human HEV is caused exclusively by the HEV-1 genotype, which causes brief hepatitis and seldom results in chronic infection[83,84]. The difference in endemicity between HEV genotypes may affect the propensity to cause symptomatic disease and viral persistence, which in turn influences the likelihood of viremia among blood donors.

Other factors influencing the reported prevalence of HEV viremia are the sensitivity and plasma pool size of the various nucleic acid test screening platforms used[85]. For example, 33 of 90 donations with a viral load of 20-750 IU/mL were positive when tested individually but missed in the pooled screening in a study by Hogema et al[57]. Delage *et al*[66] revealed a low prevalence (n = 11/50765) and viral loads of HEV-RNA in Canadian blood donors based on individual nucleic acid amplification techniques (NAT). They postulated that if pooled NAT was used, only two positive donations with viral loads > 1000 IU/mL would have been detected. The true frequency of viremia in blood donors in studies using pooled NAT could be underestimated due to a dilution effect. Vollmer et al[86] found that screening using individual NAT yielded an approximately 50% higher detection frequency compared with NAT of a mini-pool of 96 samples; nevertheless, samples exclusively positive for individual NAT had a



Table 2 Hepatitis E virus ribonucleic acid prevalence in donor, only studies include more than 1000 study subjects are included Number of Number Initial screening Prevalence Median (range) HEV genotype: n/N Ref. Country donations positive Outcome of recipient method (95%CI) viral load. IU/mL donations screened Europe Fischer *et al*[45], RT-PCR (plasma pool of 96 58915 7 0.012% 3:7/7 (2200 to 290000) N/A Austria samples) with 95% LOD 2015 11.6 IU/mL 7 Vercouter et al Belgium RT-PCR (plasma pool of 6 38137 0.018% N/A (153 to 8710) N/A [46], 2019 samples) with 95% LOD 18.6 IU/mL Harritshøj et al Denmark TMA assay on individual 25637 11 0.043% 3 (in 2 samples) 13 (unquantifiable (1) Look-back testing was performed in 7 recipients; all were tested negative for [47], 2016 plasma with 95% LOD 7.9 (0.02% to 920) HEV RNA and anti-HEV IgM; (2) No recipient developed transaminitis; and (3) IU/mL 0.07%) One patient had strongly positive anti-HEV IgG assay which may indicate recent HEV infection or secondary immune response by HEV re-exposure. Gallian et al[48], RT-PCR (plasma pool of 96 53234 22 0.045% 3c: 5/14; 3f: 8/14; 3 (468 to 5155800) N/A France 2014 samples) with 95% LOD 23 (0.043%-1/14IU/mL 0.047%). 23 0.123% Westhölter et al Germany RT-PCR (plasma pool of 24 18737 3:6/7 (120 to 11200000) (1) Retrospective analysis of 4 viremic donors showed that they were HEVsamples) with 95% LOD positive in previous donations; (2) In 3 donors, testing of the previously [49], 2018 18.6 IU/mL donated blood in pools of 24 samples failed to identify viremic donations but were tpositive in unpooled samples; (3) Fourteen recipients had received HEV RNA positive blood products; (4) One immunosuppressed recipient tested positive for HEV RNA, developed acute on chronic liver failure, and died; and (5) One immunocompetent recipient developed acute self-limited episode of hepatitis E Dreier *et al*[50], RT-PCR with 95% LOD 4.7 235524 182 0.077% 3:4/4(< 25 to 69.4) (1) Nine viremic donations were transfused to 6 different recipients; (2) Two Germany 2018 IU/ml for FFP, platelet recipients were immunocompromised (heart transplantation and leukemia); (3) concentrates, and RBC Two recipients died shortly after transfusion for reasons other than HEV supernatant; 95% LOD 8.9 infection; and (4) None of the other 4 recipients developed acute HEV infection IU/mL for RBCs. or had detectable HEV RNA / anti-HEV IgG 0.015% Corman *et al*[51], Germany RT-PCR (plasma pool of 96 93955 14 3:14/14 (3.1 to 4.8 Log₁₀ N/A 2013 samples mixed in IU/mL) metapools of 20) Vollmer *et al*[52], Germany RT-PCR (plasma pool of 48 16125 13 0.081% 3:13/13 (13 to 68100) N/A 2012 samples) with 95% LOD 4.7 IU/ml (3.26 to 5.35 log10 Baylis *et al*[53], RT-PCR (plasma pool of 96 18,100 4 0.022% 3 Donations screened positive for HEV were excluded from pharmaceutical Germany samples) with 95% LOD 2012 copies/mL) production 250 IU/mL TMA assay with 95% LOD 24985 0.020% (10 to 44550) O'Riordan et al Ireland 5 3:3/3 N/A

Cheung CKM et al. Transfusion-transmitted hepatitis E

[<mark>54</mark>], 2016		5.5 IU/mL			(0.0065%- 0.0467%)			
Spreafico <i>et al</i> [55], 2020	Italy	TMA assay on individual plasma with 95% LOD 7.9 IU/mL	9726	1	0.010% (0.00%-0.06%)	N/A	N/A	N/A
Spada <i>et al</i> [<mark>56</mark>], 2018	Italy	RT-PCR, plasma pool and sensitivity varies according to anti-HEV IgG and IgM status	10011	0	N/A	N/A	N/A	N/A
Hogema <i>et al</i> [57], 2015	Netherlands	RT-PCR (plasma pool of 96 samples) with 95% LOD 38.4 IU/mL with the EasyMag extraction method and 10.3 IU/mL using Qiagen extracts	59474	41	0.069%	3c: 15/17; 3f: 2/17	N/A	N/A
Slot <i>et al</i> [<mark>58</mark>], 2013	Netherlands	RT-PCR (plasma pool of 48 or 480 samples) with 95% LOD 25 IU/mL	40176	13	0.032%	3: 16/17	(< 25 to 470000)	N/A
Grabarczyk <i>et al</i> [<mark>59]</mark> , 2018	Poland	TMA assay on individual plasma with 95% LOD 7.9 IU/mL	12664	10	0.079% (0.043%- 0.145%)	3i: 2/3; 3c: 1/3	(16 to 6586 in 4 patients with positive results in qPCR)	N/A
Rivero-Juarez <i>et al</i> [60], 2019	Spain	RT-PCR (plasma pool of 8 samples) with sensitivity 670 IU/mL	11313	4	0.035% (0.01%-0.09%)	3: 4/4	(10788 to 2000000)	(1) Five patients received transfusions from HEV-infected donors; and (2) None of them showed an increase in alanine aminotransferase levels after transfusion
Sauleda <i>et a</i> l[<mark>61</mark>], 2015	Spain	TMA assay on individual plasma with 95% LOD 7.9 IU/mL	9998	3	0.030% (0.01%-0.09%)	3f (in 1 sample)	(250 to 2755)	N/A
Baylis <i>et al</i> [<mark>53</mark>], 2012	Sweden	RT-PCR (plasma pool of 96 samples) with 95% LOD 250 IU/mL	95835	12	0.013%	3	(3.20 to 5.68 log ₁₀ copies/mL)	Donations screened positive for HEV were excluded from pharmaceutical production
Harvala et al <mark>[62]</mark> , 2019	United Kingdom	RT-PCR (plasma pool of 24 samples) with 95% LOD 18.6 IU/mL	1838747	480	0.026%	3c: 112/149; 3e: 21/1493f: 12/149; 3a: 1/149; 2 distantly related to 3h, and 1 clustered distantly with 3a	883 (1 to 3230000)	N/A
Thom <i>et al</i> [<mark>63</mark>], 2018	United Kingdom	RT-PCR (plasma pool of 24 samples)	94302	38	0.040%	3: 10/10	N/A	N/A
Hewitt <i>et al</i> [<mark>64</mark>], 2014	United Kingdom	RT-PCR (plasma pool of 24 samples)	225000	79	0.035%	3: 79/79	3900 (50 to 2.37 × 10 ⁶)	(1) Forty-three patients who had received blood components from HEV- infected donors were followed up; (2) The overall transmission rate was 42% (18 of 43 exposed patients); (3) One recipient developed clinical hepatitis and 4 recipients developed asymptomatic transaminitis; and (4) Four heavily

immunosuppressed patients had delayed (37-38 wk) seroconversion or no antibodies detected

								antibodies detected
Cleland <i>et al</i> [<mark>65</mark>], 2013	United Kingdom	Nested PCR (plasma pool of 24 samples) with 95% LOD 201 IU/mL	43560	3	0.0069%	3:3/3	N/A	N/A
North America								
Delage <i>et al</i> [<mark>66</mark>], 2019	United States	RT-PCR on individual samples with 95% LOD 18.6 IU/mL	50724	3	0.0059%	3: 2/3; genotyping was unsuccessful in 1 patient	(23 to 1420)	N/A
	Canada		50765	11	0.022%	3 (in 1 sample)	(< 10 to 3080)	
Roth <i>et al</i> [<mark>67</mark>], 2017	United States	RT-PCR (plasma pool of 96 samples) with 95% LOD 18.6 IU/mL	128021	4	0.003%	3a: 3/3	(3.0 to 3.8 log IU/mL)	N/A
Stramer <i>et al</i> [<mark>68</mark>], 2016	United States	TMA assay on individual plasma with 95% LOD 7.9 IU/mL	18829	2	0.011% (0.0018%- 0.351%)	N/A	14 IU/mL in one sample	N/A
Xu et al <mark>[69</mark>], 2013	United States	RT-PCR (plasma pool of 7 to 8 samples) with 95% LOD 400 IU/mL and nested PCR with 95% LOD 200 IU/mL	1939	0	N/A	N/A	N/A	N/A
Baylis <i>et al</i> [<mark>53</mark>], 2012	United States	RT-PCR (plasma pool of 96 samples) with 95% LOD 250 IU/mL	51075	0	N/A	N/A	N/A	N/A
Asia								
Wen <i>et al</i> [70], 2018	China	RT-PCR on individual plasma	5345	15	0.281%	One 4h, another one clustered between genotype 2 and 4i	N/A	N/A
Tsoi <i>et al</i> [71] , 2019	Hong Kong	RT-PCR with 95% LOD 7.89 IU/mL	10000	2	0.02%	4 (in 1 sample)	N/A	N/A
Katiyar H <i>et al</i> [72], 2018	India	RT-PCR (plasma pool of 3 samples) with LOD 100 IU/mL	1799	0	N/A	N/A	N/A	N/A
Minagi T <i>et al</i> [73], 2016	Japan	RT-PCR (plasma pool of 50 or 500 samples) with 95% LOD 152 IU/mL	620140	36	0.0058%	3: 36/36	(< 1.69 to 7.22 log ₁₀ copies/mL)	N/A
Intharasongkroh et al[74], 2019	Thailand	RT-PCR (plasma pool of 6 samples) with 95% LOD 53.5 IU/mL	30115	26	0.086%	3:6/6	N/A	N/A

Cheung CKM et al. Transfusion-transmitted hepatitis E

Others								
Hoad <i>et al</i> [75], 2017	Australia	TMA (plasma pool of 6 samples)	74131	1	0.0013%	N/A	180	N/A
Shrestha <i>et al</i> [76], 2016	Australia	TMA assay on individual plasma with 95% LOD 7.9 IU/mL	14799	1	0.0068% (0.0002%- 0.0376%	3	15000	N/A
Hewitt <i>et al</i> [77], 2018	New Zealand	RT-PCR (plasma pool of 8 to 12 samples)	5000	0	N/A	N/A	N/A	N/A
Maponga <i>et al</i> [78], 2020	South Africa	TMA assay on individual plasma with 95% LOD 7.9IU/mL	10000	1	0.01%	3	79000	All donations from donors with active HEV infection were discarded

CI: Confidence interval; FFP: Fresh frozen plasma; HEV: Hepatitis E virus; Ig: Immunoglobulin; LOD: Limit of detection; RBC: Red blood cells; RNA: Ribonucleic acid; RT-PCR: Real time polymerase chain reaction; TMA: Transcription mediated amplification.

corresponding viral load of < 25 IU/mL. High-sensitivity individual NAT can yield false-positive results[55]. Whether the identification of low-level HEV-positive donors translates into clinical significance and whether a single individual NAT is adequate remain undefined.

Antibodies

In addition to direct detection of HEV RNA, another important indirect assessment of HEV burden is the prevalence of anti-HEV IgM and IgG in blood donors (Table 3)[45, 46,54-56,58,59,61,63,65,68,69,71,72,77,87-124]. HEV IgG prevalence increases with age which likely represents the cumulative effect of HEV exposure over a lifetime, especially as IgG antibodies can persist for decades[81]. The absence of detectable antibodies in donors was related to an increased risk of transfusion transmission of HEV[64]. However, the presence of anti-HEV IgG may not always be protective as multiple HEV reinfections could occur despite pre-existing antibodies[125]. Various HEV strains in serum are capable of replication in cell culture and generate infectious particles in the culture supernatant despite the coexistence of antibodies[126]. Anti-HEV IgM could be used to detect recent infection yet it failed to identify infected donors during the window period. For example, a meta-analysis of data from 28 countries found that only 26.6% of viremic blood units had positive anti-HEV antibodies[127]. In another study by Tedder *et*[128] *al*, a significant portion of viremic individuals (n = 57/79) were seronegative at the time of donation. Anti-HEV IgM sometimes exhibits unexpectedly long persistence for up to 3 years after a self-limiting acute hepatitis E episode^[129]. Only a minority of anti-HEV IgM-positive donors have detectable RNA[58,93,103,109]. All these findings suggest that detection of anti-HEV IgG or IgM alone may not provide effective screening of HEV in blood donors.

Table 3 Seroprevalence of hepatitis E in blood donors

Ref.	Country	Number of donations screened	Assay used	Number of samples positive for HEV IgG antibodies	Anti-HEV IgG prevalence (95%Cl)	Number of samples positive for HEV IgM antibodies	Anti-HEV IgM prevalence (95%CI)	Number of samples positive for HEV RNA in anti-HEV IgM positive	Viral load, IU/mL	Genotype
Europe										
Fischer <i>et al</i> [45], 2015	Austria	1203 (from HEV RNA negative donors)	Wantai	163	13.55% (11.6%- 15.5%)	N/A	N/A	0	N/A	N/A
Vercouter <i>et al</i> [46], 2019	Belgium	356 (from HEV RNA negative donors)	Wantai	31	8.71% (6.20%- 12.10%)	0	N/A	0	N/A	N/A
Miletić <i>et al</i> [<mark>87</mark>], 2019	Croatia	1036	3 commercial ELISA assays were used, only findings with highest prevalence are shown	209	20.17%	46	4.44%	0	N/A	N/A
Holm <i>et al</i> [88], 2015	Denmark	504	In-house NIH assay	54	10.7% (8.2%- 13.7%)	N/A	N/A	N/A	N/A	N/A
			Wantai	100	19.8% (16.4%- 23.6%)					
Dimeglio <i>et al</i> [89], 2018	France	300	Wantai	23	7.7% (4.9%- 11.3%)	2	0.6% (0.1%-2.4%)	0	N/A	N/A
Juhl <i>et al</i> [90], 2013	Germany	1019	RecomWell assay and Western blot	69	6.8% (5.3%-8.3%)	N/A	N/A	N/A	N/A	N/A
Dalekos <i>et al</i> [<mark>91</mark>], 1998	Greece	3016	Abbott assay and Western blot	8	0.27%	0	N/A	N/A	N/A	N/A
O'Riordan <i>et al</i> [<mark>54]</mark> , 2016	Ireland	1076	Wantai	57	5.3% (4.0%-6.8%)	2	0.19%	0	N/A	N/A
Spreafico <i>et al</i> [55], 2020	Italy	767	DiaPro	52	6.8% (5.1%-8.8%)	0	N/A	0	N/A	N/A
Spada <i>et al</i> [<mark>56</mark>], 2018	Italy	10011	Wantai	869	8.7% (8.14%- 9.25%)	46	0.4% (0.34% - 0.61%)	0	N/A	N/A
De Sabato <i>et al</i> [<mark>92</mark>], 2017	Italy	170	Bio-Chain Institute and Western blot	15	8.82%	3	1.76%	0	N/A	N/A
Lucarelli <i>et al</i> [93], 2016	Italy	313	Wantai	153	48.9% (43%-54%)	2	0.6% (0.08%-2.3%)	1	100	3
Puttini <i>et al</i> [<mark>94</mark>], 2015	Italy	132	EIAgen HEV IgG kit	12	9.1%	N/A	N/A	N/A	N/A	N/A

Cheung CKM et al. Transfusion-transmitted hepatitis E

Hogema <i>et al</i> [95], 2014	Netherlands	513	Wantai	58	11.31%	N/A	N/A	N/A	N/A	N/A
Slot <i>et al</i> [58] , 2013	Netherlands	5239	Wantai	1401	26.7% (25.6%- 28.0%)	49	0.94%	4	Range: < 25 to 3700	3
Grabarczyk <i>et al</i> [<mark>59</mark>], 2018	Poland	3079	Wantai	1340	43.52% (41.78%- 45.28%)	39	1.27% (0.93%- 1.73%)	N/A	N/A	N/A
Sauleda <i>et al</i> [<mark>61</mark>], 2015	Spain	1082	Wantai	216	19.96% (17.60%- 22.32%)	13	1.20%	0	N/A	N/A
			Mikrogen	116	10.72% (8.90%- 12.60%)					
Mateos <i>et al</i> [<mark>96</mark>], 1999	Spain	863	Abbott assay and Western blot	34	3.9%	0	N/A	N/A	N/A	N/A
Niederhauser <i>et al</i> [97], 2018	Switzerland	3609	Wantai	737	20.4% (19.1%- 21.8%)	N/A	N/A	N/A	N/A	N/A
Kaufmann <i>et al</i> [98], 2011	Switzerland	550	MP Biomedicals	27	4.9%	N/A	N/A	N/A	N/A	N/A
	United Kingdom	1714	Wantai	104	6.1% (5.0%-7.3%)	N/A	N/A	N/A	N/A	N/A
	United Kingdom	1559	Wantai	73	4.7% (3.6%-5.8%)	0	N/A	N/A	N/A	N/A
	United Kingdom	262	Wantai	31	11.8%	4	1.5%	0	N/A	N/A
North America										
	United States	5040 (from HEV	DSI	569	11.29%	146	2.90%	0	N/A	N/A
[<mark>100</mark>], 2018		RNA negative donor)	MP Biomedicals	537	10.65%	93	1.85%			
			Wantai	619	12.28%	34	0.67%			
Stramer <i>et al</i> [68], 2016	United States	4499	MP Biomedicals	329	7.3% (6.6%-8.1%)	26	0.58% (0.39%- 0.85%)	N/A	N/A	N/A
Xu et al[69], 2013	United States	1939	Wantai	364	18.8% (17.0%- 20.5%)	8	0.4% (0.1%-0.7%)	0	N/A	N/A
South America										
Di Lello <i>et al</i> [<mark>101</mark>], 2020	Argentina	391	DiaPro	44	11.3%	8	2.0%	0	N/A	N/A
Bangueses <i>et al</i> [102], 2020	Uruguay	400	DiaPro	40	10%	19	4.75%	3	N/A	3

Asia										
Nouhin <i>et al</i> [<mark>103</mark>], 2016	Cambodia	301	Wantai	85	28.2% (23.4%- 33.5%)	3	1.0% (0.01%-1.8%)	1	956	3
Chen <i>et al</i> [<mark>104</mark>], 2019	China	4044	Wantai	799	19.8% (18.6%- 21.0%)	43	1.1% (0.8%-1.4%)	2	N/A	4
Wen <i>et al</i> [70], 2018	China	5345	Wantai	1227	22.96%	38	0.71%	15	N/A	N/A
Wang <i>et al</i> [105], 2017	China	9069	Wantai	2682	29.57%	131	1.44%	5	N/A	N/A
Ma et al <mark>[106</mark>], 2015	China	816	Wantai	172	21.1%	4	0.5%	0	N/A	N/A
Ren <i>et al</i> [107], 2014	China	10741	Wantai	2945	27.42%	109	1.01%	0	N/A	N/A
Zhuang <i>et al</i> [<mark>108</mark>], 2014	China	486	ELISA based on antigen protein pB166 and MPII	113	23.3%	N/A	N/A	N/A	N/A	N/A
Tsoi <i>et al</i> [71] , 2019	Hong Kong	2000	Wantai	315	15.8% (14.2%- 17.4%)	16	0.8%	0	N/A	N/A
Tripathy <i>et al</i> [109], 2019	India	2447	Wantai	433	17.70% (16.23%- 19.26%)	5	0.20%	2	Ranged from 3.5×10^4 to 4.6×10^5 copies/mL	1
Katiyar et al [72] , 2018	India	633	Wantai	383	60.5%	N/A	N/A	0	N/A	N/A
Gajjar <i>et al</i> [<mark>110]</mark> , 2014	India	460	DiaPro	N/A	N/A	22	4.78%	N/A	N/A	N/A
Parsa <i>et al</i> [111], 2016	Iran	700	DiaPro	42	6.0%	5	0.71%	5 (only 50 seropositive blood donors were tested)	N/A	1
Hesamizadeh <i>et</i> al[<mark>112]</mark> , 2016	Iran	559	DiaPro	45	8.05%	N/A	N/A	N/A	N/A	N/A
Naeimi <i>et al</i> [<mark>113</mark>], 2015	Iran	628	HEV IgG, Pasto, Iran	105	16.72%	N/A	N/A	N/A	N/A	N/A
Ehteram <i>et al</i> [114], 2013	Iran	530	DiaPro	76	14.3%	N/A	N/A	N/A	N/A	N/A
Taremi <i>et al</i> [<mark>115</mark>], 2007	Iran	399	DiaPro	31	7.8%	N/A	N/A	N/A	N/A	N/A
Takeda <i>et al</i> [<mark>116], 2010</mark>	Japan	12600	in-house ELISA	431	3.42%	N/A	N/A	N/A	N/A	N/A
Shrestha et al	Nepal	1845	Wantai	773	41.9% (39.7%-	55	3.0% (2.2%-3.8%)	N/A	N/A	N/A

Cheung CKM et al. Transfusion-transmitted hepatitis E

[117], 2016					44.2%)					
Nasrallah <i>et al</i> [<mark>118</mark>], 2017	Qatar	5854	Wantai	1198	20.46%	34	0.58%	4	N/A	N/A
Jupattanasin <i>et al</i> [119], 2019	Thailand	630	EUROIMMUN test kit	187	29.7% (26.2%- 33.4%)	N/A	N/A	N/A	N/A	N/A
Africa										
Traoré <i>et al</i> [120], 2016	Burkina Faso	1497	DiaPro and Wantai	584	39%	13	0.87%	N/A	N/A	N/A
Ibrahim <i>et al</i> [<mark>121</mark>], 2011	Egypt	760	N/A	N/A	N/A	3	0.39%	2	N/A	N/A
Meldal <i>et al</i> [<mark>122</mark>], 2013	Ghana	239	4 commercial assays were used, findings reactive in; at least two serological assays are shown	11	4.6%	14	5.9%	0	N/A	N/A
Lopes <i>et al</i> [123], 2017	South Africa	300	Fortress Diagnostics	76	25.3%	0	N/A	N/A	N/A	N/A
Ben-Ayed <i>et al</i> [124], 2015	Tunisia	426	Globe; Diagnostics Srl ELISA	19	4.46%	N/A	N/A	N/A	N/A	N/A
Others										
Hewitt <i>et al</i> [77], 2018	New Zealand	1013	Wantai	98	9.7% (7.9%- 11.7%)	N/A	N/A	N/A	N/A	N/A
			MP Biomedicals	82	8.1% (6.5%- 10.0%)					

ALT: Alanine aminotransferase; CI: Confidence interval; DSI: Diagnostic Systems Incorporated; ELISA: Enzyme-linked immunosorbent assay; HEV: Hepatitis E virus; NIH: National Institutes of Health.

Geographical variation, racial differences, and diverse study methodology and laboratory techniques all contribute to differences in HEV seroprevalence. More than one-third of donors had evidence of past HEV infection in Poland, India, Nepal and Burkina Faso[59,72,117,120]. Lucarelli *et al*[93] reported an unexpectedly high prevalence (48.9%) of anti-HEV IgG among 313 donors in central Italy. Eating raw dried pig liver sausage was the only independent risk factor for HEV IgG in their study, but the authors speculated that the uncontrolled expansion of the wild boar population had resulted in contamination of the soil and watercourses for people living in rural areas, and this may also have also contributed to the high prevalence of HEV[93].

Caution is needed when interpreting the HEV serology results because commercial kits for serological detection show marked variation in sensitivity and specificity. Despite the relatively high sensitivity of the IgM assay, the sensitivity of IgG detection kits is highly dependent on a patient's immune status, being 80% to 90% in immuno-competent individuals, but falling dramatically to 15% to 45% in immunocom-

promised patients^[130]. In a meta-analysis conducted in Europe, the pooled anti-HEV IgG seroprevalence rates determined by different commercial assays showed large variability with reported seroprevalence rates ranging from 2% to 17% [131]. Poor concordance of test results between the Wantai, Dia.Pro and MP Diagnostics HEV enzyme-linked immunosorbent assays (ELISA) were observed[132,133]. This may partly explain the broad ranges of anti-HEV IgG prevalence (5.3% to 48.9%) reported in Italy [55,56,92-94]. In contrast, most studies conducted in China used the Wantai assay and revealed a similar seroprevalence of around 20% to 30%. This assay is believed to be more sensitive than other commercial assays in detecting anti-HEV IgG [134,135].

TRANSFUSION-TRANSMITTED HEPATITIS E

HEV transmission via transfusion has been reported since 2004[136] and there has been increasing recognition of the risk of transmitting HEV by transfusion in recent years. Cases of TT-HEV are shown in Table 4[137-150]. Identical genomic sequences were identified in most infected patients and blood donors. Table 4 Likely only represents the tip of the iceberg as other probable or possible cases have been reported in the literature[151,152]. At the same time, patients with mild symptoms of hepatitis E may have gone undiagnosed. Physicians should stay vigilant for HEV infection in patients who have received a blood transfusion.

Although blood components that contain larger plasma volumes, principally fresh frozen plasma and platelet components, are believed to transmit HEV more readily [64], a number of TT-HEV cases associated with red blood cell transfusion have also been described[138,140,141,143,144,148-150]. Red blood cell transfusion was a significant risk factor for HEV seropositivity in patients on hemodialysis in Croatia[153]. Twenty percent (n = 8/40) of multiply transfused thalassemia patients were anti-HEV IgG positive compared with 11.0% (n = 10/91) in blood donors[154]. In contrast, a study in Iran found anti-HEV antibodies in only 1.67% of patients with thalassemia, suggesting a low rate of TT-HEV in that country[155]. Results from these two studies in thalassemia patients were limited by the small sample size. Ankcorn *et al*[156] analyzed 1591 patients with hematologic malignancy and found that the more transfusions of non-HEV screened blood products the patients had received, the higher their likelihood of being IgG seroreactive was, suggesting HEV acquisition via transfusion in these patients.

A study by Hewitt *et al*^[64] indicated that a viral concentration of between 407 and 257039 IU/mL in blood products was associated with TT-HEV, and that a high viral load in donors rendered infection more likely (P < 0.0001). However, this may not be true in immunocompromised patients. In a systematic review, Dreier et al[50] calculated the median transfused viral load in HEV-infected and non-infected immunocompromised patients. Although the transfused viral load was higher in the infected than the non-infected individuals (4.80×10^5 IU vs 1.55×10^4 IU), the betweengroup difference was not statistically significant (P = 0.1006)[50]. A potential reason for this finding is that a low viral concentration (150 IU/mL) of the blood component could already be infectious [140].

Most cases of TT-HEV occur in immunocompromised recipients, such as patients with hematologic malignancies, or recipients of solid organ or hematopoietic stem cell transplants. However, patients on simple immunosuppressants like corticosteroids and cyclosporine or even immunocompetent individuals are also at risk[157]. Massive transfusion increased the risk of HEV transmission in an immunocompetent trauma patient[158]. Spontaneous resolution, viral eradication by immunosuppressant reduction and/or ribavirin are possible[159] but occasionally there are cases which have progressed into chronic hepatitis, liver cirrhosis or multi-organ failure. Transfusion recipients are more vulnerable to chronic liver injury than the general population as a result of foodborne infection [140]. More than 60% (n = 56/85) of solid organ transplant recipients infected with HEV developed chronic hepatitis, with tacrolimus use as an independent predictive factor [160]. Pas et al [161] screened 1200 solid-organ transplant recipients in the Netherlands for HEV RNA and identified 12 patients with HEV infection. Nine of these 12 patients had been treated with a tacrolimus-based regimen postoperatively. In liver transplant recipients, graft hepatitis with rapid histological disease progression and requirement of re-transplantation due to liver cirrhosis has been reported [162,163]. The rapid progression of HEV infection to advanced fibrosis and cirrhosis has also been observed in individuals receiving kidney or heart transplants[33]. In 50 patients with hematologic malignancy and clinically



Table 4 Reported cases of transfusion transmitted hepatitis E

Study	Number of patients	Comorbidity	Blood component received (<i>n</i>)	Viral load of transfused blood product IU/mL	Genotype	Treatment	Outcome
Okano <i>et al</i> [137], 2020	1	AML on chemotherapy	Plt	N/A	3b	Nil	Spontaneous resolution and developed HEV antibodies after cessation of chemotherapy for AML
Gallian <i>et al</i> [<mark>138</mark>], 2019	23	Solid organ transplant, $n = 9$; allogeneic hematopoietic stem cells transplant, $n = 4$; hematologic malignancies, $n = 5$; immunosuppressant, $n = 2$; immunocompetent, $n = 3$	RBC $n = 7$; apheresis Plt n = 3; whole blood-derived pooled Plt $n = 6$; FFP $n =$ 7	Ranged from 1.14 $\times 10^3$ to 31 × 62.10 6			Acute HEV infection, $n = 8$; spontaneous resolution, $n = 4$; ribavirin treatment, $n = 3$; immunosuppressant reduction, $n = 1$; chronic HEV infection, $n = 14$, all immunosuppressed; resolution with ribavirin, $n = 10$; resolution with immunosuppressant reduction, $n = 4$; One solid organ transplant recipient did not clear HEV infection despite ribavirin and died of multiorgan failure
Ledesma <i>et al</i> [139], 2019	2	Allogeneic BMT, $n = 1$; liver transplant, $n = 1$	Plt	3×10^4	Зе	Ribavirin, <i>n</i> = 1	The patient received BMT remained HEV-infected and IgM/IgG -negative until death; the patient with liver transplant was treated successfully with a course of ribavirin
Satake <i>et al</i> [<mark>140]^a, 2017</mark>	19	Hematologic malignancies, $n = 6$; organ transplant, $n = 2$; systemic disease, $n = 8$; no major comorbidity, $n = 3$	RBC <i>n</i> = 10; Plt <i>n</i> = 6; FFP <i>n</i> = 3	Ranged from 1.5×10^2 to 5.3×10^6	4 <i>, n</i> = 2	N/A	Two patients with malignant lymphoma and two who had received liver transplant developed chronic hepatitis E; the two liver transplant recipients were successfully cleared of HEV by ribavirin
Lhomme <i>et al</i> [141], 2017	3	Solid organ transplant	One patient received RBC; one patient received RBC and Plt; one patient received Plt and FFP	Ranged from 3.6 to 8.2 log IU	3, $n = 1$; 3f, $n = 2$	N/A	N/A
Yamazaki <i>et</i> al[<mark>142</mark>], 2017	2	Hematologic malignancies treated with chemotherapy	N/A	N/A	3b	N/A	Did not become chronic hepatitis E
Belliere <i>et al</i> [143], 2017	1	Heart transplant	RBC	1430 copies/mL	3	Ribavirin	Died from multi-organ failure despite treatment
Riveiro- Barciela <i>et al</i> [144], 2017	1	Immunocompetent, admitted for disseminated infection	RBC	75000	3	Nil	Spontaneous resolution
Hoad <i>et al</i> [145], 2017	1	Liver transplant	FFP	947	3	Ribavirin	Resolved with treatment
Matsui <i>et al</i> [<mark>146</mark>], 2015	1	AMI post CABG with hemorrhagic cardiac tamponade	Plt	10 ^{6.8} copies	3	Nil	Spontaneous resolution
Huzly <i>et al</i> [147], 2013	1	Immunocompromised	Apheresis Plt	30888-37273	3f	N/A	N/A
Coilly <i>et al</i> [148], 2013	1	Liver transplant	RBC	3.5 log ₁₀	3с	Ribavirin	Resolved with treatment
Boxall <i>et al</i> [<mark>149</mark>], 2006	1	Lymphoma on chemotherapy	RBC	N/A	3	Nil	Spontaneous resolution

Mitsui <i>et al</i> 1 Hemoc [150], 2004	dialysis RBC	N/A	3 Nil	Subclinical infection without elevated ALT
--	--------------	-----	-------	--

^aTwo cases were not confirmed by sequence identity and should only be considered as probable TT-HEV.

ALT: Alanine aminotransferase; AMI: Acute myocardial infarction; AML: Acute myeloid leukemia; BMT: Bone marrow transplant; CABG: Coronary artery bypass graft; FFP: Fresh-frozen plasma; HEV: Hepatitis E virus; Ig: Immunoglobulin; Plt: Platelet concentrates; RBC: Red blood cell.

overt hepatitis E, the mortality rate was 16% (n = 8), with liver-related death occurring in 4 patients[164]. HEV could actively suppress the cellular immune response and increase levels of immunosuppressive interleukin-10 that may perpetuate chronic infection and subsequent liver damage[165,166].

TREATMENT

The management strategy for HEV infection should be determined by the clinical presentation. Currently, there is limited information in the published literature that describes the clinical features of TT-HEV, or the optimal approach to management. Acute TT-HEV infections are usually subclinical or mild, with no severe or fulminant cases reported[140]. Therefore, most acute HEV infections should be treated conservatively, while waiting for spontaneous clearance, although a short course of ribavirin may also be considered. In 21 patients with acute HEV infection who were at high risk of liver failure, receiving immunosuppressive therapy for an autoimmune disease or undergoing chemotherapy, a short course of ribavirin for up to 3 mo was associated with rapid virological response and normalization of liver enzymes[167].

The current practice for management of chronic HEV infection is mainly based on observational data[18]; Figure 1 shows a proposed algorithm for management. In patients who are on immunosuppressants, the first-line intervention should be a dose reduction or discontinuation of the immunosuppressive drug[168,169]. In solid organ transplant recipients, reducing the dose of immunosuppressive therapies that principally target T-cells can achieve HEV clearance in nearly one third of patients [160]. Most immunosuppressive drugs such as cyclosporine and tacrolimus increase HEV replication in vitro; mycophenolate mofetil is the only immunosuppressant agent demonstrated to have an anti-viral effect[170].

If modification of the immunosuppressant regimen is not possible or is unsuccessful, pharmacological agents such as ribavirin and/or pegylated interferon-alpha (peg-IFN) can be used[171]. In a meta-analysis that included 395 patients with chronic hepatitis E, ribavirin monotherapy for a median of 3 mo achieved sustained virological response (SVR) in 76% of patients[172]. The reported dose of ribavirin in the literature ranged from 29 to 1200 mg/d, and the duration from 1 to 18 mo. Data on the optimal treatment regimen are needed[173]. HEV RNA should be assessed in the serum and in the stool before treatment discontinuation[169]. A second course of ribavirin for 6 mo

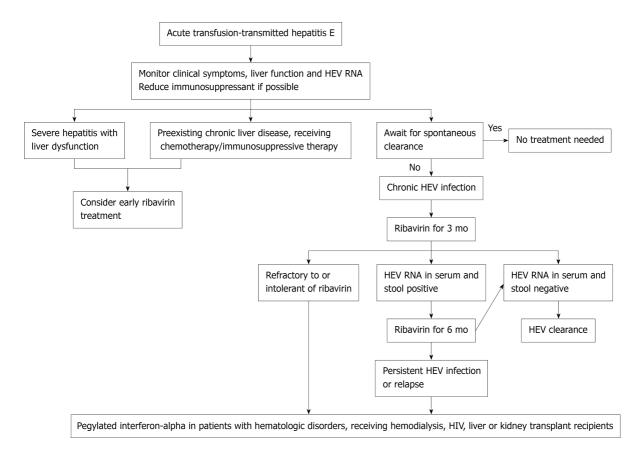


Figure 1 Recommended algorithm for management of transfusion-transmitted hepatitis E.

can be attempted in cases of treatment failure[172]. HEV RNA concentrations decrease within the first week of initiating ribavirin therapy, and a greater reduction in viral load on day 7 is an independent predictor of SVR[174]. Ribavirin failure has been linked to the presence of certain single nucleotide variants (SNVs) and in-frame insertions in the hypervariable region of open reading frame (ORF) 1 in the HEV genome[175].

For those who are refractory to, or intolerant of, ribavirin, peg-IFN can be considered. Its efficacy has been documented in patients with hematologic disorders, patients receiving hemodialysis, and in combination with ribavirin in patients with HIV[176-178]. Close monitoring is needed if it is used in transplant recipients because of an increased risk of acute humoral and cellular rejection[179,180]. Peg-IFN was thought to be safe only in liver transplant recipients until recent case reports described its successful use in a kidney transplant recipient[181-183].

Sofosbuvir is a nucleotide analog shown to decrease replication of HEV-3 *in vitro* [184]. However, in clinical studies, only modest antiviral activity was observed and SVR was not achieved[185-187]. Rescue treatment for patients who are not eligible for, or not responding to, ribavirin and/or peg-IFN remains an unmet need.

HOW TO REDUCE TRANSFUSION-TRANSMITTED HEPATITIS E

The background risk of foodborne HEV transmission to both donors and recipients of blood products is not negligible. The transfusion-related risk of infection only exceeds the annual dietary risk when more than 13 individual donor components are transfused[188]. Strategies to reduce *de novo* infection, such as modifying eating habits and eliminating HEV from pigs and other animals that are used for food production are essential[189]. The one available vaccine (HEV 239, Hecolin, Xiamen, China) is licensed only in China, and has yet to play a fundamental role in global outbreaks or pandemic control[190]. Nonetheless, the transmissibility and disease phenotype may not be the same for a person who acquires the virus orally and a person who gets infected intravenously, as there may be some protection provided by the acidic environment of the stomach and the mucosal barrier in the gut[191]. The infectivity of the non-enveloped form is different to that of enveloped HEV[9]. Data reporting

WJG | https://www.wjgnet.com

outcomes of recipients of HEV-infected blood products are sparse[47,49,50,60,64].

Policies on screening HEV in blood products differ between countries. Universal screening was adopted in the United Kingdom, Ireland, and the Netherlands. Germany and France implemented targeted screening of donated plasma intended for use in high-risk patients[192]. In Japan, the use of nucleic acid-based screening is limited to Hokkaido[193]. Blood donors are not routinely tested for HEV infection in China including Hong Kong[70,71,194]. There has been much debate on mandatory HEV screening in blood donations[195]. Key questions, such as whether or not to screen, which laboratory assay to use, which donors to screen (universal or selective screening), and which types of blood components to screen should be assessed based on risk assessment, resource availability, health economics, and political or other influences. The answers may vary considerably by geographical location[169,196]. In areas where HEV is highly endemic, most donors and/or recipients have probably been exposed to HEV previously and would have positive IgG antibodies. Therefore, the decision on serological screening should also take into consideration the prevalence of HEV infection in that particular region.

All donors should answer a questionnaire about symptoms of clinical hepatitis and potential exposure to HEV prior to blood donation. Donation should be deferred in any donors with a history of clinical hepatitis[197]. Neither alanine aminotransferase (ALT) nor anti-HEV IgM testing correlate with the presence of HEV RNA, supporting the use of NAT for screening of blood donations[60,61,105]. A simulation study by Kamp *et al*[198] reported that testing for HEV RNA by NAT with a pool size of 96, and a 95% limit of detection of 20 IU/mL will result in an 80% reduction in expected HEV transmissions as well as of consequent chronic infections with severe complications. The risk of transmission could be reduced by 90% in NAT using a mini-pool of 24 samples[198].

If opting for selective screening instead of universal screening, a clear definition of at-risk patients is warranted[199]. Targeted screening should be contemplated for blood components that will be supplied to transplant recipients, or patients with hematologic malignancies or chronic liver disease, as these individuals are at high risk of developing fulminant hepatitis, acute on chronic liver failure, or chronic hepatitis. However, it is not yet clear whether patients with rheumatologic diseases, those on low-intensity immunosuppression, or elderly individuals should only receive HEV-negative blood products. A multicenter retrospective study in Europe including 21 rheumatology and internal medicine patients found that patients with rheumatoid arthritis who were receiving methotrexate or biologics were at risk of chronic hepatitis E infection[200]. However, another study in France did not find worse hepatitis E severity or increased risk of chronicity in 23 patients with inflammatory arthritis treated with immunosuppressants[201].

Patients co-infected with HIV with CD4+ count < 200/mm³ are at risk for persistent HEV infection[29]. In HIV patients with low CD4+ count, anti-HEV IgG seroconversion was delayed until immune reconstitution occurred[202]. A recent meta-analysis found that the HEV RNA positivity rate was significantly higher in transplant recipients than in HIV-positive patients [1.2% (95%CI: 0.9-1.6) *vs* 0.39% (95%CI: 0.2-0.7); *P* = 0.0011], possibly due to better immune status in the HIV-positive individuals using anti-retroviral therapy[203].

HEV-1 and -2 infections can take a fulminant course in pregnancy, resulting in liver failure, membrane rupture, spontaneous abortions, and stillbirths[204]. HEV-3 infection in pregnancy appears to be less virulent without significant maternal, fetal, or neonatal complications[205-207]. During pregnancy, a reduced cellular immunity and a high level of steroid hormones, in particular estrogen, progesterone, and human chorionic gonadotropin, influence viral replication/expression and possibly explain the disease severity[208]. The immune response could be influenced by HEV genotype, translating into different outcomes[209]. Ribavirin and peg-IFN are contraindicated in pregnancy due to concerns of teratogenicity[210]. Further studies are needed to clarify the risk of transmission of HEV to pregnant women *via* blood transfusion; however, in view of the potentially serious disease course and absence of a safe treatment, pregnant women are a priority group for HEV-negative blood products.

Roth *et al*[67] evaluated the safety of plasma-derived medicinal products (PDMP) and found a very low prevalence of HEV RNA (0.002%) in plasma donors. Since viral reduction methods are used in the manufacturing processes of PDMP, these data do not support routine screening of all plasma pools intended for producing PDMP. Currently there is a lack of evidence to suggest that human serum albumin or coagulation factor concentrates are a major source of HEV infection[211,212].

The cost effectiveness of HEV screening of blood donations was analyzed in the Netherlands. Screening of whole blood donations in pools of 24 would prevent 4.52 of the 4.94 TT-HEV infections annually at a cost of approximately €310000 (Euro) per prevented chronic case. The estimated cost per incurable case prevented was 10-fold higher. Costs could potentially be reduced by 85% if only the blood products intended for use by immunocompromised patients were screened. Additional costs for selective screening may arise for logistic reasons and a possible increase in the number of blood products that expire before use. They concluded that preventing HEV transmission by screening of blood donations appears not excessively expensive compared with other blood-screening measures but the impact on disease burden may be small as only a minority of all HEV cases are transmitted by blood transfusion[213]. Another economic analysis performed in North America found a very low estimated risk of TT-HEV infection risk leading to severe liver disease. When compared with no screening, the costs were \$2.68 (USD) per component for a selective screening approach, and \$6.68 per component for universal screening. The respective costs per quality-adjusted life-year gained were \$225546 and \$561810, respectively, which exceeded the threshold for what is considered as "cost-effective" [66].

In addition to screening, various pathogen reduction methods have been proposed to reduce risk of TT-HEV. Solvent/detergent treatment could not eliminate nonenveloped HEV in plasma[214]. Non-enveloped HEV is also resistant to the Intercept method, which combines a synthetic psoralen amotosalen HCl treatment with ultraviolet A light illumination to block the replication of DNA and RNA[215]. However, substantial viral reduction has been demonstrated during the manufacturing process of plasma products using immunoaffinity chromatography, nanofiltration, cold ethanol fractionation and heat treatment[216]. Anti-HEV antibodies enhanced HEV removal by nanofiltration[217]. Furthermore, ultraviolet C light provided effective inactivation of HEV in platelet concentrates[218].

CONCLUSION

To conclude, TT-HEV is gaining attention worldwide. Although the overall prevalence of viremic blood donations is low, HEV can cause sinister consequences in immunocompromised recipients. Future studies are needed to define the incidence of transmission through transfusion, clinical features, outcomes, and prognosis. The decision on a screening policy in asymptomatic blood donors should be based on local risk assessment and health economics.

REFERENCES

- Khuroo MS. Chronic liver disease after non-A, non-B hepatitis. Lancet 1980; 2: 860-861 [PMID: 1 6107528]
- World Health Organization. Hepatitis E Fact sheet [DOI: 10.1097/grh.000000000000002]
- 3 Li P, Liu J, Li Y, Su J, Ma Z, Bramer WM, Cao W, de Man RA, Peppelenbosch MP, Pan Q. The global epidemiology of hepatitis E virus infection: A systematic review and meta-analysis. Liver Int 2020; 40: 1516-1528 [PMID: 32281721 DOI: 10.1111/liv.14468]
- 4 Wallace SJ, Swann R, Donnelly M, Kemp L, Guaci J, Murray A, Spoor J, Lin N, Miller M, Dalton HR, Hussaini SH, Gunson R, Simpson K, Stanley A, Fraser A. Mortality and morbidity of locally acquired hepatitis E in the national Scottish cohort: a multicentre retrospective study. Aliment Pharmacol Ther 2020; 51: 974-986 [PMID: 32285976 DOI: 10.1111/apt.15704]
- Bergløv A, Hallager S, Weis N. Hepatitis E during pregnancy: Maternal and foetal case-fatality rates and adverse outcomes-A systematic review. J Viral Hepat 2019; 26: 1240-1248 [PMID: 31095813 DOI: 10.1111/jvh.13129]
- Purdy MA, Harrison TJ, Jameel S, Meng XJ, Okamoto H, Van der Poel WHM, Smith DB; Ictv 6 Report Consortium. ICTV Virus Taxonomy Profile: Hepeviridae. J Gen Virol 2017; 98: 2645-2646 [PMID: 29022866 DOI: 10.1099/jgv.0.000940]
- 7 Primadharsini PP, Nagashima S, Okamoto H. Genetic Variability and Evolution of Hepatitis E Virus. Viruses 2019; 11 [PMID: 31109076 DOI: 10.3390/v11050456]
- 8 Sridhar S, Yip CC, Wu S, Chew NF, Leung KH, Chan JF, Zhao PS, Chan WM, Poon RW, Tsoi HW, Cai JP, Chan HS, Leung AW, Tse CW, Zee JS, Tsang OT, Cheng VC, Lau SK, Woo PC, Tsang DN, Yuen KY. Transmission of Rat Hepatitis E Virus Infection to Humans in Hong Kong: A Clinical and Epidemiological Analysis. Hepatology 2021; 73: 10-22 [PMID: 31960460 DOI: 10.1002/hep.31138]
- Yin X, Ambardekar C, Lu Y, Feng Z. Distinct Entry Mechanisms for Nonenveloped and Quasi-Enveloped Hepatitis E Viruses. J Virol 2016; 90: 4232-4242 [PMID: 26865708 DOI:



10.1128/JVI.02804-15]

- 10 Himmelsbach K, Bender D, Hildt E. Life cycle and morphogenesis of the hepatitis E virus. Emerg Microbes Infect 2018; 7: 196 [PMID: 30498191 DOI: 10.1038/s41426-018-0198-7]
- Kenney SP, Meng XJ. Hepatitis E Virus Genome Structure and Replication Strategy. Cold Spring 11 Harb Perspect Med 2019; 9 [PMID: 29530948 DOI: 10.1101/cshperspect.a031724]
- 12 Kmush B, Wierzba T, Krain L, Nelson K, Labrique AB. Epidemiology of hepatitis E in low- and middle-income countries of Asia and Africa. Semin Liver Dis 2013; 33: 15-29 [PMID: 23564386 DOI: 10.1055/s-0033-1338111]
- 13 Khuroo MS, Khuroo NS. Hepatitis E: Discovery, global impact, control and cure. World J Gastroenterol 2016; 22: 7030-7045 [PMID: 27610014 DOI: 10.3748/wjg.v22.i31.7030]
- 14 Rein DB, Stevens GA, Theaker J, Wittenborn JS, Wiersma ST. The global burden of hepatitis E virus genotypes 1 and 2 in 2005. Hepatology 2012; 55: 988-997 [PMID: 22121109 DOI: 10.1002/hep.25505]
- Hakze-van der Honing RW, van Coillie E, Antonis AF, van der Poel WH. First isolation of 15 hepatitis E virus genotype 4 in Europe through swine surveillance in the Netherlands and Belgium. PLoS One 2011; 6: e22673 [PMID: 21829641 DOI: 10.1371/journal.pone.0022673]
- 16 Garbuglia AR, Scognamiglio P, Petrosillo N, Mastroianni CM, Sordillo P, Gentile D, La Scala P, Girardi E, Capobianchi MR. Hepatitis E virus genotype 4 outbreak, Italy, 2011. Emerg Infect Dis 2013; 19: 110-114 [PMID: 23260079 DOI: 10.3201/eid1901.120983]
- Donnelly MC, Scobie L, Crossan CL, Dalton H, Hayes PC, Simpson KJ. Review article: hepatitis E-17 a concise review of virology, epidemiology, clinical presentation and therapy. Aliment Pharmacol Ther 2017; 46: 126-141 [PMID: 28449246 DOI: 10.1111/apt.14109]
- 18 Goel A, Aggarwal R. Hepatitis E: Epidemiology, Clinical Course, Prevention, and Treatment. Gastroenterol Clin North Am 2020; 49: 315-330 [PMID: 32389365 DOI: 10.1016/j.gtc.2020.01.011]
- 19 Kamar N, Izopet J, Pavio N, Aggarwal R, Labrique A, Wedemeyer H, Dalton HR. Hepatitis E virus infection. Nat Rev Dis Primers 2017; 3: 17086 [PMID: 29154369 DOI: 10.1038/nrdp.2017.86]
- 20 Dalton HR, Stableforth W, Thurairajah P, Hazeldine S, Remnarace R, Usama W, Farrington L, Hamad N, Sieberhagen C, Ellis V, Mitchell J, Hussaini SH, Banks M, Ijaz S, Bendall RP. Autochthonous hepatitis E in Southwest England: natural history, complications and seasonal variation, and hepatitis E virus IgG seroprevalence in blood donors, the elderly and patients with chronic liver disease. Eur J Gastroenterol Hepatol 2008; 20: 784-790 [PMID: 18617784 DOI: 10.1097/MEG.0b013e3282f5195a
- 21 Lhomme S, Marion O, Abravanel F, Izopet J, Kamar N. Clinical Manifestations, Pathogenesis and Treatment of Hepatitis E Virus Infections. J Clin Med 2020; 9 [PMID: 31991629 DOI: 10.3390/icm90203311
- 22 Haffar S, Shalimar, Kaur RJ, Wang Z, Prokop LJ, Murad MH, Bazerbachi F. Acute liver failure caused by hepatitis E virus genotype 3 and 4: A systematic review and pooled analysis. Liver Int 2018; 38: 1965-1973 [PMID: 29675889 DOI: 10.1111/liv.13861]
- 23 Smith DB, Simmonds P. Hepatitis E virus and fulminant hepatitis--a virus or host-specific pathology? Liver Int 2015; 35: 1334-1340 [PMID: 24974734 DOI: 10.1111/liv.12629]
- Frias M, López-López P, Rivero A, Rivero-Juarez A. Role of Hepatitis E Virus Infection in Acute-24 on-Chronic Liver Failure. Biomed Res Int 2018; 2018: 9098535 [PMID: 30050945 DOI: 10.1155/2018/9098535
- 25 Radha Krishna Y, Saraswat VA, Das K, Himanshu G, Yachha SK, Aggarwal R, Choudhuri G. Clinical features and predictors of outcome in acute hepatitis A and hepatitis E virus hepatitis on cirrhosis. Liver Int 2009; 29: 392-398 [PMID: 19267864 DOI: 10.1111/j.1478-3231.2008.01887.x]
- Kamar N, Rostaing L, Legrand-Abravanel F, Izopet J. How should hepatitis E virus infection be 26 defined in organ-transplant recipients? Am J Transplant 2013; 13: 1935-1936 [PMID: 23659713 DOI: 10.1111/ajt.12253]
- Pischke S, Stiefel P, Franz B, Bremer B, Suneetha PV, Heim A, Ganzenmueller T, Schlue J, Horn-27 Wichmann R, Raupach R, Darnedde M, Scheibner Y, Taubert R, Haverich A, Manns MP. Wedemeyer H, Bara CL. Chronic hepatitis e in heart transplant recipients. Am J Transplant 2012; 12: 3128-3133 [PMID: 22823202 DOI: 10.1111/j.1600-6143.2012.04200.x]
- Ollier L, Tieulie N, Sanderson F, Heudier P, Giordanengo V, Fuzibet JG, Nicand E. Chronic 28 hepatitis after hepatitis E virus infection in a patient with non-Hodgkin lymphoma taking rituximab. Ann Intern Med 2009; 150: 430-431 [PMID: 19293084 DOI: 10.7326/0003-4819-150-6-200903170-00026
- 29 Dalton HR, Bendall RP, Keane FE, Tedder RS, Ijaz S. Persistent carriage of hepatitis E virus in patients with HIV infection. N Engl J Med 2009; 361: 1025-1027 [PMID: 19726781 DOI: 10.1056/NEJMc0903778
- Sridhar S, Chan JFW, Yap DYH, Teng JLL, Huang C, Yip CCY, Hung IFN, Tang SCW, Lau SKP, 30 Woo PCY, Yuen KY. Genotype 4 hepatitis E virus is a cause of chronic hepatitis in renal transplant recipients in Hong Kong. J Viral Hepat 2018; 25: 209-213 [PMID: 28984015 DOI: 10.1111/jvh.12799]
- 31 Wang Y, Chen G, Pan Q, Zhao J. Chronic Hepatitis E in a Renal Transplant Recipient: The First Report of Genotype 4 Hepatitis E Virus Caused Chronic Infection in Organ Recipient. Gastroenterology 2018; 154: 1199-1201 [PMID: 29432746 DOI: 10.1053/j.gastro.2017.12.028]
- Owada Y, Oshiro Y, Inagaki Y, Harada H, Fujiyama N, Kawagishi N, Yagisawa T, Usui J, Akutsu 32 N, Itabashi Y, Saito K, Watarai Y, Ichimaru N, Imamura R, Kyakuno M, Ide K, Shibuya Y, Okabe



Y, Ono M, Sasaki K, Shiose A, Yamagishi K, Ohnishi H, Nagashima S, Takahashi M, Yuzawa K, Okamoto H, Ohkohchi N. A Nationwide Survey of Hepatitis E Virus Infection and Chronic Hepatitis in Heart and Kidney Transplant Recipients in Japan. Transplantation 2020; 104: 437-444 [PMID: 31205267 DOI: 10.1097/TP.000000000002801]

- 33 Zhou X, de Man RA, de Knegt RJ, Metselaar HJ, Peppelenbosch MP, Pan Q. Epidemiology and management of chronic hepatitis E infection in solid organ transplantation: a comprehensive literature review. Rev Med Virol 2013; 23: 295-304 [PMID: 23813631 DOI: 10.1002/rmv.1751]
- 34 Mirazo S, Arbiza J. Hepatitis E and chronic liver damage in apparently immunocompetent individuals: Now what? Ann Hepatol 2019; 18: 539-540 [PMID: 31130468 DOI: 10.1016/j.aohep.2019.05.002]
- 35 Protzer U, Böhm F, Longerich T, Seebach J, Heidary Navid M, Friemel J, Marques-Maggio E, Bawohl M, Heikenwalder M, Schirmacher P, Dutkowski P, Clavien PA, Schemmer P, Schnitzler P, Gotthardt D, Müllhaupt B, Weber A. Molecular detection of hepatitis E virus (HEV) in liver biopsies after liver transplantation. Mod Pathol 2015; 28: 523-532 [PMID: 25412844 DOI: 10.1038/modpathol.2014.147]
- Suneetha PV, Pischke S, Schlaphoff V, Grabowski J, Fytili P, Gronert A, Bremer B, Markova A, 36 Jaroszewicz J, Bara C, Manns MP, Cornberg M, Wedemeyer H. Hepatitis E virus (HEV)-specific Tcell responses are associated with control of HEV infection. Hepatology 2012; 55: 695-708 [PMID: 22006345 DOI: 10.1002/hep.24738]
- 37 Borentain P, Colson P, Bolon E, Gauchez P, Coso D, Gérolami R. Hepatocellular carcinoma complicating hepatitis E virus-related cirrhosis. Hepatology 2018; 67: 446-448 [PMID: 28873236 DOI: 10.1002/hep.29508]
- 38 Pischke S, Hartl J, Pas SD, Lohse AW, Jacobs BC, Van der Eijk AA. Hepatitis E virus: Infection beyond the liver? J Hepatol 2017; 66: 1082-1095 [PMID: 27913223 DOI: 10.1016/j.jhep.2016.11.016]
- Fousekis FS, Mitselos IV, Christodoulou DK. Extrahepatic manifestations of hepatitis E virus: An 39 overview. Clin Mol Hepatol 2020; 26: 16-23 [PMID: 31601068 DOI: 10.3350/cmh.2019.0082]
- Kamar N, Weclawiak H, Guilbeau-Frugier C, Legrand-Abravanel F, Cointault O, Ribes D, Esposito 40 L, Cardeau-Desangles I, Guitard J, Sallusto F, Muscari F, Peron JM, Alric L, Izopet J, Rostaing L. Hepatitis E virus and the kidney in solid-organ transplant patients. Transplantation 2012; 93: 617-623 [PMID: 22298032 DOI: 10.1097/TP.0b013e318245f14c]
- 41 Noble J, Jouve T, Malvezzi P, Rostaing L. Renal complications of liver diseases. Expert Rev Gastroenterol Hepatol 2018; 12: 1135-1142 [PMID: 30269605 DOI: 10.1080/17474124.2018.1530984]
- 42 Kamar N, Marion O, Abravanel F, Izopet J, Dalton HR. Extrahepatic manifestations of hepatitis E virus. Liver Int 2016; 36: 467-472 [PMID: 27005692 DOI: 10.1111/liv.13037]
- Liu H, Ma Y. Hepatitis E virus-associated Guillain-Barre syndrome: Revision of the literature. 43 Brain Behav 2020; 10: e01496 [PMID: 31828968 DOI: 10.1002/brb3.1496]
- Ripellino P, Pasi E, Melli G, Staedler C, Fraga M, Moradpour D, Sahli R, Aubert V, Martinetti G, Bihl F, Bernasconi E, Terziroli Beretta-Piccoli B, Cerny A, Dalton HR, Zehnder C, Mathis B, Zecca C, Disanto G, Kaelin-Lang A, Gobbi C. Neurologic complications of acute hepatitis E virus infection. Neurol Neuroimmunol Neuroinflamm 2020; 7 [PMID: 31806684 DOI: 10.1212/NXI.00000000000643]
- 45 Fischer C, Hofmann M, Danzer M, Hofer K, Kaar J, Gabriel C. Seroprevalence and Incidence of hepatitis E in blood donors in Upper Austria. PLoS One 2015; 10: e0119576 [PMID: 25751574 DOI: 10.1371/journal.pone.0119576]
- 46 Vercouter AS, Van Houtte F, Verhoye L, González Fraile I, Blanco L, Compernolle V, Meuleman P. Hepatitis E virus prevalence in Flemish blood donors. J Viral Hepat 2019; 26: 1218-1223 [PMID: 31194897 DOI: 10.1111/jvh.13161]
- 47 Harritshøj LH, Holm DK, Saekmose SG, Jensen BA, Hogema BM, Fischer TK, Midgley SE, Krog JS, Erikstrup C, Ullum H. Low transfusion transmission of hepatitis E among 25,637 singledonation, nucleic acid-tested blood donors. Transfusion 2016; 56: 2225-2232 [PMID: 27385646 DOI: 10.1111/trf.137001
- Gallian P, Lhomme S, Piquet Y, Sauné K, Abravanel F, Assal A, Tiberghien P, Izopet J. Hepatitis E 48 virus infections in blood donors, France. Emerg Infect Dis 2014; 20: 1914-1917 [PMID: 25340881 DOI: 10.3201/eid2011.140516]
- 49 Westhölter D, Hiller J, Denzer U, Polywka S, Ayuk F, Rybczynski M, Horvatits T, Gundlach S, Blöcker J, Schulze Zur Wiesch J, Fischer N, Addo MM, Peine S, Göke B, Lohse AW, Lütgehetmann M, Pischke S. HEV-positive blood donations represent a relevant infection risk for immunosuppressed recipients. J Hepatol 2018; 69: 36-42 [PMID: 29551705 DOI: 10.1016/j.jhep.2018.02.031]
- 50 Dreier J, Knabbe C, Vollmer T. Transfusion-Transmitted Hepatitis E: NAT Screening of Blood Donations and Infectious Dose. Front Med (Lausanne) 2018; 5: 5 [PMID: 29450199 DOI: 10.3389/fmed.2018.00005
- 51 Corman VM, Drexler JF, Eckerle I, Roth WK, Drosten C, Eis-Hübinger AM. Zoonotic hepatitis E virus strains in German blood donors. Vox Sang 2013; 104: 179-180 [PMID: 22913247 DOI: 10.1111/j.1423-0410.2012.01638.x]
- 52 Vollmer T, Diekmann J, Johne R, Eberhardt M, Knabbe C, Dreier J. Novel approach for detection of hepatitis E virus infection in German blood donors. J Clin Microbiol 2012; 50: 2708-2713



[PMID: 22675127 DOI: 10.1128/JCM.01119-12]

- 53 Baylis SA, Gärtner T, Nick S, Ovemyr J, Blümel J. Occurrence of hepatitis E virus RNA in plasma donations from Sweden, Germany and the United States. Vox Sang 2012; 103: 89-90 [PMID: 22220775 DOI: 10.1111/j.1423-0410.2011.01583.x]
- 54 O'Riordan J, Boland F, Williams P, Donnellan J, Hogema BM, Ijaz S, Murphy WG. Hepatitis E virus infection in the Irish blood donor population. Transfusion 2016; 56: 2868-2876 [PMID: 27522065 DOI: 10.1111/trf.13757]
- 55 Spreafico M, Raffaele L, Guarnori I, Foglieni B, Berzuini A, Valenti L, Gerosa A, Colli A, Prati D. Prevalence and 9-year incidence of hepatitis E virus infection among North Italian blood donors: Estimated transfusion risk. J Viral Hepat 2020; 27: 858-861 [PMID: 32196831 DOI: 10.1111/jvh.13296]
- 56 Spada E, Pupella S, Pisani G, Bruni R, Chionne P, Madonna E, Villano U, Simeoni M, Fabi S, Marano G, Marcantonio C, Pezzotti P, Ciccaglione AR, Liumbruno GM. A nationwide retrospective study on prevalence of hepatitis E virus infection in Italian blood donors. Blood Transfus 2018; 16: 413-421 [PMID: 29757135 DOI: 10.2450/2018.0033-18]
- Hogema BM, Molier M, Sjerps M, de Waal M, van Swieten P, van de Laar T, Molenaar-de Backer 57 M, Zaaijer HL. Incidence and duration of hepatitis E virus infection in Dutch blood donors. Transfusion 2016; 56: 722-728 [PMID: 26559806 DOI: 10.1111/trf.13402]
- Slot E, Hogema BM, Riezebos-Brilman A, Kok TM, Molier M, Zaaijer HL. Silent hepatitis E virus 58 infection in Dutch blood donors, 2011 to 2012. Euro Surveill 2013; 18 [PMID: 23929229 DOI: 10.2807/1560-7917.es2013.18.31.20550
- 59 Grabarczyk P, Sulkowska E, Gdowska J, Kopacz A, Liszewski G, Kubicka-Russel D, Baylis SA, Corman VM, Noceń E, Piotrowski D, Antoniewicz-Papis J, Łętowska M. Molecular and serological infection marker screening in blood donors indicates high endemicity of hepatitis E virus in Poland. Transfusion 2018; 58: 1245-1253 [PMID: 29492976 DOI: 10.1111/trf.14531]
- 60 Rivero-Juarez A, Jarilla-Fernandez M, Frias M, Madrigal-Sanchez E, López-López P, Andújar-Troncoso G, Machuca I, Camacho A, Muñoz-Valbuena P, Rivero A. Hepatitis E virus in Spanish donors and the necessity for screening. J Viral Hepat 2019; 26: 603-608 [PMID: 30661278 DOI: 10.1111/jvh.13064]
- 61 Sauleda S, Ong E, Bes M, Janssen A, Cory R, Babizki M, Shin T, Lindquist A, Hoang A, Vang L, Piron M, Casamitjana N, Koppelman M, Danzig L, Linnen JM. Seroprevalence of hepatitis E virus (HEV) and detection of HEV RNA with a transcription-mediated amplification assay in blood donors from Catalonia (Spain). Transfusion 2015; 55: 972-979 [PMID: 25403913 DOI: 10.1111/trf.12929]
- 62 Harvala H, Hewitt PE, Reynolds C, Pearson C, Haywood B, Tettmar KI, Ushiro-Lumb I, Brailsford SR, Tedder R, Ijaz S. Hepatitis E virus in blood donors in England, 2016 to 2017: from selective to universal screening. Euro Surveill 2019; 24 [PMID: 30862338 DOI: 10.2807/1560-7917.ES.2019.24.10.1800386]
- 63 Thom K, Gilhooly P, McGowan K, Malloy K, Jarvis LM, Crossan C, Scobie L, Blatchford O, Smith-Palmer A, Donnelly MC, Davidson JS, Johannessen I, Simpson KJ, Dalton HR, Petrik J. Hepatitis E virus (HEV) in Scotland: evidence of recent increase in viral circulation in humans. Euro Surveill 2018; 23 [PMID: 29589577 DOI: 10.2807/1560-7917.ES.2018.23.12.17-00174]
- Hewitt PE, Ijaz S, Brailsford SR, Brett R, Dicks S, Haywood B, Kennedy IT, Kitchen A, Patel P, 64 Poh J, Russell K, Tettmar KI, Tossell J, Ushiro-Lumb I, Tedder RS. Hepatitis E virus in blood components: a prevalence and transmission study in southeast England. Lancet 2014; 384: 1766-1773 [PMID: 25078306 DOI: 10.1016/S0140-6736(14)61034-5]
- 65 Cleland A. Smith L. Crossan C. Blatchford O. Dalton HR. Scobie L. Petrik J. Hepatitis E virus in Scottish blood donors. Vox Sang 2013; 105: 283-289 [PMID: 23763589 DOI: 10.1111/vox.12056]
- 66 Delage G, Fearon M, Gregoire Y, Hogema BM, Custer B, Scalia V, Hawes G, Bernier F, Nguyen ML, Stramer SL. Hepatitis E Virus Infection in Blood Donors and Risk to Patients in the United States and Canada. Transfus Med Rev 2019; 33: 139-145 [PMID: 31324552 DOI: 10.1016/j.tmrv.2019.05.017]
- Roth NJ, Schäfer W, Alexander R, Elliott K, Elliott-Browne W, Knowles J, Wenzel JJ, Simon TL. 67 Low hepatitis E virus RNA prevalence in a large-scale survey of United States source plasma donors. Transfusion 2017; 57: 2958-2964 [PMID: 28833188 DOI: 10.1111/trf.14285]
- 68 Stramer SL, Moritz ED, Foster GA, Ong E, Linnen JM, Hogema BM, Mak M, Chia CP, Dodd RY. Hepatitis E virus: seroprevalence and frequency of viral RNA detection among US blood donors. Transfusion 2016; 56: 481-488 [PMID: 26434952 DOI: 10.1111/trf.13355]
- Xu C, Wang RY, Schechterly CA, Ge S, Shih JW, Xia NS, Luban NL, Alter HJ. An assessment of hepatitis E virus (HEV) in US blood donors and recipients: no detectable HEV RNA in 1939 donors tested and no evidence for HEV transmission to 362 prospectively followed recipients. Transfusion 2013; 53: 2505-2511 [PMID: 23829163 DOI: 10.1111/trf.12326]
- 70 Wen GP, Chen CR, Song XY, Tang ZM, Ji WF, Wang SL, Zhang K, Zhang J, Ou SH, Zheng ZZ, Xia NS. Long-term HEV carriers without antibody seroconversion among eligible immunocompetent blood donors. Emerg Microbes Infect 2018; 7: 125 [PMID: 29977038 DOI: 10.1038/s41426-018-0125-y
- 71 Tsoi WC, Zhu X, To AP, Holmberg J. Hepatitis E virus infection in Hong Kong blood donors. Vox Sang 2020; 115: 11-17 [PMID: 31709559 DOI: 10.1111/vox.12846]
- 72 Katiyar H, Goel A, Sonker A, Yadav V, Sapun S, Chaudhary R, Aggarwal R. Prevalence of



hepatitis E virus viremia and antibodies among healthy blood donors in India. Indian J Gastroenterol 2018: 37: 342-346 [PMID: 30159666 DOI: 10.1007/s12664-018-0880-7]

- 73 Minagi T, Okamoto H, Ikegawa M, Ideno S, Takahashi K, Sakai K, Hagiwara K, Yunoki M, Wakisaka A. Hepatitis E virus in donor plasma collected in Japan. Vox Sang 2016; 111: 242-246 [PMID: 27280485 DOI: 10.1111/vox.12425]
- 74 Intharasongkroh D, Thongmee T, Sa-Nguanmoo P, Klinfueng S, Duang-In A, Wasitthankasem R, Theamboonlers A, Charoonruangrit U, Oota S, Payungporn S, Vongpunsawad S, Chirathaworn C, Poovorawan Y. Hepatitis E virus infection in Thai blood donors. Transfusion 2019; 59: 1035-1043 [PMID: 30443992 DOI: 10.1111/trf.15041]
- 75 Hoad VC, Seed CR, Fryk JJ, Harley R, Flower RLP, Hogema BM, Kiely P, Faddy HM. Hepatitis E virus RNA in Australian blood donors: prevalence and risk assessment. Vox Sang 2017; 112: 614-621 [PMID: 28833229 DOI: 10.1111/vox.12559]
- 76 Shrestha AC, Flower RL, Seed CR, Keller AJ, Harley R, Chan HT, Hoad V, Warrilow D, Northill J, Holmberg JA, Faddy HM. Hepatitis E virus RNA in Australian blood donations. Transfusion 2016; 56: 3086-3093 [PMID: 27667133 DOI: 10.1111/trf.13799]
- 77 Hewitt J, Harte D, Sutherland M, Croucher D, Fouche L, Flanagan P, Williamson D. Prevalence of hepatitis E virus antibodies and infection in New Zealand blood donors. N Z Med J 2018; 131: 38-43 [PMID: 29389927]
- 78 Maponga TG, Lopes T, Cable R, Pistorius C, Preiser W, Andersson MI. Prevalence and risks of hepatitis E virus infection in blood donors from the Western Cape, South Africa. Vox Sang 2020; 115: 695-702 [PMID: 32597542 DOI: 10.1111/vox.12966]
- Wang M, Fu P, Yin Y, He M, Liu Y. Acute, Recent and Past HEV Infection among Voluntary 79 Blood Donors in China: A Systematic Review and Meta-Analysis. PLoS One 2016; 11: e0161089 [PMID: 27597991 DOI: 10.1371/journal.pone.0161089]
- Di Cola G, Fantilli AC, Pisano MB, Ré VE. Foodborne transmission of hepatitis A and hepatitis E 80 viruses: A literature review. Int J Food Microbiol 2021; 338: 108986 [PMID: 33257099 DOI: 10.1016/i.iifoodmicro.2020.108986
- 81 Kamar N, Bendall R, Legrand-Abravanel F, Xia NS, Ijaz S, Izopet J, Dalton HR. Hepatitis E. Lancet 2012; **379**: 2477-2488 [PMID: 22549046 DOI: 10.1016/S0140-6736(11)61849-7]
- Pérez-Gracia MT, Suay B, Mateos-Lindemann ML. Hepatitis E: an emerging disease. Infect Genet 82 Evol 2014; 22: 40-59 [PMID: 24434240 DOI: 10.1016/j.meegid.2014.01.002]
- Gupta N, Sarangi AN, Dadhich S, Dixit VK, Chetri K, Goel A, Aggarwal R. Acute hepatitis E in 83 India appears to be caused exclusively by genotype 1 hepatitis E virus. Indian J Gastroenterol 2018; **37**: 44-49 [PMID: 29399748 DOI: 10.1007/s12664-018-0819-z]
- 84 Mevis FM, Sabeena S, Sanjay R, Robin S, Devadiga S, Prasad V, Oliver D, Ameen A, Arunkumar G. Currently circulating genotypes of hepatitis E virus in India, 2014-2018. Indian J Med Microbiol 2019; 37: 563-568 [PMID: 32436881 DOI: 10.4103/ijmm.IJMM_19_449]
- 85 Gallian P, Couchouron A, Dupont I, Fabra C, Piquet Y, Djoudi R, Assal A, Tiberghien P. Comparison of hepatitis E virus nucleic acid test screening platforms and RNA prevalence in French blood donors. Transfusion 2017; 57: 223-224 [PMID: 28097700 DOI: 10.1111/trf.13889]
- Vollmer T, Diekmann J, Knabbe C, Dreier J. Hepatitis E virus blood donor NAT screening: as much 86 as possible or as much as needed? Transfusion 2019; 59: 612-622 [PMID: 30548866 DOI: 10.1111/trf.15058]
- 87 Miletić M, Vuk T, Hećimović A, Stojić Vidović M, Jemeršić L, Jukić I. Estimation of the hepatitis E assay-dependent seroprevalence among Croatian blood donors. Transfus Clin Biol 2019; 26: 229-233 [PMID: 31277986 DOI: 10.1016/j.tracli.2019.06.234]
- Holm DK, Moessner BK, Engle RE, Zaaijer HL, Georgsen J, Purcell RH, Christensen PB. Declining 88 prevalence of hepatitis E antibodies among Danish blood donors. Transfusion 2015; 55: 1662-1667 [PMID: 25819381 DOI: 10.1111/trf.13028]
- 89 Dimeglio C, Beau F, Broult J, Gouy P, Izopet J, Lastère S, Abravanel F. Hepatitis E prevalence in French Polynesian blood donors. PLoS One 2018; 13: e0208934 [PMID: 30532225 DOI: 10.1371/journal.pone.0208934]
- 90 Juhl D, Baylis SA, Blümel J, Görg S, Hennig H. Seroprevalence and incidence of hepatitis E virus infection in German blood donors. Transfusion 2014; 54: 49-56 [PMID: 23441647 DOI: 10.1111/trf.12121
- Dalekos GN, Zervou E, Elisaf M, Germanos N, Galanakis E, Bourantas K, Siamopoulos KC, 91 Tsianos EV. Antibodies to hepatitis E virus among several populations in Greece: increased prevalence in an hemodialysis unit. Transfusion 1998; 38: 589-595 [PMID: 9661693 DOI: 10.1046/j.1537-2995.1998.38698326339.x]
- 92 De Sabato L, Di Bartolo I, Montomoli E, Trombetta C, Ruggeri FM, Ostanello F. Retrospective Study Evaluating Seroprevalence of Hepatitis E Virus in Blood Donors and in Swine Veterinarians in Italy (2004). Zoonoses Public Health 2017; 64: 308-312 [PMID: 27911040 DOI: 10.1111/zph.12332]
- 93 Lucarelli C, Spada E, Taliani G, Chionne P, Madonna E, Marcantonio C, Pezzotti P, Bruni R, La Rosa G, Pisani G, Dell'Orso L, Ragone K, Tomei C, Ciccaglione AR. High prevalence of antihepatitis E virus antibodies among blood donors in central Italy, February to March 2014. Euro Surveill 2016; 21 [PMID: 27494608 DOI: 10.2807/1560-7917.ES.2016.21.30.30299]
- Puttini C, Riccio ML, Redi D, Tordini G, Cenerini M, Romanello F, De Luca A, Carmellini M, 94 Fossombroni V, Cusi MG, Zanelli G. Seroprevalence of hepatitis E virus (HEV) infection in blood



donors and renal transplant recipients: a retrospective study from central Italy. Infez Med 2015; 23: 253-256 [PMID: 26397295]

- 95 Hogema BM, Molier M, Slot E, Zaaijer HL. Past and present of hepatitis E in the Netherlands. Transfusion 2014; 54: 3092-3096 [PMID: 24889277 DOI: 10.1111/trf.12733]
- 96 Mateos ML, Camarero C, Lasa E, Teruel JL, Mir N, Baquero F. Hepatitis E virus: relevance in blood donors and risk groups. Vox Sang 1999; 76: 78-80 [PMID: 10085522 DOI: 10.1159/000031024]
- 97 Niederhauser C, Widmer N, Hotz M, Tinguely C, Fontana S, Allemann G, Borri M, Infanti L, Sarraj A, Sigle J, Stalder M, Thierbach J, Waldvogel S, Wiengand T, Züger M, Gowland P. Current hepatitis E virus seroprevalence in Swiss blood donors and apparent decline from 1997 to 2016. Euro Surveill 2018; 23 [PMID: 30180927 DOI: 10.2807/1560-7917.ES.2018.23.35.1700616]
- 98 Kaufmann A, Kenfak-Foguena A, André C, Canellini G, Bürgisser P, Moradpour D, Darling KE, Cavassini M. Hepatitis E virus seroprevalence among blood donors in southwest Switzerland. PLoS One 2011; 6: e21150 [PMID: 21701586 DOI: 10.1371/journal.pone.0021150]
- Beale MA, Tettmar K, Szypulska R, Tedder RS, Ijaz S. Is there evidence of recent hepatitis E virus 99 infection in English and North Welsh blood donors? Vox Sang 2011; 100: 340-342 [PMID: 21392024 DOI: 10.1111/j.1423-0410.2010.01412.x]
- 100 Zafrullah M, Zhang X, Tran C, Nguyen M, Kamili S, Purdy MA, Stramer SL. Disparities in detection of antibodies against hepatitis E virus in US blood donor samples using commercial assays. Transfusion 2018; 58: 1254-1263 [PMID: 29520800 DOI: 10.1111/trf.14553]
- Di Lello FA, Blejer J, Alter A, Bartoli S, Vargas F, Ruiz R, Galli C, Blanco S, Carrizo LH, Gallego 101 S, Fernández R, Martínez AP, Flichman DM. Seroprevalence of hepatitis E virus in Argentinean blood donors. Eur J Gastroenterol Hepatol 2020 [DOI: 10.1097/meg.00000000001853]
- 102 Bangueses F, Abin-Carriquiry JA, Cancela F, Curbelo J, Mirazo S. Serological and molecular prevalence of hepatitis E virus among blood donors from Uruguay. J Med Virol 2021; 93: 4010-4014 [PMID: 32592500 DOI: 10.1002/jmv.26231]
- 103 Nouhin J, Prak S, Madec Y, Barennes H, Weissel R, Hok K, Pavio N, Rouet F. Hepatitis E virus antibody prevalence, RNA frequency, and genotype among blood donors in Cambodia (Southeast Asia). Transfusion 2016; 56: 2597-2601 [PMID: 27480100 DOI: 10.1111/trf.13731]
- 104 Chen X, Gong P, Wagner AL, Li Y, Wang G, Lu Y. Identification of hepatitis E virus subtype 4f in blood donors in Shanghai, China. Virus Res 2019; 265: 30-33 [PMID: 30836112 DOI: 10.1016/j.virusres.2019.03.001
- 105 Wang M, He M, Wu B, Ke L, Han T, Wang J, Shan H, Ness P, Guo N, Liu Y, Nelson KE. The association of elevated alanine aminotransferase levels with hepatitis E virus infections among blood donors in China. Transfusion 2017; 57: 273-279 [PMID: 28194856 DOI: 10.1111/trf.13991]
- Ma L, Sun P, Lin F, Wang H, Rong X, Dai Y, Liu J, Qian L, Fang M, Su N, Xiao W, Ye S, Li C. 106 Prevalence of hepatitis E virus in Chinese blood donors. J Int Med Res 2015; 43: 257-262 [PMID: 25710945 DOI: 10.1177/0300060514562054]
- 107 Ren F, Zhao C, Wang L, Wang Z, Gong X, Song M, Zhuang H, Huang Y, Shan H, Wang J, Liu Q, Ness P, Nelson KE, Wang Y. Hepatitis E virus seroprevalence and molecular study among blood donors in China. Transfusion 2014; 54: 910-917 [PMID: 24372259 DOI: 10.1111/trf.12530]
- 108 Zhuang W, Ding X, Lyu C, Xiang L, Teng H, Li J. Hepatitis E virus seroprevalence among blood donors in Jiangsu Province, East China. Int J Infect Dis 2014; 26: 9-11 [PMID: 24981426 DOI: 10.1016/j.ijid.2014.04.022]
- 109 Tripathy AS, Puranik S, Sharma M, Chakraborty S, Devakate UR. Hepatitis E virus seroprevalence among blood donors in Pune, India. J Med Virol 2019; 91: 813-819 [PMID: 30489644 DOI: 10.1002/jmv.25370]
- Gajjar MD, Bhatnagar NM, Sonani RV, Gupta S, Patel T. Hepatitis E seroprevalence among blood 110 donors: A pilot study from Western India. Asian J Transfus Sci 2014; 8: 29-31 [PMID: 24678170 DOI: 10.4103/0973-6247.126685]
- 111 Parsa R, Adibzadeh S, Behzad Behbahani A, Farhadi A, Yaghobi R, Rafiei Dehbidi GR, Haijzamani S, Rahbar S, Nikouvan N, Okhovat MA, Naderi S, Salehi S, Alizadeh M, Ranibaran R, Zarnegar G, Alavi P. Detection of Hepatitis E Virus Genotype 1 Among Blood Donors From Southwest of Iran. Hepat Mon 2016; 16: e34202 [PMID: 27630719 DOI: 10.5812/hepatmon.34202]
- Hesamizadeh K, Sharafi H, Keyvani H, Alavian SM, Najafi-Tireh Shabankareh A, Sharifi Olyaie R, 112 Keshvari M. Hepatitis A Virus and Hepatitis E Virus Seroprevalence Among Blood Donors in Tehran, Iran. Hepat Mon 2016; 16: e32215 [PMID: 27110256 DOI: 10.5812/hepatmon.32215]
- 113 Naeimi B, Mazloom Kalimani F, Pourfatolah AA, Azimzadeh M, Mankhian A, Akbarzadeh S, Hajiani G, Kooshesh F, Khamisipour G. Hepatitis E Virus Seroprevalence Among Blood Donors in Bushehr, South of Iran. Hepat Mon 2015; 15: e29219 [PMID: 26834784 DOI: 10.5812/hepatmon.29219
- Ehteram H, Ramezani A, Eslamifar A, Sofian M, Banifazl M, Ghassemi S, Aghakhani A, 114 Mashayekhi P. Seroprevalence of Hepatitis E Virus infection among volunteer blood donors in central province of Iran in 2012. Iran J Microbiol 2013; 5: 172-176 [PMID: 23825737]
- Taremi M, Gachkar L, MahmoudArabi S, Kheradpezhouh M, Khoshbaten M. Prevalence of 115 antibodies to hepatitis E virus among male blood donors in Tabriz, Islamic Republic of Iran. East Mediterr Health J 2007; 13: 98-102 [PMID: 17546911]
- 116 Takeda H, Matsubayashi K, Sakata H, Sato S, Kato T, Hino S, Tadokoro K, Ikeda H. A nationwide survey for prevalence of hepatitis E virus antibody in qualified blood donors in Japan. Vox Sang



2010; 99: 307-313 [PMID: 20576022 DOI: 10.1111/j.1423-0410.2010.01362.x]

- Shrestha AC, Flower RL, Seed CR, Rajkarnikar M, Shrestha SK, Thapa U, Hoad VC, Faddy HM. 117 Hepatitis E virus seroepidemiology: a post-earthquake study among blood donors in Nepal. BMC Infect Dis 2016; 16: 707 [PMID: 27887586 DOI: 10.1186/s12879-016-2043-8]
- 118 Nasrallah GK, Al Absi ES, Ghandour R, Ali NH, Taleb S, Hedaya L, Ali F, Huwaidy M, Husseini A. Seroprevalence of hepatitis E virus among blood donors in Qatar (2013-2016). Transfusion 2017; 57: 1801-1807 [PMID: 28453178 DOI: 10.1111/trf.14116]
- 119 Jupattanasin S, Chainuvati S, Chotiyaputta W, Chanmanee T, Supapueng O, Charoonruangrit U, Oota S, Louisirirotchanakul S. A Nationwide Survey of the Seroprevalence of Hepatitis E Virus Infections Among Blood Donors in Thailand. Viral Immunol 2019; 32: 302-307 [PMID: 31403386 DOI: 10.1089/vim.2018.0146]
- 120 Traoré KA, Ouoba JB, Rouamba H, Nébié YK, Dahourou H, Rossetto F, Traoré AS, Barro N, Roques P. Hepatitis E Virus Prevalence among Blood Donors, Ouagadougou, Burkina Faso. Emerg Infect Dis 2016; 22: 755-757 [PMID: 26982195 DOI: 10.3201/eid2204.151728]
- Ibrahim EH, Abdelwahab SF, Nady S, Hashem M, Galal G, Sobhy M, Saleh AS, Shata MT. 121 Prevalence of anti-HEV IgM among blood donors in Egypt. Egypt J Immunol 2011; 18: 47-58 [PMID: 23082470]
- 122 Meldal BH, Sarkodie F, Owusu-Ofori S, Allain JP. Hepatitis E virus infection in Ghanaian blood donors - the importance of immunoassay selection and confirmation. Vox Sang 2013; 104: 30-36 [PMID: 22845878 DOI: 10.1111/j.1423-0410.2012.01637.x]
- 123 Lopes T, Cable R, Pistorius C, Maponga T, Ijaz S, Preiser W, Tedder R, Andersson MI. Racial differences in seroprevalence of HAV and HEV in blood donors in the Western Cape, South Africa: a clue to the predominant HEV genotype? Epidemiol Infect 2017; 145: 1910-1912 [PMID: 28357965 DOI: 10.1017/S0950268817000565]
- Ben-Ayed Y, Hannachi H, Ben-Alaya-Bouafif N, Gouider E, Triki H, Bahri O. Hepatitis E virus 124 seroprevalence among hemodialysis and hemophiliac patients in Tunisia (North Africa). J Med Virol 2015; 87: 441-445 [PMID: 25331682 DOI: 10.1002/jmv.24082]
- Schemmerer M, Rauh C, Jilg W, Wenzel JJ. Time course of hepatitis E-specific antibodies in 125 adults. J Viral Hepat 2017; 24: 75-79 [PMID: 27699946 DOI: 10.1111/jvh.12621]
- 126 Takahashi M, Tanaka T, Takahashi H, Hoshino Y, Nagashima S, Jirintai, Mizuo H, Yazaki Y, Takagi T, Azuma M, Kusano E, Isoda N, Sugano K, Okamoto H. Hepatitis E Virus (HEV) strains in serum samples can replicate efficiently in cultured cells despite the coexistence of HEV antibodies: characterization of HEV virions in blood circulation. J Clin Microbiol 2010; 48: 1112-1125 [PMID: 20107086 DOI: 10.1128/JCM.02002-09]
- Goel A, Vijay HJ, Katiyar H, Aggarwal R. Prevalence of hepatitis E viraemia among blood donors: 127 a systematic review. Vox Sang 2020; 115: 120-132 [PMID: 32030767 DOI: 10.1111/vox.12887]
- 128 Tedder RS, Tettmar KI, Brailsford SR, Said B, Ushiro-Lumb I, Kitchen A, Morgan D, Lattimore S, Tossell J, Ijaz S, Hewitt PE. Virology, serology, and demography of hepatitis E viremic blood donors in South East England. Transfusion 2016; 56: 1529-1536 [PMID: 26841005 DOI: 10.1111/trf.13498]
- 129 Riveiro-Barciela M, Rando-Segura A, Barreira-Díaz A, Bes M, P Ruzo S, Piron M, Quer J, Sauleda S, Rodríguez-Frías F, Esteban R, Buti M. Unexpected long-lasting anti-HEV IgM positivity: Is HEV antigen a better serological marker for hepatitis E infection diagnosis? J Viral Hepat 2020; 27: 747-753 [PMID: 32106351 DOI: 10.1111/jvh.13285]
- 130 Kar P, Karna R. A Review of the Diagnosis and Management of Hepatitis E. Infect Dis 2020; 1-11 [DOI: 10.1007/s40506-020-00235-4]
- 131 Hartl J, Otto B, Madden RG, Webb G, Woolson KL, Kriston L, Vettorazzi E, Lohse AW, Dalton HR, Pischke S. Hepatitis E Seroprevalence in Europe: A Meta-Analysis. Viruses 2016; 8 [PMID: 27509518 DOI: 10.3390/v8080211]
- 132 Shrestha AC, Flower RL, Seed CR, Stramer SL, Faddy HM. A Comparative Study of Assay Performance of Commercial Hepatitis E Virus Enzyme-Linked Immunosorbent Assay Kits in Australian Blood Donor Samples. J Blood Transfus 2016; 2016: 9647675 [PMID: 27891290 DOI: 10.1155/2016/9647675
- Galli C, Fomiatti L, Tagliacarne C, Velati C, Zanetti AR, Castaldi S, Romanò L. Seroprevalence of 133 hepatitis E virus among blood donors in northern Italy (Sondrio, Lombardy) determined by three different assays. Blood Transfus 2017; 15: 502-505 [PMID: 29059041 DOI: 10.2450/2017.0089-17]
- 134 Park HK, Jeong SH, Kim JW, Woo BH, Lee DH, Kim HY, Ahn S. Seroprevalence of anti-hepatitis E virus (HEV) in a Korean population: comparison of two commercial anti-HEV assays. BMC Infect Dis 2012; 12: 142 [PMID: 22726615 DOI: 10.1186/1471-2334-12-142]
- 135 Bendall R, Ellis V, Ijaz S, Ali R, Dalton H. A comparison of two commercially available anti-HEV IgG kits and a re-evaluation of anti-HEV IgG seroprevalence data in developed countries. J Med Virol 2010; 82: 799-805 [PMID: 20336757 DOI: 10.1002/jmv.21656]
- 136 Matsubayashi K, Nagaoka Y, Sakata H, Sato S, Fukai K, Kato T, Takahashi K, Mishiro S, Imai M, Takeda N, Ikeda H. Transfusion-transmitted hepatitis E caused by apparently indigenous hepatitis E virus strain in Hokkaido, Japan. Transfusion 2004; 44: 934-940 [PMID: 15157263 DOI: 10.1111/j.1537-2995.2004.03300.x
- Okano H, Nakano T, Ito R, Tanaka A, Hoshi Y, Matsubayashi K, Asakawa H, Nose K, Tsuruga S, 137 Tochio T, Kumazawa H, Isono Y, Tanaka H, Matsusaki S, Sase T, Saito T, Mukai K, Nishimura A, Kawakami K, Nagashima S, Takahashi M, Okamoto H. The spontaneous clearance of hepatitis E



virus (HEV) and emergence of HEV antibodies in a transfusion-transmitted chronic hepatitis E case after completion of chemotherapy for acute myeloid leukemia. Clin J Gastroenterol 2020; 13: 252-259 [PMID: 31342463 DOI: 10.1007/s12328-019-01024-3]

- 138 Gallian P, Pouchol E, Djoudi R, Lhomme S, Mouna L, Gross S, Bierling P, Assal A, Kamar N, Mallet V, Roque-Afonso AM, Izopet J, Tiberghien P. Transfusion-Transmitted Hepatitis E Virus Infection in France. Transfus Med Rev 2019; 33: 146-153 [PMID: 31327668 DOI: 10.1016/j.tmrv.2019.06.001
- Ledesma J, Williams D, Stanford FA, Hewitt PE, Zuckerman M, Bansal S, Dhawan A, Mbisa JL, 139 Tedder R, Ijaz S. Resolution by deep sequencing of a dual hepatitis E virus infection transmitted via blood components. J Gen Virol 2019; 100: 1491-1500 [PMID: 31592753 DOI: 10.1099/jgv.0.001302]
- Satake M, Matsubayashi K, Hoshi Y, Taira R, Furui Y, Kokudo N, Akamatsu N, Yoshizumi T, 140 Ohkohchi N, Okamoto H, Miyoshi M, Tamura A, Fuse K, Tadokoro K. Unique clinical courses of transfusion-transmitted hepatitis E in patients with immunosuppression. Transfusion 2017; 57: 280-288 [PMID: 28144952 DOI: 10.1111/trf.13994]
- 141 Lhomme S, Bardiaux L, Abravanel F, Gallian P, Kamar N, Izopet J. Hepatitis E Virus Infection in Solid Organ Transplant Recipients, France. Emerg Infect Dis 2017; 23: 353-356 [PMID: 28098552 DOI: 10.3201/eid2302.161094]
- Yamazaki Y, Naganuma A, Arai Y, Takeuchi S, Kobayashi T, Takakusagi S, Hatanaka T, Hoshino 142 T, Namikawa M, Hashizume H, Takizawa D, Ohyama T, Suzuki H, Horiguchi N, Takagi H, Sato K, Kakizaki S, Kusano M, Nagashima S, Takahashi M, Okamoto H, Yamada M. Clinical and virological features of acute hepatitis E in Gunma prefecture, Japan between 2004 and 2015. Hepatol Res 2017; 47: 435-445 [PMID: 27322051 DOI: 10.1111/hepr.12765]
- 143 Belliere J, Abravanel F, Nogier MB, Martinez S, Cintas P, Lhomme S, Lavayssière L, Cointault O, Faguer S, Izopet J, Kamar N. Transfusion-acquired hepatitis E infection misdiagnosed as severe critical illness polyneuromyopathy in a heart transplant patient. Transpl Infect Dis 2017; 19 [PMID: 28963742 DOI: 10.1111/tid.12784]
- Riveiro-Barciela M, Sauleda S, Quer J, Salvador F, Gregori J, Pirón M, Rodríguez-Frías F, Buti M. 144 Red blood cell transfusion-transmitted acute hepatitis E in an immunocompetent subject in Europe: a case report. Transfusion 2017; 57: 244-247 [PMID: 27785789 DOI: 10.1111/trf.13876]
- 145 Hoad VC, Gibbs T, Ravikumara M, Nash M, Levy A, Tracy SL, Mews C, Perkowska-Guse Z, Faddy HM, Bowden S. First confirmed case of transfusion-transmitted hepatitis E in Australia. Med J Aust 2017; 206: 289-290 [PMID: 28403756 DOI: 10.5694/mja16.01090]
- 146 Matsui T, Kang JH, Matsubayashi K, Yamazaki H, Nagai K, Sakata H, Tsuji K, Maguchi H. Rare case of transfusion-transmitted hepatitis E from the blood of a donor infected with the hepatitis E virus genotype 3 indigenous to Japan: Viral dynamics from onset to recovery. Hepatol Res 2015; 45: 698-704 [PMID: 25041213 DOI: 10.1111/hepr.12390]
- 147 Huzly D, Umhau M, Bettinger D, Cathomen T, Emmerich F, Hasselblatt P, Hengel H, Herzog R, Kappert O, Maassen S, Schorb E, Schulz-Huotari C, Thimme R, Unmüssig R, Wenzel JJ, Panning M. Transfusion-transmitted hepatitis E in Germany, 2013. Euro Surveill 2014; 19 [PMID: 24906377 DOI: 10.2807/1560-7917.es2014.19.21.20812]
- 148 Coilly A, Haïm-Boukobza S, Roche B, Antonini TM, Pause A, Mokhtari C, Becq A, Farahmand H, Hauser L, Duclos-Vallée JC, Samuel D, Adam R, Roque-Afonso AM. Posttransplantation hepatitis E: transfusion-transmitted hepatitis rising from the ashes. Transplantation 2013; 96: e4-e6 [PMID: 23857003 DOI: 10.1097/TP.0b013e318296c9f7]
- 149 Boxall E, Herborn A, Kochethu G, Pratt G, Adams D, Ijaz S, Teo CG. Transfusion-transmitted hepatitis E in a 'nonhyperendemic' country. Transfus Med 2006; 16: 79-83 [PMID: 16623913 DOI: 10.1111/j.1365-3148.2006.00652.x]
- 150 Mitsui T, Tsukamoto Y, Yamazaki C, Masuko K, Tsuda F, Takahashi M, Nishizawa T, Okamoto H. Prevalence of hepatitis E virus infection among hemodialysis patients in Japan: evidence for infection with a genotype 3 HEV by blood transfusion. J Med Virol 2004; 74: 563-572 [PMID: 15484278 DOI: 10.1002/jmv.20215]
- Ticehurst JR, Pisanic N, Forman MS, Ordak C, Heaney CD, Ong E, Linnen JM, Ness PM, Guo N, 151 Shan H, Nelson KE. Probable transmission of hepatitis E virus (HEV) via transfusion in the United States. Transfusion 2019; 59: 1024-1034 [PMID: 30702157 DOI: 10.1111/trf.15140]
- 152 Andonov A, Rock G, Lin L, Borlang J, Hooper J, Grudeski E, Wu J; Members of the Canadian Apheresis Group (CAG). Serological and molecular evidence of a plausible transmission of hepatitis E virus through pooled plasma. Vox Sang 2014; 107: 213-219 [PMID: 24830322 DOI: 10.1111/vox.12156
- Mrzljak A, Dinjar-Kujundzic P, Knotek M, Kudumija B, Ilic M, Gulin M, Zibar L, Hrstic I, 153 Jurekovic Z, Kolaric B, Jemersic L, Prpic J, Tomljenovic M, Vilibic-Cavlek T. Seroepidemiology of hepatitis E in patients on haemodialysis in Croatia. Int Urol Nephrol 2020; 52: 371-378 [PMID: 31894559 DOI: 10.1007/s11255-019-02363-3]
- 154 Slavov SN, Maçonetto JDM, Martinez EZ, Silva-Pinto AC, Covas DT, Eis-Hübinger AM, Kashima S. Prevalence of hepatitis E virus infection in multiple transfused Brazilian patients with thalassemia and sickle cell disease. J Med Virol 2019; 91: 1693-1697 [PMID: 31066064 DOI: 10.1002/jmv.25498]
- 155 Dalvand N, Dalvand A, Sharifi Z, Hosseini SM. Prevalence of hepatitis E virus in thalassemia patients with hepatitis C in Tehran, Iran. Iran J Microbiol 2019; 11: 535-540 [PMID: 32148686]



- 156 Ankcorn MJ, Fox TA, Ijaz S, Nicholas C, Houston E, Longair I, Suri D, Mattes FM, Walker JL, Tedder RS, Sekhar M. Characterising the risk of Hepatitis E virus infection in haematological malignancies: a UK prospective prevalence study. Br J Haematol 2019; 186: 191-195 [PMID: 30768677 DOI: 10.1111/bjh.15796]
- Haïm-Boukobza S, Ferey MP, Vétillard AL, Jeblaoui A, Pélissier E, Pelletier G, Teillet L, Roque-157 Afonso AM. Transfusion-transmitted hepatitis E in a misleading context of autoimmunity and druginduced toxicity. J Hepatol 2012; 57: 1374-1378 [PMID: 22885386 DOI: 10.1016/j.jhep.2012.08.001]
- 158 Loyrion E, Trouve-Buisson T, Pouzol P, Larrat S, Decaens T, Payen JF. Hepatitis E Virus Infection after Platelet Transfusion in an Immunocompetent Trauma Patient. Emerg Infect Dis 2017; 23: 146-147 [PMID: 27983485 DOI: 10.3201/eid2301.160923]
- Peters van Ton AM, Gevers TJ, Drenth JP. Antiviral therapy in chronic hepatitis E: a systematic 159 review. J Viral Hepat 2015; 22: 965-973 [PMID: 25760481 DOI: 10.1111/jvh.12403]
- 160 Kamar N, Garrouste C, Haagsma EB, Garrigue V, Pischke S, Chauvet C, Dumortier J, Cannesson A, Cassuto-Viguier E, Thervet E, Conti F, Lebray P, Dalton HR, Santella R, Kanaan N, Essig M, Mousson C, Radenne S, Roque-Afonso AM, Izopet J, Rostaing L. Factors associated with chronic hepatitis in patients with hepatitis E virus infection who have received solid organ transplants. Gastroenterology 2011; 140: 1481-1489 [PMID: 21354150 DOI: 10.1053/j.gastro.2011.02.050]
- 161 Pas SD, de Man RA, Mulders C, Balk AH, van Hal PT, Weimar W, Koopmans MP, Osterhaus AD, van der Eijk AA. Hepatitis E virus infection among solid organ transplant recipients, the Netherlands. Emerg Infect Dis 2012; 18: 869-872 [PMID: 22516170 DOI: 10.3201/eid1805.111712]
- Kamar N, Selves J, Mansuy JM, Ouezzani L, Péron JM, Guitard J, Cointault O, Esposito L, 162 Abravanel F, Danjoux M, Durand D, Vinel JP, Izopet J, Rostaing L. Hepatitis E virus and chronic hepatitis in organ-transplant recipients. N Engl J Med 2008; 358: 811-817 [PMID: 18287603 DOI: 10.1056/NEJMoa0706992
- 163 Haagsma EB, van den Berg AP, Porte RJ, Benne CA, Vennema H, Reimerink JH, Koopmans MP. Chronic hepatitis E virus infection in liver transplant recipients. Liver Transpl 2008; 14: 547-553 [PMID: 18383084 DOI: 10.1002/lt.21480]
- 164 von Felden J, Alric L, Pischke S, Aitken C, Schlabe S, Spengler U, Giordani MT, Schnitzler P, Bettinger D, Thimme R, Xhaard A, Binder M, Ayuk F, Lohse AW, Cornelissen JJ, de Man RA, Mallet V. The burden of hepatitis E among patients with haematological malignancies: A retrospective European cohort study. J Hepatol 2019; 71: 465-472 [PMID: 31108159 DOI: 10.1016/i.ihep.2019.04.022]
- 165 Cao D, Cao QM, Subramaniam S, Yugo DM, Heffron CL, Rogers AJ, Kenney SP, Tian D, Matzinger SR, Overend C, Catanzaro N, LeRoith T, Wang H, Piñeyro P, Lindstrom N, Clark-Deener S, Yuan L, Meng XJ. Pig model mimicking chronic hepatitis E virus infection in immunocompromised patients to assess immune correlates during chronicity. Proc Natl Acad Sci U S A 2017; 114: 6914-6923 [PMID: 28630341 DOI: 10.1073/pnas.1705446114]
- 166 Saravanabalaji S, Tripathy AS, Dhoot RR, Chadha MS, Kakrani AL, Arankalle VA. Viral load, antibody titers and recombinant open reading frame 2 protein-induced TH1/TH2 cytokines and cellular immune responses in self-limiting and fulminant hepatitis e. Intervirology 2009; 52: 78-85 [PMID: 19401616 DOI: 10.1159/000214862]
- 167 Péron JM, Abravanel F, Guillaume M, Gérolami R, Nana J, Anty R, Pariente A, Renou C, Bureau C, Robic MA, Alric L, Vinel JP, Izopet J, Kamar N. Treatment of autochthonous acute hepatitis E with short-term ribavirin: a multicenter retrospective study. Liver Int 2016; 36: 328-333 [PMID: 26179015 DOI: 10.1111/liv.12911]
- 168 McPherson S, Elsharkawy AM, Ankcorn M, Ijaz S, Powell J, Rowe I, Tedder R, Andrews PA. Summary of the British Transplantation Society UK Guidelines for Hepatitis E and Solid Organ Transplantation. Transplantation 2018; 102: 15-20 [PMID: 28795981 DOI: 10.1097/TP.000000000001908
- European Association for the Study of the Liver. Electronic address: easloffice@easloffice.eu.; 169 European Association for the Study of the Liver. EASL Clinical Practice Guidelines on hepatitis E virus infection. J Hepatol 2018; 68: 1256-1271 [DOI: 10.1016/j.jhep.2012.09.013]
- 170 Wang Y, Zhou X, Debing Y, Chen K, Van Der Laan LJ, Neyts J, Janssen HL, Metselaar HJ, Peppelenbosch MP, Pan Q. Calcineurin inhibitors stimulate and mycophenolic acid inhibits replication of hepatitis E virus. Gastroenterology 2014; 146: 1775-1783 [PMID: 24582714 DOI: 10.1053/j.gastro.2014.02.036
- Kamar N, Lhomme S, Abravanel F, Marion O, Peron JM, Alric L, Izopet J. Treatment of HEV 171 Infection in Patients with a Solid-Organ Transplant and Chronic Hepatitis. Viruses 2016; 8 [PMID: 27537905 DOI: 10.3390/v8080222]
- Gorris M, van der Lecq BM, van Erpecum KJ, de Bruijne J. Treatment for chronic hepatitis E virus 172 infection: A systematic review and meta-analysis. J Viral Hepat 2021; 28: 454-463 [PMID: 33301609 DOI: 10.1111/jvh.13456]
- 173 De Winter BCM, Hesselink DA, Kamar N. Dosing ribavirin in hepatitis E-infected solid organ transplant recipients. Pharmacol Res 2018; 130: 308-315 [PMID: 29499270 DOI: 10.1016/j.phrs.2018.02.030]
- Kamar N, Lhomme S, Abravanel F, Cointault O, Esposito L, Cardeau-Desangles I, Del Bello A, 174 Dörr G, Lavayssière L, Nogier MB, Guitard J, Ribes D, Goin AL, Broué P, Metsu D, Sauné K, Rostaing L, Izopet J. An Early Viral Response Predicts the Virological Response to Ribavirin in



Hepatitis E Virus Organ Transplant Patients. Transplantation 2015; 99: 2124-2131 [PMID: 26214817 DOI: 10.1097/TP.000000000000850]

- 175 Todt D. Meister TL, Steinmann E. Hepatitis E virus treatment and ribavirin therapy: viral mechanisms of nonresponse. Curr Opin Virol 2018; 32: 80-87 [PMID: 30384328 DOI: 10.1016/j.coviro.2018.10.001]
- Alric L, Bonnet D, Laurent G, Kamar N, Izopet J. Chronic hepatitis E virus infection: successful 176 virologic response to pegylated interferon-alpha therapy. Ann Intern Med 2010; 153: 135-136 [PMID: 20547885 DOI: 10.7326/0003-4819-153-2-201007200-00256]
- 177 Kamar N, Abravanel F, Garrouste C, Cardeau-Desangles I, Mansuy JM, Weclawiak H, Izopet J, Rostaing L. Three-month pegylated interferon-alpha-2a therapy for chronic hepatitis E virus infection in a haemodialysis patient. Nephrol Dial Transplant 2010; 25: 2792-2795 [PMID: 20494897 DOI: 10.1093/ndt/gfq282]
- 178 Rivero-Juarez A, Lopez-Lopez P, Frias M, Rivero A. Hepatitis E Infection in HIV-Infected Patients. Front Microbiol 2019; 10: 1425 [PMID: 31297100 DOI: 10.3389/fmicb.2019.01425]
- 179 Nakano R, Ohira M, Ishiyama K, Ide K, Kobayashi T, Tahara H, Shimizu S, Arihiro K, Imamura M, Chayama K, Tanaka Y, Ohdan H. Acute Graft Rejection and Formation of De Novo Donor-Specific Antibodies Triggered by Low Cyclosporine Levels and Interferon Therapy for Recurrent Hepatitis C Infection After Liver Transplantation: A Case Report. Transplant Proc 2017; 49: 1634-1638 [PMID: 28838454 DOI: 10.1016/j.transproceed.2017.05.006]
- 180 Selzner N, Guindi M, Renner EL, Berenguer M. Immune-mediated complications of the graft in interferon-treated hepatitis C positive liver transplant recipients. J Hepatol 2011; 55: 207-217 [PMID: 21145865 DOI: 10.1016/j.jhep.2010.11.012]
- 181 Kamar N, Rostaing L, Abravanel F, Garrouste C, Esposito L, Cardeau-Desangles I, Mansuy JM, Selves J, Peron JM, Otal P, Muscari F, Izopet J. Pegylated interferon-alpha for treating chronic hepatitis E virus infection after liver transplantation. Clin Infect Dis 2010; 50: e30-e33 [PMID: 20113176 DOI: 10.1086/650488]
- 182 Haagsma EB, Riezebos-Brilman A, van den Berg AP, Porte RJ, Niesters HG. Treatment of chronic hepatitis E in liver transplant recipients with pegylated interferon alpha-2b. Liver Transpl 2010; 16: 474-477 [PMID: 20373458 DOI: 10.1002/lt.22014]
- 183 Ollivier-Hourmand I, Lebedel L, Lecouf A, Allaire M, Nguyen TTN, Lier C, Dao T. Pegylated interferon may be considered in chronic viral hepatitis E resistant to ribavirin in kidney transplant recipients. BMC Infect Dis 2020; 20: 522 [PMID: 32677900 DOI: 10.1186/s12879-020-05212-2]
- 184 Dao Thi VL, Debing Y, Wu X, Rice CM, Neyts J, Moradpour D, Gouttenoire J. Sofosbuvir Inhibits Hepatitis E Virus Replication In Vitro and Results in an Additive Effect When Combined With Ribavirin. Gastroenterology 2016; 150: 82-85.e4 [PMID: 26408347 DOI: 10.1053/i.gastro.2015.09.0111
- van der Valk M, Zaaijer HL, Kater AP, Schinkel J. Sofosbuvir shows antiviral activity in a patient 185 with chronic hepatitis E virus infection. J Hepatol 2017; 66: 242-243 [PMID: 27702641 DOI: 10.1016/j.jhep.2016.09.014]
- van Wezel EM, de Bruijne J, Damman K, Bijmolen M, van den Berg AP, Verschuuren EAM, 186 Ruigrok GA, Riezebos-Brilman A, Knoester M. Sofosbuvir Add-on to Ribavirin Treatment for Chronic Hepatitis E Virus Infection in Solid Organ Transplant Recipients Does Not Result in Sustained Virological Response. Open Forum Infect Di 2019; 6 [DOI: 10.1093/ofid/ofz346]
- 187 Cornberg M, Pischke S, Müller T, Behrendt P, Piecha F, Benckert J, Todt D, Steinmann E, Papkalla A, von Karpowitz M, Koch A, Lohse A, Hardtke S, Manns MP, Wedemeyer H. Sofosbuvir monotherapy fails to achieve HEV RNA elimination in patients with chronic hepatitis E - The HepNet SofE pilot study. J Hepatol 2020; 73: 696-699 [PMID: 32624195 DOI: 10.1016/j.jhep.2020.05.020]
- 188 Tedder RS, Ijaz S, Kitchen A, Ushiro-Lumb I, Tettmar KI, Hewitt P, Andrews N. Hepatitis E risks: pigs or blood-that is the question. Transfusion 2017; 57: 267-272 [PMID: 28194857 DOI: 10.1111/trf.13976]
- Denner J. Hepatitis E virus (HEV)-The Future. Viruses 2019; 11 [PMID: 30871152 DOI: 189 10.3390/v110302511
- Wu X, Chen P, Lin H, Hao X, Liang Z. Hepatitis E virus: Current epidemiology and vaccine. Hum 190 Vaccin Immunother 2016; 12: 2603-2610 [PMID: 27184971 DOI: 10.1080/21645515.2016.1184806
- 191 Denner J, Pischke S, Steinmann E, Blümel J, Glebe D. Why all blood donations should be tested for hepatitis E virus (HEV). BMC Infect Dis 2019; 19: 541 [PMID: 31221098 DOI: 10.1186/s12879-019-4190-11
- 192 Domanović D, Tedder R, Blümel J, Zaaijer H, Gallian P, Niederhauser C, Sauleda Oliveras S, O'Riordan J, Boland F, Harritshøj L, Nascimento MSJ, Ciccaglione AR, Politis C, Adlhoch C, Flan B, Oualikene-Gonin W, Rautmann G, Strengers P, Hewitt P. Hepatitis E and blood donation safety in selected European countries: a shift to screening? Euro Surveill 2017; 22 [PMID: 28449730 DOI: 10.2807/1560-7917.ES.2017.22.16.30514]
- 193 Matsubayashi K, Sakata H, Ikeda H. Hepatitis E virus infection and blood transfusion in Japan. ISBT Sci Ser 2011; 6: 344-349 [DOI: 10.1111/j.1751-2824.2011.01512.x]
- 194 Lee CK, Chau TN, Lim W, Tsoi WC, Lai ST, Lin CK. Prevention of transfusion-transmitted hepatitis E by donor-initiated self exclusion. Transfus Med 2005; 15: 133-135 [PMID: 15859980 DOI: 10.1111/j.0958-7578.2005.00563.x]



- 195 Pawlotsky JM. Hepatitis E screening for blood donations: an urgent need? Lancet 2014; 384: 1729-1730 [PMID: 25078305 DOI: 10.1016/S0140-6736(14)61187-9]
- 196 Boland F, Martinez A, Pomeroy L, O'Flaherty N. Blood Donor Screening for Hepatitis E Virus in the European Union. Transfus Med Hemother 2019; 46: 95-103 [PMID: 31191195 DOI: 10.1159/000499121]
- 197 American Association of Blood Banks. (2014) Hepatitis E virus [cited 20 March 2021]. Available from: https://www.aabb.org/docs/default-source/default-document-library/regulatory/eid/hepatitis-evirus.pdf?sfvrsn=9f532d0e_2
- 198 Kamp C, Blümel J, Baylis SA, Bekeredjian-Ding I, Chudy M, Heiden M, Henseler O, Keller-Stanislawski B, de Vos AS, Funk MB. Impact of hepatitis E virus testing on the safety of blood components in Germany - results of a simulation study. Vox Sang 2018; 113: 811-813 [PMID: 30318777 DOI: 10.1111/vox.12719]
- 199 Bi H, Yang R, Wu C, Xia J. Hepatitis E virus and blood transfusion safety. Epidemiol Infect 2020; 148: e158 [PMID: 32594963 DOI: 10.1017/S0950268820001429]
- 200 Pischke S, Peron JM, von Wulffen M, von Felden J, Höner Zu Siederdissen C, Fournier S, Lütgehetmann M, Iking-Konert C, Bettinger D, Par G, Thimme R, Cantagrel A, Lohse AW, Wedemeyer H, de Man R, Mallet V. Chronic Hepatitis E in Rheumatology and Internal Medicine Patients: A Retrospective Multicenter European Cohort Study. Viruses 2019; 11 [PMID: 30813268 DOI: 10.3390/v11020186]
- Bauer H, Luxembourger C, Gottenberg JE, Fournier S, Abravanel F, Cantagrel A, Chatelus E, 201 Claudepierre P, Hudry C, Izopet J, Fabre S, Lefevre G, Marguerie L, Martin A, Messer L, Molto A, Pallot-Prades B, Pers YM, Roque-Afonso AM, Roux C, Sordet C, Soubrier M, Veissier C, Wendling D, Péron JM, Sibilia J; Club Rhumatismes et Inflammation, a section of the French Society of Rheumatology. Outcome of hepatitis E virus infection in patients with inflammatory arthritides treated with immunosuppressants: a French retrospective multicenter study. Medicine (Baltimore) 2015; 94: e675 [PMID: 25860212 DOI: 10.1097/MD.00000000000675]
- Kenfak-Foguena A, Schöni-Affolter F, Bürgisser P, Witteck A, Darling KE, Kovari H, Kaiser L, 202 Evison JM, Elzi L, Gurter-De La Fuente V, Jost J, Moradpour D, Abravanel F, Izpopet J, Cavassini M; Data Center of the Swiss HIV Cohort Study, Lausanne, Switzerland. Hepatitis E Virus seroprevalence and chronic infections in patients with HIV, Switzerland. Emerg Infect Dis 2011; 17: 1074-1078 [PMID: 21749774 DOI: 10.3201/eid/1706.101067]
- 203 Buescher G, Ozga AK, Lorenz E, Pischke S, May J, Addo MM, Horvatits T. Hepatitis E seroprevalence and viremia rate in immunocompromised patients: a systematic review and metaanalysis. Liver Int 2021; 41: 449-455 [PMID: 33034121 DOI: 10.1111/liv.14695]
- 204 Navaneethan U, Al Mohajer M, Shata MT. Hepatitis E and pregnancy: understanding the pathogenesis. Liver Int 2008; 28: 1190-1199 [PMID: 18662274 DOI: 10.1111/j.1478-3231.2008.01840.x
- Anty R, Ollier L, Péron JM, Nicand E, Cannavo I, Bongain A, Giordanengo V, Tran A. First case 205 report of an acute genotype 3 hepatitis E infected pregnant woman living in South-Eastern France. J Clin Virol 2012; 54: 76-78 [PMID: 22336086 DOI: 10.1016/j.jev.2012.01.016]
- 206 Tabatabai J, Wenzel JJ, Soboletzki M, Flux C, Navid MH, Schnitzler P. First case report of an acute hepatitis E subgenotype 3c infection during pregnancy in Germany. J Clin Virol 2014; 61: 170-172 [PMID: 24996764 DOI: 10.1016/j.jcv.2014.06.008]
- Bouthry E, Benachi A, Vivanti AJ, Letamendia E, Vauloup-Fellous C, Roque-Afonso AM. 207 Autochthonous Hepatitis E during Pregnancy, France. Emerg Infect Dis 2018; 24: 1586-1587 [PMID: 30016249 DOI: 10.3201/eid2408.180105]
- 208 Jilani N, Das BC, Husain SA, Baweja UK, Chattopadhya D, Gupta RK, Sardana S, Kar P. Hepatitis E virus infection and fulminant hepatic failure during pregnancy. J Gastroenterol Hepatol 2007; 22: 676-682 [PMID: 17444855 DOI: 10.1111/j.1440-1746.2007.04913.x]
- 209 Kar P, Jilani N, Husain SA, Pasha ST, Anand R, Rai A, Das BC. Does hepatitis E viral load and genotypes influence the final outcome of acute liver failure during pregnancy? Am J Gastroenterol 2008; 103: 2495-2501 [PMID: 18785952 DOI: 10.1111/j.1572-0241.2008.02032.x]
- 210 Kar P, Sengupta A. A guide to the management of hepatitis E infection during pregnancy. Expert Rev Gastroenterol Hepatol 2019; 13: 205-211 [PMID: 30791760 DOI: 10.1080/17474124.2019.1568869]
- Horvatits T, Westhölter D, Peine S, Schulze Zur Wiesch J, Lohse AW, Lütgehetmann M, Pischke 211 S. Lack of evidence for human serum albumin as major source of HEV infections. Transfus Med 2018; 28: 470-471 [PMID: 29707836 DOI: 10.1111/tme.12536]
- Juhl D, Nowak-Göttl U, Blümel J, Görg S, Hennig H. Lack of evidence for the transmission of 212 hepatitis E virus by coagulation factor concentrates based on seroprevalence data. Transfus Med 2018; 28: 427-432 [PMID: 29280212 DOI: 10.1111/tme.12498]
- 213 de Vos AS, Janssen MP, Zaaijer HL, Hogema BM. Cost-effectiveness of the screening of blood donations for hepatitis E virus in the Netherlands. Transfusion 2017; 57: 258-266 [PMID: 28144956 DOI: 10.1111/trf.13978]
- 214 Gallian P, Lhomme S, Morel P, Gross S, Mantovani C, Hauser L, Tinard X, Pouchol E, Djoudi R, Assal A, Abravanel F, Izopet J, Tiberghien P. Risk for Hepatitis E Virus Transmission by Solvent/Detergent-Treated Plasma. Emerg Infect Dis 2020; 26: 2881-2886 [PMID: 33219652 DOI: 10.3201/eid2612.191482
- 215 Hauser L, Roque-Afonso AM, Beylouné A, Simonet M, Deau Fischer B, Burin des Roziers N,



Mallet V, Tiberghien P, Bierling P. Hepatitis E transmission by transfusion of Intercept blood system-treated plasma. Blood 2014; 123: 796-797 [PMID: 24482503 DOI: 10.1182/blood-2013-09-524348]

- 216 Farcet MR, Lackner C, Antoine G, Rabel PO, Wieser A, Flicker A, Unger U, Modrof J, Kreil TR. Hepatitis E virus and the safety of plasma products: investigations into the reduction capacity of manufacturing processes. Transfusion 2016; 56: 383-391 [PMID: 26399175 DOI: 10.1111/trf.13343]
- 217 Kapsch AM, Farcet MR, Wieser A, Ahmad MQ, Miyabayashi T, Baylis SA, Blümel J, Kreil TR. Antibody-enhanced hepatitis E virus nanofiltration during the manufacture of human immunoglobulin. Transfusion 2020; 60: 2500-2507 [PMID: 32794187 DOI: 10.1111/trf.16014]
- Praditya D, Friesland M, Gravemann U, Handke W, Todt D, Behrendt P, Müller TH, Steinmann E, 218 Seltsam A. Hepatitis E virus is effectively inactivated in platelet concentrates by ultraviolet C light. Vox Sang 2020; 115: 555-561 [PMID: 32383163 DOI: 10.1111/vox.12936]



WJG

World Journal of Gastroenterology

Submit a Manuscript: https://www.f6publishing.com

World J Gastroenterol 2022 January 7; 28(1): 76-95

DOI: 10.3748/wjg.v28.i1.76

ISSN 1007-9327 (print) ISSN 2219-2840 (online)

REVIEW

Viral hepatitis in 2021: The challenges remaining and how we should tackle them

Rebecca Dunn, Aaron Wetten, Stuart McPherson, Mhairi C Donnelly

ORCID number: Rebecca Dunn 0000-0003-0559-5755; Aaron Wetten 0000-0003-4989-9726; Stuart McPherson 0000-0002-5638-2453; Mhairi C Donnelly 0000-0001-7655-7284.

Author contributions: Dunn R and Wetten A performed the literature review and wrote the manuscript; McPherson S contributed to writing the manuscript and performed a critical review of the manuscript; Donnelly MC designed the review, contributed to writing the manuscript and performed a critical review of the manuscript; and all authors have read and approve the final manuscript.

Conflict-of-interest statement: All authors have no conflict of interest to declare.

Country/Territory of origin: United Kingdom

Specialty type: Gastroenterology and hepatology

Provenance and peer review: Invited article; Externally peer reviewed.

Peer-review model: Single blind

Peer-review report's scientific quality classification

Grade A (Excellent): 0 Grade B (Very good): B Rebecca Dunn, Gastroenterology, University Hospital of North Tees, Stockton on Tees TS198PE, United Kingdom

Aaron Wetten, Stuart McPherson, Mhairi C Donnelly, Liver Unit, Freeman Hospital, Newcastle NE77DN, United Kingdom

Aaron Wetten, Stuart McPherson, Translational and Clinical Research Institute, Newcastle University, Newcastle NE17RU, United Kingdom

Corresponding author: Mhairi C Donnelly, MBChB, Doctor, Liver Unit, Freeman Hospital, Freeman Road High Heaton, Newcastle NE77DN, United Kingdom. mhairi.donnelly@nhs.net

Abstract

Viral hepatitis results in 1.4 million deaths annually. The World Health Organization (WHO) set an ambitious target to eliminate viral hepatitis by 2030, but significant challenges remain. These include inequalities in access to healthcare, reaching at risk populations and providing access to screening and effective treatment. Stigma around viral hepatitis persists and must be addressed. The WHO goal of global elimination by 2030 is a worthy aim, but remains ambitious and the coronavirus 2019 pandemic undoubtedly has set back progress. This review article will focus on hepatitis A to E, highlighting problems that have been resolved in the field over the past decade, those that remain to be resolved and suggest directions for future problem solving and research.

Key Words: Hepatitis A; Hepatitis B; Hepatitis C; Hepatitis D; Hepatitis E; COVID-19

©The Author(s) 2022. Published by Baishideng Publishing Group Inc. All rights reserved.

Core Tip: Viral hepatitis results in 1.4 million deaths annually. The World Health Organization set an ambitious target to eliminate viral hepatitis by 2030, but significant challenges remain. These include inequalities in access to healthcare, reaching at risk populations and providing access to screening and effective treatment. In this review article, we discuss the advances in the field of viral hepatitis over the past decade. We also discuss the remaining challenges relating to viral hepatitis A to E, and suggest strategies and pathways for their resolution.



WJG | https://www.wjgnet.com

Grade C (Good): 0 Grade D (Fair): 0 Grade E (Poor): 0

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: htt ps://creativecommons.org/Licens es/by-nc/4.0/

Received: May 30, 2021 Peer-review started: May 30, 2021 First decision: June 11, 2021 Revised: June 26, 2021 Accepted: December 22, 2021 Article in press: December 22, 2021 Published online: January 7, 2022

P-Reviewer: Jackson K S-Editor: Wang JJ L-Editor: A P-Editor: Wang JJ



Citation: Dunn R, Wetten A, McPherson S, Donnelly MC. Viral hepatitis in 2021: The challenges remaining and how we should tackle them. World J Gastroenterol 2022; 28(1): 76-95

URL: https://www.wjgnet.com/1007-9327/full/v28/i1/76.htm DOI: https://dx.doi.org/10.3748/wjg.v28.i1.76

INTRODUCTION

Our understanding of the epidemiology of viral hepatitis and associated treatment strategies has advanced significantly over the past decade. Arguably, the most significant advances have occurred in the treatment of chronic hepatitis C, which is now curable with a short course of all oral antiviral therapy. Despite this, viral hepatitis still kills more than 1.4 million people a year[1]. As such, viral hepatitis has become a global health priority and a number of large-scale public health policies have been implemented. The World Health Organization (WHO) has set out an ambitious global elimination strategy for viral hepatitis, aiming to eliminate viral hepatitis as a public health threat by 2030[2]. Key interventions for viral elimination have been identified and include hepatitis B vaccination, facilitation of safe injection practices and safe blood transfusions, promotion of safe sex, hepatitis B treatment and hepatitis C cure. However, modelling studies suggest that up to 80% of high-income countries will not meet the WHO target[3].

This review article will focus on hepatitis A-E, highlighting problems that have been resolved in the field over the past decade, those that remain to be resolved and suggest directions for future problem solving and research. We will also discuss the impact of the coronavirus 2019 (COVID-19) pandemic on viral elimination.

METHODS

A PubMed search was performed using the following terms: "hepatitis A"; "hepatitis B"; "hepatitis C"; "hepatitis D"; "delta agent"; "hepatitis E"; "cirrhosis"; "direct acting antivirals"; "chronic kidney disease"; "chronic liver disease"; "functional cure"; "hepatocellular carcinoma"; "liver transplant"; "reinfection"; "ribavirin"; "viral elimination"; "viral resistance"; "virologic cure". Only English-language articles were included in this review. Reference lists of selected articles were reviewed for relevant studies. Published abstracts were included.

HEPATITIS A VIRUS

Worldwide, the incidence of hepatitis A virus (HAV) is decreasing[4,5], but with increasing globalization there are significant shifts in the epidemiology of HAV infection^[6]. Due to a large number of cases being asymptomatic and an estimated under-reporting of up to 80% of cases, it is acknowledged that the true incidence is difficult to quantify[7]. The incidence rate of HAV infection is strongly correlated with socioeconomic indicators; the incidence decreases with increasing access to clean water and sanitation. HAV infection is commonly reported in countries where conflict leads to the displacement of people, resulting in poor sanitation and overcrowding[8].

Advances in the past decade and problems now solved

Recent studies have expanded our understanding of the molecular virology and pathobiology of HAV. It is likely that multiple immune mechanisms contribute to the development of acute liver injury due to HAV infection, including decreased frequency of regulatory T-cells due to Fas-mediated apoptosis[9] and a polymorphism in TIM1[10]. Factors now recognized to influence the clinical course of HAV infection include variations in the viral nucleotide sequence within the 5'UTR[10].

The WHO estimated that HAV infection caused approximately 7134 deaths in 2016 [11]. In the United States, case-fatality estimates range from 0.3% to 0.6% for all age groups, rising to 1.8% amongst patients aged > 50 years[12]. A safe and effective inactivated vaccine has been in use for almost 30 years[13]. It was initially developed for individual prophylaxis, but now is used to control endemics[13]. A live attenuated



vaccine has been developed and licensed in China and it is used in the Chinese national vaccination program. Use of this vaccine in children has reportedly reduced the incidence of HAV infection by 80%[14]. There are now 34 countries that use or are planning to introduce HAV vaccination into routine immunization of children in specific risk groups[11]. Within the United Kingdom, persons who are considered high-risk for HAV infection and should be offered vaccination include those in close contact with someone with HAV infection, travelers who plan to travel to parts of the world where HAV is highly endemic, persons with chronic liver disease, men who have sex with other men (MSM), people who inject drugs (PWIDs) and those who are likely to be exposed to HAV from their employment, for example workers who are exposed to raw sewage such as within the construction industry.

Another advance in the past decade has been in the area of post-exposure prophylaxis (PEP) against HAV. PEP is recommended for persons who are immunocompromised and those who have chronic liver disease[15]. Immunoglobulin was previously the only recommended PEP however due to a number of factors including declining anti-HAV IgG titres in donor pools, new strategies were sought. Recent data support post-exposure immunization with an inactivated HAV vaccine as being effective in preventing infection when given within 14 d of exposure[13].

Problems remaining to be solved

Prevention of infection in high-risk populations (including targeted vaccination): With increasing numbers of forcibly displaced persons in certain parts of the world [16], endemic HAV infection will continue to be an ongoing but preventable issue that requires a global response to provide public health infrastructure, sanitation and free HAV vaccination programmes. This approach requires significant input from public health agencies and politicians alike.

Person to person transmission is described, with infection reported amongst PWIDs and homeless populations. These populations can be difficult to engage, and vaccinating these high-risk individuals needs to be a public health priority (at least in developed countries). MSM have been linked to outbreaks of cases in developed countries, with epidemiological and laboratory investigations linking genotypes between countries[17]. It is important that high-risk groups such as MSM are identified and offered vaccination to prevent outbreaks in susceptible communities where there is lack of herd immunity[18]. Improving uptake of HAV vaccination in the MSM population is a remaining challenge. Targeting these at risk populations by methods such as social media and dating apps have been shown to improve vaccination uptake[17]. Patients with chronic liver disease should also be offered HAV vaccination due to their risk of more severe infection, however doing so has not entered widespread clinical practice^[19]. In one American study of HAV vaccination in patients with chronic liver disease, 28% of patients seen in specialist centres underwent vaccination compared with 5% of patients managed in primary care[20]. In another American study of patients with hepatitis C, 7.9% of patients underwent HAV vaccination^[21]. As HAV is a vaccine-preventable disease, universal vaccination of infants would be an effective method for controlling the infection going forwards.

Treatment of severe liver injury due to HAV infection: Although rare, patients with acute HAV infection can progress to acute liver failure (ALF)[7]. Whilst these patients can recover with supportive management, a small number of patients may require transplantation. Patients progressing to ALF are typically older and may not be suitable candidates for liver transplantation, and therefore other specific treatment strategies are required. Furthermore, liver transplantation is not accessible to those most at risk in displaced communities. Ribavirin has successfully been used in treatment of acute hepatitis E infection; it has been shown to have an inhibitory effect on HAV *in vitro* but has not been assessed *in vivo* for therapeutic activity[22].

HEPATITIS B + D

Hepatitis B virus

Chronic hepatitis B infection is a global problem, but the burden of disease is mostly in low to middle income countries, with 248 million of the estimated 292 million people affected residing in Asia, Africa, the Pacific and Latin America. Chronic hepatitis B virus (HBV) accounts for approximately 47% of all viral hepatitis related deaths, the vast majority of which are secondary to complications of chronic liver disease[23,24].

Zaishideng® WJG | https://www.wjgnet.com

Advances in the past decade and problems now solved

In 2017 the nomenclature to describe the different phases of chronic HBV changed within the updated European Association for the Study of the Liver (EASL) hepatitis B guidelines[25]. This was to better reflect and highlight the two main pathological processes of chronic infection and chronic hepatitis, in particular taking into account the presence of hepatitis B e antigen (HBeAg), HBV DNA levels, alanine aminotransferase (ALT) values and the presence or absence of liver inflammation. The new definition of phases highlights the increased risk of advancing liver disease in both chronic hepatitis phases - even in HBeAg negative patients - where there is elevated HBV DNA levels and/or elevated ALT, removing the somewhat misleading term "inactive carrier". These changes in nomenclature have now been widely adopted[24].

Multiple societies now provide guidance on when to initiate treatment. Viral resistance to treatment is a problem which has now been largely overcome. The nucleos(t)ide analogues (NAs) entecavir (ETV), tenofovir disoproxil fumarate (TDF) and tenofovir alafenamide are recommended as first-line treatment in both American and European HBV guidelines[25,26]. These agents show high rates of viral suppression and high genetic barriers to resistance[27,28] and have largely replaced lamivudine (LAM) with which resistance was problematic and common. Following treatment with LAM for 1 year, 14%-32% of patients developed resistance, increasing to over 80% after 4 years[27]. Those who develop resistance to LAM and are switched to ETV are more likely to subsequently develop resistance to ETV, with resistance rates of up to 50% after 5 years of treatment compared to only 1.2% of patients developing resistance with ETV where LAM has not been previously used[27]. TDF monotherapy has been shown to be effective in patients who have previously experienced treatment failure due to LAM resistance[29] and although there have been cases reported of reduced efficacy of tenofovir, there have been very few reported cases of resistance.

Problems remaining to be solved

There remain a number of challenges in the diagnosis and management of patients with chronic hepatitis B infection - Figure 1.

Diagnosing and linking infected patients to care programmes: A significant proportion of infected persons have not been identified; current estimates suggests that only 10.5% of infected individuals have been diagnosed and only 5% of those eligible for treatment for chronic HBV infection are receiving treatment[30]. A large systemic review found that 10% of people (26 million) with HBV infection might need urgent treatment due to cirrhosis and 12%-25% of patients would also be eligible for treatment according to different international guidelines^[30]. Many countries do not have the infrastructure to deliver widespread testing, vaccination or treatment; this is particularly true in low-middle income countries where resources are limited. Detailed discussion on the challenges of such health inequalities are beyond the scope of this review. The approach to up-scaling diagnostic testing needs to vary according to the target population. In the United Kingdom and other developed countries, the majority of individuals with undiagnosed hepatitis B infection are born in countries with intermediate or high prevalence rates. Identifying these individuals may increase diagnosis rates. Case finding in high-risk groups is effective; in North-East England, individuals from the British-Chinese and South Asian communities were invited to education and screening (via dry blood spot testing) sessions in local community centres[31]. The prevalence of hepatitis B surface antigen (HBsAg) positivity was 4.6%, which is above the 2% screening threshold recommended by the Centers for Disease Control and Prevention[31]. Another study looked at the cost-effectiveness of a onetime opt out case-finding approach in a primary care setting in the United Kingdom migrant population. This approach was deemed very likely to be cost effective amongst migrant populations with HBsAg prevalence $\geq 1\%$ [32].

New point of care (POC) tests are also becoming available, making diagnosing infection easier and quicker. For example, the Determine HBsAg 2 test provides a HBsAg result in 15 min with high sensitivity and specificity[33]. POC tests allow testing and diagnosis to move out of established health care settings and may be of particular utility in resource poor settings and high-risk communities.

Increasing testing and subsequent diagnosis rates relies on public engagement to break down stereotypes and address stigma, improved interactions with health care services and addressing health inequalities arising from poverty and language barriers [34,35]. Collaboration and integration with other successful public health programs such as human immunodeficiency virus (HIV) services is also likely to be effective.

Zaishideng® WJG | https://www.wjgnet.com

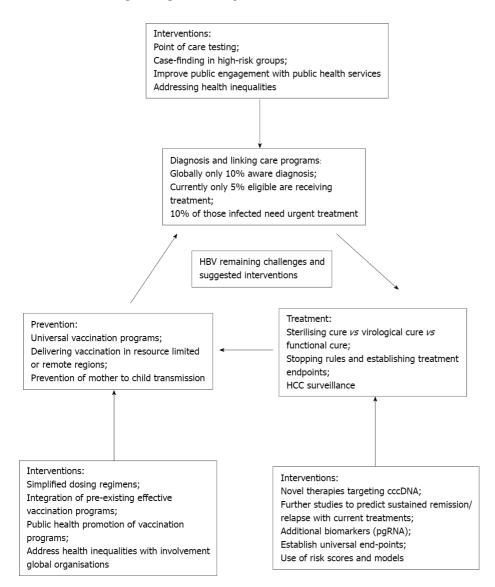


Figure 1 Remaining challenges in hepatitis B virus infection. HBV: Hepatitis B virus; cccDNA: Closed circular DNA; HCC: Hepatocellular carcinoma; pgRNA: Pregenomic RNA.

> Defining cure: A 'cure' for HBV might be considered as one where the virus is completely eliminated [undetectable HBsAg, HBeAg, HBV DNA and hepatic covalently closed circular DNA (cccDNA)] and where any (risk of) associated liver disease is also removed [36]. Consensus on definitions of cure remain contentious and as there is no current or upcoming treatment to achieve the 'holy grail' described above, there is reticence in how the word 'cure' is used. However, this is a key aspect of clinical care and research, therefore a globally accepted definition of cure needs to be obtained.

> The term 'sterilising cure' (complete eradication of the virus) has been replaced with 'functional cure'. Functional cure is currently defined as sustained HBsAg loss, undetectable HBV DNA, with or without seroconversion to hepatitis B surface antibody, following a finite course of treatment [25] and it occurs in 1% of chronically infected patients annually[37]. However, HBV genomes can persist in the liver even if HBsAg is undetectable questioning the true value of achieving a functional cure. A 'partial functional cure' is considered an intermediate goal of therapy and signifies detectable HBsAg but persistent undetectable HBV DNA 6 mo post-treatment. Virologic cure is essentially 'halting' all forms of HBV replication, however difficulties with obtaining virologic cure remain due to the persistence of cccDNA in hepatocytes. To obtain virologic cure, treatments inhibiting both cccDNA and viral replication are required[38].

> An agreed definition of cure remains elusive, however with clearly defining treatment endpoints and new therapies targeting different aspects of the HBV life



WJG https://www.wjgnet.com

cycle, virologic cure may be achievable in the future.

Striving for prevention rather than cure: To prevent HBV infection, there needs to be a focus on improving vaccination strategies. Barriers to HBV vaccination, particularly in resource limited or remote regions, can be attributed to inadequate resources to acquire vaccinations, current dosing regimens, insufficient trained health staff for administration of the vaccine and lack of facilities to keep vaccinations between 2-8 °C. One study of a two-dose regime of HBsAg-1018 (containing HBsAg plus a toll-like receptor 9 agonist adjuvant) demonstrated a higher seroprotection rate at one year compared with the standard three dose regimen[39]. Simplified regimens with fewer doses over a shorter time period (HBsAg-1018 given at 0 and 4 wk) are likely to be associated with increased uptake[39]. Many countries have now instituted effective COVID-19 vaccination programmes, and similar systems could be used to roll out simplified HBV vaccination regimens.

Preventing mother to child (vertical) transmission of hepatitis B is vital if global elimination is to be achieved[40,41]. High maternal viral load is the greatest risk factor for mother to child transmission; HBeAg positivity also increases risk[41]. In resource poor settings the WHO-recommended vaccine strategy may be difficult to deliver, and diagnostic assays for HBV testing may not be readily available. A potential strategy in these settings is POC testing to establish HBeAg status, followed by empirical treatment with tenofovir in the 3rd trimester in those who are HBeAg positive to reduce viral load and the risk of perinatal transmission[42], however such diagnostic assays are not readily available and remain costly.

Defining 'stopping rules' for HBeAg negative patients treated with NAs: Where seroconversion of HBeAg occurs, 67%-85% of patients have a sustained inactive state (HBeAg negative chronic infection); this is particularly the case where seroconversion occurs below the age of 30 years and where a low or undetectable HBV DNA level has been maintained^[43]. However, given significant relapse rates it remains controversial as to whether NA treatment can be stopped after HBeAg loss. A HBeAg negative state is associated with higher rates of regression of fibrosis but some patients will develop HBeAg negative hepatitis, the risk of which increases with time (22% at 10 years) and increases the risk of progression to advanced liver disease[44].

Given the low rate of clearance of HBsAg, HBeAg seroconversion is considered as a potential endpoint of treatment, where undetectable HBV DNA is achieved on three separate occasions in a 6[25] or 12-mo[26] period. If treatment is stopped at this endpoint, 50% will undergo HBeAg reversion requiring treatment with NAs to restart; close biochemical monitoring is therefore required. There is evidence to suggest that longer treatment with NAs results in a higher chance of persistent remission, with viral remission for 24-mo on NAs offering the most likely chance of sustained remission^[45].

Therefore, currently there is no universal stopping rule. In real-world practice, many different factors are taken in to consideration when making the decision to stop treatment with NAs, including the stage of fibrosis and family history of hepatocellular carcinoma (HCC). Further studies are needed to more clearly define the predictors of sustained remission and/or relapse to guide stopping decisions.

Establishing treatment endpoints - aiming for viral suppression vs cure: Currently, long-term suppression of HBV DNA levels is the main endpoint of treatment (+/-HBeAg loss in HBeAg positive patients). It remains a subject of debate as to whether the endpoint of treatment should be viral suppression, functional cure, partial functional cure or virologic cure. The ideal goal however would be virologic cure. In 2019 the joint EASL-American Association for the Study of Liver Diseases HBV treatment endpoints conference agreed that a "functional cure" should be the primary endpoint of phase III trials; sustained HBsAg loss in more than 30% of patients was accepted as an acceptable rate of response in phase III trials[38]. The endpoint for trials may not be the same as the endpoint for real world clinical practice however.

Biomarkers continue to be developed and may prove useful in defining future treatment endpoints. These biomarkers are likely to be used in conjunction with currently utilised clinical markers. The development of hepatitis B core-related antigen (HBcrAg) as a potential serological marker for cccDNA levels may identify patients who could discontinue NA therapy, those at risk of HCC development or of recurrence following treatment[28,46]. Pregenomic RNA may be a novel marker of viral replication; evidence is emerging that this may provide an earlier predictor for HBeAg seroconversion for those patients on NAs (an important indicator for partial immune response) and may help guide future treatment in those not achieving HBeAg seroconversion^[47].



Establishing a universally accepted endpoint of treatment along with biomarkers to help predict or confirm the achievement of this endpoint would be an important advance in the treatment of chronic HBV infection.

Risk of HCC and surveillance in patients on long term NAs: Chronic HBV infection is a leading cause of HCC; it is responsible for around 25% of liver cancer cases in developed countries and up to 60% of cases in developing countries[48]. NA therapy has been reported to decrease incidence of HCC[49,50]. While HBsAg loss after the development of advanced fibrosis minimizes the risk of the development of HCC, it does not negate it completely^[49]. A number of factors are taken into consideration when deciding which patient to survey for HCC including disease phase, age, ethnicity and family history of HCC[49]; international guidelines do not agree on the populations for surveillance however, promoting inequalities in care.

In those on NA therapy, risk scores such as the REACH-B score[51] or PAGE-B score[52] are used to identify patients who would benefit from HCC surveillance. The REACH-B scoring system was developed in a cohort of Asian patients with chronic HBV infection who were treatment naïve; no patients with cirrhosis were included in the development of this score[51]. This score does not offer good predictability in Caucasian patients with chronic HBV infection[53]. The modified REACH-B score substituted HBV DNA levels for the liver stiffness value which increased its accuracy [54]. The PAGE-B score was developed for use in Caucasian populations receiving tenofovir or ETV. A modified PAGE-B score (addition of serum albumin) has recently been tested in Asian patients on NA therapy, with an area under the receiver operating characteristic curve of 0.82[53]. The PAGE-B score is also predictive of HCC development in untreated patients[52].

Quantitative HBsAg and HBcrAg have been proposed as new biomarkers for HCC risk which might influence patient selection for HCC surveillance[55]. Risk models incorporating these biomarkers would be an advance in the field of HBV. New models could also incorporate other novel markers such as specific HBV mutations, presence of the metabolic syndrome and HBV genotype.

Identifying new treatments with finite duration and high cure rates: Most patients with chronic HBV currently require lifelong therapy, achieving viral suppression rather than cure^[25,26]. To achieve cure, combinations of therapy targeting different aspects of the HBV lifecycle are likely to be required including inhibition of cccDNA and viral replication[38].

A number of new treatments are being investigated for HBV and these are aiming to achieve clearance of HBsAg rather than just suppressing HBV DNA[36]. A detailed description of these treatments is beyond the scope of this review, but these include the development of new NAs (besifovir and metacavir), cccDNA silencers (e.g., lymphotoxin beta receptor agonist) and HBV entry inhibitors (Myrcludex B)[28,38,56]. There may also be a role for immunomodulatory therapies such as toll-like receptor agonists (acting via activating the innate immune response), check point inhibitors (helping to restore T-cell dysfunction) or therapeutic vaccines such as TherVacB[56, 57]. Gene editing strategies and RNA interference may be other potential treatment strategies[56]. Where eligible, patients should be considered for entry into clinical trials of novel therapies.

Hepatitis D virus

The current burden of hepatitis D virus (HDV) infection is unknown; estimates from a recent meta-analysis vary considerably, ranging from 12 million to 72 million individuals infected with HDV worldwide[58]. There is geographical variation in the prevalence of HDV infection. A recent systematic review and meta-analysis estimated anti-HDV prevalence to be 4.5% amongst HBsAg positive individuals globally with rates lower in Europe (3.0%) compared with Africa (5.97%)[58-60]. However, other meta-analysis estimates differ, demonstrating higher seroprevalence amongst HBsAg positive individuals worldwide (10.58%-13.02%) and within Europe (13.81%). Such differences are likely due to variation in modelling strategies and highlight the difficulties in truly identifying the burden of HDV[58-60]. Issues and challenges remaining in the field of HDV infection include identification of infected individuals, effective treatments, treatment endpoints and prevention.

Problems remaining to be solved

Identification of infected patients: A positive HDV antibody should be accompanied by detectable serum HDV RNA to detect active infection. However, some guidelines do not explicitly make recommendations for HDV testing and therefore many patients



who are HBsAg positive are not tested for HDV. One study looking at clinic-led anti-HDV testing identified that only 40% of HBV patients were tested [61]. The same study looked at a different centre offering reflex laboratory testing and found that 99.4% of first HBsAg positive samples were tested for anti-HDV. This is a potentially reliable approach to increasing detection of patients with HDV infection, as all patients who are newly diagnosed with HBsAg positivity should be tested for serological evidence of HDV infection.

There is an epidemiological association between anti-HDV seroprevalence and PWIDs, commercial sex workers, MSM and recipients of haemodialysis[58,62]. Suggested patient groups who should be prioritised for screening for HDV include: Patients who are HBsAg positive, patients with HIV, PWIDs, MSM and migrants from highly endemic regions.

Treatment for HDV infection: Pegylated-interferon (PEG-IFN) is the only treatment proven to have antiviral efficacy against chronic HDV infection, however viral suppression rates with PEG-IFN are poor in HDV infection and the adverse effects of PEG-IFN therapy are well described [62,63]. Extended duration of treatment has not been associated with a consistent or significant increase in efficacy, and the addition of NAs does not improve efficacy. New treatments are urgently required; therapies currently being evaluated include HBV/HDV entry inhibitors (Myrcludex B), virion secretion inhibitors (REP 2139) and inhibitors of the prenylation of the large HDV antigen (lonafarnib)[63]. Patients with HBV/HDV co-infection should be considered for entry into clinical trials. Ultimately, global prevention of HBV infection would be the most effective means of treating HDV infection.

Establishing treatment endpoints: Unfortunately, endpoints for HDV treatment and indicators of response to treatment have not been well established[38]. Cure may not be feasible. ALT normalization, changes in HDV RNA and qHBsAg are markers of response to treatment. Barriers to establishing treatment endpoints include lack of widespread availability of HDV diagnostics and lack of standardization of HDV RNA assays. Composite endpoints are likely to be more useful than singular end-points.

HEPATITIS C VIRUS

Perhaps the greatest advances in our understanding of virology and development of treatment strategies over the past decade have occurred in relation to hepatitis C virus (HCV) infection. Despite these advances a number of challenges remain, including targeting difficult to reach populations and expanding HCV testing and treatment programmes in resource poor countries. Addressing these areas will be critical if global elimination of HCV is to be achieved by 2030.

Advances in past decade and problems now solved

Treatment and cure: HCV treatment has evolved rapidly in the last 10 years, with the emergence of direct acting antiviral (DAA) regimens. These drugs are very well tolerated and highly effective in achieving sustained virologic response (SVR), even in patients who were previously considered 'hard to treat' or in whom interferon-based treatment was contraindicated. As a result, antiviral treatment with DAAs is recommended in all patients with active HCV infection[64] and elimination of HCV is an achievable goal if these drugs can be made widely available worldwide.

In 2011 the first protease-inhibitors (telaprevir and boceprevir) were approved for use in HCV infected individuals in combination with pegylated-interferon and ribavirin, but whilst SVR rates improved so did the frequency of side effects[65]. This was quickly followed by the approval of the first interferon-free regimens for the treatment of genotype 1 HCV infection in 2014, followed by the first pangenotypic regimen, sofosbuvir-velpatasvir, in 2016[66]. Pangenotypic regimens are advantageous because they remove the need for genotype testing prior to the commencement of treatment which simplifies treatment regimens, thus reducing the frequency of patients dropping out before they start antiviral treatment.

Presently, the availability of safe and highly effective DAA regimens supports a strategy of treating all individuals with chronic HCV infection over the age of 12, irrespective of the stage of disease [67]. Current regimens offer a number of advantages over previous interferon-containing regimens including much greater efficacy, few side-effects, oral once daily dosing and shorter duration of treatment. For current DAA regimes, SVR rates (undetectable HCV RNA at 12 or 24 wk after treatment) well exceed 90% for most patient cohorts, compared with approximately 50% of patients



treated with PEG-interferon and ribavirin. Patients with chronic kidney disease (including dialysis-dependent patients) and cirrhosis were previously considered difficult to treat but now have similar SVRs when treated with DAAs to those without chronic kidney disease and cirrhosis[68,69].

Significant improvements in SVR rates with DAAs has translated into a reduction in morbidity and mortality rates in patients with HCV. A systemic review and metaanalysis concluded that there was an 87% reduction in the incidence of HCC and a 75% reduction in all-cause mortality in those who achieved SVR when compared with those who did not[70]. By 2019 in the United Kingdom, the incidence of HCV-related end stage liver disease and HCC had fallen by 24% following the introduction of DAAs and the associated increase in the number of patients completing treatment. In Scotland, new presentations of HCV-related decompensated cirrhosis decreased by 51% in the DAA area with an estimated avoidance of 330 cases of decompensated cirrhosis^[73].

Problems remaining to be solved

Prevention: The ideal preventative treatment for HCV would be a vaccine. However, development of an HCV vaccine has been challenging due to the genetic diversity of the virus, the virus' ability to avoid the host immune response and a lack of in vitro and *in vivo* models of infection^[71]. Some progress has been made, and a recent trial of a vaccine regimen to prevent chronic HCV infection was safe and induced HCVspecific T-cell responses but it did not prevent chronic HCV infection in a cohort of patients with a recent history of intravenous drug use [72]. It is therefore unlikely that an available efficacious vaccine will be available in the short-term. Work to develop a vaccine is ongoing.

In the absence of a vaccine, improving harm reduction approaches for PWIDs is vital. Existing strategies include promotion of sterile injection equipment use through needle exchange programmes and opioid substitution therapy. These services are often poorly provided and under-utilized, but they have been shown to be highly costeffective[73]. It is been estimated that eliminating non-sterile injection techniques could prevent 43% of incident HCV infections between 2018 and 2030[74].

Difficult to reach populations: Despite advances in the medical treatment of hepatitis C, global elimination is unlikely to be achieved unless all infected patients are identified and then complete their treatment regimen. A significant proportion of people with HCV infection are unaware of their diagnosis, and our ability to find these patients is becoming increasingly challenging. Previous work has shown that HCV testing is concentrated in areas with lower risk of infection[75], commonly settings where patients are either in recovery from previous drug use or ongoing drug use is more 'controlled'. Testing needs to be expanded among 'difficult to reach' populations, especially those who may be in a more 'chaotic' phase of their drug use and are not in contact with addiction or other medical services. This group can be challenging to find and engage, but approaches such as testing and treatment in homeless hostels and food kitchens can be effective[76]. Moreover, testing delivered by peers is an approach that can increase diagnosis and subsequent treatment in patients considered hard to reach. In the United Kingdom, the hepatitis C trust run a peer-to-peer education programme, in which peer educators with personal experience of HCV deliver workshops sharing the importance of testing and treatment[77]. This has increased testing numbers and reduced attrition, whilst providing valuable education.

One important area to target to increase testing and treatment is in the prison population. Prison populations have a high prevalence of HCV infection with many studies reporting an incidence > 10 times that of the general population[78]. Drug use prior to or during imprisonment is common, yet harm reduction methods such as access to clean injecting equipment is non-existent or inadequate in the majority of prisons. Opt-out screening for blood borne viruses (BBVs) is recommended in the EASL HCV guidelines for all prison inmates[79], but even where this is practiced rates of testing are suboptimal^[78]. Opt-in testing is more commonly practiced but is a far less effective approach. BBV testing can be challenging, particularly in reception prisons (prisoners awaiting sentencing) because these typically have a very large throughput of inmates and periods of incarceration can be short. However, these challenges can be effectively overcome with investment and an organized approach to testing. Effective approaches to increasing testing for HCV and scaling up of treatment with DAAs can also be used as 'treatment as prevention'. This approach was practiced in an Australian prison population and led to a significant reduction in incidence of new HCV infections[80].



Another approach that could be considered to identify undiagnosed patients with HCV is a 'track and trace' approach by mapping the social networks of individuals with a history of injecting drug use and offering HCV testing to those in a group who may not have been tested. Whilst this may sound like a practical solution, one study showed that this was ineffective in real world clinical practice with only one participant coming forward for testing[81]. Further work is needed to determine whether this approach could be refined to increase its efficacy, particularly since people are now more aware of 'track and trace' programmes as a result of the COVID-19 pandemic.

Attrition: Increasing detection rates will only help in the strive for global elimination if these are translated into increased treatment rates. An analysis of two large national laboratory databases from 2013 to 2016 found that 89.4% of patients diagnosed with chronic HCV infection did not receive a prescription for antiviral therapy[82]. In Spain, 49.8% of those with a positive anti-HCV result were not then linked into specialist care [75]. One reason for this is that care pathways have been unnecessarily complex including multiple investigations prior to treatment, which leads to patients frequently being lost to follow up and never completing treatment. Attrition appears to occur early in the treatment cascade; in one study 57.3% of patients dropped off prior to having liver enzymes checked[83]. With the advent of pangenotypic regimens and simple non-invasive fibrosis scores [*e.g.,* fibrosis-4 (FIB-4) and aminotransferase-platelet ratio index] pathways is shown in Figure 2.

Moving care delivery out of hospital settings may improve attrition rates. One study from the North-East of England found that distance from a HCV treatment service was a major predictor of patients not commencing antivirals[84]. DAAs can be effectively delivered in non-hospital settings, increasing access to treatment. A cluster-randomized trial showed that pharmacist delivered treatment in patients on opiate substitution therapy was more effective than conventionally delivered HCV therapy with more patients initiating and completing treatment, and achieving SVR[85]. Other examples of successful non-traditional HCV services have been delivered in primary care, nurse led community clinics, addiction services and homeless hostels[86]. Empowering addiction workers and those working with the homeless to become involved in the care cascade is also likely to improve attrition rates.

Reinfections: Re-infection with HCV after SVR, detected by the presence of HCV RNA rather than HCV antibodies, is largely related to an individuals' ongoing high risk behavior, inadequate harm reduction knowledge and/or lack of availability of clean injecting equipment. The true rate of reinfection is not known and is likely to vary significantly depending on the population studied. Individuals who continue to actively inject drugs after treatment have the highest rates. Very high rates of reinfection (up to 40%) have been seen in some high-risk groups, but other studies have reported lower rates[87]. Better access to harm reduction methods is vital to reduce reinfection rates. In addition, PWIDs should be tested at least annually for HCV RNA if they have ongoing high-risk behavior to identify reinfections. It is critical that they are offered re-treatment to try and reduce the risk of onward transmission of the infection[79].

Long term impact of hepatitis C infection: HCV infection is associated with multiple extrahepatic complications including increased risk of autoimmune disorders, cryoglobulinaemia and lymphoma. In addition, there is increased risk of type II diabetes, cardiovascular disease, chronic fatigue and psychological morbidity. Many of these comorbidities persist following SVR and one study found that nearly all patients have at least one co-morbidity that remains long-term[88].

Despite individuals with HCV having a significantly increased risk of cardiovascular disease, few are actively treated to reduce their cardiovascular risk[89]. Moreover, even though quality of life improves following successful antiviral treatment, this remains significantly worse than the general population[89]. This probably relates to high rates of mental health disorders, unhealthy alcohol consumption, ongoing drug use, deprivation, type II diabetes and the metabolic syndrome. Participation in physical activity in individuals with HCV is associated with improved quality of life[90]. Taking a more holistic approach to the care of individuals with HCV rather than just focusing on treating the infection may help improve long-term outcomes and improve quality of life. Use of a holistic care bundle may help achieve this[89].

Zaishidena® WJG | https://www.wjgnet.com

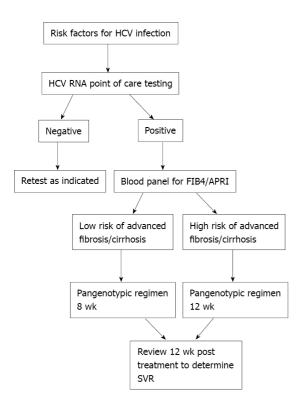


Figure 2 Proposed simplified pathway for hepatitis C virus diagnosis, staging and treatment. HCV: Hepatitis C virus; FIB-4: Fibrosis-4; APRI: Aminotransferase-platelet ratio index; SVR: Sustained virologic response.

HCC surveillance in non-cirrhotic patients: The risk of development of HCC in individuals with HCV-related cirrhosis falls following SVR, but remains approximately 2% per year and as a result, surveillance is recommended for these individuals [91]. HCC may also occur in patients with advanced fibrosis, but at a lower rate than in those with cirrhosis and it remains uncertain whether HCC surveillance is clinically effective and cost effective in this group. This is further complicated by the fact that many patients are staged using transient elastography and relevant cut-offs to identify those who are likely to benefit from HCC surveillance have not been defined. There is a clear cut need to develop better models to predict the development of HCC in individuals following SVR. International societies have different recommendations regarding HCC surveillance in those achieving SVR which reflects the overall uncertainty - Table 1.

There have been some recent studies that have attempted to more clearly define patients who would benefit from HCC surveillance post SVR[90,91]. One study developed a model to predict patients with advanced fibrosis who have a low risk of HCC and may therefore not benefit from surveillance. The model used a combination of baseline and dynamic changes in liver stiffness measurement, FIB-4 score and serum albumin after SVR and identified that nearly 20% of their cohort of patients with compensated advanced fibrosis had a very low risk of developing HCC[90]. Dynamic assessment of the FIB-4 score in isolation may also predict the risk of development of HCC after SVR[98]. In one study, no patients with a FIB-4 < 1.45 after SVR developed HCC[92]. A number of studies are underway with the aim of developing better predictive models for HCC using clinical parameters and novel biomarkers.

Increased use of hepatitis C positive donor organs: The advent of safe and highly effective DAAs for HCV infection has increased the potential to use HCV-positive organs even when the donor is viraemic, expanding the donor pool[93]. HCV positive (HCV RNA +) donor organs universally transmit HCV to the recipient[94] so prior to the widespread availability of DAAs use of these organs was restricted to those who already had HCV viraemia. However, given the efficacy of DAAs it is now possible to transplant HCV RNA + organs in to HCV negative recipients and then treat the HCV infection in the recipient.

In 2019, Kwong et al [95] assessed the outcomes from HCV treatment with DAAs in 10 non-viraemic patients who received HCV RNA + livers. Short-term outcomes were

WJG | https://www.wjgnet.com

Table 1 Recommendations for hepatocellular carcinoma surveillance in patients with hepatitis C virus achieving sustained virologic response					
Guideline	Recommendation				
EASL[114] 2020	Survey patients with advanced fibrosis (F3) or cirrhosis (F4)				
AASLD[115]	Survey cirrhotic patients				
Asia-Pacific[116]	Survey cirrhotic patients; Survey patients with any histologic stage of HCV with comorbidities, such as alcohol abuse and diabetes mellitus				

HCV: Hepatitis C virus. AASLD: American Association for the Study of Liver Diseases; EASL: European Association for the Study of the Liver.

excellent with 100% achieving SVR at 12 wk post treatment. The practice of using HCV RNA + organs with subsequent DAA treatment is now routine in some countries around the world.

HEPATITIS E VIRUS

Hepatitis E virus (HEV) is the most common cause of acute hepatitis worldwide and carries a significant global burden of disease. HEV genotypes 1 and 2 account for approximately 20.1 million HEV infections, 3.4 million symptomatic cases, 70000 deaths, and 3000 stillbirths annually [96]. Our understanding of the impact of hepatitis E infection has advanced significantly over the past decade, with the recognition of chronic infection, risk of progression to cirrhosis, risk factors for transmission and treatment strategies. Despite these advances, there are problems that remain to be resolved.

Advances in past decades and problems now solved

There are now eight recognized genotypes of HEV. Genotypes 1-4 and 7 cause human infection. Genotypes 1 and 2 are obligate human pathogens transmitted by the faecooral route and cause both sporadic infection and large outbreaks. In the developed world, sporadic infections are mainly caused by genotype 3 infection.

Transmission of HEV infection

Ingestion of raw or under-cooked meat (particularly pork products), shellfish and contaminated fruits is a significant risk factor for locally-acquired infection in the Western world. Genotype 3 and 4 HEV infection can be transmitted via transfusion of infected blood products and solid organ transplantation, and may have a significant clinical impact upon immunosuppressed individuals. A French study looked at 23 cases of reported transfusion related HEV infections in France between 2006-2016. It reported that 14 of these cases, all of whom were immunosuppressed, went on to develop chronic HEV infection[97]. The United Kingdom introduced a universal screening policy for blood products in 2017 and also screens deceased and live organ donors for HEV RNA^[98]. Other countries have a more selective strategy and only screen blood products intended for high-risk patients[99]. Universal screening has been shown to be more cost effective than selective screening if the incidence of HEV infection is above 1 in 10000 blood donations[100]. Sexual transmission in MSM has also been reported more recently[101].

Chronic infection and risk of cirrhosis

Prior to 2008, HEV was recognized to cause an acute, self-limiting illness. Genotype 3 HEV was first reported to cause chronic infection in 2008 and chronic infection has now been reported in immunocompromised individuals including solid organ transplant (SOT) recipients, patients receiving chemotherapy for haematological malignancies, HIV-1 infected patients and patients receiving immunomodulating drugs. In immunocompromised patients, the detection of HEV RNA in plasma or stool after 3 mo is defined as chronic infection[102]. Progression to cirrhosis in those with chronic hepatitis E infection occurs in 10%-15% and can occur rapidly, within 2-3 years [103]. In a study of 85 patients with chronic HEV infection in 17 transplant centres across Europe and North America, almost 66% of transplant recipients who contracted HEV developed chronic infection and 10% progressed to cirrhosis[103,104]. Chronic infection and the risk of cirrhosis is not seen with genotype 1 or 2 infection.



Treatment of chronic infection

Most published data regarding treatment of chronic HEV infection are from case series and reports in SOT recipients[105]. Reducing immunosuppression dose by around 30% has been shown to be effective in clearing HEV in around one third of patients [106]. Both PEGylated interferon and ribavirin are effective in treating chronic HEV infection. Interferon increases the risk of organ rejection in transplant recipients and therefore ribavirin monotherapy is the preferred option[107]. A systematic review has shown that 64% of patients were HEV RNA negative at 6 mo after the end of treatment with ribavirin monotherapy[108]. The optimal dose and duration of treatment is still to be determined but 3 mo courses have been used most commonly[107]. A multi-centre case series of 59 transplant recipients infected with HEV showed that ribavirin monotherapy, at a median dose of 600 mg/d for 3 mo achieved SVR in 78% of cases [107].

Problems remaining to be solved

Non-response to ribavirin: The main problem to be solved in relation to chronic HEV infection is how to manage non-response to ribavirin. Sofosbuvir has been proposed as an alternative agent to treat chronic HEV infection. It has shown promise in inhibiting HEV replication *in vitro*[109] but it had a negligible effect on improving viraemia in a case report[110]. A later study of sofosbuvir monotherapy in nine patients demonstrated a modest reduction in viral load but viral elimination was not achieved[111]. Convalescent plasma has also been trialed in a patient with persistent hepatitis E infection, and showed no effect on HEV RNA levels.

We also need to understand the relevance of HEV mutations and their effect on ribavirin resistance. Mutations have been identified in ribavirin non-responders but their impact on the treatment of these and other individuals has yet to be established. For example, the G1634R mutation does not lead to absolute ribavirin resistance and does not appear to compromise the response to a second course of treatment with ribavirin[112]. New treatments are ultimately required for those who fail treatment with ribavirin.

IMPACT OF COVID-19 ON VIRAL ELIMINATION

The COVID-19 pandemic has compromised efforts to progress towards the WHO goal of elimination of viral hepatitis. This impact of the pandemic is likely to be felt for years to come and during the initial peaks has resulted in delays in diagnosis and treatment, and reduced access to harm reduction services. In April 2020 in the United Kingdom, new diagnoses of HCV were down 85% and new treatment initiations had also fallen by 63% compared with the year prior[113]. Although there has been some recovery, pre-COVID 19 levels of testing and treatment have not yet been reached. Funding and resources have also been re-allocated to fighting the COVID-19 pandemic. In addition to the impact on global elimination, the COVID-19 pandemic has significantly impacted upon the provision of HCC surveillance programmes for patients with viral hepatitis.

However, during the pandemic many new ways of working (such as telemedicine) and care cascades have been adopted, which may in fact positively impact upon the delivery of viral hepatitis services in the years to come. For example, in some centres patients have been commenced on HCV treatment remotely using telemedicine (personal communication). The vaccination programmes and 'track and trace' systems set up during the COVID-19 pandemic could be extrapolated to viral hepatitis to improve service delivery.

SUGGESTED PUBLIC HEALTH AND RESEARCH PRIORITIES FOR THE NEXT DECADE

The global hepatology community is well placed to set public health and research priorities in viral hepatitis for the forthcoming decade, striving towards global elimination and reduced health care burden. Potential priorities for each individual virus are proposed in Table 2.

WJG | https://www.wjgnet.com

Table 2 Public health and research priorities for the next decade						
Virus	Public health priorities	Research priorities				
Hepatitis A	Increased vaccination of high-risk individuals; Improved sanitation and vaccination in camps for displaced persons	Medical treatments for those with acute liver failure				
Hepatitis B	Increase uptake of vaccination; Identifying undiagnosed individuals; Linkage to care	Establishing treatment end-points; Identifying curative treatment				
Hepatitis C	Microelimination; Reducing re-infection rates; Identifying undiagnosed individuals; Harm reduction	Vaccination; Confirming most effective HCC surveillance strategies				
Hepatitis D	Identification of infected individuals; Clarifying current disease burden of HDV	Novel therapies				
Hepatitis E	Increased screening of blood products/change in donor policies; Educating immunosuppressed patients of risk of food-borne transmission; Further understanding of sources of infection	RCT to confirm optimal dose and duration of ribavirin therapy; Novel treatments; Vaccination; Greater understanding of genetic mutations				

HDV: Hepatitis D virus; HCC: Hepatocellular carcinoma; RCT: Randomized controlled trial.

CONCLUSION

Significant advances have occurred in the field of viral hepatitis over the past decade, particularly in relation to the treatment and cure of hepatitis C. Over the next decade as we strive towards global elimination of viral hepatitis - the gastroenterology and hepatology community must focus on identifying the undiagnosed and engaging these individuals in to treatment programmes whilst continuing to develop novel treatments with the ultimate aim of cure.

REFERENCES

- Stanaway JD, Flaxman AD, Naghavi M, Fitzmaurice C, Vos T, Abubakar I, Abu-Raddad LJ, Assadi R, Bhala N, Cowie B, Forouzanfour MH, Groeger J, Hanafiah KM, Jacobsen KH, James SL, MacLachlan J, Malekzadeh R, Martin NK, Mokdad AA, Mokdad AH, Murray CJL, Plass D, Rana S, Rein DB, Richardus JH, Sanabria J, Saylan M, Shahraz S, So S, Vlassov VV, Weiderpass E, Wiersma ST, Younis M, Yu C, El Sayed Zaki M, Cooke GS. The global burden of viral hepatitis from 1990 to 2013: findings from the Global Burden of Disease Study 2013. Lancet 2016; 388: 1081-1088 [PMID: 27394647 DOI: 10.1016/S0140-6736(16)30579-7]
- World Health Organization. Global health sector stratergy on viral hepatitis 2016-2021: Towards 2 ending viral hepatitis: World Health Organisation; 2016. [cited 10 May 2021]. Available from: https://apps.who.int/iris/bitstream/handle/10665/246177/WHO-HIV-2016.06-eng.pdf?sequence=1
- 3 Razavi H, Sanchez Gonzalez Y, Yuen C, Cornberg M. Global timing of hepatitis C virus elimination in high-income countries. Liver Int 2020; 40: 522-529 [PMID: 31815353 DOI: 10.1111/liv.14324]
- 4 Jacobsen KH, Wiersma ST. Hepatitis A virus seroprevalence by age and world region, 1990 and 2005. Vaccine 2010; 28: 6653-6657 [PMID: 20723630 DOI: 10.1016/j.vaccine.2010.08.037]
- Jacobsen KH, Koopman JS. The effects of socioeconomic development on worldwide hepatitis A 5 virus seroprevalence patterns. Int J Epidemiol 2005; 34: 600-609 [PMID: 15831565 DOI: 10.1093/ije/dyi062]
- Jacobsen KH. Globalization and the Changing Epidemiology of Hepatitis A Virus. Cold Spring 6 Harb Perspect Med 2018; 8 [PMID: 29500305 DOI: 10.1101/cshperspect.a031716]
- World Health Organization. World Health Organisation Immunological basis for Immunization 7 Series. Module 18: Hepatitis A Update 2019. [cited 10 May 2021]. Available from: https://apps.who.int/iris/handle/10665/326501
- Kaddoura M, Allaham R, Abubakar A, Ezzeddine A, Barakat A, Mala P, Zaraket H. Hepatitis A Virus Genotype IB Outbreak among Internally Displaced Persons, Syria. Emerg Infect Dis 2020; 26: 369-371 [PMID: 31829918 DOI: 10.3201/eid2602.190652]
- 9 Choi YS, Lee J, Lee HW, Chang DY, Sung PS, Jung MK, Park JY, Kim JK, Lee JI, Park H, Cheong JY, Suh KS, Kim HJ, Lee JS, Kim KA, Shin EC. Liver injury in acute hepatitis A is associated with decreased frequency of regulatory T cells caused by Fas-mediated apoptosis. Gut 2015; 64: 1303-1313 [PMID: 25007815 DOI: 10.1136/gutjnl-2013-306213]
- 10 Fujiwara K, Kojima H, Yonemitsu Y, Yasui S, Imazeki F, Miki M, Suzuki K, Sakaida I, Okita K, Tanaka E, Omata M, Yokosuka O. Phylogenetic analysis of hepatitis A virus in sera from patients with hepatitis A of various severities. Liver Int 2009; 29: 838-845 [PMID: 19040539 DOI: 10.1111/j.1478-3231.2008.01919.x]
- 11 World Health Organization. Hepatitis A. 2020. [cited 27 April 2021]. Available from:



https://www.who.int/news-room/fact-sheets/detail/hepatitis-a

- 12 Advisory Committee on Immunization Practices (ACIP), Fiore AE, Wasley A, Bell BP. Prevention of hepatitis A through active or passive immunization: recommendations of the Advisory Committee on Immunization Practices (ACIP). MMWR Recomm Rep 2006; 55: 1-23 [PMID: 16708058
- 13 Herzog C, Van Herck K, Van Damme P. Hepatitis A vaccination and its immunological and epidemiological long-term effects - a review of the evidence. Hum Vaccin Immunother 2021: 17: 1496-1519 [PMID: 33325760 DOI: 10.1080/21645515.2020.1819742]
- 14 Sun X, Wang F, Zheng H, Miao N, Yuan Q, Cui F, Yin Z, Zhang G, Levine H. The impact of expanded program on immunization with live attenuated and inactivated Hepatitis A vaccines in China, 2004-2016. Vaccine 2018; 36: 1279-1284 [PMID: 29398275 DOI: 10.1016/j.vaccine.2018.01.043]
- 15 National Institute for Health and Care Excellence. Scenario: Prevention of infection with hepatitis A. [cited 20 May 2021]. London: NICE; 2021. Available from: https://cks.nice.org.uk/topics/hepatitis-a/management/prevention-of-infection-with-hepatitis-a/
- 16 United Nations High Commissioner for Refugees. Global Trends: Forced Displacement in 2019: The United Nations Refugee Agency; 2019. [cited 20 May 2021]. Available from: https://www.unhcr.org/globaltrends2019/
- 17 Zimmermann R, Faber M, Dudareva S, Ingiliz P, Jessen H, Koch J, Marcus U, Michaelis K, Rieck T, Ruscher C, Schilling B, Schumacher J, Sissolak D, Thoulass J, Wenzel JJ, Werber D, Sagebiel D. Hepatitis A outbreak among MSM in Berlin due to low vaccination coverage: Epidemiology, management, and successful interventions. Int J Infect Dis 2021; 103: 146-153 [PMID: 33207272 DOI: 10.1016/j.jjid.2020.11.1331
- 18 Hu X, Collier MG, Xu F. Hepatitis A Outbreaks in Developed Countries: Detection, Control, and Prevention. Foodborne Pathog Dis 2020; 17: 166-171 [PMID: 31829731 DOI: 10.1089/fpd.2019.2648]
- 19 Yin S, Barker L, Ly KN, Kilmer G, Foster MA, Drobeniuc J, Jiles RB. Susceptibility to Hepatitis A Virus Infection in the United States, 2007-2016. Clin Infect Dis 2020; 71: e571-e579 [PMID: 32193542 DOI: 10.1093/cid/ciaa298]
- 20 Jacobs RJ, Meyerhoff AS, Saab S. Immunization needs of chronic liver disease patients seen in primary care versus specialist settings. Dig Dis Sci 2005; 50: 1525-1531 [PMID: 16110847 DOI: 10.1007/s10620-005-2873-5
- 21 Shim M, Khaykis I, Park J, Bini EJ. Susceptibility to hepatitis A in patients with chronic liver disease due to hepatitis C virus infection: missed opportunities for vaccination. Hepatology 2005; 42: 688-695 [PMID: 16104047 DOI: 10.1002/hep.20830]
- 22 Chaudhary RK, Andonov AP. Effect of ribavirin on hepatitis A virus replication in vitro. Can J Infect Dis 1992; 3: 67-70 [PMID: 22529734 DOI: 10.1155/1992/531837]
- Howell J, Pedrana A, Schroeder SE, Scott N, Aufegger L, Atun R, Baptista-Leite R, Hirnschall G, 't 23 Hoen E, Hutchinson SJ, Lazarus JV, Olufunmilayo L, Peck R, Sharma M, Sohn AH, Thompson A, Thursz M, Wilson D, Hellard M. A global investment framework for the elimination of hepatitis B. J Hepatol 2021; 74: 535-549 [PMID: 32971137 DOI: 10.1016/j.jhep.2020.09.013]
- 24 World Health Organization. Hepatitis B. 2020. [cited 20 May 2021]. Available from: https://www.who.int/news-room/fact-sheets/detail/hepatitis-b
- 25 European Association for the Study of the Liver. EASL 2017 Clinical Practice Guidelines on the management of hepatitis B virus infection. J Hepatol 2017; 67: 370-398 [PMID: 28427875 DOI: 10.1016/j.jhep.2017.03.021]
- Terrault NA, Lok ASF, McMahon BJ, Chang KM, Hwang JP, Jonas MM, Brown RS Jr, Bzowej 26 NH, Wong JB. Update on prevention, diagnosis, and treatment of chronic hepatitis B: AASLD 2018 hepatitis B guidance. Hepatology 2018; 67: 1560-1599 [PMID: 29405329 DOI: 10.1002/hep.29800]
- 27 Zoulim F, Locarnini S. Hepatitis B virus resistance to nucleos(t)ide analogues. Gastroenterology 2009; 137: 1593-608.e1 [PMID: 19737565 DOI: 10.1053/j.gastro.2009.08.063]
- Do A, Reau NS. Chronic Viral Hepatitis: Current Management and Future Directions. Hepatol 28 Commun 2020; 4: 329-341 [PMID: 32140652 DOI: 10.1002/hep4.1480]
- 29 van Bömmel F, de Man RA, Wedemeyer H, Deterding K, Petersen J, Buggisch P, Erhardt A, Hüppe D, Stein K, Trojan J, Sarrazin C, Böcher WO, Spengler U, Wasmuth HE, Reinders JG, Möller B, Rhode P, Feucht HH, Wiedenmann B, Berg T. Long-term efficacy of tenofovir monotherapy for hepatitis B virus-monoinfected patients after failure of nucleoside/nucleotide analogues. Hepatology 2010; **51**: 73-80 [PMID: 19998272 DOI: 10.1002/hep.23246]
- 30 Tan M, Bhadoria AS, Cui F, Tan A, Van Holten J, Easterbrook P, Ford N, Han Q, Lu Y, Bulterys M, Hutin Y. Estimating the proportion of people with chronic hepatitis B virus infection eligible for hepatitis B antiviral treatment worldwide: a systematic review and meta-analysis. Lancet Gastroenterol Hepatol 2021; 6: 106-119 [PMID: 33197397 DOI: 10.1016/S2468-1253(20)30307-1]
- McPherson S, Valappil M, Moses SE, Eltringham G, Miller C, Baxter K, Chan A, Shafiq K, Saeed 31 A, Qureshi R, Hudson M, Bassendine MF. Targeted case finding for hepatitis B using dry blood spot testing in the British-Chinese and South Asian populations of the North-East of England. J Viral Hepat 2013; 20: 638-644 [PMID: 23910648 DOI: 10.1111/jvh.12084]
- 32 Martin NK, Vickerman P, Khakoo S, Ghosh A, Ramsay M, Hickman M, Williams J, Miners A. Chronic hepatitis B virus case-finding in UK populations born abroad in intermediate or high endemicity countries: an economic evaluation. BMJ Open 2019; 9: e030183 [PMID: 31256040 DOI:



10.1136/bmjopen-2019-030183]

- 33 Avellon A, Ala A, Diaz A, Domingo D, Gonzalez R, Hidalgo L, Kooner P, Loganathan S, Martin D, McPherson S, Munoz-Chimeno M, Ryder S, Slapak G, Ryan P, Valbuena M, Kennedy PT. Clinical performance of Determine HBsAg 2 rapid test for Hepatitis B detection. J Med Virol 2020 [PMID: 32270883 DOI: 10.1002/jmv.25862]
- 34 Nguyen MH, Wong G, Gane E, Kao JH, Dusheiko G. Hepatitis B Virus: Advances in Prevention, Diagnosis, and Therapy. Clin Microbiol Rev 2020; 33 [PMID: 32102898 DOI: 10.1128/CMR.00046-19
- 35 McNaughton AL, Lourenço J, Bester PA, Mokaya J, Lumley SF, Obolski U, Forde D, Maponga TG, Katumba KR, Goedhals D, Gupta S, Seeley J, Newton R, Ocama P, Matthews PC. Hepatitis B virus seroepidemiology data for Africa: Modelling intervention strategies based on a systematic review and meta-analysis. PLoS Med 2020; 17: e1003068 [PMID: 32315297 DOI: 10.1371/journal.pmed.1003068]
- 36 Revill PA, Chisari FV, Block JM, Dandri M, Gehring AJ, Guo H, Hu J, Kramvis A, Lampertico P, Janssen HLA, Levrero M, Li W, Liang TJ, Lim SG, Lu F, Penicaud MC, Tavis JE, Thimme R; Members of the ICE-HBV Working Groups; ICE-HBV Stakeholders Group Chairs; ICE-HBV Senior Advisors, Zoulim F. A global scientific strategy to cure hepatitis B. Lancet Gastroenterol Hepatol 2019; 4: 545-558 [PMID: 30981686 DOI: 10.1016/S2468-1253(19)30119-0]
- 37 Zhou K, Contag C, Whitaker E, Terrault N. Spontaneous loss of surface antigen among adults living with chronic hepatitis B virus infection: a systematic review and pooled meta-analyses. Lancet Gastroenterol Hepatol 2019; 4: 227-238 [PMID: 30679109 DOI: 10.1016/S2468-1253(18)30308-X]
- 38 Cornberg M, Lok AS, Terrault NA, Zoulim F; 2019 EASL-AASLD HBV Treatment Endpoints Conference Faculty. Guidance for design and endpoints of clinical trials in chronic hepatitis B -Report from the 2019 EASL-AASLD HBV Treatment Endpoints Conference[‡]. J Hepatol 2020; 72: 539-557 [PMID: 31730789 DOI: 10.1016/j.jhep.2019.11.003]
- Hyer R, McGuire DK, Xing B, Jackson S, Janssen R. Safety of a two-dose investigational hepatitis 39 B vaccine, HBsAg-1018, using a toll-like receptor 9 agonist adjuvant in adults. Vaccine 2018; 36: 2604-2611 [PMID: 29628151 DOI: 10.1016/j.vaccine.2018.03.067]
- 40 Wen WH, Lai MW, Chang MH. A review of strategies to prevent mother-to-infant transmission of hepatitis B virus infection. Expert Rev Gastroenterol Hepatol 2016; 10: 317-330 [PMID: 26566769 DOI: 10.1586/17474124.2016.1120667]
- Funk AL, Lu Y, Yoshida K, Zhao T, Boucheron P, van Holten J, Chou R, Bulterys M, Shimakawa 41 Y. Efficacy and safety of antiviral prophylaxis during pregnancy to prevent mother-to-child transmission of hepatitis B virus: a systematic review and meta-analysis. Lancet Infect Dis 2021; 21: 70-84 [PMID: 32805200 DOI: 10.1016/S1473-3099(20)30586-7]
- 42 World Health Organization. Prevention of Mother-To-Child Transmission of Hepatitis B virus: Guidelines on antiviral prophylaxis. [cited 20 May 2021]. Available from: https://apps.who.int/iris/bitstream/handle/10665/333391/9789240002708eng.pdf?sequence=1&isAllowed=y
- Liaw YF. HBeAg seroconversion as an important end point in the treatment of chronic hepatitis B. Hepatol Int 2009; 3: 425-433 [PMID: 19669245 DOI: 10.1007/s12072-009-9140-3]
- 44 Hsu YS, Chien RN, Yeh CT, Sheen IS, Chiou HY, Chu CM, Liaw YF. Long-term outcome after spontaneous HBeAg seroconversion in patients with chronic hepatitis B. Hepatology 2002; 35: 1522-1527 [PMID: 12029639 DOI: 10.1053/jhep.2002.33638]
- 45 Polaris Observatory Collaborators. Global prevalence, treatment, and prevention of hepatitis B virus infection in 2016: a modelling study. Lancet Gastroenterol Hepatol 2018; 3: 383-403 [PMID: 29599078 DOI: 10.1016/S2468-1253(18)30056-6]
- 46 Mak LY, Wong DK, Cheung KS, Seto WK, Lai CL, Yuen MF. Review article: hepatitis B corerelated antigen (HBcrAg): an emerging marker for chronic hepatitis B virus infection. Aliment Pharmacol Ther 2018; 47: 43-54 [PMID: 29035003 DOI: 10.1111/apt.14376]
- 47 van Bömmel F, Bartens A, Mysickova A, Hofmann J, Krüger DH, Berg T, Edelmann A. Serum hepatitis B virus RNA levels as an early predictor of hepatitis B envelope antigen seroconversion during treatment with polymerase inhibitors. Hepatology 2015; 61: 66-76 [PMID: 25132147 DOI: 10.1002/hep.27381]
- MacLachlan JH, Cowie BC. Hepatitis B virus epidemiology. Cold Spring Harb Perspect Med 48 2015; 5: a021410 [PMID: 25934461 DOI: 10.1101/cshperspect.a021410]
- 49 Papatheodoridis GV, Lampertico P, Manolakopoulos S, Lok A. Incidence of hepatocellular carcinoma in chronic hepatitis B patients receiving nucleos(t)ide therapy: a systematic review. J Hepatol 2010; 53: 348-356 [PMID: 20483498 DOI: 10.1016/j.jhep.2010.02.035]
- 50 Sherman M. Hepatocellular carcinoma: epidemiology, surveillance, and diagnosis. Semin Liver Dis 2010; **30**: 3-16 [PMID: 20175029 DOI: 10.1055/s-0030-1247128]
- Yang HI, Yuen MF, Chan HL, Han KH, Chen PJ, Kim DY, Ahn SH, Chen CJ, Wong VW, Seto 51 WK; REACH-B Working Group. Risk estimation for hepatocellular carcinoma in chronic hepatitis B (REACH-B): development and validation of a predictive score. Lancet Oncol 2011; 12: 568-574 [PMID: 21497551 DOI: 10.1016/S1470-2045(11)70077-8]
- 52 Papatheodoridis G, Dalekos G, Sypsa V, Yurdaydin C, Buti M, Goulis J, Calleja JL, Chi H, Manolakopoulos S, Mangia G, Gatselis N, Keskin O, Savvidou S, de la Revilla J, Hansen BE, Vlachogiannakos I, Galanis K, Idilman R, Colombo M, Esteban R, Janssen HL, Lampertico P. PAGE-B predicts the risk of developing hepatocellular carcinoma in Caucasians with chronic



hepatitis B on 5-year antiviral therapy. J Hepatol 2016; 64: 800-806 [PMID: 26678008 DOI: 10.1016/j.jhep.2015.11.035]

- 53 Voulgaris T, Papatheodoridi M, Lampertico P, Papatheodoridis GV. Clinical utility of hepatocellular carcinoma risk scores in chronic hepatitis B. Liver Int 2020; 40: 484-495 [PMID: 31884726 DOI: 10.1111/liv.14334]
- 54 Lee HW, Ahn SH. Prediction models of hepatocellular carcinoma development in chronic hepatitis B patients. World J Gastroenterol 2016; 22: 8314-8321 [PMID: 27729738 DOI: 10.3748/wjg.v22.i37.8314]
- 55 Suzuki Y, Maekawa S, Komatsu N, Sato M, Tatsumi A, Miura M, Matsuda S, Muraoka M, Nakakuki N, Shindo H, Amemiya F, Takano S, Fukasawa M, Nakayama Y, Yamaguchi T, Inoue T, Sato T, Sakamoto M, Yamashita A, Moriishi K, Enomoto N. Hepatitis B virus (HBV)-infected patients with low hepatitis B surface antigen and high hepatitis B core-related antigen titers have a high risk of HBV-related hepatocellular carcinoma. Hepatol Res 2019; 49: 51-63 [PMID: 30350374 DOI: 10.1111/hepr.13277]
- 56 Ligat G, Goto K, Verrier E, Baumert TF. Targeting Viral cccDNA for Cure of Chronic Hepatitis B. Cur Hepatol Rep 2020; 19: 235-44 [DOI: 10.1007/s11901-020-00534-w]
- 57 European Union's Horizon 2020 research and innovation programme. TherVacB - A Therapeutic vaccine to cure Hepatitis B. [cited 20 May 2021]. Available from: https://www.thervacb.eu/
- Stockdale AJ, Kreuels B, Henrion MYR, Giorgi E, Kyomuhangi I, de Martel C, Hutin Y, Geretti 58 AM. The global prevalence of hepatitis D virus infection: Systematic review and meta-analysis. J Hepatol 2020; 73: 523-532 [PMID: 32335166 DOI: 10.1016/j.jhep.2020.04.008]
- Chen HY, Shen DT, Ji DZ, Han PC, Zhang WM, Ma JF, Chen WS, Goyal H, Pan S, Xu HG. 59 Prevalence and burden of hepatitis D virus infection in the global population: a systematic review and meta-analysis. Gut 2019; 68: 512-521 [PMID: 30228220 DOI: 10.1136/gutjnl-2018-316601]
- Miao Z, Zhang S, Ou X, Li S, Ma Z, Wang W, Peppelenbosch MP, Liu J, Pan Q. Estimating the 60 Global Prevalence, Disease Progression, and Clinical Outcome of Hepatitis Delta Virus Infection. J Infect Dis 2020; 221: 1677-1687 [PMID: 31778167 DOI: 10.1093/infdis/jiz633]
- El Bouzidi K, Elamin W, Kranzer K, Irish DN, Ferns B, Kennedy P, Rosenberg W, Dusheiko G, 61 Sabin CA, Smith BC, Nastouli E. Hepatitis delta virus testing, epidemiology and management: a multicentre cross-sectional study of patients in London. J Clin Virol 2015; 66: 33-37 [PMID: 25866333 DOI: 10.1016/j.jcv.2015.02.011]
- 62 Ferrante ND, Lo Re V 3rd. Epidemiology, Natural History, and Treatment of Hepatitis Delta Virus Infection in HIV/Hepatitis B Virus Coinfection. Curr HIV/AIDS Rep 2020; 17: 405-414 [PMID: 32607773 DOI: 10.1007/s11904-020-00508-z]
- 63 Urban S, Neumann-Haefelin C, Lampertico P. Hepatitis D virus in 2021: virology, immunology and new treatment approaches for a difficult-to-treat disease. Gut 2021; 70: 1782-1794 [PMID: 34103404 DOI: 10.1136/gutjnl-2020-323888]
- 64 World Health Organization. Hepatitis C. 2020. [cited 20 May 2021]. Available from: https://www.who.int/news-room/fact-sheets/detail/hepatitis-c
- 65 Park C, Jiang S, Lawson KA. Efficacy and safety of telaprevir and boceprevir in patients with hepatitis C genotype 1: a meta-analysis. J Clin Pharm Ther 2014; 39: 14-24 [PMID: 24237070 DOI: 10.1111/jcpt.12106
- Martinello M, Bajis S, Dore GJ. Progress Toward Hepatitis C Virus Elimination: Therapy and 66 Implementation. Gastroenterol Clin North Am 2020; 49: 253-277 [PMID: 32389362 DOI: 10.1016/j.gtc.2020.01.005]
- 67 World Health Organization. Guidelines for the Care and Treatment of Persons Diagnosed with Chronic Hepatitis C Virus Infection. [cited 20 May 2021]. Available from: http://apps.who.int/iris/bitstream/handle/10665/273174/9789241550345-eng.pdf?ua=1
- Iliescu EL, Mercan-Stanciu A, Toma L. Safety and efficacy of direct-acting antivirals for chronic 68 hepatitis C in patients with chronic kidney disease. BMC Nephrol 2020; 21: 21 [PMID: 31948406 DOI: 10.1186/s12882-020-1687-1]
- 69 Velosa J. Why is viral eradication so important in patients with HCV-related cirrhosis? Antivir Ther 2017; 22: 1-12 [PMID: 27553973 DOI: 10.3851/IMP3077]
- 70 Bang CS, Song IH. Impact of antiviral therapy on hepatocellular carcinoma and mortality in patients with chronic hepatitis C: systematic review and meta-analysis. BMC Gastroenterol 2017; 17: 46 [PMID: 28376711 DOI: 10.1186/s12876-017-0606-9]
- 71 Stoll-Keller F, Barth H, Fafi-Kremer S, Zeisel MB, Baumert TF. Development of hepatitis C virus vaccines: challenges and progress. Expert Rev Vaccines 2009; 8: 333-345 [PMID: 19249975 DOI: 10.1586/14760584.8.3.333]
- 72 Page K, Melia MT, Veenhuis RT, Winter M, Rousseau KE, Massaccesi G, Osburn WO, Forman M, Thomas E, Thornton K, Wagner K, Vassilev V, Lin L, Lum PJ, Giudice LC, Stein E, Asher A, Chang S, Gorman R, Ghany MG, Liang TJ, Wierzbicki MR, Scarselli E, Nicosia A, Folgori A, Capone S, Cox AL. Randomized Trial of a Vaccine Regimen to Prevent Chronic HCV Infection. N Engl J Med 2021; 384: 541-549 [PMID: 33567193 DOI: 10.1056/NEJMoa2023345]
- 73 US Department of Health and Human Services (HHS). Facing addiction in America: the Surgeon General's spotlight on opioids. [cited 19 May 2021]. Available from: https://addiction.surgeongeneral.gov/sites/default/files/OC SpotlightOnOpioids.pdf
- 74 Platt L, Minozzi S, Reed J, Vickerman P, Hagan H, French C, Jordan A, Degenhardt L, Hope V,



Hutchinson S, Maher L, Palmateer N, Taylor A, Bruneau J, Hickman M. Needle and syringe programmes and opioid substitution therapy for preventing HCV transmission among people who inject drugs: findings from a Cochrane Review and meta-analysis. Addiction 2018; 113: 545-563 [PMID: 28891267 DOI: 10.1111/add.14012]

- 75 Reyes-Urueña J, Celly A, Moreno S, Majó X, Colom J, Casabona J. Hepatitis C virus: Testing rate and attrition at linkage to specialized care, Catalonia, Spain 2011-2016. J Viral Hepat 2021; 28: 288-299 [PMID: 33098176 DOI: 10.1111/jvh.13427]
- Nyamathi AM, Dixon EL, Robbins W, Smith C, Wiley D, Leake B, Longshore D, Gelberg L. Risk 76 factors for hepatitis C virus infection among homeless adults. J Gen Intern Med 2002; 17: 134-143 [PMID: 11841529 DOI: 10.1046/j.1525-1497.2002.10415.x]
- 77 The Hepatitis C Trust. Community Peer Programme. [cited 20 may 2021]. Available from: http://www.hepctrust.org.uk/services/community-peer-programme
- 78 Gallacher J, McPherson S. Progress towards micro-elimination of hepatitis C in the custodial setting. J Viral Hepat 2021; 28: 300-301 [PMID: 33131191 DOI: 10.1111/jvh.13428]
- 79 European Association for the Study of the Liver. EASL Recommendations on Treatment of Hepatitis C 2018. J Hepatol 2018; 69: 461-511 [PMID: 29650333 DOI: 10.1016/j.jhep.2018.03.026]
- 80 Hajarizadeh B, Grebely J, Byrne M, Marks P, Amin J, McManus H, Butler T, Cunningham EB, Vickerman P, Martin NK, McHutchison JG, Brainard DM, Treloar C, Chambers GM, Grant L, Mcgrath C, Lloyd AR, Dore GJ; SToP-C study group. Evaluation of hepatitis C treatment-asprevention within Australian prisons (SToP-C): a prospective cohort study. Lancet Gastroenterol Hepatol 2021; 6: 533-546 [PMID: 33965006 DOI: 10.1016/S2468-1253(21)00077-7]
- 81 Donaldson SR, Radley A, Dillon JF. Identifying the Hidden Population: Former Intravenous Drug Users Who Are No Longer in Contact with Services. "Ask a Friend". Diagnostics (Basel) 2021; 11 [PMID: 33504077 DOI: 10.3390/diagnostics11020170]
- Papaluca T, McDonald L, Craigie A, Gibson A, Desmond P, Wong D, Winter R, Scott N, Howell J, 82 Doyle J, Pedrana A, Lloyd A, Stoove M, Hellard M, Iser D, Thompson A. Outcomes of treatment for hepatitis C in prisoners using a nurse-led, statewide model of care. J Hepatol 2019; 70: 839-846 [PMID: 30654067 DOI: 10.1016/j.jhep.2019.01.012]
- 83 Rege SM, Gonzalez, Sanchez Y PhD, Marx, PharmD2 S; Reau, Nancy MD, FACG3. Patient Flow Across Physician Specialties Over the Course of the Hepatitis C Care Cascade: A Real World Analysis From the United States. Am J Gastroenterol 2019; 114: s561
- 84 Simpson H, Manley P, Lawler J, Morey S, Buchanan E, Hewett M, Knowles J, Miller C, McCarron B, Valappil M, McPherson S. Distance to treatment as a factor for loss to follow up of hepatitis C patients in North East England. J Public Health (Oxf) 2019; 41: 700-706 [PMID: 30351415 DOI: 10.1093/pubmed/fdy190]
- 85 Radley A, de Bruin M, Inglis SK, Donnan PT, Hapca A, Barclay ST, Fraser A, Dillon JF. Clinical effectiveness of pharmacist-led versus conventionally delivered antiviral treatment for hepatitis C virus in patients receiving opioid substitution therapy: a pragmatic, cluster-randomised trial. Lancet Gastroenterol Hepatol 2020; 5: 809-818 [PMID: 32526210 DOI: 10.1016/S2468-1253(20)30120-5]
- 86 Hashim A, O'Sullivan M, Williams H, Verma S. Developing a community HCV service: project ITTREAT (integrated community-based test - stage - TREAT) service for people who inject drugs. Prim Health Care Res Dev 2018; 19: 110-120 [PMID: 29199921 DOI: 10.1017/81463423617000731
- Simmons B, Saleem J, Hill A, Riley RD, Cooke GS. Risk of Late Relapse or Reinfection With Hepatitis C Virus After Achieving a Sustained Virological Response: A Systematic Review and Meta-analysis. Clin Infect Dis 2016; 62: 683-694 [PMID: 26787172 DOI: 10.1093/cid/civ948]
- 88 Louie KS, St Laurent S, Forssen UM, Mundy LM, Pimenta JM. The high comorbidity burden of the hepatitis C virus infected population in the United States. BMC Infect Dis 2012; 12: 86 [PMID: 22494445 DOI: 10.1186/1471-2334-12-86]
- 89 McPherson S, Gosrani S, Hogg S, Patel P, Wetten A, Welton R, Hallsworth K, Campbell M. Increased cardiovascular risk and reduced quality of life are highly prevalent among individuals with hepatitis C. BMJ Open Gastroenterol 2020; 7 [PMID: 32847899 DOI: 10.1136/bmjgast-2020-000470]
- 90 Alonso López S, Manzano ML, Gea F, Gutiérrez ML, Ahumada AM, Devesa MJ, Olveira A, Polo BA, Márquez L, Fernández I, Cobo JCR, Rayón L, Riado D, Izquierdo S, Usón C, Real Y, Rincón D, Fernández-Rodríguez CM, Bañares R. A Model Based on Noninvasive Markers Predicts Very Low Hepatocellular Carcinoma Risk After Viral Response in Hepatitis C Virus-Advanced Fibrosis. Hepatology 2020; 72: 1924-1934 [PMID: 33022803 DOI: 10.1002/hep.31588]
- Ioannou GN, Beste LA, Green PK, Singal AG, Tapper EB, Waljee AK, Sterling RK, Feld JJ, 91 Kaplan DE, Taddei TH, Berry K. Increased Risk for Hepatocellular Carcinoma Persists Up to 10 Years After HCV Eradication in Patients With Baseline Cirrhosis or High FIB-4 Scores. Gastroenterology 2019; 157: 1264-1278.e4 [PMID: 31356807 DOI: 10.1053/j.gastro.2019.07.033]
- 92 Toyoda H, Tada T, Yasuda S, Mizuno K, Ito T, Kumada T. Dynamic Evaluation of Liver Fibrosis to Assess the Risk of Hepatocellular Carcinoma in Patients With Chronic Hepatitis C Who Achieved Sustained Virologic Response. Clin Infect Dis 2020; 70: 1208-1214 [PMID: 31056696 DOI: 10.1093/cid/ciz359]
- 93 Kahn JA. The use of organs from hepatitis C virus-viremic donors into uninfected recipients. Curr Opin Organ Transplant 2020; 25: 620-625 [PMID: 33105203 DOI: 10.1097/MOT.0000000000826]



- 94 Ting PS, Hamilton JP, Gurakar A, Urrunaga NH, Ma M, Glorioso J, King E, Toman LP, Wesson R, Garonzik-Wang J, Ottmann S, Philosophe B, Sulkowski M, Cameron AM, Durand CM, Chen PH. Hepatitis C-positive donor liver transplantation for hepatitis C seronegative recipients. Transpl Infect Dis 2019; 21: e13194 [PMID: 31609520 DOI: 10.1111/tid.13194]
- 95 Kwong AJ, Wall A, Melcher M, Wang U, Ahmed A, Subramanian A, Kwo PY. Liver transplantation for hepatitis C virus (HCV) non-viremic recipients with HCV viremic donors. Am J Transplant 2019; 19: 1380-1387 [PMID: 30378723 DOI: 10.1111/ajt.15162]
- Rein DB, Stevens GA, Theaker J, Wittenborn JS, Wiersma ST. The global burden of hepatitis E 96 genotypes 1 and 2 in 2005. Hepatology 2012; 55: 988-997 [PMID: 22121109 DOI: 10.1002/hep.25505]
- 97 Gallian P, Pouchol E, Djoudi R, Lhomme S, Mouna L, Gross S, Bierling P, Assal A, Kamar N, Mallet V, Roque-Afonso AM, Izopet J, Tiberghien P. Transfusion-Transmitted Hepatitis E Virus Infection in France. Transfus Med Rev 2019; 33: 146-153 [PMID: 31327668 DOI: 10.1016/j.tmrv.2019.06.001]
- 98 Harvala H, Hewitt PE, Reynolds C, Pearson C, Haywood B, Tettmar KI, Ushiro-Lumb I, Brailsford SR, Tedder R, Ijaz S. Hepatitis E virus in blood donors in England, 2016 to 2017: from selective to universal screening. Euro Surveill 2019; 24 [PMID: 30862338 DOI: 10.2807/1560-7917.ES.2019.24.10.1800386]
- Dreier J, Knabbe C, Vollmer T. Transfusion-Transmitted Hepatitis E: NAT Screening of Blood 99 Donations and Infectious Dose. Front Med (Lausanne) 2018; 5: 5 [PMID: 29450199 DOI: 10.3389/fmed.2018.00005
- 100 Domanović D, Tedder R, Blümel J, Zaaijer H, Gallian P, Niederhauser C, Sauleda Oliveras S, O'Riordan J, Boland F, Harritshøj L, Nascimento MSJ, Ciccaglione AR, Politis C, Adlhoch C, Flan B, Oualikene-Gonin W, Rautmann G, Strengers P, Hewitt P. Hepatitis E and blood donation safety in selected European countries: a shift to screening? Euro Surveill 2017; 22 [PMID: 28449730 DOI: 10.2807/1560-7917.ES.2017.22.16.30514]
- 101 Lawrence D GY, Clarke A, Fisher M, Richardson D. P34 Two cases of acute hepatitis e causing a transient transaminitis in hiv infected msm. Sexually Transmitted Infections 2015; A26-A7 [DOI: 10.1136/sextrans-2015-052126.78]
- 102 European Association for the Study of the Liver. EASL Clinical Practice Guidelines on hepatitis E virus infection. J Hepatol 2018; 68: 1256-1271 [PMID: 29609832 DOI: 10.1016/j.jhep.2018.03.005]
- 103 Kamar N, Garrouste C, Haagsma EB, Garrigue V, Pischke S, Chauvet C, Dumortier J, Cannesson A, Cassuto-Viguier E, Thervet E, Conti F, Lebray P, Dalton HR, Santella R, Kanaan N, Essig M, Mousson C, Radenne S, Roque-Afonso AM, Izopet J, Rostaing L. Factors associated with chronic hepatitis in patients with hepatitis E virus infection who have received solid organ transplants. Gastroenterology 2011; 140: 1481-1489 [PMID: 21354150 DOI: 10.1053/j.gastro.2011.02.050]
- 104 Kamar N, Selves J, Mansuy JM, Ouezzani L, Péron JM, Guitard J, Cointault O, Esposito L, Abravanel F, Danjoux M, Durand D, Vinel JP, Izopet J, Rostaing L. Hepatitis E virus and chronic hepatitis in organ-transplant recipients. N Engl J Med 2008; 358: 811-817 [PMID: 18287603 DOI: 10.1056/NEJMoa0706992
- 105 Kamar N, Abravanel F, Behrendt P, Hofmann J, Pageaux GP, Barbet C, Moal V, Couzi L, Horvatits T, De Man RA, Cassuto E, Elsharkawy AM, Riezebos-Brilman A, Scemla A, Hillaire S, Donnelly MC, Radenne S, Sayegh J, Garrouste C, Dumortier J, Glowaki F, Matignon M, Coilly A, Figueres L, Mousson C, Minello A, Dharancy S, Rerolle JP, Lebray P, Etienne I, Perrin P, Choi M, Marion O, Izopet J; Hepatitis E Virus Ribavirin Study Group. Ribavirin for Hepatitis E Virus Infection After Organ Transplantation: A Large European Retrospective Multicenter Study. Clin Infect Dis 2020; 71: 1204-1211 [PMID: 31793638 DOI: 10.1093/cid/ciz953]
- 106 Kamar N, Izopet J, Tripon S, Bismuth M, Hillaire S, Dumortier J, Radenne S, Coilly A, Garrigue V, D'Alteroche L, Buchler M, Couzi L, Lebray P, Dharancy S, Minello A, Hourmant M, Roque-Afonso AM, Abravanel F, Pol S, Rostaing L, Mallet V. Ribavirin for chronic hepatitis E virus infection in transplant recipients. N Engl J Med 2014; 370: 1111-1120 [PMID: 24645943 DOI: 10.1056/NEJMoa1215246
- 107 Narayanan S, Abutaleb A, Sherman KE, Kottilil S. Clinical features and determinants of chronicity in hepatitis E virus infection. J Viral Hepat 2019; 26: 414-421 [PMID: 30636092 DOI: 10.1111/jvh.13059]
- Peters van Ton AM, Gevers TJ, Drenth JP. Antiviral therapy in chronic hepatitis E: a systematic 108 review. J Viral Hepat 2015; 22: 965-973 [PMID: 25760481 DOI: 10.1111/jvh.12403]
- 109 Dao Thi VL, Debing Y, Wu X, Rice CM, Neyts J, Moradpour D, Gouttenoire J. Sofosbuvir Inhibits Hepatitis E Virus Replication In Vitro and Results in an Additive Effect When Combined With Ribavirin. Gastroenterology 2016; 150: 82-85.e4 [PMID: 26408347 DOI: 10.1053/j.gastro.2015.09.011]
- 110 Donnelly MC, Imlach SN, Abravanel F, Ramalingam S, Johannessen I, Petrik J, Fraser AR, Campbell JD, Bramley P, Dalton HR, Hayes PC, Kamar N, Simpson KJ. Sofosbuvir and Daclatasvir Anti-Viral Therapy Fails to Clear HEV Viremia and Restore Reactive T Cells in a HEV/HCV Co-Infected Liver Transplant Recipient. Gastroenterology 2017; 152: 300-301 [PMID: 27883881 DOI: 10.1053/j.gastro.2016.05.060]
- 111 Cornberg M, Pischke S, Müller T, Behrendt P, Piecha F, Benckert J, Todt D, Steinmann E, Papkalla A, von Karpowitz M, Koch A, Lohse A, Hardtke S, Manns MP, Wedemeyer H. Sofosbuvir monotherapy fails to achieve HEV RNA elimination in patients with chronic hepatitis E - The



HepNet SofE pilot study. J Hepatol 2020; 73: 696-699 [PMID: 32624195 DOI: 10.1016/j.jhep.2020.05.020]

- 112 Lhomme S, Kamar N, Nicot F, Ducos J, Bismuth M, Garrigue V, Petitjean-Lecherbonnier J, Ollivier I, Alessandri-Gradt E, Goria O, Barth H, Perrin P, Saune K, Dubois M, Carcenac R, Lefebvre C, Jeanne N, Abravanel F, Izopet J. Mutation in the Hepatitis E Virus Polymerase and Outcome of Ribavirin Therapy. Antimicrob Agents Chemother 2015; 60: 1608-1614 [PMID: 26711757 DOI: 10.1128/AAC.02496-15]
- 113 Public Health England. Hepatitis C in Endland 2020 Working to eliminate hepatitis C as a major public health threat. [cited 20 May 2021]. Available from: https://assets.publishing.service.gov.uk/go vernment/uploads/system/uploads/attachment_data/file/898221/HCV_in_England_2020_report.pdf
- European Association for the Study of the Liver. Clinical Practice Guidelines Panel: Chair:; 114 EASL Governing Board representative:; Panel members:. EASL recommendations on treatment of hepatitis C: Final update of the series[☆]. *J Hepatol* 2020; **73**: 1170-1218 [PMID: 32956768 DOI: 10.1016/j.jhep.2020.08.018]
- AASLD-IDSA HCV Guidance Panel. Hepatitis C Guidance 2018 Update: AASLD-IDSA 115 Recommendations for Testing, Managing, and Treating Hepatitis C Virus Infection. Clin Infect Dis 2018; 67: 1477-1492 [PMID: 30215672 DOI: 10.1093/cid/ciy585]
- Omata M, Cheng AL, Kokudo N, Kudo M, Lee JM, Jia J, Tateishi R, Han KH, Chawla YK, Shiina 116 S, Jafri W, Payawal DA, Ohki T, Ogasawara S, Chen PJ, Lesmana CRA, Lesmana LA, Gani RA, Obi S, Dokmeci AK, Sarin SK. Asia-Pacific clinical practice guidelines on the management of hepatocellular carcinoma: a 2017 update. Hepatol Int 2017; 11: 317-370 [PMID: 28620797 DOI: 10.1007/s12072-017-9799-9]



WJG

World Journal of Gastroenterology

Submit a Manuscript: https://www.f6publishing.com

World J Gastroenterol 2022 January 7; 28(1): 96-107

DOI: 10.3748/wjg.v28.i1.96

ISSN 1007-9327 (print) ISSN 2219-2840 (online)

MINIREVIEWS

Risk of hepatocellular carcinoma after hepatitis C virus cure

Maria Alejandra Luna-Cuadros, Hao-Wei Chen, Hira Hanif, Mukarram Jamat Ali, Muzammil Muhammad Khan, Daryl Tan-Yeung Lau

ORCID number: Maria Alejandra Luna-Cuadros 0000-0001-9259-2565; Hao-Wei Chen 0000-0003-0279-4214; Hira Hanif 0000-0002-1888-0056; Mukarram Jamat Ali 0000-0003-3313-9933; Muzammil Muhammad Khan 0000-0003-1324-3637; Daryl Tan-Yeung Lau 0000-0003-4139-1987.

Author contributions: Luna-Cuadros MA, Chen HW contributed equally to this work; Luna-Cuadros MA, Chen HW organized and wrote significant sections and revision of the manuscript; Hanif H, Khan MM contributed to the literature search and manuscript writing; Ali AJ designed the figures and contributed to the edit and revision of the manuscript; Lau DTY provided guidance on the overall concept and execution of the manuscript.

Conflict-of-interest statement:

Authors declare no conflict of interests for this article.

Country/Territory of origin: United States

Specialty type: Gastroenterology and hepatology

Provenance and peer review: Invited article; Externally peer reviewed.

Peer-review model: Single blind

Maria Alejandra Luna-Cuadros, Hao-Wei Chen, Hira Hanif, Mukarram Jamat Ali, Muzammil Muhammad Khan, Daryl Tan-Yeung Lau, Liver Center, Division of Gastroenterology and Hepatology, Beth Israel Deaconess Medical Center, Harvard Medical School, Boston, MA 02215, United States

Corresponding author: Daryl Tan-Yeung Lau, MD, MSc, Associate Professor, Liver Center, Division of Gastroenterology and Hepatology, Beth Israel Deaconess Medical Center, Harvard Medical School, 110 Francis Street, Suite 4A, Boston, MA 02215, United States. dlau@bidmc.harvard.edu

Abstract

Hepatitis C virus (HCV) is a significant cause of hepatocellular carcinoma (HCC). The direct-acting antivirals marked a new era of HCV therapy and are associated with greater than 95% cure rate. Successful treatment of chronic hepatitis C greatly reduces the risk of HCC. A proportion of patients, especially those with pre-existing cirrhosis, remain at risk for HCC despite sustained virologic response (SVR). Diabetes mellitus, hepatic steatosis, alcohol consumption and lack of fibrosis regression are associated with risks of HCC after HCV cure. Noninvasive modalities such as aspartate aminotransferase to platelet ratio index and fibrosis-4 index and transient elastography have been used to monitor hepatic fibrosis. More recently, various fibrosis scores have been combined with clinical parameters and other novel biomarkers to predict risks of HCC for patients who achieved SVR. These models still need to be validated and standardized prior to applying to routine clinical care.

Key Words: Hepatitis C virus cure; Hepatocellular carcinoma; Hepatocellular carcinoma risk models; Fibrosis markers; Transient elastography

©The Author(s) 2022. Published by Baishideng Publishing Group Inc. All rights reserved.

Core Tip: Direct-acting antivirals (DAA) therapy has revolutionized the treatment for chronic hepatitis C. However, the development of hepatocellular carcinoma (HCC) after achieving DAA-induced sustained virologic response remains a significant concern, especially those with advanced fibrosis. It is critically important to monitor hepatic fibrosis and continue HCC surveillance for patients with pre-existing cirrhosis. Lack of hepatic regression and several comorbid conditions are associated with HCC



WJG | https://www.wjgnet.com

Peer-review report's scientific quality classification

Grade A (Excellent): 0 Grade B (Very good): B Grade C (Good): C Grade D (Fair): 0 Grade E (Poor): 0

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: htt p://creativecommons.org/License s/by-nc/4.0/

Received: May 29, 2021 Peer-review started: May 29, 2021 First decision: June 22, 2021 Revised: July 12, 2021 Accepted: December 25, 2021 Article in press: December 25, 2021 Published online: January 7, 2022

P-Reviewer: Xu J, Yang L S-Editor: Wang JJ L-Editor: A P-Editor: Wang JJ



risks. Some promising models for predicting HCC risks after hepatitis C virus cure are in development.

Citation: Luna-Cuadros MA, Chen HW, Hanif H, Ali MJ, Khan MM, Lau DTY. Risk of hepatocellular carcinoma after hepatitis C virus cure. *World J Gastroenterol* 2022; 28(1): 96-107

URL: https://www.wjgnet.com/1007-9327/full/v28/i1/96.htm **DOI:** https://dx.doi.org/10.3748/wjg.v28.i1.96

INTRODUCTION

Hepatitis C virus (HCV) is a global health issue affecting 160-170 million people worldwide[1]. According to recent National Health and Nutrition Examination Survey data, there are approximately 2.4 million people with chronic hepatitis C (CHC) in the United States[2]. There are 6 major genotypes of HCV[3]. Globally, G1 is most common accounting for 49.1% of all infections among adults, followed by G3 (17.9%), G4 (16.8%), G2 (11.0%), G5 (2.0%) and G6 (1.4%)[3]. There are significant geographic variations in the 6 HCV genotypes (Table 1). G1 is the predominant HCV genotype, for example, in North America, Europe, Caribbean and Latin America. G4 is most common in North Africa especially Egypt and the Middle East. The high prevalence of G3 in Asia is largely contributed by South Asia in particular India and Pakistan[3].

HCV-related hepatocellular carcinoma (HCC) is the fifth most common cancer worldwide, accounting for 85%-90% of primary liver cancers[4]. Advanced stage liver fibrosis (Metavir stage F3) carries an increased risk of HCC, and patients with cirrhosis (Metavir stage F4) have an annual HCC incidence of approximately 4%[4]. With the advent of direct-acting antivirals (DAA) therapy, over 95% of the treated patients were able to achieve sustained virologic response (SVR) or HCV cure[5]. HCV cure reduces the HCC risk but those with preexisting cirrhosis remain at risk[6,7]. This review focused on the pathogenesis and risk factors of HCC after HCV cure, and the applications of noninvasive modalities and models to predict HCC.

NATURAL HISTORY OF HCV INFECTION

The transmission of HCV occurs mainly *via* blood with the majority due to unsafe injection use (intravenous drug use, healthcare workers in underdeveloped countries) and blood transfusion recipients before 1992[8]. Moreover, sexual transmission of HCV has significantly increased in human immunodeficiency virus-infected MSM in recent years[9,10].

After the virus transmission, HCV RNA reaches a detectable level in the serum in 7 to 21 d[11,12]. HCV RNA levels rise rapidly during acute infection but it generally takes 4-12 wk for the elevation of alanine aminotransferase (ALT) (indicative of hepatic injury) with an associated increase of serum bilirubin[13]. HCV itself is not cytolytic, but it generates potent innate and adaptive immune responses with cytotoxic cytokines production and hepatic injury[14]. Acute liver failure due to HCV is rare, but its incidence increases especially in patients with pre-existing chronic liver diseases[12].

Spontaneous eradication of HCV with recovery occurs only in only 15%-25% of patients with acute hepatitis C. The presence of homozygous rs12979860-C alleles in the interferon lambda gene, however, is associated with about 80% of spontaneous recovery[15,16].

CHC is defined as the persistence of HCV RNA six months after the initial infection. CHC can lead to progressive fibrosis, cirrhosis, end-stage liver disease and complicated with HCC (Figure 1). It is estimated that 20%-30% of patients with CHC will develop cirrhosis after a period of 20 years[17]. Monitoring the development of fibrosis over time can provide a more accurate progression to cirrhosis. A study of paired liver biopsies scored by the same pathologists suggested the time to develop cirrhosis from diagnosis is about 30 to 40 years[17].

Zaishidene® WJG | https://www.wjgnet.com

Luna-Cuadros MA et al. Risk of HCC after HCV cure

Table 1 Regional prevalence of hepatitis C virus genotypes								
Regions	G1 (%)	G2 (%)	G3 (%)	G4 (%)	G5 (%)	G6 (%)	Mixed	
Africa	26.3	23.7	6.3	28.1	12.2	-	3.4	
North Africa/Middle East	27.3	0.8	6.3	65.3	0.3	-	-	
North America	66.3	13.1	15.7	4.3	-	0.6	-	
Caribbean	83	7.2	2.1	0.6	-	0.1	7.0	
Central Latin America	74.6	21.6	3.3	0.1	0.1	-	0.3	
Central Asia	70.4	8.6	19.6	-	-	-	1.4	
South Asia	15.5	1.9	66.7	3.7	0.1	0.5	11.6	
Europe	64.4	5.5	25.5	3.7	0.1	0.1	0.7	
Australasia	55.0	6.5	36.0	1.2	-	1.3	-	

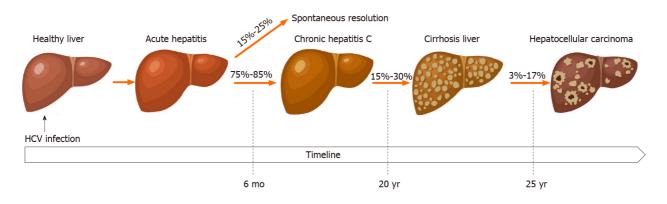


Figure 1 Natural history of chronic hepatitis. HCV: Hepatitis C virus.

After the progression to cirrhosis, patients are at increased risk of decompensated liver disease with associated complications such as ascites, spontaneous bacterial peritonitis, variceal bleeding and hepatic encephalopathy. The development of any of these complications is an indicator of increased risk of death or need for liver transplantation. Among patients with compensated cirrhosis, the 5-year and 10-year survival was 85%-91% and 60%-79% respectively[18]. The rate of clinical decompensation was 2%-5% per year and incidence of HCC was 1%-4% in these patients[18]. Generally, the risk for HCC and death increases significantly once decompensation develops[18].

HCV CURE

Treatment for HCV has revolutionized in the last decade. Before 2011, interferon was the mainstay of the therapy for HCV. Pegylated interferon combined with ribavirin had a success rate of 70% and 80% for genotype 2 and 3, respectively. However, the efficacy of interferon in HCV genotype 1 was low at 10%-20% only[19]. The advent of DAA marked the new era of HCV cure (Table 2). Boceprevir (Victrelis®) and Telaprevir (Incivek®) were the first DAA agents approved for the treatment of genotype 1 HCV infection and multiple other regimens obtained approval in the ensuing years. Since 2016, there are three pangenotypic combination therapies against genotype 1 to 6 with potent efficacy.

HCV cure or SVR is characterized by the absence of detectable HCV RNA in the serum 12 wk after the completion of DAA therapy[20]. In a meta-analysis of 43 studies, the risk of relapse or reinfection in the low-risk patients was 0.95% [95% confidence interval (CI): 0.35%-1.69%] over a 5-year period. Among the high-risk populations, such as injecting drug users or prisoners, the reinfection rate increased to 10.67% (95%CI: 6.38%-15.66%) in 5 years[21].

Zaishidene® WJG | https://www.wjgnet.com

Table 2 Current therapies for treatment of chronic hepatitis C							
Year approved	FDA approved therapy	Genotype	Trade name				
2011	PegIFN/RBV + Boceprevir	Genotype-1	Victrelis®				
2011	Telaprevir + PegIFNα/RBV	Genotype-1	Incivek®				
2013	Sofosbuvir + RBV or Sofosbuvir + PegIFNa/RBV	Genotype-1, 2, 3 and 4	Sovaldi [®]				
2014	Ledipasvir + Sofosbuvirwith or without RBV	Genotype-1, 4, 5 and 6	Harvoni [®]				
2015	Daclatasvir + Sofosbuvir with or without RBV	Genotype-1, and 3	Daklinza™ + Sovaldi [®]				
2016	Grazoprevir + Elbasvir + RBV	Genotype-1, and 4	Zepatier TM				
2016	Velpatasvir + Sofosbuvir	Genotype 1 to 6	Epclusa®				
2017	Glecaprevir + Pibrentasvir	Genotype 1 to 6	Mavyret™				
2017	Sofosbuvir + Velpatasvir + Voxilaprevir	Genotype 1 to 6	Vosevi®				

FDA: Food and Drug Administration; RBV: Ribavirin; IFN: Interferon.

REGRESSION OF FIBROSIS AFTER DAA THERAPY

Liver biopsy is the gold standard to estimate liver fibrosis regression after DAA therapy. In a study by Cheng *et al*[22], the Metavir fibrosis score decreased from F3-F4 to F0-2 in more than 50% of the patients from baseline to post-therapy. Since liver biopsy is an invasive procedure that can be associated with potential adverse events, non-invasive modalities have been developed to monitor hepatic fibrosis[23,24].

Fibrosis markers: Fibrosis-4 and aminotransferase to platelet ratio index

Aminotransferase to platelet ratio index (APRI) and fibrosis-4 (FIB-4) are non-invasive serum fibrosis markers. FIB-4 and APRI values have been shown to decrease significantly during the first four weeks of DAA therapy[22]. The initial reduction in fibrosis may be related to a decrease in hepatic inflammation. They reported that aspartate aminotransferase (AST) and ALT values significantly decreased by 50.8% and 64.1% respectively after 4 wk of DAA therapy and ultimately reaching normal values[22].

Fibroscan or vibration-controlled transient elastography

Vibration-controlled transient elastography is a non-invasive and accurate measuring tool of liver fibrosis. Liver stiffness scores significantly decreased in patients who responded to DAA. Several studies have shown long-term regression of fibrosis over a follow-up period of 2 years.

Rout *et al*[25] reported that high baseline liver stiffness measurements (LSM), low platelet count, and low body mass index (BMI) were independently associated with improvement of LSM values one year after successful therapy. Furthermore, the levels of serum transaminases were not significantly associated with a reduction of LSM on multivariate analysis.

Chan *et al*[26] monitored a cohort of patients for at least a year after completion of DAA therapy to exclude the confounding effect of liver inflammation on LSM. They observed the median intra-patient LSM reduction was 0.5 kPa between the end of therapy and 12 mo after treatment.

Stasi *et al*[27] observed the greatest reduction in stiffness values at end of DAA therapy. The reduction in fibrosis was more gradual thereafter. In this group of patients, the liver stiffness values reduced progressively at 1 year, 2 years after treatment, respectively. Their findings suggested a continued reduction of fibrosis beyond the initial resolution of inflammation.

Several studies reported that patients with advanced fibrosis had significant fibrosis regression after achieving SVR. The reduction was approximately 3.1 kPa in 6-12 mo after achieving HCV cure, and the median decline in liver stiffness was 28.2% (interquartile range of 21.8% to 34.8%)[28]. Despite a reduction from baseline LSM, more than half of the patients remained cirrhotic at week 24 after treatment completion[29]. This result is consistent with previous observations that advanced fibrosis often persists after SVR[30,31].

Zaishidena® WJG | https://www.wjgnet.com

RISKS OF HCC AFTER HCV CURE

Lack of fibrosis regression

It is crucial to explore the relationship between the lack of fibrosis regression and HCC risk especially in patients with advanced fibrosis and cirrhosis^[32,33]. In a study by Ravaioli et al[34], 139 patients with HCV-related cirrhosis who achieved SVR after DAA treatment were included to evaluate their HCC risk by comparing LSM at baseline to end of treatment. The majority of the patients were male (65.5%) and genotype 1b (58.3%). Those who developed HCC had in average an 18% reduction in LSM compared to 28.9% among those without HCC (P = 0.005). At multivariate analysis, a less than 30% reduction in LSM was an independent HCC risk factor.

In another study, Kawagishi et al[35] evaluated fibrosis regression by LSM in 110 HCV patients who achieved SVR. Regression of liver fibrosis was defined as: A decrease by > 1 stage after DAA therapy in patients with liver fibrosis stage F2 to F4; and no deterioration of fibrosis in patients with liver fibrosis F0/1. They found the rate of regression was lower at 96 wk after SVR among those with higher baseline fibrosis stages.

Hepatic steatosis and non-alcoholic fatty liver disease

Hepatic steatosis is one of the histopathologic features of CHC[36]. Both in vitro and in vivo studies have shown that HCV core protein expression either in cell cultures or in transgenic mice led to the development of hepatic steatosis, contributing to carcinogenesis[37-39]. Cholet et al[40] in their study demonstrated a significant relationship between steatosis and hepatic fibrosis in CHC highlighting the important role played by steatosis in liver disease progression in CHC. This relationship remained significant in multivariate analysis as well[40].

Hepatic steatosis is among the factors associated with increased risk of developing HCC in HCV patients after DAA therapy[41,42]. In a large retrospective study conducted by Peleg et al[43] on 515 CHC patients treated with interferon-free DAA regimens, baseline liver steatosis (LS) was significantly associated with all-cause mortality and the development of HCC after treatment. Patients with LS had higher incidence rates of HCC (5.23 cases per 100 person-years, 95% CI: 4.85-5.71) compared to patients with advanced fibrosis (3.51 cases per 100 persons-years, 95%CI: 3.33-3.67). Moreover, patients with LS without advanced fibrosis had higher rates of mortality and HCC compared to those with advanced fibrosis but without steatosis[42]. Kono et al[44] concluded in their study of 286 CHC patients that fatty liver along with advanced liver fibrosis is associated with sustained liver damage with abnormal alpha-feto protein (AFP) and ALT levels even after HCV cure. In a prospective study conducted by Noureddin et al[45], 47.5% of the HCV patients with SVR had evidence of LS. Long-term follow-up of these patients is critically important to monitor progressive liver disease.

Diabetes mellitus

Diabetes mellitus (DM) is identified as a significant risk factor for HCC in HCV patients after SVR but the mechanism remains unclear [46-48]. There is some evidence suggesting hyperinsulinemia and insulin-dependent signaling pathways are linked to the pathogenesis and progression of HCC. Insulin resistance increases the rate of fibrosis progression in HCV infected patients. Hyperinsulinemia and insulin resistance as a result of cirrhosis can further promote the development of HCC[49]. HCC risk after interferon-induced SVR in patients with DM and cirrhosis had been reported. Subsequently, this association was also noted after DAA therapy. A 3-year follow-up study including 565 CHC patients with cirrhosis treated with DAAs identified diabetes as an independent predictor of de novo HCC[50-54]. Degasperi et al[47] identified diabetes as a strong independent predictor for de novo HCC development and also HCC recurrence in a cohort of 546 HCV patients treated with DAA. On the multivariate analysis, diabetes [hazard ratio (HR): 2.52, 95% CI: 1.08-5.87, P = 0.03] predicted *de novo* HCC as well as HCC recurrence (HR: 4.12, 95%CI: 1.55-10.93, P = 0.004)[47]. Similarly, in another study, Lu et al[48] also found that DM had a significant effect on the risk of HCC [adjusted HR (aHR): 1.65, 95% CI: 1.09-2.49]. In contrast, DM was not associated with an increased risk of developing HCC after DAA-induced SVR in studies by Kanwal *et al*[41].

Alcohol

Alcohol is an important HCC risk factor regardless of the presence of HCV. The annual incidence of HCC is higher among patients with alcohol use compared to those



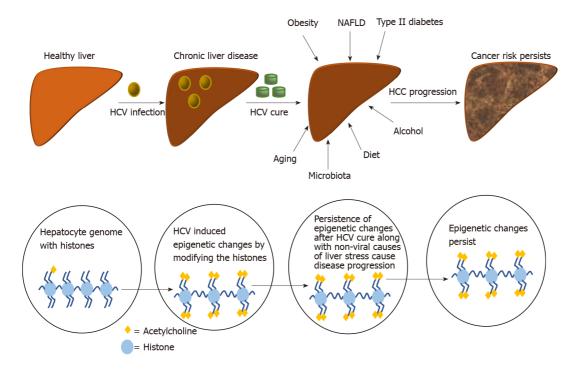


Figure 2 Pathogenesis of hepatocellular carcinoma. HCV: Hepatitis C virus; NAFLD: Non-alcoholic fatty liver disease; HCC: Hepatocellular carcinoma.

without (aHR: 4.73, 95%CI: 3.34-6.68)[41]. Alcohol-induced oxidative stress and the hepatic metabolism of ethanol could increase the conversion of pro-carcinogens to active carcinogens that results in HCC[42]. Caldwell *et al*[55] found that history of heavy alcohol consumption, defined as consumption of more than 2 drinks per day or 14 drinks per week for female; 3 drinks per day or 21 drinks per week for male, had a direct impact on FIB-4 score. It was significantly higher in the group with heavy alcohol abuse compared to no alcohol abuse. A daily intake of \geq 80 g of ethanol for > 10 years is thought to increase the risk of HCC by approximately five-fold and women are more susceptible to alcohol toxicity than men[56,57]. Alcohol acts synergistically with HCV in accelerating the progression to cirrhosis and liver-related complications [58]. The ethanol's effects on hepatic fibrogenesis persist after HCV cure for those who continue to consume alcohol. A study by Kanwal *et al*[41] reported a higher annual incidence of HCC among patients with alcohol use (1.01%, 95%CI: 0.83-1.19) compared to those without (0.72%, 95%CI: 0.54%-0.91%; aHR: 1.56, 95%CI: 1.11-2.18) after achieving SVR post DAA therapy[41] (Figure 2).

PATHOGENESIS OF HCC AFTER HCV CURE

A number of key pathways are involved in the development of HCV-related HCC: (1) Fibrosis due to continuous necrosis; (2) Immune-surveillance failures attributable to persistent viral replication with immune system escape mechanisms; and (3) Direct carcinogenic effect of HCV proteins which deregulate host cell cycle checkpoints leading to DNA mutations in liver cells[59]. The pathogenesis of HCC after HCV cure remains elusive. A 186-gene expression signature in liver tissue of CHC patients with HCC suggested virus-induced transcriptional reprogramming in the liver leading to carcinogenesis[60,61]. Epigenetic modifications of histones, for example, can lead to chromatin opening and compacting which, in turn, affect gene regulation[62]. Hamdane et al[63] investigated HCV-induced epigenetic alterations that might increase HCC risk after DAA treatment in patients and mice with humanized livers. They found that chronic HCV infection induced specific genome-wide changes in H3K27ac. The 5318 modified genes associated with CHC correlated with changes in the expression of mRNAs and proteins. A number of the altered pathways resulting from epigenetic changes persisted after HCV cure with DAAs. Namely, molecular pathways involving tumor necrosis factor α signaling, inflammatory response, G2M checkpoint, epithelial-mesenchymal transition, phosphoinositide 3-kinase, Akt, and mammalian target of rapamycin^[63]. This analysis showed that H3K27ac changes observed in HCV-infected patients were partly reversed after cure for those with stage



F2-3 fibrosis. This group shared only 42.5% of the HCV-modified genes. In contrast, in DAA-cured patients with cirrhosis (stage F4), 96.6% of the HCV-induced H3K27ac changes persisted[63]. By performing chromatin immunoprecipitation followed by next-generation sequencing of histone post-translational modifications that are epigenetic markers for active and repressed chromatin, Perez et al[64] also demonstrated that HCV infection induces genome-wide epigenetic changes. The "epigenetic signature" persisted after achieving DAA- related cure. Santangelo et al[65] examined the impact of DAAs on the ability of exosomal microRNAs (miRs) to modulate the innate immune response in patients with CHC. miR-122 was selectively studied as it is involved in HCV replication and its loss has been associated with HCC development. The study showed that miR-122-5p, miR-222-3p, miR146-5p, miR-150-5p, miR-30C-5p, miR-378a-3p, miR-20a5p were enriched in exosomes derived from the HCV-infected cells. The liver-specific miR-122 levels and the expression of the aforementioned miRs significantly decreased after DAAs therapy[65]. Human HCC cells express vascular endothelial growth factor (VEGF) that functions as a cytokine and affects cancer cell growth and survival[66]. The VEGF expression correlates with liver cancer angiogenesis and proliferative activity. Villani et al[66] studied the effect of DAA treatmentinduced VEGF on HCC angiogenesis. In this study on 117 cirrhotic patients treated with DAA, a 4-fold increase in VEGF was observed compared to baseline. This significant increase in VEGF could potentially lead to an acceleration of cancer cell proliferation prior to HCV cure and the carcinogenesis remained after DAA even though the VEGF decreased to normal levels 12 wk after DAA treatment (Figure 2).

IDENTIFYING PATIENTS WITH HCC RISK AFTER HCV CURE

Although achieving SVR is the goal of HCV treatment, the risk of developing HCC remains high particularly in patients with advanced fibrosis and cirrhosis[67]. This risk ranges between 1.8% and 2.5% annually. The current guidelines suggest that these patients should undergo HCC surveillance every six months by ultrasound with or without alfa-fetoprotein indefinitely. On the contrary, patients with no or moderate fibrosis who achieved SVR and have no risk behavior could be discharged from specialty care[68]. Methods to identify patients with differential HCC risks can be challenging

APRI and FIB-4 have been used to assess the HCC risks. These scores, however, were not developed specifically for HCC indication; thus, their accuracy is limited. Transient elastography, similarly, was not designed to detect HCC[68]. Specific score systems designed to predict HCC after HCV cure remain an unmet need.

A group in Japan developed a simple score to identify HCV patients at risk of HCC after achieving SVR[69]. The majority were HCV serotype 1 or 2 patients. They use multivariate analysis to identify predictive variables. They found that age (cutoff 75 years) and post-treatment AFP (cutoff 6 ng/mL) values were independent factors for HCC. Thus, they used a score with 0 and 1 point for each factor: < 75 and > 75 years were set as 0 and 1 point; < 6 and > 6 ng/mL were set as 0 and 1 points respectively. The sum of each factor was considered as the final score. HCC incidence increased significantly with higher scores. In the 0-point group, the incidence of HCC was 0% at 6 mo; 0.3% at 12, 18 and 24 mo; and only 1.26% at 36 mo. In contrast, the risk increased in the 2-point group: 2.88% at 6 mo; 4.92% at 12 mo; 11.61% at 18 mo; and up to 18.37% after 24 mo. This scoring system is simple to apply but needs to be validated prospectively in different patient populations.

In Egypt, Shiha et al[70] conducted a prospective study to develop an HCC risk model after SVR. Their model used clinical variables to create scores for low, intermediate and high HCC risk. Each variable was given a score according to its HR. This General Evaluation Score included age (< 54 = 0; > 54 = 1), gender (male = 3.5; female = 0), fibrosis stage (F3 = 1.5; F4 = 3), albumin (> 3.8 g/dL = 0; < 3.8 g/dL = 2) and alpha-fetoprotein levels (< 20 ng/mL = 0; > 20 ng/mL = 3). The score range was between 0 and 12.5. The low-risk group (score < 6) had a 1-year HCC incidence of 0.1%, 1.2% at 2 years and 1.9% at 3 years. The intermediate-risk group (score 6-7.5) had a 1-year incidence of 0.7%, 3.3% at 2 years and 5.8% at 3 years. Finally, the high-risk group (score > 7.5) had a 1-year HCC incidence of 1.2% which increased to 7.1% at 2 years and 9.5% at 3 years. The advantage of this tool is that it uses commonly available clinical variables that can be applied in different settings including low and mediumincome populations. This study included only patients with HCV genotype 4. If it is validated in other HCV genotypes and populations, it can be a cost-effective tool for HCC surveillance.

WJG https://www.wjgnet.com

Ioannou et al^[71] developed different sets of models according to treatment modalities for CHC. For those with DAA-induced HCV cure, the regression model showed that age > 60, platelet count < 61×10^4 , serum AST/ALT ratio > 8.8 in noncirrhotic and > 11.01 in cirrhotic; and albumin < 2.9 were major predictive variables for the development of HCC. By applying these variables in the models, the cirrhotic/non-SVR group was predicted to have a 13.1% HCC risk at 2.6-year follow-up; the cirrhotic/SVR group had a 4.5% incidence at 2-year follow-up; the non-cirrhotic/non-SVR had a 4.2% incidence at 3.7-year follow-up; whereas the non-cirrhotic/SVR group had only a low 0.7% HCC risk at 2.3-year follow-up. Given the differential risks according to the clinical characteristics, the HCC screening guidelines could potentially be narrowed to specific risk groups. Although this model was internally validated and is easily available as a web-calculator tool, external validation would be necessary since it was performed using the Veterans Affairs healthcare data only and the majority of patients had HCV genotype 1.

Recently, a model using transient elastography was developed in Spain by Alonso L ópez et al[72], they built two dynamic models for patients with advanced fibrosis and cirrhosis who achieved SVR. Their objective was to identify very low HCC risk patients who may not require continued HCC surveillance despite the presence of advanced fibrosis prior to therapy. The first model included baseline albumin, baseline and 1-year follow-up elastography. Given that elastography may not be available in every setting, the second model included serological markers only: Baseline albumin, baseline and 1-year follow-up FIB-4 and 1-year gamma-glutamyl transferase. They found that both models were useful as predictors of HCC. Moreover, after stratification of risk assigned by scoring each variable in both models, the ones who scored 0 had 0%-0.4% risk of developing HCC. The ability to accurately identify those at very low HCC risk could effectively stratify patients for HCC surveillance.

Alpha-fetoprotein is the most available HCC biomarker. Its sensitivity and specificity are very variable^[73]. Recent studies have shown sphingolipids as potential biomarkers to detect hepatic decompensation in cirrhotic patients[74]. Two types of sphingolipids - C16-ceramide and sphingosine-1-phosphate - have been applied as HCC biomarkers in cirrhotic patients. Mücke et al [75] in Germany evaluated sphingolipids as early predictive HCC biomarkers in HCV patients with cirrhosis who had achieved SVR. They identified C16Cer as an independent biomarker for early detection of *de novo* HCC in both AFP-positive or AFP-negative patients. Although this finding seems novel and promising, prospective studies are needed to clarify the association between sphingolipids and carcinogenesis.

In the area of deep learning, Ioannou et al[76] utilized recurrent neural network (RNN) models to identify patients at high risk of developing HCC for at least a 3-year follow-up period after HCV cure. They used two types of variables: Baseline and longitudinal ones to evaluate the risk progression. They compared three models: Cross-sectional logistic regression (LR), longitudinal LR and RNN. The area under the receiver operating characteristic curve for these groups was 0.67, 0.70 and 0.80 respectively. The RNN model was superior to the conventional LR models and could be a promising tool after computational refi-nement.

CONCLUSION

In the DAA era, the development of HCC remains a significant concern especially among those with advanced hepatic fibrosis. A number of factors including diabetes mellitus, underlying non-alcoholic fatty liver disease and alcohol consumption have been associated with progression to HCC after HCV cure. Promising HCC predictive models are being developed but most require validation and standardization. The pathogenesis of HCC after HCV cure remains poorly understood. The understanding of the molecular mechanisms leading to HCC could facilitate the identification of novel biomarkers for early HCC detection.

REFERENCES

- Lavanchy D. Evolving epidemiology of hepatitis C virus. Clin Microbiol Infect 2011; 17: 107-115 1 [PMID: 21091831 DOI: 10.1111/j.1469-0691.2010.03432.x]
- Hofmeister MG, Rosenthal EM, Barker LK, Rosenberg ES, Barranco MA, Hall EW, Edlin BR, Mermin J, Ward JW, Ryerson AB. Estimating Prevalence of Hepatitis C Virus Infection in the United States, 2013-2016. Hepatology 2019; 69: 1020-1031 [PMID: 30398671 DOI: 10.1002/hep.30297]



- 3 Petruzziello A, Marigliano S, Loquercio G, Cozzolino A, Cacciapuoti C. Global epidemiology of hepatitis C virus infection: An up-date of the distribution and circulation of hepatitis C virus genotypes. World J Gastroenterol 2016; 22: 7824-7840 [PMID: 27678366 DOI: 10.3748/wjg.v22.i34.7824]
- 4 Hoshida Y, Fuchs BC, Bardeesy N, Baumert TF, Chung RT. Pathogenesis and prevention of hepatitis C virus-induced hepatocellular carcinoma. J Hepatol 2014; 61: S79-S90 [PMID: 25443348 DOI: 10.1016/j.jhep.2014.07.010
- Falade-Nwulia O, Suarez-Cuervo C, Nelson DR, Fried MW, Segal JB, Sulkowski MS. Oral Direct-5 Acting Agent Therapy for Hepatitis C Virus Infection: A Systematic Review. Ann Intern Med 2017; 166: 637-648 [PMID: 28319996 DOI: 10.7326/M16-2575]
- Singal AK, Singh A, Jaganmohan S, Guturu P, Mummadi R, Kuo YF, Sood GK. Antiviral therapy 6 reduces risk of hepatocellular carcinoma in patients with hepatitis C virus-related cirrhosis. Clin Gastroenterol Hepatol 2010; 8: 192-199 [PMID: 19879972 DOI: 10.1016/j.cgh.2009.10.026]
- 7 van der Meer AJ, Veldt BJ, Feld JJ, Wedemeyer H, Dufour JF, Lammert F, Duarte-Rojo A, Heathcote EJ, Manns MP, Kuske L, Zeuzem S, Hofmann WP, de Knegt RJ, Hansen BE, Janssen HL. Association between sustained virological response and all-cause mortality among patients with chronic hepatitis C and advanced hepatic fibrosis. JAMA 2012; 308: 2584-2593 [PMID: 23268517 DOI: 10.1001/jama.2012.1448781
- Grebely J, Prins M, Hellard M, Cox AL, Osburn WO, Lauer G, Page K, Lloyd AR, Dore GJ; International Collaboration of Incident HIV and Hepatitis C in Injecting Cohorts (InC3). Hepatitis C virus clearance, reinfection, and persistence, with insights from studies of injecting drug users: towards a vaccine. Lancet Infect Dis 2012; 12: 408-414 [PMID: 22541630 DOI: 10.1016/S1473-3099(12)70010-5
- 9 López-Diéguez M, Montes ML, Pascual-Pareja JF, Quereda C, Von Wichmann MA, Berenguer J, Tural C, Hernando A, González-García J, Serrano L, Arribas JR; GESIDA 37/03-FIPSE 36465/03-NEAT IG5 Study Group. The natural history of liver cirrhosis in HIV-hepatitis C virus-coinfected patients. AIDS 2011; 25: 899-904 [PMID: 21330908 DOI: 10.1097/QAD.0b013e3283454174]
- 10 Hagan H, Jordan AE, Neurer J, Cleland CM. Incidence of sexually transmitted hepatitis C virus infection in HIV-positive men who have sex with men. AIDS 2015; 29: 2335-2345 [PMID: 26258525 DOI: 10.1097/QAD.00000000000834]
- 11 Mosley JW, Operskalski EA, Tobler LH, Andrews WW, Phelps B, Dockter J, Giachetti C, Busch MP. Viral and host factors in early hepatitis C virus infection. Hepatology 2005; 42: 86-92 [PMID: 15954090 DOI: 10.1002/hep.20742]
- 12 Hajarizadeh B, Grebely J, Dore GJ. Epidemiology and natural history of HCV infection. Nat Rev Gastroenterol Hepatol 2013; 10: 553-562 [PMID: 23817321 DOI: 10.1038/nrgastro.2013.107]
- 13 Shin EC, Sung PS, Park SH. Immune responses and immunopathology in acute and chronic viral hepatitis. Nat Rev Immunol 2016; 16: 509-523 [PMID: 27374637 DOI: 10.1038/nri.2016.69]
- 14 Negro F. Natural History of Hepatic and Extrahepatic Hepatitis C Virus Diseases and Impact of Interferon-Free HCV Therapy. Cold Spring Harb Perspect Med 2020; 10 [PMID: 31636094]
- Ge D, Fellay J, Thompson AJ, Simon JS, Shianna KV, Urban TJ, Heinzen EL, Qiu P, Bertelsen AH, 15 Muir AJ, Sulkowski M, McHutchison JG, Goldstein DB. Genetic variation in IL28B predicts hepatitis C treatment-induced viral clearance. Nature 2009; 461: 399-401 [PMID: 19684573 DOI: 10.1038/nature083091
- 16 Tanaka Y, Nishida N, Sugiyama M, Kurosaki M, Matsuura K, Sakamoto N, Nakagawa M, Korenaga M, Hino K, Hige S, Ito Y, Mita E, Tanaka E, Mochida S, Murawaki Y, Honda M, Sakai A, Hiasa Y, Nishiguchi S, Koike A, Sakaida I, Imamura M, Ito K, Yano K, Masaki N, Sugauchi F, Izumi N, Tokunaga K, Mizokami M. Genome-wide association of IL28B with response to pegylated interferonalpha and ribavirin therapy for chronic hepatitis C. Nat Genet 2009; 41: 1105-1109 [PMID: 19749757 DOI: 10.1038/ng.449]
- 17 Ryder SD, Irving WL, Jones DA, Neal KR, Underwood JC; Trent Hepatitis C Study Group. Progression of hepatic fibrosis in patients with hepatitis C: a prospective repeat liver biopsy study. Gut 2004; 53: 451-455 [PMID: 14960533 DOI: 10.1136/gut.2003.021691]
- 18 Lingala S, Ghany MG. Natural History of Hepatitis C. Gastroenterol Clin North Am 2015; 44: 717-734 [PMID: 26600216 DOI: 10.1016/j.gtc.2015.07.003]
- 19 Zając M, Muszalska I, Sobczak A, Dadej A, Tomczak S, Jelińska A. Hepatitis C New drugs and treatment prospects. Eur J Med Chem 2019; 165: 225-249 [PMID: 30685524 DOI: 10.1016/j.ejmech.2019.01.025]
- 20 Pearlman BL, Traub N. Sustained virologic response to antiviral therapy for chronic hepatitis C virus infection: a cure and so much more. Clin Infect Dis 2011; 52: 889-900 [PMID: 21427396 DOI: 10.1093/cid/cir076]
- Simmons B, Saleem J, Hill A, Riley RD, Cooke GS. Risk of Late Relapse or Reinfection With Hepatitis C Virus After Achieving a Sustained Virological Response: A Systematic Review and Metaanalysis. Clin Infect Dis 2016; 62: 683-694 [PMID: 26787172 DOI: 10.1093/cid/civ948]
- 22 Cheng CH, Chu CY, Chen HL, Lin IT, Wu CH, Lee YK, Hu PJ, Bair MJ. Direct-acting antiviral therapy of chronic hepatitis C improves liver fibrosis, assessed by histological examination and laboratory markers. J Formos Med Assoc 2021; 120: 1259-1268 [PMID: 33339709 DOI: 10.1016/j.jfma.2020.11.018]
- 23 Sharma S, Khalili K, Nguyen GC. Non-invasive diagnosis of advanced fibrosis and cirrhosis. World J Gastroenterol 2014; 20: 16820-16830 [PMID: 25492996 DOI: 10.3748/wjg.v20.i45.16820]



- Papastergiou V, Tsochatzis E, Burroughs AK. Non-invasive assessment of liver fibrosis. Ann 24 Gastroenterol 2012; 25: 218-231 [PMID: 24714123]
- 25 Rout G, Nayak B, Patel AH, Gunjan D, Singh V, Kedia S, Shalimar. Therapy with Oral Directly Acting Agents in Hepatitis C Infection Is Associated with Reduction in Fibrosis and Increase in Hepatic Steatosis on Transient Elastography. J Clin Exp Hepatol 2019; 9: 207-214 [PMID: 31024203 DOI: 10.1016/j.jceh.2018.06.009]
- 26 Chan J, Gogela N, Zheng H, Lammert S, Ajayi T, Fricker Z, Kim AY, Robbins GK, Chung RT. Direct-Acting Antiviral Therapy for Chronic HCV Infection Results in Liver Stiffness Regression Over 12 Months Post-treatment. Dig Dis Sci 2018; 63: 486-492 [PMID: 28887750 DOI: 10.1007/s10620-017-4749-x]
- 27 Stasi C, Sadalla S, Carradori E, Monti M, Petraccia L, Madia F, Gragnani L, Zignego AL. Longitudinal evaluation of liver stiffness and outcomes in patients with chronic hepatitis C before and after short- and long-term IFN-free antiviral treatment. Curr Med Res Opin 2020; 36: 245-249 [PMID: 31702411 DOI: 10.1080/03007995.2019.1691517]
- 28 Singh S, Facciorusso A, Loomba R, Falck-Ytter YT. Magnitude and Kinetics of Decrease in Liver Stiffness After Antiviral Therapy in Patients With Chronic Hepatitis C: A Systematic Review and Meta-analysis. Clin Gastroenterol Hepatol 2018; 16: 27-38.e4 [PMID: 28479504 DOI: 10.1016/j.cgh.2017.04.038]
- Dolmazashvili E, Abutidze A, Chkhartishvili N, Karchava M, Sharvadze L, Tsertsvadze T. Regression of liver fibrosis over a 24-week period after completing direct-acting antiviral therapy in patients with chronic hepatitis C receiving care within the national hepatitis C elimination program in Georgia: results of hepatology clinic HEPA experience. Eur J Gastroenterol Hepatol 2017; 29: 1223-1230 [PMID: 28857900 DOI: 10.1097/MEG.000000000000964]
- 30 Balart LA, Lisker-Melman M, Hamzeh FM, Kwok A, Lentz E, Rodriguez-Torres M; LATINO study investigators. Peginterferon α-2a plus ribavirin in Latino and Non-Latino Whites with HCV genotype 1: Histologic outcomes and tolerability from the LATINO Study. Am J Gastroenterol 2010; 105: 2177-2185 [PMID: 20389293 DOI: 10.1038/ajg.2010.157]
- Wei H, Song B. Elastography for Longitudinal Assessment of Liver Fibrosis after Antiviral Therapy: 31 A Review. J Clin Transl Hepatol 2020; 8: 445-453 [PMID: 33447528 DOI: 10.14218/JCTH.2020.00033]
- Macek Jílková Z, Seigneurin A, Coppard C, Ouaguia L, Aspord C, Marche PN, Leroy V, Decaens T. 32 Circulating IL-13 Is Associated with De Novo Development of HCC in HCV-Infected Patients Responding to Direct-Acting Antivirals. Cancers (Basel) 2020; 12 [PMID: 33352852 DOI: 10.3390/cancers12123820]
- 33 Hamoir C, Horsmans Y, Stärkel P, Dahlqvist G, Negrin Dastis S, Lanthier N. Risk of hepatocellular carcinoma and fibrosis evolution in hepatitis C patients with severe fibrosis or cirrhosis treated with direct acting antiviral agents. Acta Gastroenterol Belg 2021; 84: 25-32 [PMID: 33639690 DOI: 10.51821/84.1.420
- 34 Ravaioli F, Conti F, Brillanti S, Andreone P, Mazzella G, Buonfiglioli F, Serio I, Verrucchi G, Bacchi Reggiani ML, Colli A, Marasco G, Colecchia A, Festi D. Hepatocellular carcinoma risk assessment by the measurement of liver stiffness variations in HCV cirrhotics treated with direct acting antivirals. Dig Liver Dis 2018; 50: 573-579 [PMID: 29567413 DOI: 10.1016/j.dld.2018.02.010
- Kawagishi N, Suda G, Kimura M, Maehara O, Yamada R, Tokuchi Y, Kubo A, Kitagataya T, 35 Shigesawa T, Suzuki K, Ohara M, Nakai M, Sho T, Natsuizaka M, Morikawa K, Ogawa K, Kudo Y, Nishida M, Sakamoto N. Baseline elevated serum angiopoietin-2 predicts long-term non-regression of liver fibrosis after direct-acting antiviral therapy for hepatitis C. Sci Rep 2021; 11: 9207 [PMID: 33911145 DOI: 10.1038/s41598-021-88632-7]
- 36 Kralj D, Virović Jukić L, Stojsavljević S, Duvnjak M, Smolić M, Čurčić IB. Hepatitis C Virus, Insulin Resistance, and Steatosis. J Clin Transl Hepatol 2016; 4: 66-75 [PMID: 27047774 DOI: 10.14218/JCTH.2015.00051]
- 37 Moradpour D, Englert C, Wakita T, Wands JR. Characterization of cell lines allowing tightly regulated expression of hepatitis C virus core protein. Virology 1996; 222: 51-63 [PMID: 8806487 DOI: 10.1006/viro.1996.0397]
- 38 Barba G, Harper F, Harada T, Kohara M, Goulinet S, Matsuura Y, Eder G, Schaff Z, Chapman MJ, Miyamura T, Bréchot C. Hepatitis C virus core protein shows a cytoplasmic localization and associates to cellular lipid storage droplets. Proc Natl Acad Sci US A 1997; 94: 1200-1205 [PMID: 9037030 DOI: 10.1073/pnas.94.4.1200]
- 39 Moriya K, Fujie H, Shintani Y, Yotsuyanagi H, Tsutsumi T, Ishibashi K, Matsuura Y, Kimura S, Miyamura T, Koike K. The core protein of hepatitis C virus induces hepatocellular carcinoma in transgenic mice. Nat Med 1998; 4: 1065-1067 [PMID: 9734402 DOI: 10.1038/2053]
- 40 Cholet F, Nousbaum JB, Richecoeur M, Oger E, Cauvin JM, Lagarde N, Robaszkiewicz M, Gouérou H. Factors associated with liver steatosis and fibrosis in chronic hepatitis C patients. Gastroenterol Clin Biol 2004; 28: 272-278 [PMID: 15094677 DOI: 10.1016/s0399-8320(04)94918-4]
- Kanwal F, Kramer J, Asch SM, Chayanupatkul M, Cao Y, El-Serag HB. Risk of Hepatocellular 41 Cancer in HCV Patients Treated With Direct-Acting Antiviral Agents. Gastroenterology 2017; 153: 996-1005.e1 [PMID: 28642197 DOI: 10.1053/j.gastro.2017.06.012]
- Lieber CS, Seitz HK, Garro AJ, Worner TM. Alcohol-related diseases and carcinogenesis. Cancer 42 Res 1979; 39: 2863-2886 [PMID: 221110]



- Peleg N, Issachar A, Sneh Arbib O, Cohen-Naftaly M, Harif Y, Oxtrud E, Braun M, Leshno M, 43 Barsheshet A, Shlomai A. Liver steatosis is a major predictor of poor outcomes in chronic hepatitis C patients with sustained virological response. J Viral Hepat 2019; 26: 1257-1265 [PMID: 31243878 DOI: 10.1111/jvh.13167]
- Kono M, Nishida N, Hagiwara S, Minami T, Chishina H, Arizumi T, Minaga K, Kamata K, Komeda 44 Y, Sakurai T, Takenaka M, Takita M, Yada N, Ida H, Minami Y, Ueshima K, Watanabe T, Kudo M. Unique Characteristics Associated with Sustained Liver Damage in Chronic Hepatitis C Patients Treated with Direct Acting Antivirals. Dig Dis 2017; 35: 556-564 [PMID: 29040988 DOI: 10.1159/000480148
- 45 Noureddin M, Wong MM, Todo T, Lu SC, Sanyal AJ, Mena EA. Fatty liver in hepatitis C patients post-sustained virological response with direct-acting antivirals. World J Gastroenterol 2018; 24: 1269-1277 [PMID: 29568207 DOI: 10.3748/wjg.v24.i11.1269]
- Hedenstierna M, Nangarhari A, Weiland O, Aleman S. Diabetes and Cirrhosis Are Risk Factors for 46 Hepatocellular Carcinoma After Successful Treatment of Chronic Hepatitis C. Clin Infect Dis 2016; 63: 723-729 [PMID: 27282709 DOI: 10.1093/cid/ciw362]
- Degasperi E, D'Ambrosio R, Iavarone M, Sangiovanni A, Aghemo A, Soffredini R, Borghi M, 47 Lunghi G, Colombo M, Lampertico P. Factors Associated With Increased Risk of De Novo or Recurrent Hepatocellular Carcinoma in Patients With Cirrhosis Treated With Direct-Acting Antivirals for HCV Infection. Clin Gastroenterol Hepatol 2019; 17: 1183-1191.e7 [PMID: 30613002 DOI: 10.1016/j.cgh.2018.10.038]
- 48 Lu M, Li J, Rupp LB, Holmberg SD, Moorman AC, Spradling PR, Teshale EH, Zhou Y, Boscarino JA, Schmidt MA, Lamerato LE, Trinacty C, Trudeau S, Gordon SC; CHeCS Investigators. Hepatitis C treatment failure is associated with increased risk of hepatocellular carcinoma. J Viral Hepat 2016; 23: 718-729 [PMID: 27028626 DOI: 10.1111/jvh.12538]
- 49 Kukla M, Piotrowski D, Waluga M, Hartleb M. Insulin resistance and its consequences in chronic hepatitis C. Clin Exp Hepatol 2015; 1: 17-29 [PMID: 28856251 DOI: 10.5114/ceh.2015.51375]
- Polesel J, Zucchetto A, Montella M, Dal Maso L, Crispo A, La Vecchia C, Serraino D, Franceschi S, 50 Talamini R. The impact of obesity and diabetes mellitus on the risk of hepatocellular carcinoma. Ann Oncol 2009; 20: 353-357 [PMID: 18723550 DOI: 10.1093/annonc/mdn565]
- Regimbeau JM, Colombat M, Mognol P, Durand F, Abdalla E, Degott C, Degos F, Farges O, 51 Belghiti J. Obesity and diabetes as a risk factor for hepatocellular carcinoma. Liver Transpl 2004; 10: S69-S73 [PMID: 14762843 DOI: 10.1002/lt.20033]
- 52 Karagozian R, Derdák Z, Baffy G. Obesity-associated mechanisms of hepatocarcinogenesis. Metabolism 2014; 63: 607-617 [PMID: 24629562 DOI: 10.1016/j.metabol.2014.01.011]
- Khan MM, Saito S, Takagi S, Ohnishi H, Izumi H, Sakauchi F, Washio M, Sonoda T, Nagata Y, 53 Asakura S, Kobayashi K, Mori M, Shimamoto K. Relationship between hepatocellular carcinoma and impaired glucose tolerance among Japanese. Hepatogastroenterology 2006; 53: 742-746 [PMID: 17086880]
- Baffy G. Hepatocellular Carcinoma in Non-alcoholic Fatty Liver Disease: Epidemiology, 54 Pathogenesis, and Prevention. J Clin Transl Hepatol 2013; 1: 131-137 [PMID: 26355775 DOI: 10.14218/JCTH.2013.00005
- 55 Caldwell SH, Li X, Rourk RM, Millar A, Sosnowski KM, Sue M, Barritt AS, McCallum RW, Schiff ER. Hepatitis C infection by polymerase chain reaction in alcoholics: false-positive ELISA results and the influence of infection on a clinical prognostic score. Am J Gastroenterol 1993; 88: 1016-1021 [PMID: 8391209]
- 56 El-Serag HB, Mason AC. Risk factors for the rising rates of primary liver cancer in the United States. Arch Intern Med 2000; 160: 3227-3230 [PMID: 11088082 DOI: 10.1001/archinte.160.21.3227]
- Allen NE, Beral V, Casabonne D, Kan SW, Reeves GK, Brown A, Green J; Million Women Study 57 Collaborators. Moderate alcohol intake and cancer incidence in women. J Natl Cancer Inst 2009; 101: 296-305 [PMID: 19244173 DOI: 10.1093/jnci/djn514]
- 58 Lieber CS. Mechanism of ethanol induced hepatic injury. Pharmacol Ther 1990; 46: 1-41 [PMID: 2181486 DOI: 10.1016/0163-7258(90)90032-w]
- Wirth TC, Manns MP. The impact of the revolution in hepatitis C treatment on hepatocellular 59 carcinoma. Ann Oncol 2016; 27: 1467-1474 [PMID: 27226385 DOI: 10.1093/annonc/mdw219]
- Nakagawa S, Wei L, Song WM, Higashi T, Ghoshal S, Kim RS, Bian CB, Yamada S, Sun X, 60 Venkatesh A, Goossens N, Bain G, Lauwers GY, Koh AP, El-Abtah M, Ahmad NB, Hoshida H, Erstad DJ, Gunasekaran G, Lee Y, Yu ML, Chuang WL, Dai CY, Kobayashi M, Kumada H, Beppu T, Baba H, Mahajan M, Nair VD, Lanuti M, Villanueva A, Sangiovanni A, Iavarone M, Colombo M, Llovet JM, Subramanian A, Tager AM, Friedman SL, Baumert TF, Schwarz ME, Chung RT, Tanabe KK, Zhang B, Fuchs BC, Hoshida Y; Precision Liver Cancer Prevention Consortium. Molecular Liver Cancer Prevention in Cirrhosis by Organ Transcriptome Analysis and Lysophosphatidic Acid Pathway Inhibition. Cancer Cell 2016; 30: 879-890 [PMID: 27960085 DOI: 10.1016/j.ccell.2016.11.004]
- Hoshida Y, Villanueva A, Sangiovanni A, Sole M, Hur C, Andersson KL, Chung RT, Gould J, 61 Kojima K, Gupta S, Taylor B, Crenshaw A, Gabriel S, Minguez B, Iavarone M, Friedman SL, Colombo M, Llovet JM, Golub TR. Prognostic gene expression signature for patients with hepatitis C-related early-stage cirrhosis. Gastroenterology 2013; 144: 1024-1030 [PMID: 23333348 DOI: 10.1053/j.gastro.2013.01.021]
- Jones PA, Issa JP, Baylin S. Targeting the cancer epigenome for therapy. Nat Rev Genet 2016; 17: 62



630-641 [PMID: 27629931 DOI: 10.1038/nrg.2016.93]

- Hamdane N, Jühling F, Crouchet E, El Saghire H, Thumann C, Oudot MA, Bandiera S, Saviano A, 63 Ponsolles C, Roca Suarez AA, Li S, Fujiwara N, Ono A, Davidson I, Bardeesy N, Schmidl C, Bock C, Schuster C, Lupberger J, Habersetzer F, Doffoël M, Piardi T, Sommacale D, Imamura M, Uchida T, Ohdan H, Aikata H, Chayama K, Boldanova T, Pessaux P, Fuchs BC, Hoshida Y, Zeisel MB, Duong FHT, Baumert TF. HCV-Induced Epigenetic Changes Associated With Liver Cancer Risk Persist After Sustained Virologic Response. Gastroenterology 2019; 156: 2313-2329.e7 [PMID: 30836093 DOI: 10.1053/j.gastro.2019.02.038]
- 64 Perez S, Kaspi A, Domovitz T, Davidovich A, Lavi-Itzkovitz A, Meirson T, Alison Holmes J, Dai CY, Huang CF, Chung RT, Nimer A, El-Osta A, Yaari G, Stemmer SM, Yu ML, Haviv I, Gal-Tanamy M. Hepatitis C virus leaves an epigenetic signature post cure of infection by direct-acting antivirals. PLoS Genet 2019; 15: e1008181 [PMID: 31216276 DOI: 10.1371/journal.pgen.1008181]
- Santangelo L, Bordoni V, Montaldo C, Cimini E, Zingoni A, Battistelli C, D'Offizi G, Capobianchi 65 MR, Santoni A, Tripodi M, Agrati C. Hepatitis C virus direct-acting antivirals therapy impacts on extracellular vesicles microRNAs content and on their immunomodulating properties. Liver Int 2018; 38: 1741-1750 [PMID: 29359389 DOI: 10.1111/liv.13700]
- Villani R, Facciorusso A, Bellanti F, Tamborra R, Piscazzi A, Landriscina M, Vendemiale G, Serviddio G. DAAs Rapidly Reduce Inflammation but Increase Serum VEGF Level: A Rationale for Tumor Risk during Anti-HCV Treatment. PLoS One 2016; 11: e0167934 [PMID: 27997563 DOI: 10.1371/journal.pone.0167934]
- Setiawan VW, Rosen HR. Stratification of Residual Risk of HCC Following HCV Clearance With 67 Direct-Acting Antivirals in Patients With Advanced Fibrosis and Cirrhosis. Hepatology 2020; 72: 1897-1899 [PMID: 33205438 DOI: 10.1002/hep.31639]
- European Association for the Study of the Liver. EASL Recommendations on Treatment of 68 Hepatitis C 2018. J Hepatol 2018; 69: 461-511 [PMID: 29650333 DOI: 10.1016/j.jhep.2018.03.026]
- 69 Tani J, Morishita A, Sakamoto T, Takuma K, Nakahara M, Fujita K, Oura K, Tadokoro T, Mimura S, Nomura T, Yoneyama H, Kobara H, Himoto T, Tsutsui A, Senoh T, Nagano T, Ogawa C, Moriya A, Deguchi A, Takaguchi K, Masaki T. Simple scoring system for prediction of hepatocellular carcinoma occurrence after hepatitis C virus eradication by direct-acting antiviral treatment: All Kagawa Liver Disease Group Study. Oncol Lett 2020; 19: 2205-2212 [PMID: 32194718 DOI: 10.3892/ol.2020.11341
- Shiha G, Waked I, Soliman R, Elbasiony M, Gomaa A, Mikhail NNH, Eslam M. GES: A validated 70 simple score to predict the risk of HCC in patients with HCV-GT4-associated advanced liver fibrosis after oral antivirals. Liver Int 2020; 40: 2828-2833 [PMID: 32946647 DOI: 10.1111/liv.14666]
- Ioannou GN, Green PK, Beste LA, Mun EJ, Kerr KF, Berry K. Development of models estimating 71 the risk of hepatocellular carcinoma after antiviral treatment for hepatitis C. J Hepatol 2018; 69: 1088-1098 [PMID: 30138686 DOI: 10.1016/j.jhep.2018.07.024]
- Alonso López S, Manzano ML, Gea F, Gutiérrez ML, Ahumada AM, Devesa MJ, Olveira A, Polo 72 BA, Márquez L, Fernández I, Cobo JCR, Rayón L, Riado D, Izquierdo S, Usón C, Real Y, Rincón D, Fernández-Rodríguez CM, Bañares R. A Model Based on Noninvasive Markers Predicts Very Low Hepatocellular Carcinoma Risk After Viral Response in Hepatitis C Virus-Advanced Fibrosis. Hepatology 2020; 72: 1924-1934 [PMID: 33022803 DOI: 10.1002/hep.31588]
- Farinati F, Marino D, De Giorgio M, Baldan A, Cantarini M, Cursaro C, Rapaccini G, Del Poggio P, Di Nolfo MA, Benvegnù L, Zoli M, Borzio F, Bernardi M, Trevisani F. Diagnostic and prognostic role of alpha-fetoprotein in hepatocellular carcinoma: both or neither? Am J Gastroenterol 2006; 101: 524-532 [PMID: 16542289 DOI: 10.1111/j.1572-0241.2006.00443.x]
- Grammatikos G, Ferreiròs N, Waidmann O, Bon D, Schroeter S, Koch A, Herrmann E, Zeuzem S, 74 Kronenberger B, Pfeilschifter J. Serum Sphingolipid Variations Associate with Hepatic Decompensation and Survival in Patients with Cirrhosis. PLoS One 2015; 10: e0138130 [PMID: 26382760 DOI: 10.1371/journal.pone.0138130]
- Mücke VT, Thomas D, Mücke MM, Waidmann O, Zeuzem S, Sarrazin C, Pfeilschifter J, Vermehren 75 J, Finkelmeier F, Grammatikos G. Serum sphingolipids predict de novo hepatocellular carcinoma in hepatitis C cirrhotic patients with sustained virologic response. Liver Int 2019; 39: 2174-2183 [PMID: 31207039 DOI: 10.1111/liv.14178]
- Ioannou GN, Tang W, Beste LA, Tincopa MA, Su GL, Van T, Tapper EB, Singal AG, Zhu J, Waljee 76 AK. Assessment of a Deep Learning Model to Predict Hepatocellular Carcinoma in Patients With Hepatitis C Cirrhosis. JAMA Netw Open 2020; 3: e2015626 [PMID: 32870314 DOI: 10.1001/jamanetworkopen.2020.15626]

WJG

World Journal of Gastroenterology

Submit a Manuscript: https://www.f6publishing.com

World J Gastroenterol 2022 January 7; 28(1): 108-122

DOI: 10.3748/wjg.v28.i1.108

ISSN 1007-9327 (print) ISSN 2219-2840 (online)

MINIREVIEWS

Artificial intelligence in the diagnosis and management of colorectal cancer liver metastases

Gianluca Rompianesi, Francesca Pegoraro, Carlo DL Ceresa, Roberto Montalti, Roberto Ivan Troisi

ORCID number: Gianluca Rompianesi 0000-0003-0756-8013; Francesca Pegoraro 0000-0003-2162-7315; Carlo DL Ceresa 0000-0002-8702-3744; Roberto Montalti 0000-0002-3915-3851: Roberto Ivan Troisi 0000-0001-6280-810X.

Author contributions: Rompianesi G conceptualized the manuscript; Rompianesi G and Montalti R wrote the manuscript; Pegoraro F performed the literature search and the data analysis and extraction; Ceresa CDL and Troisi RI reviewed and edited the manuscript; and all authors have read and approve the final manuscript.

Conflict-of-interest statement: No Author has any conflict of interest to disclose.

Country/Territory of origin: Italy

Specialty type: Gastroenterology and hepatology

Provenance and peer review: Invited article; Externally peer reviewed.

Peer-review model: Single blind

Peer-review report's scientific quality classification

Grade A (Excellent): 0 Grade B (Very good): B Grade C (Good): 0 Grade D (Fair): 0

Gianluca Rompianesi, Francesca Pegoraro, Roberto Ivan Troisi, Division of Hepato-Bilio-Pancreatic, Minimally Invasive and Robotic Surgery, Department of Clinical Medicine and Surgery, Federico II University Hospital, Naples 80125, Italy

Carlo DL Ceresa, Department of Hepato-Pancreato-Biliary Surgery, Oxford University Hospitals NHS Foundation Trust, Oxford OX3 9ES, United Kingdom

Roberto Montalti, Division of Hepato-Bilio-Pancreatic, Minimally Invasive and Robotic Surgery, Department of Public Health, Federico II University Hospital, Naples 80125, Italy

Corresponding author: Gianluca Rompianesi, FEBS, MD, PhD, Assistant Professor, Surgeon, Division of Hepato-Bilio-Pancreatic, Minimally Invasive and Robotic Surgery, Department of Clinical Medicine and Surgery, Federico II University Hospital, via Pansini 5, Naples 80125, Italy. gianlucarompianesi@gmail.com

Abstract

Colorectal cancer (CRC) is the third most common malignancy worldwide, with approximately 50% of patients developing colorectal cancer liver metastasis (CRLM) during the follow-up period. Management of CRLM is best achieved via a multidisciplinary approach and the diagnostic and therapeutic decision-making process is complex. In order to optimize patients' survival and quality of life, there are several unsolved challenges which must be overcome. These primarily include a timely diagnosis and the identification of reliable prognostic factors. Furthermore, to allow optimal treatment options, a precision-medicine, personalized approach is required. The widespread digitalization of healthcare generates a vast amount of data and together with accessible high-performance computing, artificial intelligence (AI) technologies can be applied. By increasing diagnostic accuracy, reducing timings and costs, the application of AI could help mitigate the current shortcomings in CRLM management. In this review we explore the available evidence of the possible role of AI in all phases of the CRLM natural history. Radiomics analysis and convolutional neural networks (CNN) which combine computed tomography (CT) images with clinical data have been developed to predict CRLM development in CRC patients. AI models have also proven themselves to perform similarly or better than expert radiologists in detecting CRLM on CT and magnetic resonance scans or identifying them from the noninvasive analysis of patients' exhaled air. The application of AI and machine learning (ML) in diagnosing CRLM has also been extended to histopathological examination in order to rapidly and accurately identify CRLM tissue and



Grade E (Poor): 0

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: htt ps://creativecommons.org/Licens es/by-nc/4.0/

Received: August 13, 2021 Peer-review started: August 13, 2021

First decision: October 2, 2021 Revised: October 12, 2021 Accepted: December 25, 2021 Article in press: December 25, 2021 Published online: January 7, 2022

P-Reviewer: Liu Y S-Editor: Wang JJ L-Editor: Kerr C P-Editor: Wang JJ



its different histopathological growth patterns. ML and CNN have shown good accuracy in predicting response to chemotherapy, early local tumor progression after ablation treatment, and patient survival after surgical treatment or chemotherapy. Despite the initial enthusiasm and the accumulating evidence, AI technologies' role in healthcare and CRLM management is not yet fully established. Its limitations mainly concern safety and the lack of regulation and ethical considerations. AI is unlikely to fully replace any human role but could be actively integrated to facilitate physicians in their everyday practice. Moving towards a personalized and evidence-based patient approach and management, further larger, prospective and rigorous studies evaluating AI technologies in patients at risk or affected by CRLM are needed.

Key Words: Colorectal cancer; Liver metastases; Artificial intelligence; Machine learning; Deep learning; Neural networks; Radiomics

©The Author(s) 2022. Published by Baishideng Publishing Group Inc. All rights reserved.

Core tip: The digitalization of healthcare generating huge amount of data set the ground for the progressive ubiquitous application of artificial intelligence (AI) technologies in healthcare. AI analyses can assist clinicians in all phases of colorectal liver metastases natural history: From predicting their occurrence, to increasing diagnostic accuracy or estimating recurrence risk after treatment and patient outcome. The implementation of AI resources supports the contemporary paradigm shift that sees healthcare focus moving from a generalized, disease-oriented to an individual, patient-centered, precision medicine approach.

Citation: Rompianesi G, Pegoraro F, Ceresa CD, Montalti R, Troisi RI. Artificial intelligence in the diagnosis and management of colorectal cancer liver metastases. World J Gastroenterol 2022; 28(1): 108-122

URL: https://www.wjgnet.com/1007-9327/full/v28/i1/108.htm DOI: https://dx.doi.org/10.3748/wjg.v28.i1.108

INTRODUCTION

Colorectal cancer liver metastases

Colorectal cancer (CRC) is the most common gastrointestinal cancer, the third most frequently diagnosed malignancy (10.0%) overall, and the second highest cause of cancer-related deaths (9.4%), with incidences varying significantly worldwide[1,2]. CRC development is predominantly sporadic, with patient age, environmental and genetic factors associated with a significantly increased risk[3,4]. Over 20% of newly diagnosed CRC patients have distant metastases at presentation [5], with estimated 5year survival dropping from 80%-90% in patients with local disease to a dismal 10%-15% in those with metastatic spread [6]. The liver is the preferential metastatic site, due to its anatomical proximity and the portal systemic circulation. This results in 25%-50% of CRC patients developing liver metastasis during the course of the disease [7,8]. In cases of synchronous resectable colorectal cancer liver metastasis (CRLM), the treatment options range from the traditional staged approach, where the primary tumor is resected prior to systemic chemotherapy and liver metastasis resection, to the combined approach of bowel and liver resection during the same procedure, or the "liver first" approach[9]. Irrespective of the timing of the surgical resection, surgery in combination with chemotherapy is the optimal treatment for CRLM, but only 25% of patients are suitable candidates for resection at diagnosis[8,10]. In patients not amenable to surgery, chemotherapy is the usual treatment of choice, with the potential to render 10%-30% of tumors technically resectable through a good response and downsizing[11]. CRLM management is multidisciplinary, with oncologists, surgeons, radiologists and pathologists playing pivotal roles in the complex diagnostic and therapeutic decision-making processes aimed to achieve the best possible outcome for the patient[12]. In such a complex oncological scenario, with unsolved challenges in



timely diagnosis, reliable prognostic factor identification and optimal treatment selection, there is a strong need for a precision-medicine, personalized approach in order to optimize patients' survival and quality of life. The recent progressive implementation of artificial intelligence (AI) in healthcare has been welcomed with enthusiasm by both healthcare professionals and the general public; however, there remain several issues which are yet to be solved. AI has the potential to overcome some of the current practice limitations, and to play a crucial role in all steps of the management of CRLM but its clinical benefits have yet to be clearly established and validated.

The aim of this review is to summarize and analyze the available evidence on the application of AI technologies in the diagnosis and management of patients affected by CRLM.

AI

The term AI encompasses all the possible applications of technologies in simulating and replicating human intelligence^[13]. These endless applications range from everyday life to finance and economics[14] or various medical fields, thanks to the advances in computational power and the collection and storage of large amounts of data in healthcare. After being adequately programmed and trained, AI has the potential to outperform clinicians in some tasks in terms of accuracy, speed of execution and reduced biases[15]. AI has therefore progressively demonstrated its potential across all human lifespan; from the optimization of embryo selection during *in vitro* fertilization^[16] to the prediction of all-cause mortality^[17]. The revolutionary potential of these technologies in healthcare has generated great interest in researchers, professionals and industries, with currently over 450 AI-based medical devices approved in Europe or the United States[18]. Nevertheless, the surge of AI and its implemen-tation in clinical practice has been accompanied by several issues including legal considerations regarding security and data, software transparency, flawed algorithms and inherent bias in the input data[13,19].

Machine learning

The replication of human intelligence by AI with the utilization of data-driven algorithms that have been instructed and self-train through experience and data analysis is generally defined as machine learning (ML)[13]. After been programmed, ML can find recurrent patterns in large amount of appropriately engineered data and progressively learn and independently improve performance accuracy without human intervention. The ML algorithms are generally classified in supervised learning (the most frequent one, which utilizes classified data), unsupervised learning (where algorithms can independently identify patterns in data without previous classification), semi-supervised learning (can use a combination of both labelled and unlabeled data) and reinforcement learning (uses estimated errors as proportional rewards or penalties to teach algorithms). Deep learning (DL) is a class of ML techniques that has the ability to directly process raw data and perform detection or classification tasks automatically without the need for human intervention. The sets of algorithms utilized by DL are generally artificial neural networks (ANNs) constituted by several layers that elaborate inputs with weights, biases (or thresholds) and deliver an output. ML models can be combined with the large amount of qualitative and quantitative information mined from medical images (radiomics) and clinical data to assist clinicians in evidence-based decision making processes[20].

PREDICTIVE AI MODELS FOR THE DEVELOPMENT OF CRLM

A significant proportion of patients affected by CRC will develop CRLM during the follow-up period^[21], but only about a quarter of them will be eligible for surgical resection and therefore potential cure[22]. Being able to identify the subgroup of patients at higher risk of CRLM development could allow the adoption of individualized and more intense screening protocols and adjuvant therapies.

The Radiomics Intelligent Analysis Toolkit-based analysis platform built by Li et al [23] allowed the construction of individualized nomograms able to combine maximum-level enhanced computed tomography (CT) images in the portal venous phase and patients' clinical information [age, sex, carcinoembryonic antigen (CEA) and carbohydrate antigen 19-9] to predict the development of CRLM in patients with CRC. The area under the receiver operating characteristic (AUROC) score obtained from the analysis of 100 patients (50 with CRLM and 50 controls) was 0.899 [95% confidence



interval (CI): 0.761-1.000] on the test set in a total execution time of 270 s. The ML predictive models built by Taghavi[24] including radiomics and a com-bination of radiomics with clinical features (contrast-enhanced portal venous phase CT of the liver or abdomen with age, sex, primary tumor site, tumor stage, nodal stage, CEA at primary diagnosis, administration of adjuvant/neoadjuvant chemotherapy) of 91 patients (24 of which developed metachronous CRLM), both presented an area under the curve (AUC) in the validation cohort of 86% (95%CI: 85%-87%) in predicting the development of CRLM within 24 mo. The convolutional neural network (CNN) model developed by Lee et al^[25] in their retrospective, cross-sectional study in 2019 patients who underwent curative colectomy for stage I-III CRC was able to predict 5-year metachronous liver metastasis occurrence with a mean AUC of 0.747 when combining the analysis of the abdominal CT scan taken before the colectomy for clinical staging and clinical features (age, sex, tumor stage, nodal stage).

AI MODELS FOR THE DIAGNOSIS OF CRLM

Prompt diagnosis of CRLM at an early stage gives patients the best chances of effective treatment and a superior outcome. One of the key steps in the diagnostic process is tumor segmentation, with nodule volume being a better predictor than diameter[26]. This process is usually done manually but requires a significant expertise, is operatordependent and time-consuming. In this setting, semiautomatic tumor segmentation methods based on texture analysis have been developed[26] in order to take full advantage of AI's unique potential to increase sensitivity and specificity of metastatic tumor detection[27].

CT radiomics models

Starting with a manual tumor/nontumor class prediction voxel classification, a deformable surface model fitting the tumor boundaries is instigated[27]. A multilayer perceptron feed-forward neural network model concurrently learns per-voxel image features and classifications and, after being trained, it performs a semiautomatic pertumor segmentation on CT scans. The accuracy of the model resulted in 0.88 ± 0.11 , with a sensitivity of 0.84 ± 0.13 and a specificity of 0.92 ± 0.16 . The same group in 2019 published the results of a retrospective analysis of a fully CNN for liver lesion detection and segmentation on CT scans with a sensitivity of 71% and 85% and a positive predictive value of 83% and 94% for lesions bigger than 10 mm and 20 mm in diameter, respectively^[28]. CRLM is most commonly diagnosed in the venous phase of contrast-enhanced CT scan, as it appears hypodense, with or without peripheral rim enhancement and calcification. Portal-venous phase scans are most reliable in the detection of CRLM, with a sensitivity of approximately 85% for helical CT[29], and such diagnostic power lies in an optimal timing of image acquisition after a delay following contrast intravenous injection. Different equipment, protocols, patient's body habitus and cardiovascular system function result in high variability and impact on measurement accuracy in the absence of reliable automatic timing quantification. Ma et al[30] designed a fully automatic DL CNN that in a 3-s timespan can recognize the optimal portal venous phase acquisitions on CT scans with an AUC of 0.837 (95% CI: 0.765-0.890) in the validation set and an AUC of 0.844 (95% CI: 0.786-0.889) in the external validation set. This is aimed to improve image quality, which is crucial for the detection and characterization of liver lesions and the evaluation of parameters identified as predictors of treatment response and outcome, such as the tumor size, enhancement and vascularity[30]. The DL-based algorithm of Kim et al[31] aimed at detecting CRLM without human manipulation and fed by raw data from CT images, showed a sensitivity of 81.82%, comparable to that of radiologists (80.81%, P = 0.80), but with significantly more false positives per patient (1.330 vs 0.357, P < 0.001).

A challenging scenario that can occur in 16%-26% of patients with CRC is when the staging CT scan shows small hypoattenuating hepatic nodules defined as too small to characterize. Further imaging such as magnetic resonance (MR), repeat CT after a time interval, or performing a biopsy can delay treatment, increase costs, remain inconclusive, or have the risk of complications and tumor seeding. However, obtaining a diagnosis is of paramount importance given that 9%-14% of these nodules will prove to be malignant[32,33]. CNN could represent a useful adjunct in the characterization of small hypoattenuating liver lesions, and the model developed by Khalili et al[34] presents an AUROC similar to the one of expert radiologists, with better diagnostic confidence (significantly lower proportion of nodules rated in the low confidence zone, 19.6 vs 38.4%).



MR radiomics models

Despite CT imaging being the most widely used modality in detecting metastatic liver tumors, it can still miss up to 25% of CRLM[35] and MR has progressively gained an established role thanks to the high sensitivity and specificity and absence of ionizing radiations[36,37]. AI utilizing CNN for liver segmentation and CRLM detection could assist radiologists in this complex task and potentially reduce the manual liver lesion detection failure rate of 5%-13% [38]. The CRLM detection method developed by Jansen *et al*[38] is based on a fully CNN with an automatic liver segmentation and the analysis of both dynamic contrast-enhanced and diffusion-weighted MR images in 121 patients. It resulted in an impressive a high sensitivity of 99.8% and a low number of false positives.

Volatile-organic-compound-based models

Interestingly, a ML model has been used by Steenhuis et al[39] to analyze data from a retrospective cohort of 62 patients following curative CRC resection to detect CRLM development or local recurrence. The volatile organic compounds (VOCs) from patients' exhaled air are gaseous products of metabolism known to be altered by pathological processes, such as abnormal cell growth, necrosis or intestinal microbioma alteration, and have been evaluated by ML techniques for pattern recognition. This pilot study, despite the limitations due to the small sample size and lack of histological confirmation in about a quarter of patients, showed that the noninvasive, repeatable, and easily applicable eNose analysis was able to identify CRLM or local recurrence with a sensitivity of 0.88 (95%CI: 0.69-0.97), specificity of 0.75 (95%CI: 0.57-0.87), and an overall accuracy of 0.81. Miller-Atkins et al[40] combined VOC analysis and demographic data (age and sex) in a predictive model developed using random forest ML and cross-validation that was able to identify patients with CRLM from healthy controls with a classification accuracy of 0.86, specificity of 0.94 but a sensitivity limited to 0.51.

Histology-based models

The applications of AI and ML in diagnosing CRLM have been extended to histopathological examination in order to rapidly and accurately identify CRLM tissue. A probe electrospray ionization-mass spectrometry and ML model was able to distinguish CRLM (103 samples) from noncancer liver parenchyma (80 control samples) with an accuracy rate of 99.5% and a AUROC of 0.9999[41]. CRLM patients are a heterogeneous group with considerable variations, including histopathological growth patterns (HGPs) and corresponding microvasculature[42]. The two predominant types of HGPs are the desmoplastic and replacement, with the pushing and mixed types being far less common. Once accurately determined by analyzing the interface between the tumor cells and the nearby normal liver, HGPs can represent a useful prognostic and predictive biomarker for response to therapy and overall survival[43-46]. The MR-based radiomics model developed by Han *et al*[47] aims at preoperatively identifying HGP of CRLM with an AUC of 0.906 in the internal validation cohort when the analysis is performed on the tumor-liver interface zone.

AI MODELS FOR TREATED CRLM

Surgical resection offers patients presenting with synchronous or metachronous CRLM the only potential for cure and a superior long-term survival^[48] but unfortunately only a fraction of newly diagnosed patients are suitable for surgery. Liverdirected ablative therapies have progressively gained a role in treating nonsurgical candidates with acceptable safety and efficacy profiles [49]. In spite of this, recurrence after CRLM treatment represents a major problem, with an overall risk of local or distant tumor development after surgical resection or ablation as high as 70%-80%, with early recurrences being associated with a poorer prognosis [50,51]. Chemotherapy is of paramount importance in determining outcome of patients with either resectable or unresectable CRLM[8] and can convert up to one third of initially unresectable patients to receive potentially curative treatment[52].

Al models predicting response to chemotherapy

A reliable assessment of response to chemotherapy is of paramount importance for the personalized treatment decision-making process to determine eligibility for surgery, or the need for second-line treatments^[53]. Discriminating responsive from unresponsive



nodules or new lesions on the CT scan often represents a challenging task for radiologists, therefore Maaref et al [54] developed a fully automated framework based on DL CNN that achieved an accuracy of 0.91 (95% CI: 0.88-0.93) for differentiating treated and untreated lesions, and 0.78 (95%CI: 0.74-0.83) for predicting the response to a FOLFOX + bevacizumab-based chemotherapy regimen. Similarly, the DL ra-diomics model by Wei et al^[55] was able to predict response to chemotherapy (CAPEOX, mFOLFOX6, FOLFIRI or XELIRI regimens) of CRLM based on contrast-enhanced CT according to the response evaluation criteria in solid tumors with an AUC in the validation cohort of 0.820 (95% CI: 0.681-0.959) that increases to 0.830 (95% CI: 0.688-0.973) combining the DL-based model with the CEA serum level. Human epidermal growth factor receptor 2 amplification or overexpression is found in 2%-6% of stage 2/3 CRC patients and treatment with trastuzumab and lapatinib has proven to be beneficial in the 70% of metastatic cases[56]. Giannini et al[57] published the results of an ML algorithm predicting the therapeutic response in such a subgroup of patients with an overall sensitivity of 92% (95%CI: 75%-99%) and specificity of 86% (95%CI: 42%-100%). The radiomics-based prediction model for the response of CRLM to oxaliplatin-based chemotherapy developed by Nakanishi et al[58] with radiomics features extracted from the pre-treatment CT scans, significantly discriminated good responders (AUC: 0.7792, 95%CI: 0.618-0.941).

Al models predicting recurrence after local ablative therapies

In order to predict early local tumor progression after ablation treatment of up to five nodules per patient with a maximum diameter of 30 mm, Taghavi et al[59] developed a ML-based radiomics analysis of the pretreatment CT scan combined with patients' clinical features that showed a concordance index in the validation cohort of 0.79 (95%CI: 0.78-0.80).

AI MODELS PREDICTING SURVIVAL IN CRLM PATIENTS

Al models predicting overall survival

The systematic comparative analysis of quantitative imaging biomarkers based on the geometric and radiomics analysis of the liver tumor burden by Mühlberg et al[60], performed on a retrospective cohort of 103 patients with CRLM with automated segmentation of baseline contrast-enhanced CT images, showed that the tumor burden score (TBS) had the best discriminative performance for 1-year survival (AUC: 0.70; 95%CI: 0.56-0.90). The TBS[61] is calculated combining tumor number and maximum diameter through the Pythagorean theorem [TBS² = (maximum tumor diameter)² + (number of liver lesions)²]. An ML method has been used by Hao *et al*[62] to analyze whole-genome methylation data to predict cancer versus normal tissue of four common tumors (including 29 of 30 CRLMs) with > 95% accuracy and patient prognosis and survival through DNA methylation analysis.

Al models predicting survival after chemotherapy

Anti-epidermal growth factor receptor (EGFR) therapies are an effective option for RAS wild-type mutational status CRLM, but there is a need for reliable biomarkers that can estimate the balance between risks and clinical benefits of such therapies in individual patients[63]. Dercle et al[64] developed an AI model that through ML could create a signature that evaluated a change in tumor phenotype on interval CT scan images (baseline to 8 wk). The resultant model was able to successfully predict both sensitivity to anti-EGFR therapy (0.80; 95% CI: 0.69-0.94) and overall survival (P < 0.05).

AI models predicting survival after surgical resection of CRLM

The ANN model constructed by Spelt et al[65] retrospectively analyzed a single-center cohort of 241 patients who underwent liver resection for CRLM. Six of the 28 potential risk variables (age, preoperative chemotherapy, size of largest metastasis, hemorrhagic complications, preoperative CEA level and number of metastases) were selected by the ANN model to predict survival more accurately than the Cox regression model, with C-index of 0.72 versus 0.66. Paredes et al[66] in 2020 published the results of their ML recurrence-free prediction model for patients with CRLM undergoing curative-intent resection using clinical, pathological and morphological tumor characteristics with genetic Kirsten rat sarcoma 2 viral oncogene homolog information. The model, built on the analysis of 1406 multi-institutional patients undergoing liver resection, showed a discriminative ability to predict the recurrence risk at 1, 3 and 5 years (AUROC of



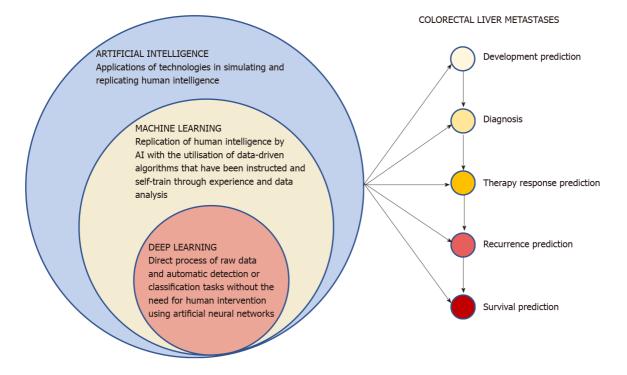


Figure 1 Possible applications of artificial intelligence technologies in the diagnosis and management of colorectal liver metastases. Al: Artificial intelligence.

0.693, 0.669 and 0.669, respectively) more accurate than the ones of Fong[67] and Vauthey[68] scores.

LIMITATIONS

In spite of AI's clear potential there remain several unresolved issues and limitations. These include the potential for artefacts in radiomics analyses to affect the results, the ethical and legal considerations, the definition of minimal accuracy rates and safeguards necessary to ensure public safety. Privacy, sensitive data protection and confidentiality need to remain the unmovable cornerstone of patient rights even in the digitalized era, but at the same time, some limitations on data utilization may affect the necessary linkages to prevent biases or errors in AI-driven analyses. There is a strong need from regulatory bodies for clear guidance during the AI-driven transformation of healthcare in order to take full advantage of the potential major improvements in individual and public health, while ensuring trust, safety and transparency. There is a significant variability in the algorithms investigated so far, as well as heterogeneity in the relatively small sample size of the population on which they have been trained and tested (Table 1). Analyses on large registries or national and international collaborations with data sharing could overcome part of the current limitations that limit the formal recognition of AI as a reliable and reproducible application in clinical scenarios.

CONCLUSION

The progressive widespread availability of high-performance computing, together with the accessibility to a large amount of data constantly generated as the result of the increase in the digitalization, set the ground for the ubiquitous implementation of AI technologies in contemporary healthcare. The fields of medical and surgical oncology have welcomed with enthusiasm the advent of augmented medicine with numerous studies investigating its potential, also given the high complexity and diversity of cancer patients. CRC makes no exception and still represents a leading cause of cancerrelated death due to its high incidence, rapid progression potential and biological heterogeneity that advocate the need for reliable and individualized diagnostic, prognostic and treatment selection tools. Recent years have seen AI technologies tested



Table 1 Summary of the studies considered in this review											
Author	Study design	Al model type	Data source	Total sample size/training cohort/validation cohort	AUC training/AUC validation	Sensitivity/specificity	PPV/NPV	Accuracy			
CRLM deve	elopment										
Li <i>et al</i> [<mark>23</mark>] (2020)	Retrospective; Single center	Radiomics/ML	CT images ± clinical data	100/NA/80	0.90/0.906	81%/84%	85%/79%	NA			
Taghavi <i>et</i> al <mark>[24]</mark> (2021)	Retrospective; Multicenter	Radiomics/ML	CT images ± clinical data	91/70/21	0.95 ² -0.68 ³ - 0.95 ⁴ /0.86 ² - 0.71 ³ -0.86 ⁴	NA/NA	NA/NA	NA			
Lee <i>et al</i> [<mark>25</mark>] (2020)	Retrospective; Single center	Radiomics/CNN	CT images ± clinical data	2019/1413/606	NA/0.606 ² - 0.709 ³ -0.747 ⁴	NA/NA	NA/NA	NA			
Diagnosis											
Vorontsov <i>et al</i> [26] (2017)	Retrospective; Single center	Radiomics/CNN	CT images	40/32/8	NA/NA	84%/92%	NA/NA	88%			
Vorontsov <i>et al</i> [28] (2019)	Retrospective; Single center	Radiomics/CNN	CT images	156/115/15	NA/NA	59% ⁵ /NA	80% ⁵ /NA	NA			
Ma et al [<mark>30</mark>] (2020)	Retrospective; Multicenter	CNN	CT images	909/479/202 (228 ⁶)	NA/0.837- 0.844 ⁶	82% ⁶ /74% ⁵	75% ⁶ /81% ⁶	NA			
Kim <i>et al</i> [<mark>31</mark>] (2021)	Retrospective; Single center	DL	CT images	587/502/85	NA/0.631	81.82%/22.22%	NA/NA	NA			
Khalili et al[<mark>34</mark>] (2020)	Retrospective; Single center	CNN	CT images ± liver metastatic status	199/150/49	NA/0.84-0.95 ⁷	(81.5%-81.5% ⁷)/(76.2%- 96.4% ⁷)	NA/NA	78.3%; 90.6% ⁶			
Jansen <i>et</i> al[<mark>38</mark>] (2019)	Retrospective; Single center	CNN	MRI images	121/334 ¹ /86 ¹	NA/NA	99.8%/NA	NA/NA	NA			
Steenhuis <i>et al</i> [<mark>39]</mark> (2020)	Retrospective; Single center	ML	VOCs	62/NA/NA	NA/0.86	88%/75%	72%/90%	81%			
Miller- Atkins <i>et</i> <i>al</i> [40] (2020)	Prospective; Single center	ML	VOCs	296/284/NA	NA/NA	51%/94%	NA/NA	86%			
Kiritani <i>et</i> al <mark>[41]</mark> (2021)	Retrospective; Single center	ML	Histologic markers	183/NA/40	NA/0.999	100%/99%	NA/NA	99.5%			
Han <i>et al</i> [47] (2020)	Retrospective; Single center	Radiomics/ML	MRI images ± clinical data	107/61 ¹ /31 ¹	0.974 ² -0.659 ³ - 0.971 ⁴ /0.912 ² - 0.676 ³ -0.909 ⁴	95.2% ² -57.1% ³ -95.2% ⁴ /80.0% ² -70.0% ³ -70.0% ⁴	NA/NA	90.3% ² ; 61.3% ³ ; 87.1% ⁴			
Chemother	apy response										
Maaref <i>et</i> al[<mark>54]</mark> (2020)	Retrospective; Single center	DL CNN	CT images	202/70%/10%	0.97/0.88	98%/54%	NA/NA	91% ⁸ ; 78% ⁹			
Wei <i>et al</i> [55] (2021)	Retrospective; Single center	Radiomics/DL	CT images ± CEA	192/144/48	0.903 ¹⁰ -0.935 ¹¹ /0.820 ¹⁰ - 0.830 ¹¹	90.9%/73.3%	88.2%/78.6%	85.4%			
Giannini <i>et al</i> [<mark>57</mark>] (2020)	Retrospective; Multicenter	Radiomics/ML	CT images	38/28/10	NA/NA	92%/86%	96%/75%	NA			
Nakanishi <i>et al</i> [<mark>58]</mark> (2021)	Retrospective; Single center	Radiomics	CT images	42/94 ¹ /32 ¹	0.8512/0.7792	NA/NA	NA/NA	NA			
Local ablative therapies efficacy											
Taghavi et	Retrospective;	Radiomics/ML	CT images	90/63/27	NA/0.78 ² -	NA/NA	NA/NA	NA			

Jaisbideng® WJG | https://www.wjgnet.com

Rompianesi G et al. AI in CRLM diagnosis and management

al[<mark>59</mark>] (2021)	Single center				0.56 ³ -0.79 ⁴							
Survival prediction												
Mühlberg <i>et al</i> [60] (2021)	Retrospective; Single center	Radiomics/ML	CT images ± WLTB ± TBS	103/NA/NA	NA/0.70 ¹² -0.73 ¹³ -0.76 ¹⁴	NA/NA	NA/NA	NA				
Hao <i>et al</i> [62] (2017)	Retrospective; Multicenter	ML	DNA methylation	1792 ¹ /NA/884 ¹ (718 ^{1,6})	NA/NA	NA/NA	NA/NA	98.4%				
Dercle <i>et al</i> [64] (2020)	Retrospective; Multicenter	ML	CT images	667/438/229	0.83/0.80	80%/78%	NA/NA	NA				
Spelt <i>et al</i> [65] (2013)	Retrospective; Single center	ANN	Clinical variables	241/NA/NA	NA/NA	NA/NA	NA/NA	72%				
Paredes <i>et</i> <i>al</i> [66] (2020)	Retrospective; Multicenter	ML	Clinical variables	1406/703/703	$\begin{array}{r} 0.527^{15} \text{-} 0.525^{16} \text{-} \\ 0.693^{17} / 0.524^{15} \text{-} \\ 0.501^{16} \text{-} 0.642^{17} \end{array}$	NA/NA	NA/NA	NA				

¹Number of lesions.

²Model based on radiomics data only.

³Model based on clinical data only.

⁴Model based on both radiomics and clinical data.

⁵Per patient values.

⁶Values calculated on the external validation set.

⁷Model based on both convolutional neural network and liver metastatic status.

⁸For differentiating treated and untreated lesions.

⁹For predicting the response to a FOLFOX + bevacizumab-based chemotherapy regimen.

¹⁰Model based on both deep learning and radiomics signature.

¹¹Model based on deep learning and radiomics signature considering carcinogenic embryonic antigen values.

¹²Model based on tumor burden score.

¹³Model based on geometric metastatic spread of whole liver tumor burden.

¹⁴Model based on the Aerts radiomics prior model.

¹⁵Model based on Fong/Blumgart clinical risk score for predicting 1-year recurrence.

¹⁶Model based on Brudvik-Vauthey clinical risk score for predicting 1-year recurrence.

¹⁷Model based on Paredes–Pawlik clinical risk score for predicting 1-year recurrence.

AI: Artificial intelligence; ANN: Artificial neural network; AUC: Area under the curve; CEA: Carcinogenic embryonic antigen; CNN: Convolutional neural network; CRLM: Colorectal cancer liver metastases; DL: Deep learning; ML: Machine learning; NPV: Negative predictive value; PPV: Positive predictive value; TBS: Tumor burden score; VOCs: Volatile organic compounds; WLTB: Whole liver tumor burden.

> by researchers in all phases of the CRLM natural history, aiming at overcoming the current difficulties and limitations faced by the multidisciplinary team responsible of the patients' care (Figure 1). The possibility of identifying the subgroup of patients at higher risk of CRLM development before the occurrence of the disease from the radiomics baseline CT scan analysis with high accuracy (AUC \ge 0.75) and in less than 5 min could give such patients the best chances of an early diagnosis, more effective treatment, and therefore, a better outcome thanks to a personalized approach[23-25]. Radiomics has also demonstrated a great potential in assisting the radiologists in diagnosing CRLM from CT and MRI scans also by optimizing the identification of the optimal phases for lesions recognition and characterizing small nodules of uncertain nature[27-31,34,38]. A more efficient diagnostic process would help reduce timings and costs, resulting in a potential benefit for both patients and healthcare systems. AI application in order to rapidly and accurately identify CRLM tissue and its different histopathological growth patterns[41,47] could give a significant contribution towards a rapid oncological individualized approach and treatments. AI technologies have also shown potential as a prognostic and outcome tool, predicting with good accuracy response to chemotherapy[54,55,57,58], early local tumor progression after ablation treatment^[59], and patient survival after surgery or chemotherapy^[60,64-66].

> The possibility of reducing human factors and error, increase accuracy and contain timings and costs while adopting a personalized medicine approach is undoubtedly fascinating and appealing, but despite showing promising results, the role of AI in CRLM patients has not yet been fully elucidated. The implementation of AI resources supports the contemporary paradigm shift that sees healthcare focus moving from a generalized, disease-oriented to an individual, patient-centered, precision medicine approach. The effectiveness of ML models lie on a rigid framework in which a welldefined problem and ground truth along with quantitative objective measures to train and validate the algorithm are needed, making the process efficient but rigid. There is



also a balance to be struck between the accuracy and artificial logic and the risk of AI becoming less intelligible and explainable. On the other hand, AI medical technologies could represent a way to enable patients to take ownership of their own care, increasing participation and autonomy for a more personalized approach.

AI will likely affect the immediate future of medicine and patients' management, but rather than replacing the human roles, it will probably be aimed to assist and facilitate physicians in their practice, while being supervised to ensure maximum safety. This could be in the context of diagnostic uncertainty or to assist in planning optimal treatment strategies. A possible future development would be to improve diagnosis and management through the AI analysis and integration of clinical information, radiomic and genetic data thanks to the recent developments in gene sequencing and liquid biopsies, that have showed great potential in gastrointestinal tumors including CRLM[69-72]. A personalized holistic approach providing reliable data for the diagnosis, management and outcome estimation of cancer patients would assist clinicians in the prevention as well as selecting the most appropriate individualized treatment that would grant the patient the best outcome as well as helping patients to make fully informed decisions.

In order to continue to pursue the ambitious goal of improving patients' care through AI healthcare technologies, further larger, prospective, randomized controlled and rigorous studies are needed.

REFERENCES

- Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, Bray F. Global Cancer Statistics 2020: GLOBOCAN Estimates of Incidence and Mortality Worldwide for 36 Cancers in 185 Countries. CA Cancer J Clin 2021; 71: 209-249 [PMID: 33538338 DOI: 10.3322/caac.21660]
- Global Burden of Disease Cancer Collaboration, Fitzmaurice C, Abate D, Abbasi N, Abbastabar H, Abd-Allah F, Abdel-Rahman O, Abdelalim A, Abdoli A, Abdollahpour I, Abdulle ASM, Abebe ND, Abraha HN, Abu-Raddad LJ, Abualhasan A, Adedeji IA, Advani SM, Afarideh M, Afshari M, Aghaali M, Agius D, Agrawal S, Ahmadi A, Ahmadian E, Ahmadpour E, Ahmed MB, Akbari ME, Akinyemiju T, Al-Aly Z, AlAbdulKader AM, Alahdab F, Alam T, Alamene GM, Alemnew BTT, Alene KA, Alinia C, Alipour V, Aljunid SM, Bakeshei FA, Almadi MAH, Almasi-Hashiani A, Alsharif U, Alsowaidi S, Alvis-Guzman N, Amini E, Amini S, Amoako YA, Anbari Z, Anber NH, Andrei CL, Anjomshoa M, Ansari F, Ansariadi A, Appiah SCY, Arab-Zozani M, Arabloo J, Arefi Z, Aremu O, Areri HA, Artaman A, Asayesh H, Asfaw ET, Ashagre AF, Assadi R, Ataeinia B, Atalay HT, Ataro Z, Atique S, Ausloos M, Avila-Burgos L, Avokpaho EFGA, Awasthi A, Awoke N, Ayala Quintanilla BP, Ayanore MA, Ayele HT, Babaee E, Bacha U, Badawi A, Bagherzadeh M, Bagli E, Balakrishnan S, Balouchi A, Bärnighausen TW, Battista RJ, Behzadifar M, Bekele BB, Belay YB, Belayneh YM, Berfield KKS, Berhane A, Bernabe E, Beuran M, Bhakta N, Bhattacharyya K, Biadgo B, Bijani A, Bin Sayeed MS, Birungi C, Bisignano C, Bitew H, Bjørge T, Bleyer A, Bogale KA, Bojia HA, Borzì AM, Bosetti C, Bou-Orm IR, Brenner H, Brewer JD, Briko AN, Briko NI, Bustamante-Teixeira MT, Butt ZA, Carreras G, Carrero JJ, Carvalho F, Castro C, Castro F, Catalá-López F, Cerin E, Chaiah Y, Chanie WF, Chattu VK, Chaturvedi P, Chauhan NS, Chehrazi M, Chiang PP, Chichiabellu TY, Chido-Amajuoyi OG, Chimed-Ochir O, Choi JJ, Christopher DJ, Chu DT, Constantin MM, Costa VM, Crocetti E, Crowe CS, Curado MP, Dahlawi SMA, Damiani G, Darwish AH, Daryani A, das Neves J, Demeke FM, Demis AB, Demissie BW, Demoz GT, Denova-Gutiérrez E, Derakhshani A, Deribe KS, Desai R, Desalegn BB, Desta M, Dey S, Dharmaratne SD, Dhimal M, Diaz D, Dinberu MTT, Djalalinia S, Doku DT, Drake TM, Dubey M, Dubljanin E, Duken EE, Ebrahimi H, Effiong A, Eftekhari A, El Sayed I, Zaki MES, El-Jaafary SI, El-Khatib Z, Elemineh DA, Elkout H, Ellenbogen RG, Elsharkawy A, Emamian MH, Endalew DA, Endries AY, Eshrati B, Fadhil I, Fallah Omrani V, Faramarzi M, Farhangi MA, Farioli A, Farzadfar F, Fentahun N, Fernandes E, Feyissa GT, Filip I, Fischer F, Fisher JL, Force LM, Foroutan M, Freitas M, Fukumoto T, Futran ND, Gallus S, Gankpe FG, Gayesa RT, Gebrehiwot TT, Gebremeskel GG, Gedefaw GA, Gelaw BK, Geta B, Getachew S, Gezae KE, Ghafourifard M, Ghajar A, Ghashghaee A, Gholamian A, Gill PS, Ginindza TTG, Girmay A, Gizaw M, Gomez RS, Gopalani SV, Gorini G, Goulart BNG, Grada A, Ribeiro Guerra M, Guimaraes ALS, Gupta PC, Gupta R, Hadkhale K, Haj-Mirzaian A, Hamadeh RR, Hamidi S, Hanfore LK, Haro JM, Hasankhani M, Hasanzadeh A, Hassen HY, Hay RJ, Hay SI, Henok A, Henry NJ, Herteliu C, Hidru HD, Hoang CL, Hole MK, Hoogar P, Horita N, Hosgood HD, Hosseini M, Hosseinzadeh M, Hostiuc M, Hostiuc S, Househ M, Hussen MM, Ileanu B, Ilic MD, Innos K, Irvani SSN, Iseh KR, Islam SMS, Islami F, Jafari Balalami N, Jafarinia M, Jahangiry L, Jahani MA, Jahanmehr N, Jakovljevic M, James SL, Javanbakht M, Jayaraman S, Jee SH, Jenabi E, Jha RP, Jonas JB, Jonnagaddala J, Joo T, Jungari SB, Jürisson M, Kabir A, Kamangar F, Karch A, Karimi N, Karimian A, Kasaeian A, Kasahun GG, Kassa B, Kassa TD, Kassaw MW, Kaul A, Keiyoro PN, Kelbore AG, Kerbo AA, Khader YS, Khalilarjmandi M, Khan EA, Khan G, Khang YH, Khatab K, Khater A, Khayamzadeh M, Khazaee-Pool M, Khazaei S, Khoja AT, Khosravi MH, Khubchandani J, Kianipour N, Kim D, Kim YJ, Kisa A, Kisa S, Kissimova-Skarbek K, Komaki



H, Koyanagi A, Krohn KJ, Bicer BK, Kugbey N, Kumar V, Kuupiel D, La Vecchia C, Lad DP, Lake EA, Lakew AM, Lal DK, Lami FH, Lan Q, Lasrado S, Lauriola P, Lazarus JV, Leigh J, Leshargie CT, Liao Y, Limenih MA, Listl S, Lopez AD, Lopukhov PD, Lunevicius R, Madadin M, Magdeldin S, El Razek HMA, Majeed A, Maleki A, Malekzadeh R, Manafi A, Manafi N, Manamo WA, Mansourian M, Mansournia MA, Mantovani LG, Maroufizadeh S, Martini SMS, Mashamba-Thompson TP, Massenburg BB, Maswabi MT, Mathur MR, McAlinden C, McKee M, Meheretu HAA, Mehrotra R, Mehta V, Meier T, Melaku YA, Meles GG, Meles HG, Melese A, Melku M, Memiah PTN, Mendoza W, Menezes RG, Merat S, Meretoja TJ, Mestrovic T, Miazgowski B, Miazgowski T, Mihretie KMM, Miller TR, Mills EJ, Mir SM, Mirzaei H, Mirzaei HR, Mishra R, Moazen B, Mohammad DK, Mohammad KA, Mohammad Y, Darwesh AM, Mohammadbeigi A, Mohammadi H, Mohammadi M, Mohammadian M, Mohammadian-Hafshejani A, Mohammadoo-Khorasani M, Mohammadpourhodki R, Mohammed AS, Mohammed JA, Mohammed S, Mohebi F, Mokdad AH, Monasta L, Moodley Y, Moosazadeh M, Moossavi M, Moradi G, Moradi-Joo M, Moradi-Lakeh M, Moradpour F, Morawska L, Morgado-da-Costa J, Morisaki N, Morrison SD, Mosapour A, Mousavi SM, Muche AA, Muhammed OSS, Musa J, Nabhan AF, Naderi M, Nagarajan AJ, Nagel G, Nahvijou A, Naik G, Najafi F, Naldi L, Nam HS, Nasiri N, Nazari J, Negoi I, Neupane S, Newcomb PA, Nggada HA, Ngunjiri JW, Nguyen CT, Nikniaz L, Ningrum DNA, Nirayo YL, Nixon MR, Nnaji CA, Nojomi M, Nosratnejad S, Shiadeh MN, Obsa MS, Ofori-Asenso R, Ogbo FA, Oh IH, Olagunju AT, Olagunju TO, Oluwasanu MM, Omonisi AE, Onwujekwe OE, Oommen AM, Oren E, Ortega-Altamirano DDV, Ota E, Otstavnov SS, Owolabi MO, P A M, Padubidri JR, Pakhale S, Pakpour AH, Pana A, Park EK, Parsian H, Pashaei T, Patel S, Patil ST, Pennini A, Pereira DM, Piccinelli C, Pillay JD, Pirestani M, Pishgar F, Postma MJ, Pourjafar H, Pourmalek F, Pourshams A, Prakash S, Prasad N, Qorbani M, Rabiee M, Rabiee N, Radfar A, Rafiei A, Rahim F, Rahimi M, Rahman MA, Rajati F, Rana SM, Raoofi S, Rath GK, Rawaf DL, Rawaf S, Reiner RC, Renzaho AMN, Rezaei N, Rezapour A, Ribeiro AI, Ribeiro D, Ronfani L, Roro EM, Roshandel G, Rostami A, Saad RS, Sabbagh P, Sabour S, Saddik B, Safiri S, Sahebkar A, Salahshoor MR, Salehi F, Salem H, Salem MR, Salimzadeh H, Salomon JA, Samy AM, Sanabria J, Santric Milicevic MM, Sartorius B, Sarveazad A, Sathian B, Satpathy M, Savic M, Sawhney M, Sayyah M, Schneider IJC, Schöttker B, Sekerija M, Sepanlou SG, Sepehrimanesh M, Seyedmousavi S, Shaahmadi F, Shabaninejad H, Shahbaz M, Shaikh MA, Shamshirian A, Shamsizadeh M, Sharafi H, Sharafi Z, Sharif M, Sharifi A, Sharifi H, Sharma R, Sheikh A, Shirkoohi R, Shukla SR, Si S, Siabani S, Silva DAS, Silveira DGA, Singh A, Singh JA, Sisay S, Sitas F, Sobngwi E, Soofi M, Soriano JB, Stathopoulou V, Sufiyan MB, Tabarés-Seisdedos R, Tabuchi T, Takahashi K, Tamtaji OR, Tarawneh MR, Tassew SG, Taymoori P, Tehrani-Banihashemi A, Temsah MH, Temsah O, Tesfay BE, Tesfay FH, Teshale MY, Tessema GA, Thapa S, Tlaye KG, Topor-Madry R, Tovani-Palone MR, Traini E, Tran BX, Tran KB, Tsadik AG, Ullah I, Uthman OA, Vacante M, Vaezi M, Varona Pérez P, Veisani Y, Vidale S, Violante FS, Vlassov V, Vollset SE, Vos T, Vosoughi K, Vu GT, Vujcic IS, Wabinga H, Wachamo TM, Wagnew FS, Waheed Y, Weldegebreal F, Weldesamuel GT, Wijeratne T, Wondafrash DZ, Wonde TE, Wondmieneh AB, Workie HM, Yadav R, Yadegar A, Yadollahpour A, Yaseri M, Yazdi-Feyzabadi V, Yeshaneh A, Yimam MA, Yimer EM, Yisma E, Yonemoto N, Younis MZ, Yousefi B, Yousefifard M, Yu C, Zabeh E, Zadnik V, Moghadam TZ, Zaidi Z, Zamani M, Zandian H, Zangeneh A, Zaki L, Zendehdel K, Zenebe ZM, Zewale TA, Ziapour A, Zodpey S, Murray CJL. Global, Regional, and National Cancer Incidence, Mortality, Years of Life Lost, Years Lived With Disability, and Disability-Adjusted Life-Years for 29 Cancer Groups, 1990 to 2017: A Systematic Analysis for the Global Burden of Disease Study. JAMA Oncol 2019; 5: 1749-1768 [PMID: 31560378 DOI: 10.1001/jamaoncol.2019.2996]

- 3 Chan AT, Giovannucci EL. Primary prevention of colorectal cancer. Gastroenterology 2010; 138: 2029-2043.e10 [PMID: 20420944 DOI: 10.1053/j.gastro.2010.01.057]
- Kastrinos F, Syngal S. Inherited colorectal cancer syndromes. Cancer J 2011; 17: 405-415 [PMID: 4 22157284 DOI: 10.1097/PPO.0b013e318237e408]
- van der Pool AE, Damhuis RA, Ijzermans JN, de Wilt JH, Eggermont AM, Kranse R, Verhoef C. 5 Trends in incidence, treatment and survival of patients with stage IV colorectal cancer: a populationbased series. Colorectal Dis 2012; 14: 56-61 [PMID: 21176063 DOI: 10.1111/j.1463-1318.2010.02539.x
- Miller KD, Nogueira L, Mariotto AB, Rowland JH, Yabroff KR, Alfano CM, Jemal A, Kramer JL, 6 Siegel RL. Cancer treatment and survivorship statistics, 2019. CA Cancer J Clin 2019; 69: 363-385 [PMID: 31184787 DOI: 10.3322/caac.21565]
- 7 Sheth KR, Clary BM. Management of hepatic metastases from colorectal cancer. Clin Colon Rectal Surg 2005; 18: 215-223 [PMID: 20011304 DOI: 10.1055/s-2005-916282]
- 8 Chow FC, Chok KS. Colorectal liver metastases: An update on multidisciplinary approach. World J Hepatol 2019; 11: 150-172 [PMID: 30820266 DOI: 10.4254/wjh.v11.i2.150]
- Lillemoe HA, Vauthey JN. Surgical approach to synchronous colorectal liver metastases: staged, combined, or reverse strategy. Hepatobiliary Surg Nutr 2020; 9: 25-34 [PMID: 32140476 DOI: 10.21037/hbsn.2019.05.14]
- 10 Arshad U, Sutton PA, Ashford MB, Treacher KE, Liptrott NJ, Rannard SP, Goldring CE, Owen A. Critical considerations for targeting colorectal liver metastases with nanotechnology. Wiley Interdiscip Rev Nanomed Nanobiotechnol 2020; 12: e1588 [PMID: 31566913 DOI: 10.1002/wnan.1588]
- Ismaili N. Treatment of colorectal liver metastases. World J Surg Oncol 2011; 9: 154 [PMID: 11



22115124 DOI: 10.1186/1477-7819-9-154]

- Weledji EP. Centralization of Liver Cancer Surgery and Impact on Multidisciplinary Teams Working 12 on Stage IV Colorectal Cancer. Oncol Rev 2017; 11: 331 [PMID: 28814999 DOI: 10.4081/oncol.2017.331]
- 13 Topol EJ. High-performance medicine: the convergence of human and artificial intelligence. Nat Med 2019; 25: 44-56 [PMID: 30617339 DOI: 10.1038/s41591-018-0300-7]
- 14 Moloi T, Marwala, T. Artificial Intelligence in Economics and Finance Theories. In: Introduction to Artificial Intelligence in Economics and Finance Theories. 2020: 1-12 [DOI: 10.1007/978-3-030-42962-1_1]
- 15 Matheny ME, Whicher D, Thadaney Israni S. Artificial Intelligence in Health Care: A Report From the National Academy of Medicine. JAMA 2020; 323: 509-510 [PMID: 31845963 DOI: 10.1001/jama.2019.21579]
- 16 Xi Q, Yang Q, Wang M, Huang B, Zhang B, Li Z, Liu S, Yang L, Zhu L, Jin L. Individualized embryo selection strategy developed by stacking machine learning model for better in vitro fertilization outcomes: an application study. Reprod Biol Endocrinol 2021; 19: 53 [PMID: 33820565 DOI: 10.1186/s12958-021-00734-z]
- Ulloa Cerna AE, Jing L, Good CW, vanMaanen DP, Raghunath S, Suever JD, Nevius CD, Wehner 17 GJ, Hartzel DN, Leader JB, Alsaid A, Patel AA, Kirchner HL, Pfeifer JM, Carry BJ, Pattichis MS, Haggerty CM, Fornwalt BK. Deep-learning-assisted analysis of echocardiographic videos improves predictions of all-cause mortality. Nat Biomed Eng 2021; 5: 546-554 [PMID: 33558735 DOI: 10.1038/s41551-020-00667-9
- Muehlematter UJ, Daniore P, Vokinger KN. Approval of artificial intelligence and machine 18 learning-based medical devices in the USA and Europe (2015-20): a comparative analysis. Lancet Digit Health 2021; 3: e195-e203 [PMID: 33478929 DOI: 10.1016/S2589-7500(20)30292-2]
- Benjamens S, Dhunnoo P, Meskó B. The state of artificial intelligence-based FDA-approved medical 19 devices and algorithms: an online database. NPJ Digit Med 2020; 3: 118 [PMID: 32984550 DOI: 10.1038/s41746-020-00324-0
- 20 Gillies RJ, Kinahan PE, Hricak H. Radiomics: Images Are More than Pictures, They Are Data. Radiology 2016; 278: 563-577 [PMID: 26579733 DOI: 10.1148/radiol.2015151169]
- 21 Hackl C, Neumann P, Gerken M, Loss M, Klinkhammer-Schalke M, Schlitt HJ. Treatment of colorectal liver metastases in Germany: a ten-year population-based analysis of 5772 cases of primary colorectal adenocarcinoma. BMC Cancer 2014; 14: 810 [PMID: 25369977 DOI: 10.1186/1471-2407-14-810
- 22 Engstrand J, Nilsson H, Strömberg C, Jonas E, Freedman J. Colorectal cancer liver metastases a population-based study on incidence, management and survival. BMC Cancer 2018; 18: 78 [PMID: 29334918 DOI: 10.1186/s12885-017-3925-x]
- Li M, Li X, Guo Y, Miao Z, Liu X, Guo S, Zhang H. Development and assessment of an 23 individualized nomogram to predict colorectal cancer liver metastases. Quant Imaging Med Surg 2020; 10: 397-414 [PMID: 32190566 DOI: 10.21037/qims.2019.12.16]
- Taghavi M, Trebeschi S, Simões R, Meek DB, Beckers RCJ, Lambregts DMJ, Verhoef C, Houwers 24 JB, van der Heide UA, Beets-Tan RGH, Maas M. Machine learning-based analysis of CT radiomics model for prediction of colorectal metachronous liver metastases. Abdom Radiol (NY) 2021; 46: 249-256 [PMID: 32583138 DOI: 10.1007/s00261-020-02624-1]
- 25 Lee S, Choe EK, Kim SY, Kim HS, Park KJ, Kim D. Liver imaging features by convolutional neural network to predict the metachronous liver metastasis in stage I-III colorectal cancer patients based on preoperative abdominal CT scan. BMC Bioinformatics 2020; 21: 382 [PMID: 32938394 DOI: 10.1186/s12859-020-03686-0
- 26 Vorontsov E, Tang A, Roy D, Pal CJ, Kadoury S. Metastatic liver tumour segmentation with a neural network-guided 3D deformable model. Med Biol Eng Comput 2017; 55: 127-139 [PMID: 27106756 DOI: 10.1007/s11517-016-1495-81
- Zheng Q, Yang L, Zeng B, Li J, Guo K, Liang Y, Liao G. Artificial intelligence performance in 27 detecting tumor metastasis from medical radiology imaging: A systematic review and meta-analysis. EclinicalMedicine 2021; 31: 100669 [PMID: 33392486 DOI: 10.1016/j.eclinm.2020.100669]
- 28 Vorontsov E, Cerny M, Régnier P, Di Jorio L, Pal CJ, Lapointe R, Vandenbroucke-Menu F, Turcotte S, Kadoury S, Tang A. Deep Learning for Automated Segmentation of Liver Lesions at CT in Patients with Colorectal Cancer Liver Metastases. Radiol Artif Intell 2019; 1: 180014 [PMID: 33937787 DOI: 10.1148/ryai.2019180014]
- 29 Schima W, Kulinna C, Langenberger H, Ba-Ssalamah A. Liver metastases of colorectal cancer: US, CT or MR? Cancer Imaging 2005; 5 Spec No A: S149-S156 [PMID: 16361131 DOI: 10.1102/1470-7330.2005.0035
- Ma J, Dercle L, Lichtenstein P, Wang D, Chen A, Zhu J, Piessevaux H, Zhao J, Schwartz LH, Lu L, 30 Zhao B. Automated Identification of Optimal Portal Venous Phase Timing with Convolutional Neural Networks. Acad Radiol 2020; 27: e10-e18 [PMID: 31151901 DOI: 10.1016/j.acra.2019.02.024]
- Kim K, Kim S, Han K, Bae H, Shin J, Lim JS. Diagnostic Performance of Deep Learning-Based 31 Lesion Detection Algorithm in CT for Detecting Hepatic Metastasis from Colorectal Cancer. Korean J Radiol 2021; 22: 912-921 [PMID: 33686820 DOI: 10.3348/kjr.2020.0447]
- Jang HJ, Lim HK, Lee WJ, Lee SJ, Yun JY, Choi D. Small hypoattenuating lesions in the liver on 32 single-phase helical CT in preoperative patients with gastric and colorectal cancer: prevalence, significance, and differentiating features. J Comput Assist Tomogr 2002; 26: 718-724 [PMID:



12439304 DOI: 10.1097/00004728-200209000-00009]

- Lim GH, Koh DC, Cheong WK, Wong KS, Tsang CB. Natural history of small, "indeterminate" 33 hepatic lesions in patients with colorectal cancer. Dis Colon Rectum 2009; 52: 1487-1491 [PMID: 19617765 DOI: 10.1007/DCR.0013e3181a74d5e]
- 34 Khalili K, Lawlor RL, Pourafkari M, Lu H, Tyrrell P, Kim TK, Jang HJ, Johnson SA, Martel AL. Convolutional neural networks versus radiologists in characterization of small hypoattenuating hepatic nodules on CT: a critical diagnostic challenge in staging of colorectal carcinoma. Sci Rep 2020; 10: 15248 [PMID: 32943654 DOI: 10.1038/s41598-020-71364-5]
- Xu LH, Cai SJ, Cai GX, Peng WJ. Imaging diagnosis of colorectal liver metastases. World J 35 Gastroenterol 2011; 17: 4654-4659 [PMID: 22180707 DOI: 10.3748/wjg.v17.i42.4654]
- Böttcher J, Hansch A, Pfeil A, Schmidt P, Malich A, Schneeweiss A, Maurer MH, Streitparth F, 36 Teichgräber UK, Renz DM. Detection and classification of different liver lesions: comparison of Gd-EOB-DTPA-enhanced MRI versus multiphasic spiral CT in a clinical single centre investigation. Eur J Radiol 2013; 82: 1860-1869 [PMID: 23932636 DOI: 10.1016/j.ejrad.2013.06.013]
- Mao Y, Chen B, Wang H, Zhang Y, Yi X, Liao W, Zhao L. Diagnostic performance of magnetic 37 resonance imaging for colorectal liver metastasis: A systematic review and meta-analysis. Sci Rep 2020; 10: 1969 [PMID: 32029809 DOI: 10.1038/s41598-020-58855-1]
- Jansen MJA, Kuijf HJ, Niekel M, Veldhuis WB, Wessels FJ, Viergever MA, Pluim JPW. Liver 38 segmentation and metastases detection in MR images using convolutional neural networks. J Med Imaging (Bellingham) 2019; 6: 044003 [PMID: 31620549 DOI: 10.1117/1.JMI.6.4.044003]
- 39 Steenhuis EGM, Schoenaker IJH, de Groot JWB, Fiebrich HB, de Graaf JC, Brohet RM, van Dijk JD, van Westreenen HL, Siersema PD, de Vos Tot Nederveen Cappel WH. Feasibility of volatile organic compound in breath analysis in the follow-up of colorectal cancer: A pilot study. Eur J Surg Oncol 2020; 46: 2068-2073 [PMID: 32778485 DOI: 10.1016/j.ejso.2020.07.028]
- Miller-Atkins G, Acevedo-Moreno LA, Grove D, Dweik RA, Tonelli AR, Brown JM, Allende DS, 40 Aucejo F, Rotroff DM. Breath Metabolomics Provides an Accurate and Noninvasive Approach for Screening Cirrhosis, Primary, and Secondary Liver Tumors. Hepatol Commun 2020; 4: 1041-1055 [PMID: 32626836 DOI: 10.1002/hep4.1499]
- 41 Kiritani S, Yoshimura K, Arita J, Kokudo T, Hakoda H, Tanimoto M, Ishizawa T, Akamatsu N, Kaneko J, Takeda S, Hasegawa K. A new rapid diagnostic system with ambient mass spectrometry and machine learning for colorectal liver metastasis. BMC Cancer 2021; 21: 262 [PMID: 33691644 DOI: 10.1186/s12885-021-08001-5]
- 42 Vermeulen PB, Colpaert C, Salgado R, Royers R, Hellemans H, Van Den Heuvel E, Goovaerts G, Dirix LY, Van Marck E. Liver metastases from colorectal adenocarcinomas grow in three patterns with different angiogenesis and desmoplasia. J Pathol 2001; 195: 336-342 [PMID: 11673831 DOI: 10.1002/path.966]
- Nielsen K, Rolff HC, Eefsen RL, Vainer B. The morphological growth patterns of colorectal liver 43 metastases are prognostic for overall survival. Mod Pathol 2014; 27: 1641-1648 [PMID: 24851832 DOI: 10.1038/modpathol.2014.4]
- van Dam PJ, van der Stok EP, Teuwen LA, Van den Eynden GG, Illemann M, Frentzas S, Majeed AW, Eefsen RL, Coebergh van den Braak RRJ, Lazaris A, Fernandez MC, Galjart B, Laerum OD, Rayes R, Grünhagen DJ, Van de Paer M, Sucaet Y, Mudhar HS, Schvimer M, Nyström H, Kockx M, Bird NC, Vidal-Vanaclocha F, Metrakos P, Simoneau E, Verhoef C, Dirix LY, Van Laere S, Gao ZH, Brodt P, Reynolds AR, Vermeulen PB. International consensus guidelines for scoring the histopathological growth patterns of liver metastasis. Br J Cancer 2017; 117: 1427-1441 [PMID: 28982110 DOI: 10.1038/bjc.2017.334]
- Stremitzer S, Vermeulen P, Graver S, Kockx M, Dirix L, Yang D, Zhang W, Stift J, Wrba F, 45 Gruenberger T, Lenz HJ, Scherer SJ. Immune phenotype and histopathological growth pattern in patients with colorectal liver metastases. Br J Cancer 2020; 122: 1518-1524 [PMID: 32205863 DOI: 10.1038/s41416-020-0812-z
- Frentzas S, Simoneau E, Bridgeman VL, Vermeulen PB, Foo S, Kostaras E, Nathan M, Wotherspoon 46 A, Gao ZH, Shi Y, Van den Eynden G, Daley F, Peckitt C, Tan X, Salman A, Lazaris A, Gazinska P, Berg TJ, Eltahir Z, Ritsma L, Van Rheenen J, Khashper A, Brown G, Nystrom H, Sund M, Van Laere S, Loyer E, Dirix L, Cunningham D, Metrakos P, Reynolds AR. Vessel co-option mediates resistance to anti-angiogenic therapy in liver metastases. Nat Med 2016; 22: 1294-1302 [PMID: 27748747 DOI: 10.1038/nm.4197]
- 47 Han Y, Chai F, Wei J, Yue Y, Cheng J, Gu D, Zhang Y, Tong T, Sheng W, Hong N, Ye Y, Wang Y, Tian J. Identification of Predominant Histopathological Growth Patterns of Colorectal Liver Metastasis by Multi-Habitat and Multi-Sequence Based Radiomics Analysis. Front Oncol 2020; 10: 1363 [PMID: 32923388 DOI: 10.3389/fonc.2020.01363]
- 48 Abdalla EK, Vauthey JN, Ellis LM, Ellis V, Pollock R, Broglio KR, Hess K, Curley SA. Recurrence and outcomes following hepatic resection, radiofrequency ablation, and combined resection/ablation for colorectal liver metastases. Ann Surg 2004; 239: 818-825; discussion 825-827 [PMID: 15166961 DOI: 10.1097/01.sla.0000128305.90650.71]
- Di Martino M, Rompianesi G, Mora-Guzmán I, Martín-Pérez E, Montalti R, Troisi RI. Systematic 49 review and meta-analysis of local ablative therapies for resectable colorectal liver metastases. Eur J Surg Oncol 2020; 46: 772-781 [PMID: 31862133 DOI: 10.1016/j.ejso.2019.12.003]
- 50 Sorbye H. Recurrence patterns after resection of liver metastases from colorectal cancer. Recent Results Cancer Res 2014; 203: 243-252 [PMID: 25103010 DOI: 10.1007/978-3-319-08060-4 17]



- Bredt LC, Rachid AF. Predictors of recurrence after a first hepatectomy for colorectal cancer liver 51 metastases: a retrospective analysis. World J Surg Oncol 2014; 12: 391 [PMID: 25528650 DOI: 10.1186/1477-7819-12-391
- 52 Folprecht G, Gruenberger T, Bechstein W, Raab HR, Weitz J, Lordick F, Hartmann JT, Stoehlmacher-Williams J, Lang H, Trarbach T, Liersch T, Ockert D, Jaeger D, Steger U, Suedhoff T, Rentsch A, Köhne CH. Survival of patients with initially unresectable colorectal liver metastases treated with FOLFOX/cetuximab or FOLFIRI/cetuximab in a multidisciplinary concept (CELIM study). Ann Oncol 2014; 25: 1018-1025 [PMID: 24585720 DOI: 10.1093/annonc/mdu088]
- 53 Bonanni L, de'Liguori Carino N, Deshpande R, Ammori BJ, Sherlock DJ, Valle JW, Tam E, O'Reilly DA. A comparison of diagnostic imaging modalities for colorectal liver metastases. Eur J Surg Oncol 2014; 40: 545-550 [PMID: 24491289 DOI: 10.1016/j.ejso.2013.12.023]
- 54 Maaref A, Romero FP, Montagnon E, Cerny M, Nguyen B, Vandenbroucke F, Soucy G, Turcotte S, Tang A, Kadoury S. Predicting the Response to FOLFOX-Based Chemotherapy Regimen from Untreated Liver Metastases on Baseline CT: a Deep Neural Network Approach. J Digit Imaging 2020; **33**: 937-945 [PMID: 32193665 DOI: 10.1007/s10278-020-00332-2]
- Wei J, Cheng J, Gu D, Chai F, Hong N, Wang Y, Tian J. Deep learning-based radiomics predicts 55 response to chemotherapy in colorectal liver metastases. Med Phys 2021; 48: 513-522 [PMID: 33119899 DOI: 10.1002/mp.14563]
- Meric-Bernstam F, Hurwitz H, Raghav KPS, McWilliams RR, Fakih M, VanderWalde A, Swanton 56 C, Kurzrock R, Burris H, Sweeney C, Bose R, Spigel DR, Beattie MS, Blotner S, Stone A, Schulze K, Cuchelkar V, Hainsworth J. Pertuzumab plus trastuzumab for HER2-amplified metastatic colorectal cancer (MyPathway): an updated report from a multicentre, open-label, phase 2a, multiple basket study. Lancet Oncol 2019; 20: 518-530 [PMID: 30857956 DOI: 10.1016/S1470-2045(18)30904-5]
- 57 Giannini V, Rosati S, Defeudis A, Balestra G, Vassallo L, Cappello G, Mazzetti S, De Mattia C, Rizzetto F, Torresin A, Sartore-Bianchi A, Siena S, Vanzulli A, Leone F, Zagonel V, Marsoni S, Regge D. Radiomics predicts response of individual HER2-amplified colorectal cancer liver metastases in patients treated with HER2-targeted therapy. Int J Cancer 2020; 147: 3215-3223 [PMID: 32875550 DOI: 10.1002/ijc.33271]
- Nakanishi R, Oki E, Hasuda H, Sano E, Miyashita Y, Sakai A, Koga N, Kuriyama N, Nonaka K, 58 Fujimoto Y, Jogo T, Hokonohara K, Hu Q, Hisamatsu Y, Ando K, Kimura Y, Yoshizumi T, Mori M. Radiomics Texture Analysis for the Identification of Colorectal Liver Metastases Sensitive to First-Line Oxaliplatin-Based Chemotherapy. Ann Surg Oncol 2021; 28: 2975-2985 [PMID: 33454878 DOI: 10.1245/s10434-020-09581-51
- Taghavi M, Staal F, Gomez Munoz F, Imani F, Meek DB, Simões R, Klompenhouwer LG, van der 59 Heide UA, Beets-Tan RGH, Maas M. CT-Based Radiomics Analysis Before Thermal Ablation to Predict Local Tumor Progression for Colorectal Liver Metastases. Cardiovasc Intervent Radiol 2021; 44: 913-920 [PMID: 33506278 DOI: 10.1007/s00270-020-02735-8]
- 60 Mühlberg A, Holch JW, Heinemann V, Huber T, Moltz J, Maurus S, Jäger N, Liu L, Froelich MF, Katzmann A, Gresser E, Taubmann O, Sühling M, Nörenberg D. The relevance of CT-based geometric and radiomics analysis of whole liver tumor burden to predict survival of patients with metastatic colorectal cancer. Eur Radiol 2021; 31: 834-846 [PMID: 32851450 DOI: 10.1007/s00330-020-07192-y
- 61 Sasaki K, Margonis GA, Andreatos N, Zhang XF, Buettner S, Wang J, Deshwar A, He J, Wolfgang CL, Weiss M, Pawlik TM. The prognostic utility of the "Tumor Burden Score" based on preoperative radiographic features of colorectal liver metastases. J Surg Oncol 2017; 116: 515-523 [PMID: 28543544 DOI: 10.1002/jso.24678]
- 62 Hao X, Luo H, Krawczyk M, Wei W, Wang W, Wang J, Flagg K, Hou J, Zhang H, Yi S, Jafari M, Lin D, Chung C, Caughey BA, Li G, Dhar D, Shi W, Zheng L, Hou R, Zhu J, Zhao L, Fu X, Zhang E, Zhang C, Zhu JK, Karin M, Xu RH, Zhang K. DNA methylation markers for diagnosis and prognosis of common cancers. Proc Natl Acad Sci USA 2017; 114: 7414-7419 [PMID: 28652331 DOI: 10.1073/pnas.1703577114
- Martinelli E, Ciardiello D, Martini G, Troiani T, Cardone C, Vitiello PP, Normanno N, Rachiglio 63 AM, Maiello E, Latiano T, De Vita F, Ciardiello F. Implementing anti-epidermal growth factor receptor (EGFR) therapy in metastatic colorectal cancer: challenges and future perspectives. Ann Oncol 2020; 31: 30-40 [PMID: 31912793 DOI: 10.1016/j.annonc.2019.10.007]
- Dercle L, Lu L, Schwartz LH, Qian M, Tejpar S, Eggleton P, Zhao B, Piessevaux H. Radiomics 64 Response Signature for Identification of Metastatic Colorectal Cancer Sensitive to Therapies Targeting EGFR Pathway. J Natl Cancer Inst 2020; 112: 902-912 [PMID: 32016387 DOI: 10.1093/jnci/djaa017]
- Spelt L, Nilsson J, Andersson R, Andersson B. Artificial neural networks-a method for prediction of 65 survival following liver resection for colorectal cancer metastases. Eur J Surg Oncol 2013; 39: 648-654 [PMID: 23514791 DOI: 10.1016/j.ejso.2013.02.024]
- 66 Paredes AZ, Hyer JM, Tsilimigras DI, Moro A, Bagante F, Guglielmi A, Ruzzenente A, Alexandrescu S, Makris EA, Poultsides GA, Sasaki K, Aucejo FN, Pawlik TM. A Novel Machine-Learning Approach to Predict Recurrence After Resection of Colorectal Liver Metastases. Ann Surg Oncol 2020; 27: 5139-5147 [PMID: 32779049 DOI: 10.1245/s10434-020-08991-9]
- Fong Y, Fortner J, Sun RL, Brennan MF, Blumgart LH. Clinical score for predicting recurrence after 67 hepatic resection for metastatic colorectal cancer: analysis of 1001 consecutive cases. Ann Surg 1999: **230**: 309-318; discussion 318-321 [PMID: 10493478 DOI: 10.1097/00000658-199909000-00004]



- Brudvik KW, Jones RP, Giuliante F, Shindoh J, Passot G, Chung MH, Song J, Li L, Dagenborg VJ, 68 Fretland ÅA, Røsok B, De Rose AM, Ardito F, Edwin B, Panettieri E, Larocca LM, Yamashita S, Conrad C, Aloia TA, Poston GJ, Bjørnbeth BA, Vauthey JN. RAS Mutation Clinical Risk Score to Predict Survival After Resection of Colorectal Liver Metastases. Ann Surg 2019; 269: 120-126 [PMID: 28549012 DOI: 10.1097/SLA.00000000002319]
- Qi ZH, Xu HX, Zhang SR, Xu JZ, Li S, Gao HL, Jin W, Wang WQ, Wu CT, Ni QX, Yu XJ, Liu L. 69 The Significance of Liquid Biopsy in Pancreatic Cancer. J Cancer 2018; 9: 3417-3426 [PMID: 30271504 DOI: 10.7150/jca.24591]
- Mason MC, Tzeng CD, Tran Cao HS, Aloia TA, Newhook TE, Overman MJ, Kopetz SE, Vauthey 70 JN, Chun YS. Preliminary Analysis of Liquid Biopsy after Hepatectomy for Colorectal Liver Metastases. J Am Coll Surg 2021; 233: 82-89.e1 [PMID: 33667566 DOI: 10.1016/j.jamcollsurg.2021.02.011]
- 71 Saini A, Pershad Y, Albadawi H, Kuo M, Alzubaidi S, Naidu S, Knuttinen MG, Oklu R. Liquid Biopsy in Gastrointestinal Cancers. Diagnostics (Basel) 2018; 8 [PMID: 30380690 DOI: 10.3390/diagnostics8040075]
- Rompianesi G, Di Martino M, Gordon-Weeks A, Montalti R, Troisi R. Liquid biopsy in 72 cholangiocarcinoma: Current status and future perspectives. World J Gastrointest Oncol 2021; 13: 332-350 [PMID: 34040697 DOI: 10.4251/wjgo.v13.i5.332]



WŨ

World Journal of Gastroenterology

Submit a Manuscript: https://www.f6publishing.com

World J Gastroenterol 2022 January 7; 28(1): 123-139

ISSN 1007-9327 (print) ISSN 2219-2840 (online)

DOI: 10.3748/wjg.v28.i1.123

ORIGINAL ARTICLE

Basic Study Focal adhesion kinase-related non-kinase ameliorates liver fibrosis by inhibiting aerobic glycolysis via the FAK/Ras/c-myc/ENO1 pathway

Tao Huang, Yuan-Qing-Xiao Li, Ming-Yu Zhou, Rui-Han Hu, Gao-Liang Zou, Jian-Chao Li, Shu Feng, Yong-Mei Liu, Chang-Qin Xin, Xue-Ke Zhao

ORCID number: Tao Huang 0000-0001-6849-5378; Yuan-Qing-Xiao Li 0000-0003-3189-1830; Ming-Yu Zhou 0000-0002-51305950; Rui-Han Hu 0000-0003-0668-2025; Gao-Liang Zou 0000-0002-9460-0802; Jian-Chao Li 0000-0003-2274-3369; Shu Feng 0000-0002-8615-1840; Yong-Mei Liu 0000-0002-0435-0409; Chang-Qin Xin 0000-0003-0635-1066; Xue-Ke Zhao 0000-0002-3032-4933.

Author contributions: Zhao XK designed the study; Huang T performed most of the experiments and wrote the article; all authors contributed to the design and interpretation of the study.

Institutional review board

statement: This study was reviewed and approved by the Institutional Review Board of the Affiliated Hospital of Guizhou Medical University (Approval 2018 Ethics Review No. 032).

Institutional animal care and use committee statement: Animal care and experimental procedures were authorized by the Animal Ethics Committee of Guizhou Medical University (No. 1801109).

Conflict-of-interest statement: The authors have no conflicts of

Tao Huang, Yuan-Qing-Xiao Li, Ming-Yu Zhou, Rui-Han Hu, Gao-Liang Zou, Jian-Chao Li, Shu Feng, Xue-Ke Zhao, Department of Infectious Diseases, Affiliated Hospital of Guizhou Medical University, Guizhou Medical University, Guiyang 550004, Guizhou Province, China

Yong-Mei Liu, Clinical Laboratory Center, Affiliated Hospital of Guizhou Medical University, Guiyang 550004, Guizhou Province, China

Chang-Qin Xin, Department of Infectious Diseases, People's Hospital of Weining Yi, Hui and Miao Autonomous County, Weining 553100, Guizhou Province, China

Corresponding author: Xue-Ke Zhao, PhD, Professor, Department of Infectious Diseases, Affiliated Hospital of Guizhou Medical University, No. 9 Beijing Road, Guiyang 550004, Guizhou Province, China. zhaoxueke1@163.com

Abstract

BACKGROUND

Hepatic stellate cell (HSC) hyperactivation is a central link in liver fibrosis development. HSCs perform aerobic glycolysis to provide energy for their activation. Focal adhesion kinase (FAK) promotes aerobic glycolysis in cancer cells or fibroblasts, while FAK-related non-kinase (FRNK) inhibits FAK phosphorylation and biological functions.

AIM

To elucidate the effect of FRNK on liver fibrosis at the level of aerobic glycolytic metabolism in HSCs.

METHODS

Mouse liver fibrosis models were established by administering CCl_{μ} and the effect of FRNK on the degree of liver fibrosis in the model was evaluated. Transforming growth factor-β1 was used to activate LX-2 cells. Tyrosine phosphorylation at position 397 (pY397-FAK) was detected to identify activated FAK, and the expression of the glycolysis-related proteins monocarboxylate transporter 1 (MCT-1) and enolase1 (ENO1) was assessed. Bioinformatics analysis was performed to predict putative binding sites for c-myc in the ENO1 promoter region, which were validated with chromatin immunoprecipitation (ChIP) and dual-



interest to declare.

Data sharing statement: No additional data are available.

ARRIVE guidelines statement: The authors have read the ARRIVE guidelines, and the manuscript was prepared and revised according to the ARRIVE guidelines.

Supported by the National Natural Science Foundation of China, No. 81860115, No. 82060116 and No. 81960118; and the Science and Technology Support Project of Guizhou Province, No. [2021] 094.

Country/Territory of origin: China

Specialty type: Gastroenterology and hepatology

Provenance and peer review:

Unsolicited article; Externally peer reviewed.

Peer-review model: Single blind

Peer-review report's scientific quality classification

Grade A (Excellent): 0 Grade B (Very good): B, B Grade C (Good): C Grade D (Fair): 0 Grade E (Poor): 0

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: htt p://creativecommons.org/License s/by-nc/4.0/

Received: July 14, 2021 Peer-review started: July 14, 2021 First decision: October 3, 2021 Revised: October 22, 2021 Accepted: December 22, 2021 Article in press: December 22, 2021 Published online: January 7, 2022

luciferase reporter assays.

RESULTS

The pY397-FAK level was increased in human fibrotic liver tissue. FRNK knockout promoted liver fibrosis in mouse models. It also increased the activation, migration, proliferation and aerobic glycolysis of primary hepatic stellate cells (pHSCs) but inhibited pHSC apoptosis. Nevertheless, opposite trends for these phenomena were observed after exogenous FRNK treatment in LX-2 cells. Mechanistically, the FAK/Ras/c-myc/ENO1 pathway promoted aerobic glycolysis, which was inhibited by exogenous FRNK.

CONCLUSION

FRNK inhibits aerobic glycolysis in HSCs by inhibiting the FAK/Ras/c-myc/ ENO1 pathway, thereby improving liver fibrosis. FRNK might be a potential target for liver fibrosis treatment.

Key Words: Liver fibrosis; Hepatic stellate cells; Focal adhesion kinase; Focal adhesion kinase-related non-kinase; Aerobic glycolysis; Enolase1

©The Author(s) 2022. Published by Baishideng Publishing Group Inc. All rights reserved.

Core Tip: We show that focal adhesion kinase-related non-kinase (FRNK) limits hepatic stellate cell (HSC) activation, proliferation, and migration and promotes HSC apoptosis by inhibiting aerobic glycolysis, thereby ameliorating liver fibrosis. FRNK may represent a potential therapeutic candidate for liver fibrosis treatment.

Citation: Huang T, Li YQX, Zhou MY, Hu RH, Zou GL, Li JC, Feng S, Liu YM, Xin CQ, Zhao XK. Focal adhesion kinase-related non-kinase ameliorates liver fibrosis by inhibiting aerobic glycolysis via the FAK/Ras/c-myc/ENO1 pathway. World J Gastroenterol 2022; 28(1): 123-139

URL: https://www.wjgnet.com/1007-9327/full/v28/i1/123.htm DOI: https://dx.doi.org/10.3748/wjg.v28.i1.123

INTRODUCTION

Long-term damage to liver function by hepatitis viruses, alcohol, and diet may cause chronic hepatic injuries leading to liver fibrosis and cirrhosis[1-4], which is characterized by the activation of hepatic stellate cells (HSCs) and their transformation into myofibroblasts^[5]. This process continuously damages the liver and disrupts the balance of liver self-repair, causing increased cell proliferation and migration and a reduced apoptosis rate[6-8]. At present, the pathological changes associated with chronic liver injuries in individuals without cirrhosis can be reversed after removing the etiological agent, such as in a small proportion of patients with hepatitis B and alcoholic fatty liver disease and most patients with hepatitis C[1,9], but the remaining patients with hepatic fibrosis develop irreversible cirrhosis due to an inability to completely and effectively reverse the pathogenesis and a lack of effective antifibrotic drugs[10]. Therefore, studies of the treatment of liver fibrosis are particularly critical, among which the regulation of the relevant biological functions of HSCs is the most important antifibrotic approach[11,12].

Focal adhesion kinases (FAKs) are a class of nonreceptor cytosolic protein tyrosine kinases that belong to the protein tyrosine kinase superfamily[13,14]. FAK plays an important role in cellular signal transduction and enhances biological behaviors such as proliferation, migration, wound healing and angiogenesis in cells and tissues after integrating signals from integrins, growth factors and mechanical stimuli[15,16]. FAK binds to extracellular matrix (ECM) proteins through an accumulation of integrin receptors to form FAK dimers, which further induce tyrosine phosphorylation at position 397 (pY397-FAK); pY397-FAK regulates these biological functions in cells[17, 18] and therefore plays an important role in a variety of malignant tumor cells[18,19]. FAK-related non-kinase (FRNK), which has a nucleotide sequence corresponding to



P-Reviewer: Morozov S, Truong NH S-Editor: Wang LL L-Editor: A P-Editor: Wang LL



the C-terminus of FAK but lacks the N-terminal functional site of FAK, is an independently expressed protein[20] with the main function of inhibiting FAK phosphorylation, thereby inversely regulating the function of FAK after cell activation [21,22]. FRNK negatively regulates FAK signaling axis function, thereby improving pulmonary fibrosis in an experimental mouse model[23].

FAK is also overexpressed in pancreatic ductal adenocarcinoma cells[24,25], promoting the conversion of pyruvate into lactate by increasing enolase1 (ENO1), pyruvate kinase 2, lactate dehydrogenase, and monocarboxylate transporter (MCT)-1 expression and lactate transport, enhancing aerobic glycolysis in cancer cells, and inhibiting mitochondrial oxidative phosphorylation in cancer cells[16]. This switch to aerobic glycolysis is an important mechanism by which tumor cells acquire energy [17, 26], as shown by the fact that oxidative phosphorylation simultaneously provides energy to cells performing aerobic glycolysis even in the presence of sufficient oxygen and normal mitochondrial function[27,28]. HSCs also exhibit increased aerobic glycolysis, resulting in lactate accumulation and gluconeogenesis inhibition when they differentiate into myofibroblasts[8,28]. This phenomenon also occurs in individuals with congenital pulmonary fibrosis[29]. Therefore, the inhibition of FAK-related pathways by FRNK may reduce energy acquisition through aerobic glycolysis during HSC activation and could be used as a targeted therapy to ameliorate liver fibrosis. Nevertheless, few studies have focused on the physiological or pathological role of FRNK in obtaining energy during hepatic fibrosis, and its mechanism remains unclear.

In the present study, we first showed that FRNK was downregulated in human liver fibrotic tissues. Then, we verified that FRNK knockout *in vivo* and *in vitro* promoted aerobic glycolysis and hepatic fibrosis. Exogenous FRNK inhibited aerobic glycolysis by inhibiting the FAK/Ras/c-myc/ENO1 pathway, limiting HSC activation, migration, and proliferation and increasing apoptosis to ameliorate liver fibrosis. Together, these data provide a detailed mechanism through which FRNK functions and suggest that FRNK represents a potential target to inhibit aerobic glycolysis in HSCs and treat liver fibrosis.

MATERIALS AND METHODS

Human liver samples

Paraffin blocks of liver tissues from 15 patients with liver fibrosis were collected from the Department of Infectious Diseases, Affiliated Hospital of Guizhou Medical University (Guiyang, China) between March 2019 and September 2019; none of the patients had any other organ-specific or systemic diseases, and liver fibrosis was diagnosed by pathological biopsy. Fifteen healthy liver samples were obtained from distal hepatocarcinoma liver tissue without any abnormalities in specimens surgically resected from patients at the Department of Hepatobiliary Surgery, Affiliated Hospital of Guizhou Medical University. None of the aforementioned subjects had contraindications to liver biopsy, and the study was approved by the Ethics Committee of the Affiliated Hospital of Guizhou Medical University (Approval 2018 Ethics Review No. 032) and conducted in accordance with the ethical guidelines of the 1975 Declaration of Helsinki. Written informed consent was obtained from the patients.

Animals

FRNK knockout (FRNK^{-/-}) mice were a gift from the Respiratory and Critical Care Medicine Center, School of Medicine, University of Alabama at Birmingham, Birmingham, AL, United States. All mouse interventions were approved by the Animal Care Committee (IACUC) of Guizhou Medical University (No. 1801109), and the methods and experimental procedures were performed in accordance with the relevant guidelines and regulations. Wild-type (WT) mice of the same genotype were used as controls, and all experimental mice were on the C57BL/6 background.

Mice were maintained under pathogen-free conditions at a controlled temperature (22 ± 2 °C) with a consistent photoperiod (12:12 h light-dark cycle); five mice were housed in each cage, with cages containing soft bedding. The mice were habituated to these conditions for 2 d before inclusion in an experiment. Healthy male mice (aged 8-11 wk, weighing 20 ± 3 g) were selected and intraperitoneally injected with 1.5 μ L/g of a 10% Carbon tetrachloride (CCl₄) in corn oil solution three times a week to establish a liver fibrosis model. Mice in the control group were injected with a 1.5 μ L/g solution of corn oil three times a week. Livers were harvested at each time point, namely, 0, 2, 4 and 6 wk, for experiments. Six mice per group were used.

Zaishidene® WJG | https://www.wjgnet.com

Reagents and antibodies

CCl₄, corn oil, and OptiPrep were purchased from Sigma-Aldrich (St. Louis, MO, United States). Transforming growth factor- β 1 (TGF- β 1) was purchased from R&D Systems (Minneapolis, MN, United States). Primary antibodies specific for the following proteins were purchased from Abcam (Cambridge, United Kingdom): Desmin rabbit monoclonal antibodies (ab32362), FAK rabbit monoclonal antibodies (ab40794), ENO1 mouse monoclonal antibodies (ab190365), alpha SMA rabbit polyclonal antibodies (ab5694), k-ras rabbit monoclonal antibodies (ab275876) and glyceraldehyde 3-phosphate dehydrogenase (GAPDH) rabbit polyclonal antibodies (ab9485). Anti-cmyc (13987) rabbit monoclonal antibodies were purchased from Cell Signaling Technology (Shanghai, China). MCT-1 rabbit polyclonal antibodies (20139-1-AP) were purchased from Proteintech (Wuhan, China). pY397-FAK rabbit polyclonal antibodies (AF3398) were purchased form Affinity Bioscience (Cincinnati, OH, United States). All other chemicals and reagents were purchased from Sigma-Aldrich (St. Louis, MO, United States) and Fisher Scientific (Waltham, MA, United States).

Immunohistochemistry (IHC), hematoxylin & eosin (H&E), Masson's trichrome and Sirius Red staining and hydroxyproline assay

H&E staining kits, Masson's trichrome staining solution and Sirius Red staining solution were purchased from Solarbio Biotechnology Co., Ltd. (Beijing, China) and used according to the manufacturer's guidelines. A hydroxyproline assay was performed using a Nanjing Jiancheng Biotechnology (Nanjing, China) hydroxyproline kit. All kits were used according to the instructions for use. Liver samples were fixed with neutral buffered formalin and embedded in paraffin for IHC. Briefly, sections were incubated with the indicated antibodies. Horseradish peroxidase-conjugated antibodies were used as the secondary antibodies. Finally, a diaminobenzidine colorimetric reagent solution was applied, followed by hematoxylin counterstaining. The slides were then scanned, and representative images were acquired.

Cells and cell culture

LX-2 cells were purchased from Zhongqiao Xinzhou (Shanghai, China). Human HSCs (ZQ0026) and LX-2 cells were cultured in Dulbecco's modified Eagle's medium (DMEM) containing 10% fetal bovine serum (FBS; Biological Industries, Kibbutz Beit-Haemek, Israel). Primary hepatic stellate cells (pHSCs) were extracted from C57BL/6 WT or FRNK^{-/-}mice aged 8-11 wk, as previously described[30,31]. Briefly, the abdominal cavity was opened with a "cross" incision, an 18-gauge trocar was inserted from the left ventricle to inject the preperfusate, and the blood in the liver was flushed by exsanguination until the tissue turned yellow. Then, the preperfusate was replaced with pronase and collagenase for 15-20 min, and the liver was removed and washed with normal saline. The liver capsule and connective tissue were removed, fully digested with a digestion solution at 37 °C with shaking and ground to generate a single-cell suspension. The supernatant was discarded after centrifugation at 1500 rpm for 5 min, and the pellet was resuspended in D-Hank's solution. The hepatocytes were removed by centrifugation, and a gradient lymphocyte separation solution was directly added. HSCs were isolated in one step using monolayer gradient centrifugation, and the cells were washed twice with DMEM and cultured with DMEM containing 10% FBS. Cell survival was evaluated by performing trypan blue staining, and cell purity was identified by desmin immunocytochemical staining. All cells were cultured in an incubator containing 5% CO₂ at 37 °C.

Recombinant FRNK adenoviral vector transfection and HSC activation

An adenovirus-mediated gene delivery system was used to effectively deliver the FRNK cDNA into HSCs. An adenoviral vector carrying the FRNK protein and green fluorescent protein (Ad-FRNK) as well as a green fluorescent protein-carrying adenovirus (Ad-GFP) were purchased from Jikai Gene (Shanghai, China). All transfections were performed according to the manufacturer's instructions, and cells in serum-free medium (DMEM with 1% BSA) were transfected with Ad-FRNK or the control vector (Ad-GFP) 24 h before TGF-B1 treatment. Twenty-four hours later, the cells were cultured with complete medium containing 2 ng/mL TGF-B1 and treated for 36 h[31].

Transwell, cell counting kit-8 (CCK-8) and flow cytometry assays

Transwell migration experiments used 8.0-µm pore size membranes (Corning, United States) according to the manufacturer's protocol. A total of 10⁵ cells were seeded in the upper chamber of each well in 100 μ L of serum-free medium, while 600 μ L of complete



medium was added to the lower chamber as a chemoattractant. After a 6-h incubation at 37 °C, the cells remaining on the upper surface of the membrane were removed with a cotton swab, and the cells on the lower surface of the membrane were considered migrated cells. After fixation with 4% paraformaldehyde and staining with a 0.1% crystal violet solution, images were acquired under an inverted microscope. Cell Counting Kit-8 (CCK-8) was purchased from Dojindo (Shanghai, China), and 10⁴ cells (100 μ L/well) were seeded in 96-well plates. After placing the culture plate in an incubator for preincubation (37 °C, 5% CO₂), 10 µL of CCK-8 solution was added to each well, and then the culture plate was evaluated with a microplate reader to detect the absorbance at 450 nm. A total of 10⁵ cells in each group were stained with an Annexin V-PE/7-AAD apoptosis kit (Hangzhou Lianke, Hangzhou, Zhejiang Province, China) according to the instructions for use, sorted with a flow cytometer (Beckman, United States) and analyzed using Flow Jo software (Tree Star); dead cells were excluded based on forward scatter and side scatter data.

Western blot analysis

Western blot analysis was performed as previously described [30]. Briefly, 1% NP-40treated whole-liver tissue lysates or whole-cell lysates were used for Western blot analysis. Protein levels were quantified using a BCA protein assay kit (Thermo Fisher Scientific, Waltham, MA) after total protein extraction. Twenty milligrams of each protein sample was separated by sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE). GAPDH was used as a loading control for all blots. Proteins were transferred to polyvinylidene difluoride (PVDF) membranes, which were incubated with primary antibodies overnight at 4 °C. The next day, after an incubation with an appropriate secondary antibody, signals were generated with an electrochemiluminescence detection kit.

Glucose consumption, 2-NBDG uptake and lactate assays

The lactate level in culture medium was detected with the Lactate Colorimetric Assay Kit (BioVision, Milpitas, CA, United States) according to the manufacturer's instructions. The 2-NBDG Glucose Uptake Kit (BioVision, Milpitas, CA, United States) was used to detect the cellular uptake of glucose, and the Glucose Colorimetric Assay Kit (BioVision, Milpitas, CA, United States) was used to detect the glucose concentration in culture medium and thus measure the cellular consumption of glucose. The 2-NBDG Glucose Uptake Kit and the Glucose Colorimetric Assay Kit were used according to the manufacturer's protocol.

Chromatin immunoprecipitation (ChIP) assay

JASPAR (http://jaspar.genereg.net) and PROMO (http://alggen.lsi.upc.es) database analyses predicted two putative c-myc binding sites in the ENO1 promoter region. A total of 10^7 cells fixed with formaldehyde were collected in 500 µL of lysis buffer from the Magna ChIP HiSens Kit (Millipore, Bedford, Massachusetts, United States) according to the manufacturer's manual. Cells were then sonicated for 25 cycles with a 6-s power-on interval of 30 s and an intensity of 200 W. Afterward, the supernatant was diluted and thoroughly mixed with Protein A/G beads. Then, $5 \mu g$ of IgG or an anti-c-myc antibody was added and incubated with the mixture overnight at 4 °C. After washing the beads the next day, the mixture was incubated with elution buffer at 62 °C for 2 h and then 95 °C for 10 min. The eluted DNA was then purified and subjected to a PCR assay to assess the binding sequence. Specific primer sequences were used to perform PCR.

Dual-luciferase reporter assays

The effect of c-myc on the ENO1 promoter was determined by cotransfecting pcDNAc-myc or pcDNA-vector (NC) into LX-2 cells with pGL3-based constructs containing an empty sequence (NC) or the WT or MT1/MT2 ENO1 promoter sequences, and Renilla luciferase reporter plasmids. Twenty-four hours after transfection, firefly and Renilla luciferase activities were measured with a luciferase reporter assay kit (Genomeditech, Shanghai, China). Fluorescence detection was performed according to the instructions of the instrument, the parameters were set, the measurement time was 10 s, and the measurement interval was 2 s. Each sample was added into a measuring tube in a total volume of 20 µL (the sample volume was consistent in each measurement), and then 20 µL of Firefly Luciferase Assay Reagent was added, mixed well 2-3 times (without vortexing), mixed well again and evaluated to determine relative light unit (RLU) 1. Cell lysis buffer was set as the blank control well. The tested samples were mixed with 20 µL of prepared Renilla Luciferase Assay working solution



2-3 times and mixed well before measuring RLU2. The measured RLU1 value was compared to the corresponding RLU2 value, and the resulting ratio determined the degree of reporter activation. The ratio of firefly luciferase activity to Renilla luciferase activity was calculated for each sample.

Statistical analysis

Data were statistically analyzed using GraphPad Prism 5.0 software, and a two-tailed Student's *t* test was used for comparisons between different groups. P < 0.05 was considered statistically significant. Data are presented as the mean ± SD.

RESULTS

The level of the pY397-FAK protein was increased while the level of the FRNK protein was decreased in human fibrotic liver tissue

We first investigated the level of pY397-FAK in fibrotic liver tissue to explore the role of pY397-FAK in liver fibrosis. Compared with normal liver tissue, liver tissue samples from patients in the liver fibrosis group showed typical pathological features, including significant steatosis, inflammatory necrosis, significant collagen deposition, hepatic fibrosis and hepatocyte loosening. Masson's trichrome staining showed less collagen deposition and a normal cell morphology in the normal group, while a large number of blue-stained collagen fibers was observed in the liver fibrosis group, and the tissue had accumulated a wide band of collagen fibers that extended into and was distributed in the hepatic lobules. Sirius Red staining showed less collagen deposition in normal subjects and a normal cell morphology but substantial red staining indicating collagen deposition in portal areas in fibrotic liver tissues. Notably, IHC showed higher α -smooth muscle actin (α -SMA) and pY397-FAK expression in fibrotic liver tissue than in normal liver tissues (Figure 1A and B). Western blot analysis showed higher levels of the pY397-FAK protein in fibrotic liver tissues than in normal liver tissues. Conversely, in fibrotic liver tissue, FRNK was expressed at lower levels than that in normal tissues (P < 0.05, Figure 1C and D). These results suggest that pY397-FAK protein expression is increased and FRNK protein expression is decreased in fibrotic liver tissue.

Exacerbation of liver fibrosis and aerobic glycolysis in mice after FRNK knockout in vivo

We established a fibrosis model by injecting CCl₄ into WT mice and FRNK^{-/-} mice. After two fortnights, the expression of the pY397-FAK protein peaked, while the FRNK protein was expressed at a low level (P < 0.05, Figure 2A and B). Therefore, mouse models with four weeks of injection were used in subsequent experiments. By performing H&E, Masson's trichrome and Sirius Red staining, we found a greater liver fibrosis area and more extensive liver fibrosis in FRNK^{-/-} mice than in WT mice after the CCl₄ intervention (P < 0.05, Figure 2C and D), while the hydroxyproline content in FRNK^{-/-} mice with fibrosis was greater than that in WT mice with fibrosis (P < 0.05, Figure 2E). Western blot analysis revealed higher levels of the pY397-FAK, MCT-1, ENO1 and α-SMA proteins in the liver tissues from FRNK^{-/-} mice treated with CCl₄ than in WT mice (P < 0.05, Figure 2F and G). Based on these results, FRNK^{-/-}mice develop more severe liver fibrosis after the CCl₄ intervention, along with increased expression of the aerobic glycolysis-related proteins MCT-1 and ENO1. It suggests that there may be more active aerobic glycolysis in the liver.

FRNK knockout promotes liver fibrosis and aerobic glycolysis in vitro

We extracted pHSCs from WT mice and FRNK-/-mice for in vitro experiments (Supplementary Figure 1). After 36 h of TGF- β 1 treatment, the migration of pHSCs from the FRNK^{-/-} groups in a Transwell chamber was increased (P < 0.05, Figure 3A). As indicated by the level of cell proliferation, pHSCs from the FRNK^{-/-} group exhibited increased cellular activity (P < 0.05, Figure 3B). After adding TGF- β 1 to pHSCs from FRNK^{-/-}mice for 36 h, the apoptosis rate was lower than that of pHSCs from the control mice (P < 0.05, Figure 3C). Moreover, pHSCs from FRNK^{-/-}mice showed increased glucose uptake and consumption compared with control pHSCs. Additionally, the lactate level in the medium of pHSCs from FRNK^{-/-}mice was increased compared with the lactate level in the medium of pHSCs from the control group (P <0.05, Figure 3D and E). Western blot analysis showed higher levels of the MCT-1, ENO1 and α -SMA proteins in the pHSCs from FRNK^{-/-}mice was higher than in the



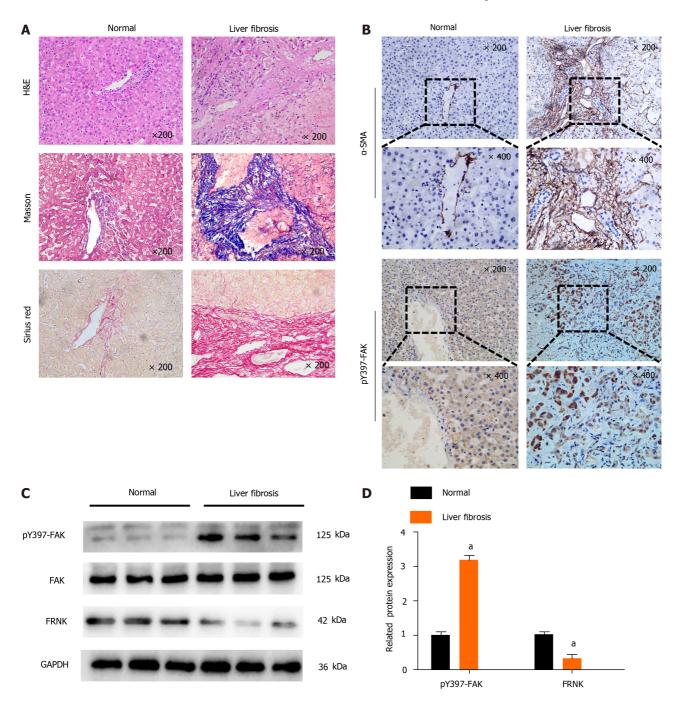


Figure 1 Tyrosine phosphorylation at position 397 of FAK is upregulated, while FRNK expression is downregulated in human fibrotic liver tissue. A: H&E, Masson's trichrome and Sirius Red staining were performed after liver biopsy to assess the tissues of normal subjects and patients with liver fibrosis under a light microscope at 200× magnification; B: Immunohistochemistry showed changes in the expression of α-smooth muscle actin (α-SMA) and tyrosine phosphorylation at position 397 of FAK(pY397-FAK) in the livers of normal subjects compared with patients with liver fibrosis under a light microscope at 200× or 400× magnification; C and D: Protein expression in biopsy tissues was analyzed using Western blotting. Representative results from three independent replicate assays are shown. aP < 0.05. Data are presented as the mean ± SD. FAK: Focal adhesion kinase; FRNK: Focal adhesion kinase-related non-kinase; GAPDH: Glyceraldehyde 3-phosphate dehydrogenase.

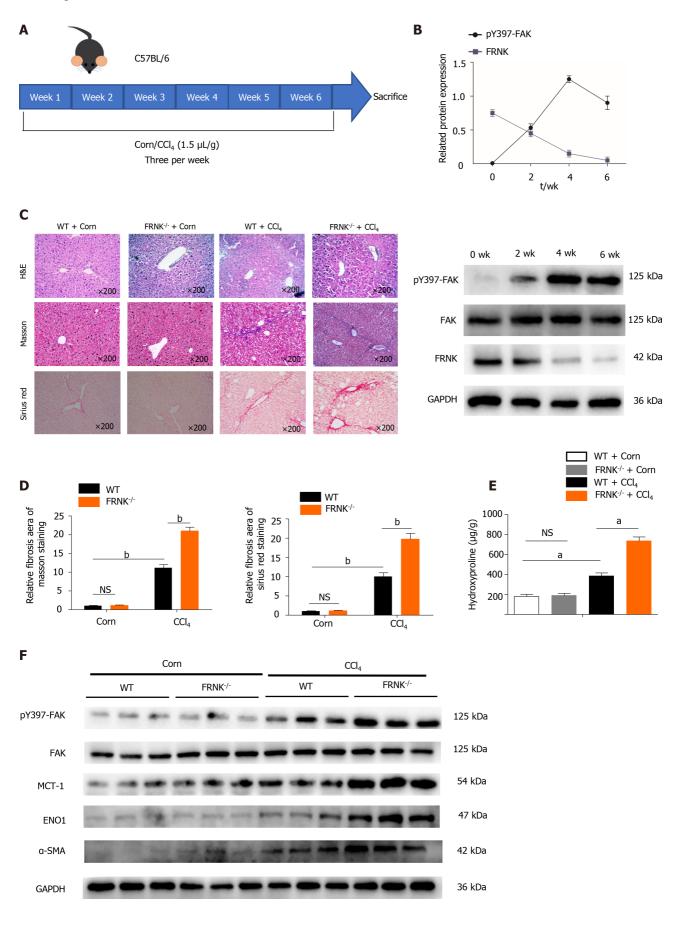
pHSCs from WT mice (*P* < 0.05, Figure 3F and G). The above results illustrate that FRNK knockout in mice increases the activation, migration, and proliferation of pHSCs and attenuates pHSC apoptosis while enhancing their aerobic glycolytic capacity in vitro.

Exogenous FRNK ameliorates experimental liver fibrosis and aerobic glycolysis in vitro

We transfected LX-2 cells with an adenovirus containing FRNK, induced the expression of the exogenous FRNK gene and incubated the cells with TGF-B1 for 36 h. We then performed Transwell, CCK-8 and flow cytometry assays with the transfected



Tao Huang et al. FRNK ameliorates liver fibrosis





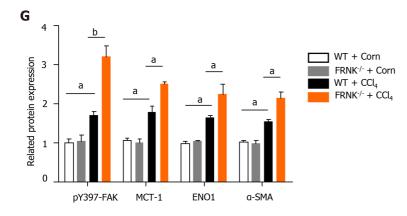


Figure 2 Liver fibrosis in mice was aggravated after FRNK knockout. A and B: WT mice were modeled for 6 wk, pY397-FAK and FRNK protein expression levels *in vivo* was measured using Western blotting every fortnight; C and D: FRNK⁺ and WT mice were used to establish a liver fibrosis model by administering CCl₄ (1.5 μ L/g), and liver tissues from these mice were stained with H&E, Masson's trichrome, and Sirius Red after 4 wk and observed under a light microscope × 200 magnification. The relative fibrotic areas were analyzed; E: The hydroxyproline content in liver tissues from the liver fibrosis model was also measured; F and G: Western blotting was used to detect the relative expression of proteins in the liver fibrosis model established with FRNK⁺ mice and WT mice. Representative results from three independent replicate assays are shown (*n* = 6). ^a*P* < 0.05 and ^b*P* < 0.01. Data are presented as the mean ± SD. MCT-1: Monocarboxylate transporter-1; ENO1: Enolase1.

cells to evaluate migration, proliferation and apoptosis. The migration of LX-2 cells was inhibited and apoptosis was increased after the introduction of exogenous FRNK compared to the control treatment (P < 0.05, Figure 4A and C). In addition, proliferation was also inhibited (P < 0.01, Figure 4B). Based on these results, exogenous FRNK inhibits cell migration and proliferation and promotes apoptosis. The abilities of cells in the Ad-FRNK group to take up and consume glucose were reduced, and the lactate level in the cell culture medium was reduced (P < 0.05, Figure 4D and E). Subsequently, cellular proteins were extracted, and the relative levels of intracellular pY397-FAK, MCT-1, ENO1 and α -SMA proteins were detected by Western blotting. The relative expression of the aforementioned proteins in the Ad-FRNK group was lower than that in the control group (P < 0.05, Figure 4F and G). Thus, the introduction of exogenous FRNK into HSCs inhibits cell proliferation and migration and promotes apoptosis. It also inhibits cellular aerobic glycolysis and thus inhibits cellular energy generation *in vitro*.

FRNK does not directly target ENO1

To explore the precise molecular mechanism by which FRNK regulates the ENO1 protein, TGF-β1 was used to stimulate LX-2 cells to activate FAK. The level of pY397-FAK was increased after stimulation with TGF- β 1. The expression of the K-Ras, c-myc and ENO1 proteins downstream of FAK was also examined (P < 0.05, Figure 5A and B). While examining whether FRNK directly inhibits ENO1 protein expression, increased ENO1 protein expression was observed after introducing exogenous c-myc into LX-2 cells, but ENO1 protein expression was not reduced after the continued introduction of exogenous FRNK (P < 0.05, Figure 5C and D), suggesting that exogenous FRNK does not directly inhibit ENO1 protein expression to exert its biological function. As a method to investigate whether c-myc directly regulates ENO1, a bioinformatics analysis of the ENO1 promoter was performed to predict the putative binding site for c-myc in the ENO1 promoter, followed by ChIP and dualluciferase reporter assays to verify that c-myc transcriptionally activates the ENO1 promoter (P < 0.05, Figure 5E and F). The results suggested that TGF- β 1 activates FAK by inducing FAK phosphorylation at position 397. Then, pY397-FAK increased the expression of downstream K-Ras and c-myc proteins, followed by transcriptional activation of ENO1 expression by c-myc, promoting aerobic glycolysis and activation in HSCs. pY397-FAK and its biological functions were inhibited by the introduction of exogenous FRNK, limiting aerobic glycolysis and activation in HSCs and ameliorating liver fibrosis (Figure 6).

DISCUSSION

The imbalance between liver injury and self-repair is the key to the development of liver fibrosis, and restoring the balance from an imbalanced state is a potential



Tao Huang et al. FRNK ameliorates liver fibrosis

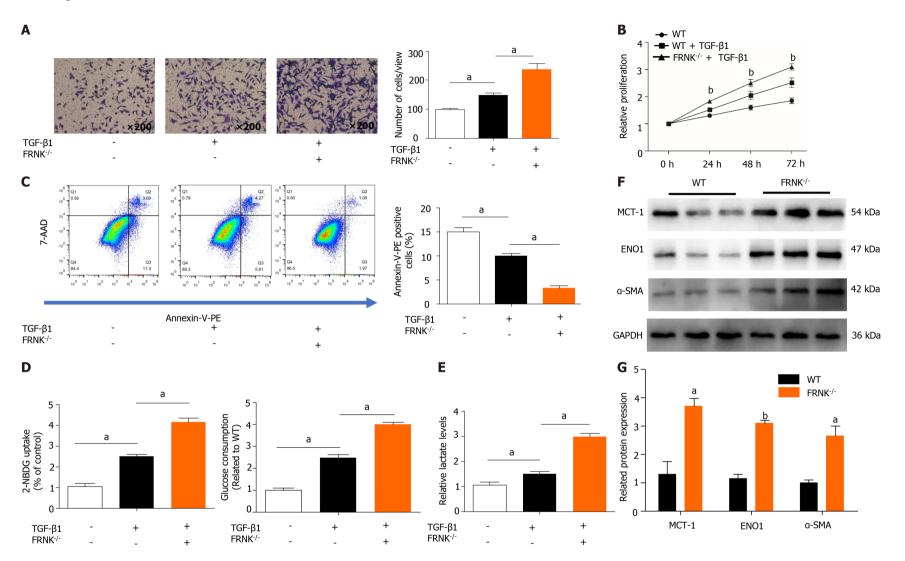


Figure 3 Knockout of FRNK promotes liver fibrosis and aerobic glycolysis *in vitro*. A: After 36 h of culture with TGF- β 1 (2 ng/mL), the migratory ability of primary hepatic stellate cells(pHSCs) was measured under a light microscope at × 200 magnification (10⁵ cells per well); B: The proliferation of pHSCs was assessed with a CCK-8 assay; C: The apoptosis of pHSCs was analyzed using flow cytometry after 36 h of intervention; D and E: pHSCs cultured under the same intervention conditions were examined for glucose uptake and consumption, and lactate levels in the cell culture medium were also assessed; F and G: MCT-1, ENO1 and α -SMA levels in pHSCs were assessed using Western blotting. Representative results from three independent replicate assays are shown. ^aP < 0.05 and ^bP < 0.01 Results are presented as the mean ± SD.

Tao Huang et al. FRNK ameliorates liver fibrosis

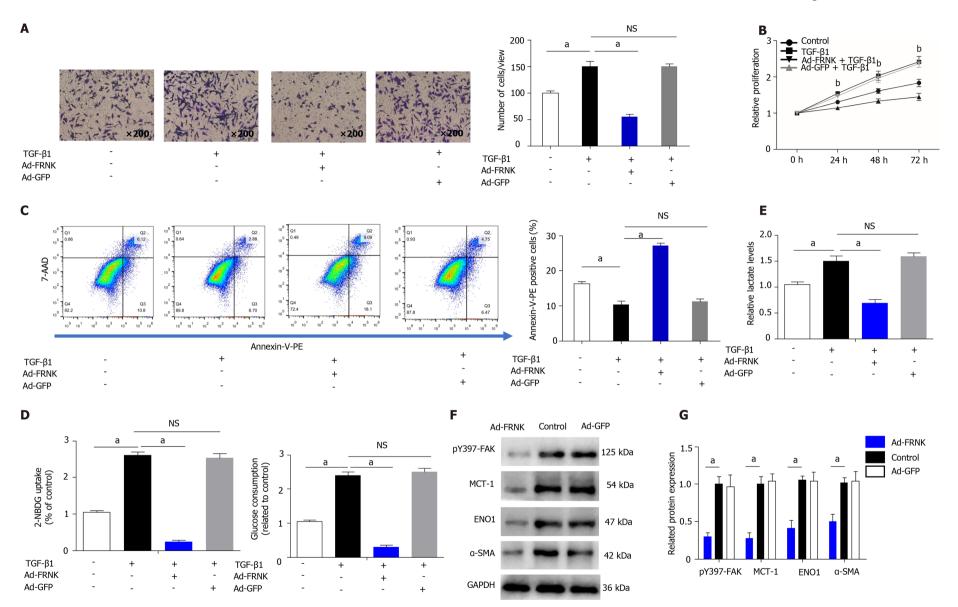


Figure 4 Exogenous FRNK ameliorates experimental liver fibrosis and aerobic glycolysis *in vitro*. LX-2 cells were transfected with a green fluorescent protein-carrying adenoviral vector (Ad-GFP) also encoding the FRNK gene. A: Cultured with TGF-β1 (2 ng/mL) for 36 h (10⁵ cells per well). Migration was measured by analyzing cells under a light microscope × 200 magnification; B: The proliferation of LX-2 cells cultured under the intervention conditions is presented; C: The

133

apoptosis of LX-2 cells was analyzed after 36 h of intervention using flow cytometry; D and E: LX-2 cells cultured under the same intervention conditions were examined for glucose consumption and uptake abilities, and lactate levels in the cell culture medium were also assessed; F and G: Levels of pY397-FAK, MCT-1, ENO1 and α -SMA in LX-2 cells were detected using Western blotting. Representative results from three independent replicate assays are shown . ^aP < 0.05 and ^bP < 0.01. Results are presented as the mean ± SD.

treatment for liver diseases. In the present study, we verified that FRNK alters the activation, proliferation, migration and apoptosis of HSCs by regulating aerobic glycolysis during liver fibrosis. FRNK inhibits aerobic glycolysis in HSCs by suppressing ENO1 activation through the FAK/Ras/c-myc/ENO1 pathway.

Early studies by our group verified that FAK plays important roles in the activation of HSCs and the development of liver fibrosis and that inhibition of FAK gene expression inhibits liver fibrogenesis[30]. Ding *et al*[23] verified that FRNK negatively regulates pulmonary fibrosis induced by FAK phosphorylation during pulmonary fibrosis. If FRNK inhibits the biological function of FAK in pulmonary fibrosis and uses a similar mechanism to repress liver fibrosis, it may represent a potential therapeutic target in liver fibrosis. Previous studies on FRNK have focused on the inhibition of the migratory function of vascular smooth muscle[32,33], combined with the presence of extracellular lactate accumulation during HSC activation[28,34] and FAK activation of aerobic glycolytic function in tumor cells[35-37]. FRNK may improve liver fibrosis by inhibiting aerobic glycolysis and inhibiting FAK activation in HSCs, but the mechanism by which FRNK exerts this effect remains unclear.

In the current study, we first observed increased expression of the pY397-FAK protein and decreased expression of the FRNK protein in tissues from patients with liver fibrosis (Figure 1). Subsequent experiments using CCl₄ to replicate liver fibrosis in a mouse model yielded the same results (Figure 2B). Therefore, we speculated that a correlation between the occurrence of liver fibrosis and the downregulation of FRNK expression may exist and subsequently performed experiments in WT mice and FRNK^{-/-}mice. After the CCl₄ intervention, the degree of liver fibrosis in WT mice was lower than that in $FRNK^{-/-}$ mice (Figure 2C and D). The expression of the aerobic glycolysis-related proteins MCT-1 and ENO1 in the liver tissue of FRNK-/- mice was increased (Figure 2F and G), suggesting that FRNK gene deletion may promote intrahepatic aerobic glycolysis and aggravate the occurrence and development of liver fibrosis. We extracted pHSCs from WT mice and FRNK^{-/-} mice for *in vitro* experiments to further explore the effect of FRNK on the biological functions of HSCs. After treatment with TGF- β 1, the biological functions of pHSCs from FRNK^{-/-}mice were more active than those of pHSCs from WT mice, as evidenced by the increased migration and proliferation and reduced apoptosis rate (Figure 3A-C). Furthermore, the uptake and consumption of glucose and extracellular lactate levels of FRNK^{-/-} pHSCs were increased (Figure 3D and E), suggesting that HSCs lacking FRNK exhibited more active aerobic glycolysis, which supplied energy for their biological functions, such as activation, proliferation and migration. On the other hand, we transfected LX-2 cells with an adenovirus carrying the FRNK gene to verify whether exogenous FRNK is a promising therapeutic target in liver fibrosis. After FRNK

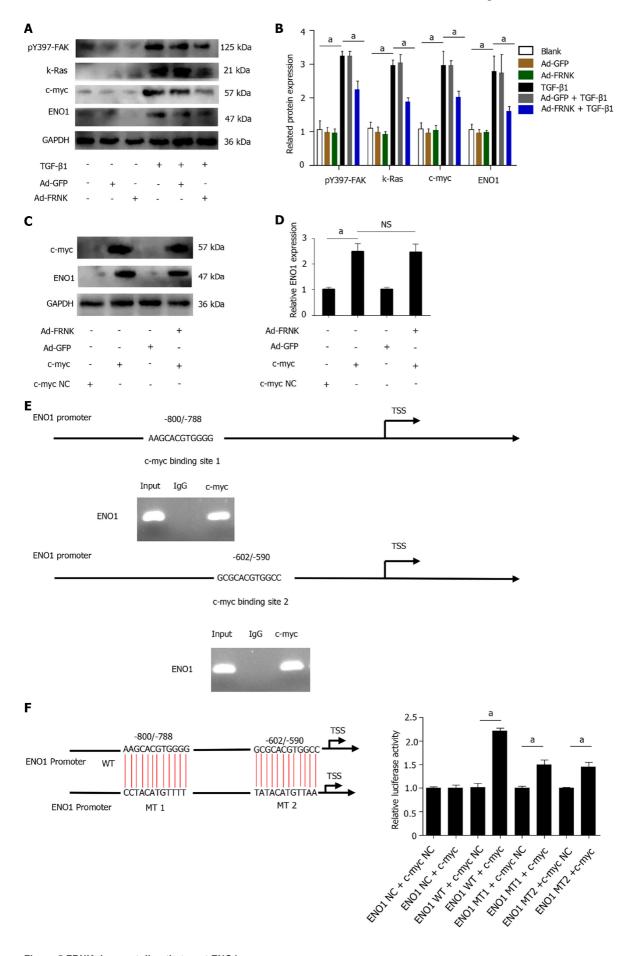


Figure 5 FRNK does not directly target ENO1. A and B: Proteins were extracted from LX-2 cells, and relative levels of the pY397-FAK, K-Ras, c-myc and

Raishideng® WJG | https://www.wjgnet.com

ENO1 proteins were determined; C and D: After transfection of LX-2 cells with pcDNA-c-myc or pcDNA-vector (NC), adenoviral vectors containing the FRNK gene (Ad-FRNK) or the negative control (Ad-GFP) were transfected, and the extracted protein was used to evaluate ENO1 expression through Western blotting; E: Schematic representation of the structure of the putative c-myc binding site in the human ENO1 promoter and chromatin immunoprecipitation (ChIP) assays with antic-myc or IgG; F: A dual-luciferase reporter assay showed the luciferase activity of WT, mutation (MT)1 and MT2 ENO1 promoters in LX-2 cells transfected with the cmyc or NC plasmid. Representative results from three independent replicate assays are shown. ^aP < 0.05 and ^bP < 0.01. Results are presented as the mean ± SD.

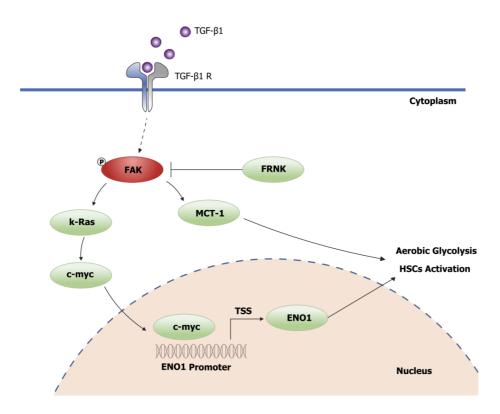


Figure 6 Schematic diagram of FRNK inhibition of the FAK/Ras/c-myc/ENO1 pathway to ameliorate liver fibrosis.

overexpression, cell proliferation and migration decreased, while the percentage of apoptotic cells increased (Figure 4A-C). The uptake and utilization of glucose and the extracellular lactate level in LX-2 cells were also decreased (Figure 4D and E), and pY397-FAK expression in these cells was decreased (Figure 4F and G). These results further indicated that increasing exogenous FRNK expression prevented HSCs from obtaining energy through aerobic glycolysis and reduced cell activation and the energy supply required for a series of biological functions after activation, thereby inhibiting liver fibrosis. While investigating the mechanism by which FRNK regulates aerobic glycolysis in HSCs, we found that FAK is phosphorylated in LX-2 cells stimulated with TGF- β 1 and that the downstream proteins K-Ras, c-myc, and ENO1 are activated. Following the introduction of FRNK, ENO1 protein expression was reduced (Figure 5A and B). We transfected both the c-myc and FRNK genes into LX-2 cells to determine whether FRNK directly inhibited ENO1 expression and found that ENO1 protein expression was not reduced upon increased exogenous FRNK expression (Figure 5C and D), thus suggesting that FRNK does not directly inhibit ENO1 protein expression. Therefore, we further hypothesized that c-myc directly alters ENO1 protein expression. Early studies revealed that c-myc is involved in the regulation of various biological functions, including metabolism, cell growth, cell cycle regulation and apoptosis[38,39], consistent with the results of our study. An increasing number of studies have shown that c-myc is involved in the regulation of promoters as a transcription factor. Hence, we predicted the putative c-myc binding site in the ENO1 promoter region by performing a bioinformatics analysis, followed by confirmation of our hypothesis that c-myc transcriptionally activates ENO1 and subsequently promotes liver fibrosis in HSCs by performing dual-luciferase reporter and ChIP assays (Figure 5E and F). Therefore, our study revealed that FRNK alleviated hepatic fibrosis via the FAK/Ras/c-myc/ENO1 pathway. The molecular mechanism by which FRNK regulates ENO1 and MCT-1 expression should be confirmed by conducting more complicated investigations in the future, and our group will be



dedicated to studying this pathway.

CONCLUSION

In conclusion, this study is the first to reveal the effect of FRNK on liver fibrosis at the metabolic level. The experimental results suggest that the FAK/FRNK genes are potentially useful therapeutic targets in liver fibrosis and provide some rationale for the development of related drugs in the future.

ARTICLE HIGHLIGHTS

Research background

Hepatic stellate cell (HSC) hyperactivation is a central link in liver fibrosis development. HSCs perform aerobic glycolysis to provide energy for their activation.

Research motivation

Focal adhesion kinase (FAK) promotes aerobic glycolysis in cancer cells or fibroblasts, while FAK-related non-kinase (FRNK) inhibits FAK phosphorylation and biological functions.

Research objectives

To elucidate the effect of FRNK on liver fibrosis at the level of aerobic glycolytic metabolism in HSCs.

Research methods

Mouse liver fibrosis models were established by administering CCl_4 and the effect of FRNK on the degree of liver fibrosis in the model was evaluated. Transforming growth factor- β 1 was used to activate LX-2 cells. Tyrosine phosphorylation at position 397 (pY397-FAK) was detected to identify activated FAK, and the expression of the glycolysis-related proteins monocarboxylate transporter 1 (MCT-1) and enolase1 (ENO1) was assessed. Bioinformatics analysis was performed to predict putative binding sites for c-myc in the ENO1 promoter region, which were validated with chromatin immunoprecipitation (ChIP) and dual-luciferase reporter assays.

Research results

The pY397-FAK level was increased in human fibrotic liver tissue. FRNK knockout promoted liver fibrosis in mouse models. It also increased the activation, migration, proliferation and aerobic glycolysis of primary hepatic stellate cells (pHSCs) but inhibited pHSC apoptosis. Nevertheless, opposite trends for these phenomena were observed after exogenous FRNK treatment in LX-2 cells. Mechanistically, the FAK/ Ras/c-myc/ENO1 pathway promoted aerobic glycolysis, which was inhibited by exogenous FRNK.

Research conclusions

FRNK inhibits aerobic glycolysis in HSCs by inhibiting the FAK/Ras/c-myc/ENO1 pathway, thereby improving liver fibrosis. FRNK might be a potential target for liver fibrosis treatment.

Research perspectives

The molecular mechanism by which FRNK regulates ENO1 and MCT-1 expression should be confirmed by conducting more complicated investigations in the future, and our group will be dedicated to studying this pathway.

ACKNOWLEDGEMENTS

The authors thank Professor Qiang Ding's laboratory at the University of Alabama at Birmingham, School of Medicine, Birmingham, AL, United States, for the gift of FRNK^{-/-} mice and providing experimental technical guidance.

REFERENCES

- Testino G, Leone S, Fagoonee S, Pellicano R. Alcoholic liver fibrosis: detection and treatment. 1 Minerva Med 2018; 109: 457-471 [PMID: 30221911 DOI: 10.23736/S0026-4806.18.05844-5]
- 2 Stål P. Liver fibrosis in non-alcoholic fatty liver disease - diagnostic challenge with prognostic significance. World J Gastroenterol 2015; 21: 11077-11087 [PMID: 26494963 DOI: 10.3748/wjg.v21.i39.11077
- 3 Sebastiani G, Gkouvatsos K, Pantopoulos K. Chronic hepatitis C and liver fibrosis. World J Gastroenterol 2014; 20: 11033-11053 [PMID: 25170193 DOI: 10.3748/wjg.v20.i32.11033]
- Seki E, Brenner DA. Recent advancement of molecular mechanisms of liver fibrosis. J Hepatobiliary 4 Pancreat Sci 2015; 22: 512-518 [PMID: 25869468 DOI: 10.1002/jhbp.245]
- Parola M, Pinzani M. Liver fibrosis: Pathophysiology, pathogenetic targets and clinical issues. Mol 5 Aspects Med 2019; 65: 37-55 [PMID: 30213667 DOI: 10.1016/j.mam.2018.09.002]
- 6 Bataller R, Brenner DA. Liver fibrosis. J Clin Invest 2005; 115: 209-218 [PMID: 15690074 DOI: 10.1172/JCI24282]
- 7 Campana L, Iredale JP. Regression of Liver Fibrosis. Semin Liver Dis 2017; 37: 1-10 [PMID: 28201843 DOI: 10.1055/s-0036-1597816]
- 8 Tsuchida T, Friedman SL. Mechanisms of hepatic stellate cell activation. Nat Rev Gastroenterol Hepatol 2017; 14: 397-411 [PMID: 28487545 DOI: 10.1038/nrgastro.2017.38]
- 9 Rocco A, Compare D, Angrisani D, Sanduzzi Zamparelli M, Nardone G. Alcoholic disease: liver and beyond. World J Gastroenterol 2014; 20: 14652-14659 [PMID: 25356028 DOI: 10.3748/wjg.v20.i40.14652]
- Schuppan D, Ashfaq-Khan M, Yang AT, Kim YO. Liver fibrosis: Direct antifibrotic agents and targeted therapies. Matrix Biol 2018; 68-69: 435-451 [PMID: 29656147 DOI: 10.1016/j.matbio.2018.04.006
- 11 Hernandez-Gea V, Friedman SL. Pathogenesis of liver fibrosis. Annu Rev Pathol 2011; 6: 425-456 [PMID: 21073339 DOI: 10.1146/annurev-pathol-011110-130246]
- 12 Dhar A, Sadiq F, Anstee QM, Levene AP, Goldin RD, Thursz MR. Thrombin and factor Xa link the coagulation system with liver fibrosis. BMC Gastroenterol 2018; 18: 60 [PMID: 29739329 DOI: 10.1186/s12876-018-0789-8]
- 13 Lai IR, Chu PY, Lin HS, Liou JY, Jan YJ, Lee JC, Shen TL. Phosphorylation of focal adhesion kinase at Tyr397 in gastric carcinomas and its clinical significance. Am J Pathol 2010; 177: 1629-1637 [PMID: 20724588 DOI: 10.2353/ajpath.2010.100172]
- 14 Tapial Martínez P, López Navajas P, Lietha D. FAK Structure and Regulation by Membrane Interactions and Force in Focal Adhesions. Biomolecules 2020; 10 [PMID: 31991559 DOI: 10.3390/biom10020179
- 15 Sakurai M, Ohtake J, Ishikawa T, Tanemura K, Hoshino Y, Arima T, Sato E. Distribution and Y397 phosphorylation of focal adhesion kinase on follicular development in the mouse ovary. Cell Tissue Res 2012; 347: 457-465 [PMID: 22322421 DOI: 10.1007/s00441-011-1307-2]
- 16 Mikuriya K, Kuramitsu Y, Ryozawa S, Fujimoto M, Mori S, Oka M, Hamano K, Okita K, Sakaida I, Nakamura K. Expression of glycolytic enzymes is increased in pancreatic cancerous tissues as evidenced by proteomic profiling by two-dimensional electrophoresis and liquid chromatographymass spectrometry/mass spectrometry. Int J Oncol 2007; 30: 849-855 [PMID: 17332923]
- 17 Li L, Liang Y, Kang L, Liu Y, Gao S, Chen S, Li Y, You W, Dong Q, Hong T, Yan Z, Jin S, Wang T, Zhao W, Mai H, Huang J, Han X, Ji Q, Song Q, Yang C, Zhao S, Xu X, Ye Q. Transcriptional Regulation of the Warburg Effect in Cancer by SIX1. Cancer Cell 2018; 33: 368-385.e7 [PMID: 29455928 DOI: 10.1016/j.ccell.2018.01.010]
- 18 Liberti MV, Locasale JW. The Warburg Effect: How Does it Benefit Cancer Cells? Trends Biochem Sci 2016; 41: 211-218 [PMID: 26778478 DOI: 10.1016/j.tibs.2015.12.001]
- 19 Sulzmaier FJ, Jean C, Schlaepfer DD. FAK in cancer: mechanistic findings and clinical applications. Nat Rev Cancer 2014; 14: 598-610 [PMID: 25098269 DOI: 10.1038/nrc3792]
- 20 Sayers RL, Sundberg-Smith LJ, Rojas M, Hayasaka H, Parsons JT, Mack CP, Taylor JM. FRNK expression promotes smooth muscle cell maturation during vascular development and after vascular injury. Arterioscler Thromb Vasc Biol 2008; 28: 2115-2122 [PMID: 18787183 DOI: 10.1161/ATVBAHA.108.175455]
- Heidkamp MC, Bayer AL, Kalina JA, Eble DM, Samarel AM. GFP-FRNK disrupts focal adhesions 21 and induces anoikis in neonatal rat ventricular myocytes. Circ Res 2002; 90: 1282-1289 [PMID: 12089066 DOI: 10.1161/01.res.0000023201.41774.ea]
- Hayasaka H, Martin KH, Hershey ED, Parsons JT. Disruption of FRNK expression by gene targeting 22 of the intronic promoter within the focal adhesion kinase gene. J Cell Biochem 2007; 102: 947-954 [PMID: 17440961 DOI: 10.1002/jcb.21329]
- 23 Ding Q, Cai GQ, Hu M, Yang Y, Zheng A, Tang Q, Gladson CL, Hayasaka H, Wu H, You Z, Southern BD, Grove LM, Rahaman SO, Fang H, Olman MA. FAK-related nonkinase is a multifunctional negative regulator of pulmonary fibrosis. Am J Pathol 2013; 182: 1572-1584 [PMID: 23499373 DOI: 10.1016/j.ajpath.2013.01.026]
- Jiang H, Liu X, Knolhoff BL, Hegde S, Lee KB, Jiang H, Fields RC, Pachter JA, Lim KH, DeNardo DG. Development of resistance to FAK inhibition in pancreatic cancer is linked to stromal depletion. Gut 2020; 69: 122-132 [PMID: 31076405 DOI: 10.1136/gutjnl-2018-317424]
- 25 Jiang H, Hegde S, Knolhoff BL, Zhu Y, Herndon JM, Meyer MA, Nywening TM, Hawkins WG,



Shapiro IM, Weaver DT, Pachter JA, Wang-Gillam A, DeNardo DG. Targeting focal adhesion kinase renders pancreatic cancers responsive to checkpoint immunotherapy. Nat Med 2016; 22: 851-860 [PMID: 27376576 DOI: 10.1038/nm.4123]

- 26 Yang J, Ren B, Yang G, Wang H, Chen G, You L, Zhang T, Zhao Y. The enhancement of glycolysis regulates pancreatic cancer metastasis. Cell Mol Life Sci 2020; 77: 305-321 [PMID: 31432232 DOI: 10.1007/s00018-019-03278-z]
- Vander Heiden MG, Cantley LC, Thompson CB. Understanding the Warburg effect: the metabolic 27 requirements of cell proliferation. Science 2009; 324: 1029-1033 [PMID: 19460998 DOI: 10.1126/science.1160809]
- 28 Chen Y, Choi SS, Michelotti GA, Chan IS, Swiderska-Syn M, Karaca GF, Xie G, Moylan CA, Garibaldi F, Premont R, Suliman HB, Piantadosi CA, Diehl AM. Hedgehog controls hepatic stellate cell fate by regulating metabolism. Gastroenterology 2012; 143: 1319-1329.e11 [PMID: 22885334 DOI: 10.1053/j.gastro.2012.07.115]
- 29 Chen Z, Liu M, Li L, Chen L. Involvement of the Warburg effect in non-tumor diseases processes. J Cell Physiol 2018; 233: 2839-2849 [PMID: 28488732 DOI: 10.1002/jcp.25998]
- 30 Zhao XK, Yu L, Cheng ML, Che P, Lu YY, Zhang Q, Mu M, Li H, Zhu LL, Zhu JJ, Hu M, Li P, Liang YD, Luo XH, Cheng YJ, Xu ZX, Ding Q. Focal Adhesion Kinase Regulates Hepatic Stellate Cell Activation and Liver Fibrosis. Sci Rep 2017; 7: 4032 [PMID: 28642549 DOI: 10.1038/s41598-017-04317-0]
- Zou GL, Zuo S, Lu S, Hu RH, Lu YY, Yang J, Deng KS, Wu YT, Mu M, Zhu JJ, Zeng JZ, Zhang 31 BF, Wu X, Zhao XK, Li HY. Bone morphogenetic protein-7 represses hepatic stellate cell activation and liver fibrosis via regulation of TGF-β/Smad signaling pathway. World J Gastroenterol 2019; 25: 4222-4234 [PMID: 31435175 DOI: 10.3748/wjg.v25.i30.4222]
- Koshman YE, Engman SJ, Kim T, Iyengar R, Henderson KK, Samarel AM. Role of FRNK tyrosine 32 phosphorylation in vascular smooth muscle spreading and migration. Cardiovasc Res 2010; 85: 571-581 [PMID: 19793767 DOI: 10.1093/cvr/cvp322]
- Koshman YE, Kim T, Chu M, Engman SJ, Iyengar R, Robia SL, Samarel AM. FRNK inhibition of 33 focal adhesion kinase-dependent signaling and migration in vascular smooth muscle cells. Arterioscler Thromb Vasc Biol 2010; 30: 2226-2233 [PMID: 20705914 DOI: 10.1161/ATVBAHA.110.212761
- Wang YH, Israelsen WJ, Lee D, Yu VWC, Jeanson NT, Clish CB, Cantley LC, Vander Heiden MG, 34 Scadden DT. Cell-state-specific metabolic dependency in hematopoiesis and leukemogenesis. Cell 2014; 158: 1309-1323 [PMID: 25215489 DOI: 10.1016/j.cell.2014.07.048]
- 35 Demircioglu F, Wang J, Candido J, Costa ASH, Casado P, de Luxan Delgado B, Reynolds LE, Gomez-Escudero J, Newport E, Rajeeve V, Baker AM, Roy-Luzarraga M, Graham TA, Foster J, Wang Y, Campbell JJ, Singh R, Zhang P, Schall TJ, Balkwill FR, Sosabowski J, Cutillas PR, Frezza C, Sancho P, Hodivala-Dilke K. Cancer associated fibroblast FAK regulates malignant cell metabolism. Nat Commun 2020; 11: 1290 [PMID: 32157087 DOI: 10.1038/s41467-020-15104-3]
- Cao D, Qi Z, Pang Y, Li H, Xie H, Wu J, Huang Y, Zhu Y, Shen Y, Dai B, Hu X, Ye D, Wang Z. 36 Retinoic Acid-Related Orphan Receptor C Regulates Proliferation, Glycolysis, and Chemoresistance via the PD-L1/ITGB6/STAT3 Signaling Axis in Bladder Cancer. Cancer Res 2019; 79: 2604-2618 [PMID: 30808674 DOI: 10.1158/0008-5472.CAN-18-3842]
- 37 Siu MKY, Jiang YX, Wang JJ, Leung THY, Han CY, Tsang BK, Cheung ANY, Ngan HYS, Chan KKL. Hexokinase 2 Regulates Ovarian Cancer Cell Migration, Invasion and Stemness via FAK/ERK1/2/MMP9/NANOG/SOX9 Signaling Cascades. Cancers (Basel) 2019; 11 [PMID: 31212816 DOI: 10.3390/cancers11060813]
- Luo W, Chen J, Li L, Ren X, Cheng T, Lu S, Lawal RA, Nie Q, Zhang X, Hanotte O. c-Myc inhibits 38 myoblast differentiation and promotes myoblast proliferation and muscle fibre hypertrophy by regulating the expression of its target genes, miRNAs and lincRNAs. Cell Death Differ 2019; 26: 426-442 [PMID: 29786076 DOI: 10.1038/s41418-018-0129-0]
- Xiao ZD, Han L, Lee H, Zhuang L, Zhang Y, Baddour J, Nagrath D, Wood CG, Gu J, Wu X, Liang H, Gan B. Energy stress-induced lncRNA FILNC1 represses c-Myc-mediated energy metabolism and inhibits renal tumor development. Nat Commun 2017; 8: 783 [PMID: 28978906 DOI: 10.1038/s41467-017-00902-z]



 \mathcal{N}

World Journal of Gastroenterology

Submit a Manuscript: https://www.f6publishing.com

World J Gastroenterol 2022 January 7; 28(1): 140-153

ISSN 1007-9327 (print) ISSN 2219-2840 (online)

DOI: 10.3748/wjg.v28.i1.140

ORIGINAL ARTICLE

Retrospective Cohort Study

Dynamics of cytokines predicts risk of hepatocellular carcinoma among chronic hepatitis C patients after viral eradication

Ming-Ying Lu, Ming-Lun Yeh, Ching-I Huang, Shu-Chi Wang, Yi-Shan Tsai, Pei-Chien Tsai, Yu-Min Ko, Ching-Chih Lin, Kuan-Yu Chen, Yu-Ju Wei, Po-Yao Hsu, Cheng-Ting Hsu, Tyng-Yuan Jang, Ta-Wei Liu, Po-Cheng Liang, Ming-Yen Hsieh, Zu-Yau Lin, Shinn-Cherng Chen, Chung-Feng Huang, Jee-Fu Huang, Chia-Yen Dai, Wan-Long Chuang, Ming-Lung Yu

ORCID number: Ming-Ying Lu 0000-0002-1317-9586; Ming-Lun Yeh 0000-0003-3728-7618; Ching-I Huang 0000-0002-5467-0398; Shu-Chi Wang 0000-0001-9691-7507; Yi-Shan Tsai 0000-0002-5251-1318: Pei-Chien Tsai 0000-0002-5044-6727; Yu-Min Ko 0000-0002-8424-9289; Ching-Chih Lin 0000-0002-4829-308X; Kuan-Yu Chen 0000-0001-9350-2699; Yu-Ju Wei 0000-0003-1266-7796; Po-Yao Hsu 0000-0002-5443-7203; Cheng-Ting Hsu 0000-0002-9057-3536; Tyng-Yuan Jang 0000-0003-2961-130X; Ta-Wei Liu 0000-0002-6978-9922; Po-Cheng Liang 0000-0001-9189-6604; Ming-Yen Hsieh 0000-0002-8019-3011; Zu-Yau Lin 0000-0002-8489-7147; Shinn-Cherng Chen 0000-0002-5925-4078; Chung-Feng Huang 0000-0002-3367-068X; Jee-Fu Huang 0000-0002-2752-7051; Chia-Yen Dai 0000-0003-2296-3054; Wan-Long Chuang 0000-0002-2376-421X; Ming-Lung Yu 0000-0001-8145-1900.

Author contributions: Lu MY

analyzed the data and wrote the manuscript; Tsai PC confirmed the statistical analysis; Wang SC, Tsai YS, Ko YM, Lin CC, and Chen KY performed the experiments; Yeh ML, Huang CI, Wei YJ, Hsu PY, Hsu CT, Jang TY, Liu TW, Liang PC, Hsieh MY, Lin ZY, Chen SC, Huang CF, Huang JF, and Dai CY

Ming-Ying Lu, Ming-Lun Yeh, Ching-I Huang, Yi-Shan Tsai, Pei-Chien Tsai, Yu-Min Ko, Ching-Chih Lin, Kuan-Yu Chen, Yu-Ju Wei, Po-Yao Hsu, Cheng-Ting Hsu, Tyng-Yuan Jang, Ta-Wei Liu, Po-Cheng Liang, Ming-Yen Hsieh, Zu-Yau Lin, Shinn-Cherng Chen, Chung-Feng Huang, Jee-Fu Huang, Chia-Yen Dai, Wan-Long Chuang, Ming-Lung Yu, Hepatitis Center and Hepatobiliary Division, Department of Internal Medicine, Kaohsiung Medical University Hospital, Kaohsiung 80708, Taiwan

Ming-Lun Yeh, Ching-I Huang, Zu-Yau Lin, Shinn-Cherng Chen, Chung-Feng Huang, Jee-Fu Huang, Chia-Yen Dai, Wan-Long Chuang, Ming-Lung Yu, School of Medicine and Hepatitis Research Center, College of Medicine, Center for Cancer Research and Center for Liquid Biopsy, Kaohsiung Medical University, Kaohsiung 80708, Taiwan

Shu-Chi Wang, Department of Medical Laboratory Science and Biotechnology, Kaohsiung Medical University, Kaohsiung 80708, Taiwan

Chia-Yen Dai, Health Management Center, Department of Community Medicine, Kaohsiung Medical University Hospital, Kaohsiung 80708, Taiwan

Ming-Lung Yu, Institute of Biomedical Sciences, National Sun Yat-Sen University, Kaohsiung 80708, Taiwan

Corresponding author: Ming-Lung Yu, MD, PhD, Chief Doctor, Full Professor, Hepatitis Center and Hepatobiliary Division, Department of Internal Medicine, Kaohsiung Medical University Hospital, No. 100 Shih-Chuan 1st Road, Sanmin District, Kaohsiung 80708, Taiwan. fish6069@gmail.com

Abstract

BACKGROUND

Chronic hepatitis C virus (HCV) infection induces profound alterations in the cytokine and chemokine signatures in peripheral blood. Clearance of HCV by antivirals results in host immune modification, which may interfere with immune-mediated cancer surveillance. Identifying HCV patients who remain at risk of hepatocellular carcinoma (HCC) following HCV eradication remains an unmet need. We hypothesized that antiviral therapy-induced immune reconstruc-



collected the clinical data; Yu ML and Chuang WL designed the study, interpreted data, and supervised the manuscript; all authors had read and approved the final manuscript.

Institutional review board

statement: This study was reviewed and approved by the Institutional Review Board of Kaohsiung Medical University Hospital [No. KMUHIRB-E(I)-20180307&KMUHIRB-G(II)-20170020].

Informed consent statement: All

study participants provided informed written consent prior to study enrollment.

Conflict-of-interest statement: The authors declare that they have no competing interests.

Data sharing statement: Technical appendix, statistical code, and dataset available from the corresponding author at fish6069@gmail.com. Participants gave informed consent for data sharing.

STROBE statement: The authors have read the STROBE Statement checklist of items, and the manuscript was prepared and revised according to the STROBE Statement -checklist of items.

Supported by Kaohsiung Medical University and Kaohsiung Medical University Hospital (KMU-KMUH Co-Project of Key Research), No. KMU-DK107004.

Country/Territory of origin: Taiwan

Specialty type: Gastroenterology and hepatology

Provenance and peer review: Unsolicited article; Externally peer reviewed.

Peer-review model: Single blind

Peer-review report's scientific quality classification

Grade A (Excellent): 0 Grade B (Very good): B Grade C (Good): C Grade D (Fair): 0

tion may be relevant to HCC development.

AIM

To investigate the impact of differential dynamics of cytokine expression on the development of HCC following successful antiviral therapy.

METHODS

One hundred treatment-naïve HCV patients with advanced fibrosis (F3/4) treated with direct-acting antivirals (DAAs) or peginterferon/ribavirin who achieved sustained virologic response [SVR, defined as undetectable HCV RNA throughout 12 wk (SVR12) for the DAA group or 24 wk (SVR24) for the interferon group after completion of antiviral therapy] were enrolled since 2003. The primary endpoint was the development of new-onset HCC. Standard HCC surveillance (abdominal ultrasound and α -fetoprotein) was performed every six months during the followup. Overall, 64 serum cytokines were detected by the multiplex immunoassay at baseline and 24 wk after end-of-treatment.

RESULTS

HCC developed in 12 of the 97 patients over 459 person-years after HCV eradication. In univariate analysis, the Fibrosis-4 index (FIB-4), hemoglobin A1c (HbA1c), the dynamics of tumor necrosis factor- α (TNF- α), and TNF-like weak inducer of apoptosis (TWEAK) after antiviral therapy were significant HCC predictors. The multivariate Cox regression model showed that Δ TNF- α (\leq -5.7 pg/mL) was the most important risk factor for HCC (HR = 11.54, 95%CI: 2.27-58.72, P = 0.003 in overall cases; HR = 9.98, 95%CI: 1.88-52.87, P = 0.007 in the interferon group). An HCC predictive model comprising FIB-4, HbA1c, Δ TNF- α , and Δ TWEAK had excellent performance, with 3-, 5-, 10-, and 13-year areas under the curve of 0.882, 0.864, 0.903, and 1.000, respectively. The 5-year accumulative risks of HCC were 0%, 16.9%, and 40.0% in the low-, intermediate-, and high-risk groups, respectively.

CONCLUSION

Downregulation of serum TNF- α significantly increases the risk of HCC after HCV eradication. A predictive model consisting of cytokine kinetics could ameliorate personalized HCC surveillance strategies for post-SVR HCV patients.

Key Words: Hepatitis C virus; Hepatocellular carcinoma; Sustained virologic response; Tumor necrosis factor-a; Tumor necrosis factor-like weak inducer of apoptosis; Cytokine

©The Author(s) 2022. Published by Baishideng Publishing Group Inc. All rights reserved.

Core Tip: Successful hepatitis C virus (HCV) eradication does not eliminate hepatocellular carcinoma (HCC) development. Clearance of HCV by antiviral agents results in host immune modification, which might interfere with immune-mediated cancer surveillance. We attempted to identify immune biomarkers to predict HCC occurrence after antiviral therapy. The dynamics of serum tumor necrosis factor- α (TNF- α) and TNF-like weak inducer of apoptosis were associated with HCC occurrence after HCV clearance. We established a predictive model to assess the risk of HCC among HCV patients after HCV eradication. Our findings provide a clue for the pathogenesis of hepatocarcinogenesis and a strategy for HCC surveillance based on risk stratification.

Citation: Lu MY, Yeh ML, Huang CI, Wang SC, Tsai YS, Tsai PC, Ko YM, Lin CC, Chen KY, Wei YJ, Hsu PY, Hsu CT, Jang TY, Liu TW, Liang PC, Hsieh MY, Lin ZY, Chen SC, Huang CF, Huang JF, Dai CY, Chuang WL, Yu ML. Dynamics of cytokines predicts risk of hepatocellular carcinoma among chronic hepatitis C patients after viral eradication. World J Gastroenterol 2022; 28(1): 140-153

URL: https://www.wjgnet.com/1007-9327/full/v28/i1/140.htm DOI: https://dx.doi.org/10.3748/wjg.v28.i1.140



Grade E (Poor): 0

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: htt p://creativecommons.org/License s/by-nc/4.0/

Received: September 8, 2021 Peer-review started: September 8, 2021

First decision: October 16, 2021 Revised: October 27, 2021 Accepted: December 23, 2021 Article in press: December 23, 2021 Published online: January 7, 2022

P-Reviewer: Emran TB, Sergi CM S-Editor: Yan JP L-Editor: A P-Editor: Yan JP



INTRODUCTION

Chronic hepatitis C virus (HCV) infection is a major cause of liver cirrhosis and hepatocellular carcinoma (HCC). As HCV treatment evolves from an interferon (IFN)based regimen to a therapy based on direct-acting antiviral agents (DAAs), it yields a sustained virologic response (SVR) rate of more than 97% in chronic hepatitis C patients[1,2]. However, successful antiviral therapy does not eliminate HCC development. In a meta-analysis of observational studies, IFN therapy decreased the risk of HCC by 76% in patients with bridging fibrosis or cirrhosis who achieved SVR[3]. Recent studies have reported that HCC occurrence and recurrence rates are potentially increased in HCV patients treated with DAAs[4-6]. This concern remains controversial due to the heterogeneous cohorts, variations in the inclusion criteria, and short duration of follow-up.

Persistent inflammation is a hallmark of chronic hepatic injury. HCV infection induces endogenous type I and III IFN activation, which activates natural killer (NK) cells^[7] and leads to the expression of IFN-stimulated genes (ISGs)^[8]. It causes profound alterations in the cytokine and chemokine signature in peripheral blood. HCV-specific CD8+ T cells play a central role in viral clearance. Chronic HCV infection is characterized by impaired HCV-specific CD8+ T cell responses resulting from viral escape and T cell exhaustion[9]. IFN-based therapy failed to recover the function of HCV-specific CD8+ T cells. This result suggested that the damage to CD8+ T cells might be permanent even after virus elimination^[10]. In contrast, the combination of deleobuvir and faldaprevir resulted in the downregulation of programmed death-1, which led to rapid restoration of HCV-specific CD8+ T cell functions[11]. DAAmediated HCV clearance is correlated with mitigation of the IFN-α-induced immune response, followed by the downregulation of CXCL10 and CXCL11 and normalization of the phenotype and function of NK cells[12].

It is unclear whether host immunological modification after viral eradication influences the development of HCC. Although DAAs are the first choice for HCV clearance, they are not sufficient to abolish hepatic inflammation. Long-term inflammatory responses may change the liver microenvironment and cause irreversible hepatocyte damage. A rapid decline in HCV viral load induced by DAAs results in the reconstitution of immune surveillance[4]. HCV eradication during DAA treatment is accompanied by downregulation of type II and III IFN, their receptors, and downstream ISGs[13], which may affect the antitumor activity of immune cells. IFNs have immunomodulatory properties that regulate various immune cells to inhibit tumor proliferation and angiogenesis. Unlike IFNs, DAAs have neither antiproliferative nor antiangiogenic properties, which may allow the proliferation of malignant cells.

The identification of HCV patients who maintain a high risk of HCC following successful antiviral therapy remains an unmet need. Hepatocarcinogenesis despite HCV clearance is still unclear. First, this study aimed to investigate the impact of differential cytokine expression profiles on the development of HCC among chronic hepatitis C patients with advanced fibrosis who achieved SVR. Second, we attempted to identify immune biomarkers to predict the risk of HCC after successful antiviral therapy.

MATERIALS AND METHODS

Subjects

One hundred treatment-naïve chronic hepatitis C patients with advanced fibrosis treated with either pegylated IFN/ribavirin or IFN-free DAA who achieved SVR were recruited from Kaohsiung Medical University Hospital since 2003. Patients were required to satisfy any one of the following criteria to be diagnosed with advanced fibrosis (F3/4): Fibrosis-4 (FIB-4) index > 3.25[14], transient elastography (Fibroscan) > 9.1 kPa, or acoustic radiation force impulse elastography > 1.81 m/s. The exclusion criteria were coinfection with hepatitis B, hepatitis D or human immunodeficiency virus; history of liver transplantation; prior presence of HCC; decompensated liver cirrhosis; malignancy; alcoholism; primary biliary cholangitis; a1-antitrypsin deficiency; autoimmune hepatitis; renal function impairment; and psychiatric conditions.

Treatment

In the IFN group, the patients were subcutaneously administered peginterferon α -2a (180 µg/wk) plus weight-based ribavirin (1200 mg/d for weights \geq 75 kg or 1000 mg/d for weights < 75 kg) for 24 to 48 wk depending on the HCV genotype. In the



DAA group, the physician selected IFN-free DAA regimens for 12 to 24 wk that were discreetly based on the HCV international treatment guidelines (The Asian Pacific Association for the Study of the Liver, European Association for the Study of the Liver and American Association for The Study of Liver Diseases).

Outcome assessment

SVR was defined as undetectable HCV RNA throughout 12 wk (SVR12) for the DAA group or 24 wk (SVR24) for the IFN group after completion of antiviral therapy[15,16]. The primary endpoint was the occurrence of new-onset HCC. Standard HCC surveillance [abdominal ultrasound and α -fetoprotein (AFP) every six months] was performed during the follow-up[17]. HCC development within six months of initiation of antiviral treatment was excluded. Proof of HCC was directly linked to the National Cancer Registration of Taiwan in Health and Welfare Data Science Center (Taiwan). This study was approved by the Institutional Review Board of Kaohsiung Medical University Hospital (No. KMUHIRB-E(I)-20180307 & KMUHIRB-G(II)-20170020). Written informed consent was acquired from all participants.

HCV genotyping and quantification

Anti-HCV antibodies were identified by a third-generation commercially available enzyme-linked immunosorbent assay (Abbott Laboratories, Chicago, IL, United States). HCV RNA was quantified by real-time polymerase chain reaction assay with a lower limit of detection of 12 IU/mL (RealTime HCV; Abbott Molecular, Des Plaines IL, United States)[18]. HCV genotypes were determined using a commercial kit (Abbott RealTime HCV Genotype II; Abbott Molecular, Des Plaines, IL, United States).

Cytokine measurement

Serum samples were collected from the participants at baseline and SVR24. In total, 64 serum cytokines and chemokines (Supplementary Tables 1-3) were measured by the magnetic bead multiplex immunoassay according to the manufacturer's instructions (Merck KGaA, Darmstadt, Germany)[19,20]. In brief, a calibration curve based on 1:3 dilutions of the highest standard was used for quantification. Beads were premixed and put into wells containing diluted serum and reagents. After fixation of the antigen on the capture antibody linked with the microspheres, a biotinylated detection antibody was added. The concentration of the analyte was quantified based on the bead color and the intensity of the fluorescent signal using the multiplex Luminex-200 (Luminex Corporation, Austin, TX, United States). All samples were analyzed in duplicate.

Statistical analysis

Student's t test and the Mann-Whitney U test were performed to compare the continuous variables. The chi-square (χ^2) test with Yates correction or Fisher's exact test was used to assess the categorical variables. Differences in the cumulative incidence of HCC between groups were analyzed by Kaplan-Meier survival analysis and the log-rank test. The risk factors for HCC were evaluated using multivariate Cox regression analysis. In conjunction with receiver operating characteristic area (ROC) analysis^[21], the optimum cutoff value to distinguish between the risk strata was calculated by the Youden index^[22]. The performance of biomarkers to predict the risk of HCC was calculated by time-dependent ROC curve analysis. The area under the ROC area (AUROC) was assessed by the timeROC package of R software (http: //www.r-project.org). The statistical power for the comparison of survival curves between two groups under the Cox proportional hazards model was calculated by the powerSurvEpi package of R software. A two-tailed P value < 0.05 was considered statistically significant. The statistical analysis was conducted by the Statistic Packages for Social Science Program (SPSS v19.0 for Windows, SPSS Inc., United States). The statistical methods of this study were reviewed by Dr. Tsai PC from Kaohsiung Medical University.

RESULTS

Baseline demographics

The baseline demographics of the study subjects are shown in Table 1. There were no significant differences in age, sex, HCV genotype, FIB-4 index, or AFP levels between the DAA and IFN groups. HCV RNA was significantly higher in the DAA group than



Lu MY et al. Cytokines predict HCC after HCV eradication

Table 1 Baseline demograp	ohics of study subjects			
Group	Total	DAA	IFN	P value (DAA vs IFN)
n	100	50	50	
Age (yr)	63.8 ± 7.2	64.9 ± 7.9	62.6 ± 6.3	0.100
Sex, n (%)				
Female	66 (66.0)	38 (76.0)	28 (56.0)	0.057
Male	34 (34.0)	12 (24.0)	22 (44.0)	
HCV genotype, n (%)				
1	65 (65.0)	37 (74.0)	28 (56.0)	0.098
2	23 (23.0)	10 (20.0)	13 (26.0)	
Mixed	12 (12.0)	3 (6.0)	9 (18.0)	
HCV RNA (log IU/mL)	2.47 ± 0.89	2.67 ± 0.84	2.28 ± 0.91	0.027
FIB-4	6.14 ± 3.28	6.55 ± 3.69	5.73 ± 2.80	0.213
AFP (ng/mL)	26.3 ± 56.7	28.8 ± 74.0	23.6 ± 29.3	0.662
Platelet (k/µL)	119.8 ± 34.7	115.2 ± 35.9	124.4 ± 33.2	0.186
AST (IU/L)	136.9 ± 79.9	115.2 ± 64.7	158.7 ± 87.9	0.006
ALT (IU/L)	177.5 ± 138.4	127.9 ± 83.9	227.0 ± 163.2	2.7×10^{-4}
γ-GT (IU/L)	67.6 ± 48.8	57.2 ± 42.6	76.7 ± 52.4	0.053
Cholesterol (mg/dL)	161.6 ± 34.8	158.6 ± 37.1	165.2 ± 32.0	0.388
Triglyceride (mg/dL)	97.7 ± 41.4	99.2 ± 40.5	96.0 ± 42.8	0.722
HDL (mg/dL)	47.0 ± 13.3	49.0 ± 11.6	44.1 ± 15.1	0.107
LDL (mg/dL)	90.6 ± 26.4	86.0 ± 24.4	97.2 ± 28.1	0.068
Cr (mg/dL)	0.79 ± 0.24	0.77 ± 0.27	0.82 ± 0.20	0.222
HbA1c (%)	5.8 ± 1.2	5.5 ± 0.7	6.0 ± 1.5	0.016
BMI (kg/m ²)	24.4 ± 4.7	24.0 ± 6.0	24.8 ± 2.9	0.381

DAA: Direct-acting antiviral agent; IFN: Interferon; HCV: Hepatitis C virus; FIB-4: Fibrosis-4 index; AFP: Alpha-fetoprotein; AST: Aspartate aminotransferase; ALT: Alanine aminotransferase; γ-GT: γ-glutamyltransferase; HDL: High-density lipoprotein; LDL: Low-density lipoprotein; Cr: Creatinine; HbA1c: Hemoglobin A1c; BMI: Body mass index.

> in the IFN group. Aspartate aminotransferase, alanine aminotransferase and hemoglobin A1c (HbA1c) were significantly elevated in the IFN group compared to the DAA group.

Cumulative probability of HCC development

HCC developed in 12 (IFN group n = 11, DAA group n = 1) of the 97 patients over 459 person-years of follow-up. Three patients were excluded because HCC occurred within six months of initiation of the antiviral treatment. The mean follow-up time was 7.46 years [interquartile range (IQR) = 3.65-12.23] in the IFN group and 1.84 years (IQR = 1.19-2.43) in the DAA group. The annual incidence of HCC was 2.95% in the IFN group and 1.16% in the DAA group. The Kaplan-Meier survival analysis showed no statistical significance in the accumulative probability of HCC between the IFN and DAA groups (log-rank *P* value = 0.712) (Figure 1).

Cytokines associated with HCC development

In total, 64 cytokines were used to analyze the relationship with HCC (Supplementary Table 1). Seven of the sixty-four cytokines were excluded from subsequent analysis because more than 80% of the samples were below the limit of detection. Members of the tumor necrosis factor (TNF) superfamily, including TNF- α and TNF-like weak inducer of apoptosis (TWEAK), were associated with the development of HCC. The baseline TNF- α level was significantly elevated in the HCC group compared

Zaishidena® WJG https://www.wjgnet.com

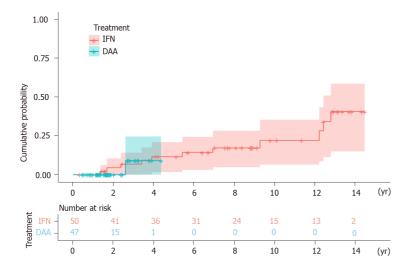
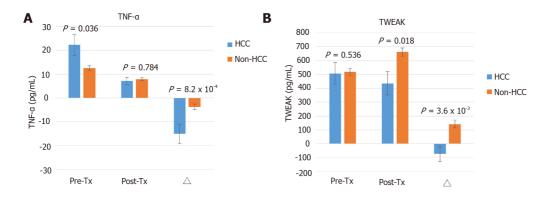
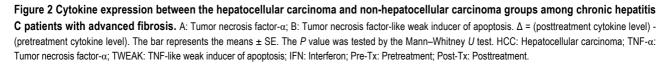


Figure 1 Kaplan-Meier survival analysis. HCC: Hepatocellular carcinoma; IFN: Interferon; DAA: Direct-acting antiviral agent.





to the non-HCC group ($22.22 \pm 4.33 vs 12.53 \pm 1.03 pg/mL$, P = 0.036). There was no significant difference in the posttreatment TNF- α levels between the HCC and non-HCC groups. The change in TNF- α levels (Δ TNF- α) before and after treatment significantly differed between the HCC and non-HCC groups (-15.11 ± 3.93 vs -3.78 ± 1.16 pg/mL, $P = 8.2 \times 10^4$) (Figure 2A). The baseline TWEAK expression was similar in both groups. The posttreatment TWEAK level was significantly lower in the HCC group than in the non-HCC group (434.82 ± 84.18 vs 660.65 ± 30.34 pg/mL, P = 0.018). Δ TWEAK showed a reciprocal change between the HCC and non-HCC groups and achieved statistical significance (-71.78 ± 54.56 vs 142.81 ± 27.69 pg/mL, $P = 3.6 \times 10^3$) (Figure 2B).

Among the HCV patients treated with pegIFN/ribavirin, the baseline TNF-α level was significantly higher in the HCC group than in the non-HCC group (22.55 ± 4.72 *vs* 9.13 ± 7.79 pg/mL, P = 0.017). The posttreatment TNF-α concentration was comparable between the HCC and non-HCC groups. Δ TNF-α levels significantly declined in HCC compared to non-HCC patients (-15.86 ± 4.22 *vs* -4.56 ± 1.85 pg/mL, P = 0.007) (Supplementary Figure 1A). The dynamic change in TWEAK did not show significant variations between the HCC and non-HCC groups (Supplementary Figure 1B).

Cox regression analysis of the relationship between the differentially expressed cytokines and HCC

In univariate Cox regression analysis, FIB-4 ($\geq 9 vs < 9$, crude HR = 4.04, 95%CI: 1.27-12.86, P = 0.018), HbA1c ($\geq 7 vs < 7\%$, crude HR = 5.38, 95%CI: 1.38-20.99, P = 0.015), pretreatment TNF- α ($\geq 18 vs < 18 pg/mL$, crude HR = 5.15, 95%CI: 1.57-16.87, P = 0.007), Δ TNF- α ($\leq -5.7 vs > -5.7 pg/mL$, crude HR = 11.07, 95%CI: 2.27-53.87, P = 0.003), and Δ TWEAK ($\leq -70 vs > -70 pg/mL$, crude HR = 4.01, 95%CI: 1.20-13.40, P = 0.024)

were significant predictors of HCC. Multivariate stepwise Cox regression analysis revealed that $\Delta TNF-\alpha$ was the only independent risk factor for HCC (HR = 11.54, 95%CI: 2.27-58.72, P = 0.003) (Table 2).

Among the HCV patients treated with pegIFN/ribavirin, univariate Cox regression showed that the significant predictors of HCC included sex, FIB-4 ($\geq 9 vs < 9$), HbA1c, baseline TNF- α (\geq 18 vs < 18 pg/mL) and Δ TNF- α (\geq -5.7 vs < -5.7 pg/mL). The association between ∆TWEAK (≤ -70 vs > -70 pg/mL) and HCC was borderline statistically significant. Stepwise multivariate Cox regression revealed that the Δ TNF- α level was the only independent risk factor for HCC in the IFN group (HR = 9.98, 95%CI: 1.88-52.87, *P* = 0.007 (Supplementary Table 2).

Subgroup analysis for the association between TNF-α and HCC

Since age and diabetes mellitus were important risk factors for HCC, the subjects were further stratified by age and HbA1c. The high- and low-risk groups were dichotomized based on Δ TNF- α with a cutoff value of -5.7 pg/mL. The multivariate Cox regression analysis revealed that the high-risk group (Δ TNF- $\alpha \leq -5.7$ pg/mL) had an 11-fold cumulative probability of HCC compared to that of the low-risk group (HR = 11.02, 95% CI: 1.86-65.17, P = 0.008) among HCV patients with HbA1c less than 7%. In the younger population (age < 65 years old), the HCC risk was borderline significant between the high- and low-risk groups (HR = 8.51, 95%CI: 0.78-92.86, P = 0.079). Among patients with both HbA1c < 7% and age below 65 years old, the high-risk group had a 20-fold cumulative probability of HCC in comparison with the low-risk group (HR = 19.99, 95% CI: 0.90-443.91, P = 0.058) (Figure 3). The level of Δ TNF- α did not influence the development of HCC in either the patients aged \geq 65 years old or with HbA1c \geq 7% (Supplementary Figure 2).

HCC predictive model

Based on previous analyses, the FIB-4 index, HbA1c, Δ TNF- α , and Δ TWEAK were selected as parameters to predict the risk of HCC. The HCC predictive model was as follows: Score = $4 \times FIB-4$ (≥ 9 , yes = 1, no = 0) + $5 \times HbA1c$ (≥ 7 , yes = 1, no = 0) + 11×10^{-1} Δ TNF (\leq -5.7, yes = 1, no = 0) + 4 × Δ TWEAK (\leq -70, yes = 1, no = 0).

The weighting coefficient for each parameter was derived from the crude hazard ratio of the univariate Cox proportional hazards model. The performance of this HCC predictive model was assessed by time-dependent ROC curve analysis. In overall cases, the 3-year, 5-year, 10-year, and 13-year areas under the ROC curve (AUCs) were 0.882, 0.864, 0.903, and 1.000, respectively (Figure 4A). In the IFN group, the 3-year, 5year, 10-year, and 13-year areas under the ROC curve (AUCs) were 0.782, 0.802, 0.870, and 1.000, respectively (Figure 4B).

Kaplan-Meier analysis for HCV patients stratified by risk scores

To classify the predictive score according to the risk of HCC, the patients were further stratified into low- (score = 0-7), intermediate- (score = 8-15), and high-risk groups (score > 15). In the high-risk group, the 3-year, 5-year, and 10-year cumulative risks of HCC were 20.0%, 40.0%, and 60.0%, respectively. In the intermediate-risk group, the 3year, 5-year, and 10-year cumulative probabilities of HCC were 11.4%, 16.9%, and 31.0%, respectively. In contrast, none of the low-risk patients had HCC within 14 years of follow-up after successful viral eradication among the overall cases (log-rank P value = $6.8 \times 10^{\circ}$) (Figure 5A). Likewise, the Kaplan-Meier survival analysis revealed a significant difference in the cumulative probability of HCC among the IFN group stratified by the risk scores (log-rank P value = 9.6 × 10⁻⁵) (Figure 5B).

DISCUSSION

This study revealed that there was no significant difference in the risk of HCC between the DAA and IFN groups after successful antiviral therapy. Downregulation of TNF- α and TWEAK increased the risk of hepatic carcinogenesis. Δ TNF- α was identified as an independent predictor of new-onset HCC among HCV patients with SVR. The effect of TNF-α was more prominent in young adults with normoglycemia. An HCC predictive model comprising FIB-4, HbA1c, Δ TNF- α , and Δ TWEAK had excellent performance, with 3-, 5-, 10-, and 13-year areas under the curve of 0.882, 0.864, 0.903, and 1.000, respectively. The 5-year accumulative risk of HCC was 0.0%, 16.9%, and 40.0% in the low-, intermediate-, and high-risk groups, respectively. These findings remained statistically significant among the HCV patients treated with pegIFN/ribavirin. Because there was only one HCC in the DAA group, the role of TNF- α in HCC should



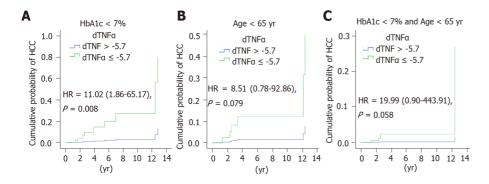
Table 2 Factors associated with the onset of hepatocellular carcinoma: Univariate and multivariate Cox regression models				
Mariahlar	Univariate Cox regression		Multivariate Cox regress	ion
Variables	Crude HR (95%CI)	<i>P</i> value	Adjusted HR (95%CI)	Adjusted P value
Age (yr)	1.07 (0.98-1.18)	0.149	-	-
Sex (male vs female)	3.19 (0.96-10.65)	0.059	-	-
HCV genotype	1.00 (0.47-2.13)	0.995	-	-
HCV RNA (log IU/mL)	0.68 (0.37-1.25)	0.213	-	-
FIB-4	1.13 (0.98-1.31)	0.089	-	-
FIB-4 ($\geq 9 vs < 9$)	4.04 (1.27-12.86)	0.018	-	-
Platelet (k/µL)	0.99 (0.98-1.01)	0.408	-	-
AFP (ng/mL)	1.00 (1.00-1.01)	0.308	-	-
HbA1c (%)	1.28 (1.01-1.62)	0.041	-	-
HbA1c (≥ 7 <i>vs</i> < 7%)	5.38 (1.38-20.99)	0.015	-	-
BMI (kg/m²)	1.00 (0.83-1.20)	0.993	-	-
Treatment (DAA vs IFN)	0.66 (0.07-6.15)	0.713	-	-
TNF-α (pg/mL)				
Pre-Tx TNF- $\alpha \ge 18$	5.15 (1.57-16.87)	0.007	-	-
Post-Tx TNF- $\alpha \ge 6$	0.79 (0.25-2.46)	0.683	-	-
$\Delta TNF-\alpha \leq -5.7$	11.07 (2.27-53.87)	0.003	11.54 (2.27-58.72)	0.003
TWEAK (pg/mL)				
Pre-Tx TWEAK ≥ 500	2.18 (0.64-7.39)	0.213	-	-
Post-Tx TWEAK ≥ 600	0.80 (0.20-3.11)	0.744	-	-
ΔTWEAK ≤ -70	4.01 (1.20-13.40)	0.024	-	-

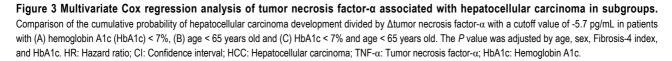
The forward stepwise multivariate Cox regression model was adjusted by age, sex, hepatitis C virus (HCV) genotypes, HCV RNA, Fibrosis-4 index (FIB-4), platelet, alpha-fetoprotein, hemoglobin A1c, body mass index, treatment, tumor necrosis factor-a and tumor necrosis factor-like weak inducer of apoptosis (pretreatment, posttreatment, Δ). The cut-off value for each cytokine and FIB-4 was determined by Youden index of receiver operating characteristic curve. Δ = (posttreatment cytokine level) - (pretreatment cytokine level). DAA: Direct-acting antiviral agent; IFN: Interferon; HCV: Hepatitis C virus; FIB-4: Fibrosis-4 index; AFP: Alpha-fetoprotein; HbA1c: Hemoglobin A1c; BMI: Body mass index; HCV: Hepatitis C virus; TNF-a: tumor necrosis factor-a; TWEAK: TNF-like weak inducer of apoptosis; Pre-Tx: Pretreatment; Post-Tx: Posttreatment; HR: Hazard ratio; CI: Confidence interval.

> be further verified in this population. The HCC risk could be modified by the preexisting host background and adjusted by the immune signatures after viral eradication. This predictive model helps clinicians adopt appropriate surveillance strategies for chronic hepatitis C patients following successful antiviral therapy according to the risk of HCC.

> Our study showed that elevation of pretreatment TNF-a levels raised the possibility of new-onset HCC. Consistent with our study, Tarhuni et al[23] found that HCVrelated cirrhotic patients carrying TNF-a 308 G>A had higher basal TNF-a production and exhibited a higher risk of HCC. Elevated basal TNF-α indicates sustained hepatic inflammation accompanied by persistent liver damage, which is susceptible to carcinogenesis. A systematic review showed that TNF-a was one of the strongest host genetic predictors for HCC in HCV-infected patients[24]. These findings suggested that the immune background was affected before antiviral therapy.

> Interestingly, the abrupt decline in $TNF-\alpha$ levels after successful antiviral therapy increased the risk of HCC in our study. This implies a potential modification of the immune milieu by antiviral therapy that may trigger HCC development. Stimulation of the immune system effectively protects tissues from malignant cell invasion. Both cytotoxic T lymphocytes and NK cells are potent effectors in immune surveillance. TNF- α mediates the immune response against tumor cells by creating a microenvironment toward immunogenic activation rather than suppression[25]. Suppression of TNF signaling enables tumor cells to evade attack by cytotoxic T lymphocytes and attenuate *in vivo* antitumor responses^[26]. Antiviral therapy may disrupt the balance





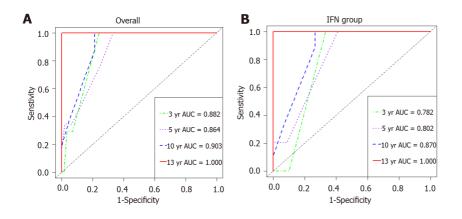


Figure 4 Time-dependent receiver operating characteristic curve analysis for the hepatocellular carcinoma predictive model. IFN: Interferon; AUC: Area under the curve.

from TNF- α activation to inhibition in immune surveillance. Alternatively, Debes *et al* [27] found that HCV patients with early-onset or recurrent HCC within 18 mo maintained stable or even higher TNF- α levels after DAA therapy. This implied that those patients might exhibit precarcinogenic or ongoing carcinogenic activity induced by TNF- α in response to occult HCC.

Both TNF- α and TWEAK belong to members of the TNF superfamily. These cytokines are mainly produced by macrophages, monocytes, and lymphocytes. TWEAK is a multifunctional cytokine that regulates a variety of cellular activities, including cell proliferation, differentiation, apoptosis, inflammation, and angiogenesis, *via* the fibroblast growth factor-inducible 14 receptor[28]. TWEAK appears to attenuate the innate response switch to adaptive immunity[29]. In addition, TWEAK is a weak inducer of apoptosis and also participates in tissue repair[30]. In chronic liver injury and repair, TWEAK appears to initiate liver progenitor cell expansion and ductal proliferation[31]. Hyperstimulation of inflammatory cells simultaneously results in excessive matrix deposition by activated hepatic stellate cells and myofibroblasts *via* the lymphotoxin- β signaling pathway[32]. Our study showed that posttreatment TWEAK expression was upregulated in the non-HCC group. Viral clearance alleviates the inflammatory status in the liver. It provides a microenvironment to facilitate the reconstruction of hepatocytes aided by TWEAK, which may further delay HCC development.

Our study confirms the consensus that DAA treatment does not markedly increase the risk of HCC compared to IFN treatment[33]. Most evidence has shown a decline in HCC risk regardless of whether SVR was achieved by IFN alone, DAA-only, or combined regimens[34]. However, successful antiviral therapy cannot eliminate the risk of HCC. The standard surveillance strategy (ultrasound and AFP every six months) advocates for all HCV patients. However, interindividual variations in HCC risk raise the question of whether the recommendations for HCC screening should be adjusted. Age, male sex, diabetes mellitus, and advanced fibrosis are well-known independent predictors of HCC after viral eradication[35-37]. In the absence of



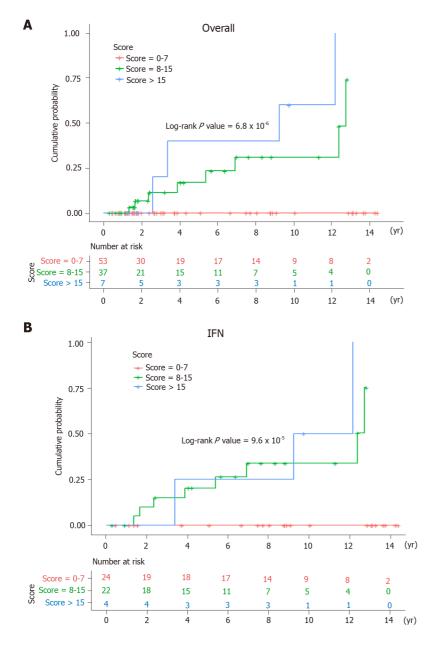


Figure 5 Kaplan-Meier survival analysis for chronic hepatitis C patients stratified by the risk scores. IFN: Interferon.

diabetes mellitus and old age (> 65 years old), the presence of Δ TNF- α ≤ -5.7 pg/mL increased the risk of HCC in patients with advanced fibrosis by 20-fold after HCV clearance. According to this HCC predictive model, patients with scores exceeding 15 should be closely monitored, since the 5-year cumulative risk of HCC reaches up to 40.0%. Nevertheless, none of the HCC cases had been identified over 14 years of follow-up in patients with a score of less than 7. The surveillance intervals may be extended among HCV patients achieving SVR in the absence of concurrent risk factors. In the post-DAA era, the risk model-based algorithm provides a cost-effective surveillance strategy for HCC.

The advancement of high-throughput technology makes early HCC detection more feasible. Currently, integrating multiomics data for HCC screening is also frequently observed[38]. The GALAD score consists of clinical factors (sex, age) and biomarkers (AFP, AFP-L3, and Des-carboxyprothrombin) that have an excellent performance to predict HCC, with an AUROC up to 0.97[39]. Using a miRNA panel (miR-22, miR-199a-3p) with AFP provided high diagnostic accuracy (AUROC = 0.988) for the early detection of HCC in HCV patients[39]. The methylation pattern of circulating cell-free DNA (APC, SFRP1, GSTP1, and RASSF1A) has demonstrated sufficient detection value to distinguish HCC patients from healthy controls[40]. Nevertheless, a majority of studies collected a cohort with a small sample size, and the analytical methods varied even in the same testing platform. These factors have limited the clinical application of these biomarkers.



Even though the sample size was limited in this pilot study, the statistical power was sufficient to be reliable. In overall cases, the statistical power of the association between Δ TNF- α and HCC was 99.9% to reject the null hypothesis at a *P* value < 0.05 under a hazard ratio of 11.54. In the IFN group, the statistical power of Δ TNF- α on HCC risk was 94.6% to reject the null hypothesis at a *P* value < 0.05 under a hazard ratio of 9.98. There are several limitations to this study. Although expensive IFN-free DAAs have been on the market since 2014, the National Health Insurance in Taiwan has reimbursed DAAs for HCV patients with advanced fibrosis since 2017. The followup time in most HCV patients treated with DAAs was less than 3 years. Owing to the small sample size and short follow-up time of the DAA group, a larger study cohort is necessary to validate the performance of this predictive model. IFN may induce distinct host immune alterations in comparison with DAA. As only one HCC case was identified in the DAA group throughout the follow-up period, it was unable to compare the diversity of cytokine profiles regarding HCC between the IFN and DAA groups. This predictive model was restricted to HCV patients with advanced fibrosis following successful antiviral therapy. Additionally, the optimal cutoff value should be further verified in other populations. The parameters of this predictive model were composed of serum cytokines involving the TNF superfamily. Host inflammation elicited by other etiologies may interfere with the predictive accuracy. Serum cytokines may not reflect the microenvironment within hepatocytes. To interpret this HCC predictive model, more care should be given to HCV patients presenting coinfection with other viruses, inflammatory disease, or malignancies.

CONCLUSION

This study revealed that downregulation of TNF-a increases the risk of HCC among HCV patients following successful antiviral therapy. Inhibition of TNF- α may attenuate host immune surveillance against tumor cells. Our findings provide a clue for the pathogenesis of hepatocarcinogenesis and a strategy for HCC surveillance based on risk stratification. With the development of high-throughput molecular technology, it is believed that more novel biomarkers will be applied in the early detection of HCC in the future.

ARTICLE HIGHLIGHTS

Research background

Successful hepatitis C virus (HCV) eradication cannot eliminate hepatocellular carcinoma (HCC) development. Chronic HCV infection induces profound alterations in cytokine and chemokine signatures. Clearance of HCV results in host immune modification, which may interfere with immune-mediated cancer surveillance.

Research motivation

The mechanism of hepatocarcinogenesis despite HCV clearance is still unclear.

Research objectives

To investigate the impact of differential cytokine expression on the development of HCC following HCV eradication.

Research methods

One hundred treatment-naïve HCV patients with advanced fibrosis who received antiviral therapy and achieved sustained virologic response (SVR) were enrolled. The primary endpoint was the development of new-onset HCC. In total, 64 serum cytokines were detected by the multiplex immunoassay at baseline and 24 wk after end-of-treatment.

Research results

The dynamics of serum tumor necrosis factor-α (TNF-α) and TNF-like weak inducer of apoptosis (TWEAK) were associated with HCC occurrence after HCV clearance. Multivariate Cox regression analysis showed that Δ TNF- $\alpha \leq -5.7$ pg/mL was an independent risk factor for HCC. An HCC predictive model comprising the Fibrosis-4 index, hemoglobin A1c, Δ TNF- α , and Δ TWEAK had excellent performance in stra-



tifying the risk of HCC among HCV patients with SVR.

Research conclusions

Downregulation of serum TNF-a significantly increased the risk of HCC after HCV eradication.

Research perspectives

Our findings provide a clue for the pathogenesis of hepatocarcinogenesis and a strategy for HCC surveillance based on risk stratification.

REFERENCES

- 1 Yu ML. Hepatitis C treatment from "response-guided" to "resource-guided" therapy in the transition era from interferon-containing to interferon-free regimens. J Gastroenterol Hepatol 2017; 32: 1436-1442 [PMID: 28124463 DOI: 10.1111/jgh.13747]
- 2 Falade-Nwulia O, Suarez-Cuervo C, Nelson DR, Fried MW, Segal JB, Sulkowski MS. Oral Direct-Acting Agent Therapy for Hepatitis C Virus Infection: A Systematic Review. Ann Intern Med 2017; 166: 637-648 [PMID: 28319996 DOI: 10.7326/M16-2575]
- Morgan RL, Baack B, Smith BD, Yartel A, Pitasi M, Falck-Ytter Y. Eradication of hepatitis C virus 3 infection and the development of hepatocellular carcinoma: a meta-analysis of observational studies. Ann Intern Med 2013; 158: 329-337 [PMID: 23460056 DOI: 10.7326/0003-4819-158-5-201303050-00005]
- Reig M, Mariño Z, Perelló C, Iñarrairaegui M, Ribeiro A, Lens S, Díaz A, Vilana R, Darnell A, Varela M, Sangro B, Calleja JL, Forns X, Bruix J. Unexpected high rate of early tumor recurrence in patients with HCV-related HCC undergoing interferon-free therapy. J Hepatol 2016; 65: 719-726 [PMID: 27084592 DOI: 10.1016/j.jhep.2016.04.008]
- 5 Meringer H, Shibolet O, Deutsch L. Hepatocellular carcinoma in the post-hepatitis C virus era: Should we change the paradigm? World J Gastroenterol 2019; 25: 3929-3940 [PMID: 31413528 DOI: 10.3748/wjg.v25.i29.3929]
- Hernáez-Alsina T, Caballol-Oliva B, Díaz-González Á, Guedes-Leal C, Reig M. Risk of recurrence 6 of hepatocellular carcinoma in patients treated with interferon-free antivirals. Gastroenterol Hepatol 2019; 42: 502-511 [PMID: 31472990 DOI: 10.1016/j.gastrohep.2019.05.003]
- Ahlenstiel G, Titerence RH, Koh C, Edlich B, Feld JJ, Rotman Y, Ghany MG, Hoofnagle JH, Liang TJ, Heller T, Rehermann B. Natural killer cells are polarized toward cytotoxicity in chronic hepatitis C in an interferon-alfa-dependent manner. Gastroenterology 2010; 138: 325-35.e1 [PMID: 19747917 DOI: 10.1053/j.gastro.2009.08.066]
- Sarasin-Filipowicz M, Oakeley EJ, Duong FH, Christen V, Terracciano L, Filipowicz W, Heim MH. Interferon signaling and treatment outcome in chronic hepatitis C. Proc Natl Acad Sci USA 2008; 105: 7034-7039 [PMID: 18467494 DOI: 10.1073/pnas.0707882105]
- 9 Wherry EJ. T cell exhaustion. Nat Immunol 2011; 12: 492-499 [PMID: 21739672 DOI: 10.1038/ni.2035]
- 10 Missale G, Pilli M, Zerbini A, Penna A, Ravanetti L, Barili V, Orlandini A, Molinari A, Fasano M, Santantonio T, Ferrari C. Lack of full CD8 functional restoration after antiviral treatment for acute and chronic hepatitis C virus infection. Gut 2012; 61: 1076-1084 [PMID: 22337949 DOI: 10.1136/gutjnl-2011-300515]
- 11 Martin B, Hennecke N, Lohmann V, Kayser A, Neumann-Haefelin C, Kukolj G, Böcher WO, Thimme R. Restoration of HCV-specific CD8+ T cell function by interferon-free therapy. J Hepatol 2014; 61: 538-543 [PMID: 24905492 DOI: 10.1016/j.jhep.2014.05.043]
- 12 Serti E, Chepa-Lotrea X, Kim YJ, Keane M, Fryzek N, Liang TJ, Ghany M, Rehermann B. Successful Interferon-Free Therapy of Chronic Hepatitis C Virus Infection Normalizes Natural Killer Cell Function. Gastroenterology 2015; 149: 190-200.e2 [PMID: 25754160 DOI: 10.1053/j.gastro.2015.03.004]
- 13 Meissner EG, Wu D, Osinusi A, Bon D, Virtaneva K, Sturdevant D, Porcella S, Wang H, Herrmann E, McHutchison J, Suffredini AF, Polis M, Hewitt S, Prokunina-Olsson L, Masur H, Fauci AS, Kottilil S. Endogenous intrahepatic IFNs and association with IFN-free HCV treatment outcome. J Clin Invest 2014; 124: 3352-3363 [PMID: 24983321 DOI: 10.1172/JCI75938]
- 14 Kayadibi H. Letter: changes in FIB-4 cut-off points for viral hepatitis. Aliment Pharmacol Ther 2017; 45: 1007-1008 [PMID: 28256081 DOI: 10.1111/apt.13956]
- 15 Burgess SV, Hussaini T, Yoshida EM. Concordance of sustained virologic response at weeks 4, 12 and 24 post-treatment of hepatitis c in the era of new oral direct-acting antivirals: A concise review. Ann Hepatol 2016; 15: 154-159 [PMID: 26845592 DOI: 10.5604/16652681.1193693]
- Lin CP, Liang PC, Huang CI, Yeh ML, Hsu PY, Hsu CT, Wei YJ, Liu TW, Hsieh MY, Hou NJ, Jang 16 TY, Lin YH, Wang CW, Lin ZY, Chen SC, Huang CF, Huang JF, Dai CY, Chuang WL, Yu ML. Concordance of SVR12, SVR24 and SVR durability in Taiwanese chronic hepatitis C patients with direct-acting antivirals. PLoS One 2021; 16: e0245479 [PMID: 33539408 DOI: 10.1371/journal.pone.0245479]



- 17 Marrero JA, Kulik LM, Sirlin CB, Zhu AX, Finn RS, Abecassis MM, Roberts LR, Heimbach JK. Diagnosis, Staging, and Management of Hepatocellular Carcinoma: 2018 Practice Guidance by the American Association for the Study of Liver Diseases. Hepatology 2018; 68: 723-750 [PMID: 29624699 DOI: 10.1002/hep.29913]
- 18 Vermehren J, Yu ML, Monto A, Yao JD, Anderson C, Bertuzis R, Schneider G, Sarrazin C. Multicenter evaluation of the Abbott RealTime HCV assay for monitoring patients undergoing antiviral therapy for chronic hepatitis C. J Clin Virol 2011; 52: 133-137 [PMID: 21803650 DOI: 10.1016/j.jcv.2011.07.007]
- 19 McKay HS, Margolick JB, Martínez-Maza O, Lopez J, Phair J, Rappocciolo G, Denny TN, Magpantay LI, Jacobson LP, Bream JH. Multiplex assay reliability and long-term intra-individual variation of serologic inflammatory biomarkers. Cytokine 2017; 90: 185-192 [PMID: 27940218 DOI: 10.1016/j.cvto.2016.09.018]
- Dressen K, Hermann N, Manekeller S, Walgenbach-Bruenagel G, Schildberg FA, Hettwer K, Uhlig 20 S, Kalff JC, Hartmann G, Holdenrieder S. Diagnostic Performance of a Novel Multiplex Immunoassay in Colorectal Cancer. Anticancer Res 2017; 37: 2477-2486 [PMID: 28476816 DOI: 10.21873/anticanres.11588]
- Zweig MH, Campbell G. Receiver-operating characteristic (ROC) plots: a fundamental evaluation tool in clinical medicine. Clin Chem 1993; 39: 561-577 [PMID: 8472349 DOI: 10.1016/0009-9120(93)90037-7
- Schisterman EF, Perkins NJ, Liu A, Bondell H. Optimal cut-point and its corresponding Youden 22 Index to discriminate individuals using pooled blood samples. Epidemiology 2005; 16: 73-81 [PMID: 15613948 DOI: 10.1097/01.ede.0000147512.81966.ba]
- Tarhuni A, Guyot E, Rufat P, Sutton A, Bourcier V, Grando V, Ganne-Carrié N, Ziol M, Charnaux 23 N, Beaugrand M, Moreau R, Trinchet JC, Mansouri A, Nahon P. Impact of cytokine gene variants on the prediction and prognosis of hepatocellular carcinoma in patients with cirrhosis. J Hepatol 2014; 61: 342-350 [PMID: 24751829 DOI: 10.1016/j.jhep.2014.04.011]
- 24 Walker AJ, Peacock CJ, Pedergnana V; STOP-HCV Consortium, Irving WL. Host genetic factors associated with hepatocellular carcinoma in patients with hepatitis C virus infection: A systematic review. J Viral Hepat 2018; 25: 442-456 [PMID: 29397014 DOI: 10.1111/jvh.12871]
- Showalter A, Limaye A, Oyer JL, Igarashi R, Kittipatarin C, Copik AJ, Khaled AR. Cytokines in 25 immunogenic cell death: Applications for cancer immunotherapy. Cytokine 2017; 97: 123-132 [PMID: 28648866 DOI: 10.1016/j.cyto.2017.05.024]
- Kearney CJ, Vervoort SJ, Hogg SJ, Ramsbottom KM, Freeman AJ, Lalaoui N, Pijpers L, Michie J, 26 Brown KK, Knight DA, Sutton V, Beavis PA, Voskoboinik I, Darcy PK, Silke J, Trapani JA, Johnstone RW, Oliaro J. Tumor immune evasion arises through loss of TNF sensitivity. Sci Immunol 2018; 3 [PMID: 29776993 DOI: 10.1126/sciimmunol.aar3451]
- Debes JD, van Tilborg M, Groothuismink ZMA, Hansen BE, Schulze Zur Wiesch J, von Felden J, de 27 Knegt RJ, Boonstra A. Levels of Cytokines in Serum Associate With Development of Hepatocellular Carcinoma in Patients With HCV Infection Treated With Direct-Acting Antivirals. Gastroenterology 2018; 154: 515-517.e3 [PMID: 29102620 DOI: 10.1053/j.gastro.2017.10.035]
- 28 Burkly LC, Michaelson JS, Hahm K, Jakubowski A, Zheng TS. TWEAKing tissue remodeling by a multifunctional cytokine: role of TWEAK/Fn14 pathway in health and disease. Cytokine 2007; 40: 1-16 [PMID: 17981048 DOI: 10.1016/j.cyto.2007.09.007]
- 29 Maecker H, Varfolomeev E, Kischkel F, Lawrence D, LeBlanc H, Lee W, Hurst S, Danilenko D, Li J, Filvaroff E, Yang B, Daniel D, Ashkenazi A. TWEAK attenuates the transition from innate to adaptive immunity. Cell 2005; 123: 931-944 [PMID: 16325585 DOI: 10.1016/j.cell.2005.09.022]
- 30 Shafritz DA. To TWEAK, or not to TWEAK: that is the question. Hepatology 2010; 52: 13-15 [PMID: 20578125 DOI: 10.1002/hep.23750]
- Jakubowski A, Ambrose C, Parr M, Lincecum JM, Wang MZ, Zheng TS, Browning B, Michaelson 31 JS, Baetscher M, Wang B, Bissell DM, Burkly LC. TWEAK induces liver progenitor cell proliferation. J Clin Invest 2005; 115: 2330-2340 [PMID: 16110324 DOI: 10.1172/JCI23486]
- Dwyer BJ, Olynyk JK, Ramm GA, Tirnitz-Parker JE. TWEAK and LTß Signaling during Chronic 32 Liver Disease. Front Immunol 2014; 5: 39 [PMID: 24592262 DOI: 10.3389/fimmu.2014.00039]
- 33 Li DK, Ren Y, Fierer DS, Rutledge S, Shaikh OS, Lo Re V 3rd, Simon T, Abou-Samra AB, Chung RT, Butt AA. The short-term incidence of hepatocellular carcinoma is not increased after hepatitis C treatment with direct-acting antivirals: An ERCHIVES study. Hepatology 2018; 67: 2244-2253 [PMID: 29205416 DOI: 10.1002/hep.29707]
- 34 Gigi E, Lagopoulos VI, Bekiari E. Hepatocellular carcinoma occurrence in DAA-treated hepatitis C virus patients: Correlated or incidental? World J Hepatol 2018; 10: 595-602 [PMID: 30310537 DOI: 10.4254/wjh.v10.i9.595]
- Yu ML, Huang CF, Yeh ML, Tsai PC, Huang CI, Hsieh MH, Hsieh MY, Lin ZY, Chen SC, Huang 35 JF, Dai CY, Chuang WL. Time-Degenerative Factors and the Risk of Hepatocellular Carcinoma after Antiviral Therapy among Hepatitis C Virus Patients: A Model for Prioritization of Treatment. Clin Cancer Res 2017; 23: 1690-1697 [PMID: 27733478 DOI: 10.1158/1078-0432.CCR-16-0921]
- Huang CF, Yeh ML, Huang CY, Tsai PC, Ko YM, Chen KY, Lin ZY, Chen SC, Dai CY, Chuang 36 WL, Huang JF, Yu ML. Pretreatment glucose status determines HCC development in HCV patients with mild liver disease after curative antiviral therapy. Medicine (Baltimore) 2016; 95: e4157 [PMID: 27399135 DOI: 10.1097/MD.000000000004157]
- Hajarizadeh B, Grebely J, Dore GJ. Epidemiology and natural history of HCV infection. Nat Rev 37



Gastroenterol Hepatol 2013; 10: 553-562 [PMID: 23817321 DOI: 10.1038/nrgastro.2013.107]

- Liu XN, Cui DN, Li YF, Liu YH, Liu G, Liu L. Multiple "Omics" data-based biomarker screening for 38 hepatocellular carcinoma diagnosis. World J Gastroenterol 2019; 25: 4199-4212 [PMID: 31435173 DOI: 10.3748/wjg.v25.i30.4199]
- 39 Johnson PJ, Pirrie SJ, Cox TF, Berhane S, Teng M, Palmer D, Morse J, Hull D, Patman G, Kagebayashi C, Hussain S, Graham J, Reeves H, Satomura S. The detection of hepatocellular carcinoma using a prospectively developed and validated model based on serological biomarkers. Cancer Epidemiol Biomarkers Prev 2014; 23: 144-153 [PMID: 24220911 DOI: 10.1158/1055-9965.EPI-13-0870]
- 40 Huang ZH, Hu Y, Hua D, Wu YY, Song MX, Cheng ZH. Quantitative analysis of multiple methylated genes in plasma for the diagnosis and prognosis of hepatocellular carcinoma. Exp Mol Pathol 2011; 91: 702-707 [PMID: 21884695 DOI: 10.1016/j.yexmp.2011.08.004]



WJG

World Journal of Gastroenterology

Submit a Manuscript: https://www.f6publishing.com

World J Gastroenterol 2022 January 7; 28(1): 154-175

DOI: 10.3748/wjg.v28.i1.154

ISSN 1007-9327 (print) ISSN 2219-2840 (online)

SYSTEMATIC REVIEWS

Current guidelines for the management of celiac disease: A systematic review with comparative analysis

Alberto Raiteri, Alessandro Granito, Alice Giamperoli, Teresa Catenaro, Giulia Negrini, Francesco Tovoli

ORCID number: Alberto Raiteri 0000-0003-3051-8487; Alessandro Granito 0000-0002-0637-739X; Alice Giamperoli 0000-0003-3717-8104; Teresa Catenaro 0000-0001-5333-8089; Giulia Negrini 0000-0002-9261-2316; Francesco Tovoli 0000-0002-8350-1155.

Author contributions: Raiteri A and Tovoli F designed the research; Granito A, and Catenaro T performed the research; Raiteri A, Giamperoli A and Negrini G analysed the data; Raiteri A and Tovoli F wrote the paper.

Conflict-of-interest statement: The authors declare that they have no conflict of interest.

PRISMA 2009 Checklist statement:

The authors have read the PRISMA 2009 Checklist, and the manuscript was prepared and revised according to the PRISMA 2009 Checklist.

Country/Territory of origin: Italy

Specialty type: Gastroenterology and hepatology

Provenance and peer review: Invited article; Externally peer reviewed.

Peer-review model: Single blind

Peer-review report's scientific quality classification

Alberto Raiteri, Alessandro Granito, Alice Giamperoli, Teresa Catenaro, Giulia Negrini, Francesco Tovoli, Division of Internal Medicine, Hepatobiliary and Immunoallergic Diseases, IRCCS Azienda Ospedaliero-Universitaria di Bologna, Bologna 40138, Italy

Corresponding author: Francesco Tovoli, MD, Assistant Professor, Research Fellow, Division of Internal Medicine, Hepatobiliary and Immunoallergic Diseases, IRCCS Azienda Ospedaliero-Universitaria di Bologna, via Albertoni 15, Bologna 40138, Italy. francesco.tovoli2@unibo.it

Abstract

BACKGROUND

Wheat and other gluten-containing grains are widely consumed, providing approximately 50% of the caloric intake in both industrialised and developing countries. The widespread diffusion of gluten-containing diets has rapidly led to a sharp increase in celiac disease prevalence. This condition was thought to be very rare outside Europe and relatively ignored by health professionals and the global media. However, in recent years, the discovery of important diagnostic and pathogenic milestones has led to the emergence of celiac disease (CD) from obscurity to global prominence. These modifications have prompted experts worldwide to identify effective strategies for the diagnosis and follow-up of CD. Different scientific societies, mainly from Europe and America, have proposed guidelines based on CD's most recent evidence.

AIM

To identify the most recent scientific guidelines on CD, aiming to find and critically analyse the main differences.

METHODS

We performed a database search on PubMed selecting papers published between January 2010 and January 2021 in the English language. PubMed was lastly accessed on 1 March 2021.

RESULTS

We distinguished guidelines from 7 different scientific societies whose reputation is worldwide recognized and representative of the clinical practice in different geographical regions. Differences were noted in the possibility of a no-biopsy diagnosis, HLA testing, follow-up protocols, and procedures.

CONCLUSION



Grade A (Excellent): 0 Grade B (Very good): B, B, B, B Grade C (Good): C Grade D (Fair): 0 Grade E (Poor): 0

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is non-commercial. See: htt p://creativecommons.org/License s/by-nc/4.0/

Received: March 2, 2021 Peer-review started: March 2, 2021 First decision: July 14, 2021 Revised: August 8, 2021 Accepted: December 25, 2021 Article in press: December 25, 2021 Published online: January 7, 2022

P-Reviewer: Poddighe D, Sabelnikova EA, Sahin Y, Vasudevan A S-Editor: Wang LL L-Editor: A P-Editor: Wang LL



We found a relatively high concordance between the guidelines for CD. Important modifications have occurred in the last years, especially about the possibility of a no-biopsy diagnosis in children. Other modifications are expected in the next future and will probably involve the extension of the non-invasive diagnosis to the adult population and the follow-up modalities.

Key Words: Celiac disease; Gluten; Gluten-free diet; Gluten sensitivity; Clinical guidelines; Non-invasive diagnosis; Histopathological findings; Serological markers; Genetics

©The Author(s) 2022. Published by Baishideng Publishing Group Inc. All rights reserved.

Core Tip: Once considered a rare condition, celiac disease (CD) is becoming a significant health issue globally. An increasing number of studies have investigated this condition. International scientific societies have proposed guidelines for the management of CD to translate this evidence into clinical practice. In this review, we critically analyse both the converging and diverging points in the current clinical guidelines of CD, focusing on the diagnostic aspects and follow-up procedures.

Citation: Raiteri A, Granito A, Giamperoli A, Catenaro T, Negrini G, Tovoli F. Current guidelines for the management of celiac disease: A systematic review with comparative analysis. World J Gastroenterol 2022; 28(1): 154-175

URL: https://www.wjgnet.com/1007-9327/full/v28/i1/154.htm DOI: https://dx.doi.org/10.3748/wjg.v28.i1.154

INTRODUCTION

Celiac disease (CD) is an immune-mediated reaction to gluten characterised by an inflammatory injury to the small bowel in genetically predisposed subjects as a result of an inappropriate T cell-mediated immune response^[1]. The epidemiology of CD is well known, with an estimated worldwide prevalence of 0.6%-1% of the general population^[2]. However, CD remains largely underdiagnosed in developing countries and has a higher impact on children[3,4]. Simultaneously, the misdiagnosis of CD is becoming an emergent problem worldwide^[5].

An evidence-based approach is needed to optimise diagnostic accuracy to avoid lifethreatening complications (including small bowel carcinoma and lymphoma)[6] resulting from unrecognised CD on the one hand, and unnecessary cost burden and impact on the quality of life due to incorrect prescription of a life-long gluten-free diet (GFD) on the other hand.

Simultaneously, follow-up of patients with CD who are on a GFD is of critical importance to assess the responsiveness to the GFD, detect complicated CD, find associated autoimmune diseases, and identify metabolic alterations induced by the GFD[7].

Thus, an increasing number of scientific societies have proposed guidelines for diagnosing and managing CD. In our systematic review, we identified the most recent and significant national and international guidelines and compared their recommendations. We also underlined the most apparent differences among these guidelines to identify 'hot topics' on CD and possible future developments.

MATERIALS AND METHODS

The primary aim of this review was to identify the most recent national and international guidelines for CD by means of a systematic review and to compare their main recommendations.

We performed a database search on PubMed and selected papers published between January 2010 and January 2021 in the English language. PubMed was last accessed on 1 March 2021. The following keywords and terms were used: (1) Coeliac Diseaseor Celiac Disease; (2) Guideline; and (3) Management. The following string was used: (("coeliac disease"[All Fields] OR "celiac disease"[MeSH Terms] OR ("celiac"[All



Fields] AND "disease"[All Fields]) OR "celiac disease"[All Fields] OR ("coeliac disease"[All Fields] OR "celiac disease"[MeSH Terms] OR ("celiac"[All Fields] AND "disease"[All Fields]) OR "celiac disease"[All Fields])) AND ("guideline"[Publication Type] OR "guidelines as topic"[MeSH Terms] OR "guideline"[All Fields] OR ("manage"[All Fields] OR "managed"[All Fields] OR "managed"[All Fields] OR "manage"[All Fields] OR "managed"[All Fields] OR "managers"[All Fields] OR "managers"[All Fields] OR "manages"[All Fields] OR "managers"[All Fields] OR "managers"[All Fields] OR "managers"[All Fields] OR "managers"[All Fields] OR "management"[All Fields] OR "management"[All Fields] OR "managers"[All Fields] OR "managers"[All Fields] OR "managers"[All Fields] OR "managers"[All Fields] OR "management"[MeSH Terms] OR ("organization"[All Fields] OR "management"[All Fields] OR "disease management"[All Fields]))).

A total of 415 papers were identified with no duplicates, and, as a first step, no papers were excluded for other reasons (PRISMA flow diagram reported in Figure 1). However, twenty-one records were unavailable, leaving 396 papers for further evaluation. As a second step, we excluded papers that were not pertinent to any of the following criteria: (1) Clinical guidelines related to diagnosis and management of CD; and (2) Clinical guidelines published by governmental agencies and scientific associations. We included only the last version of the guidelines, excluding the previous updated versions.

According to the selection criteria, out of the 396 results of PubMed research assessed for eligibility, seven guidelines were finally included in this analysis. These guidelines strictly focus on the diagnosis and management of CD. These papers are presented in order of publication (newest to oldest): (1) European Society Paediatric Gastroenterology, Hepatology and Nutrition (ESPGHAN) 2020[8]; (2) European Society for the Study of Coeliac Disease (ECD) 2019[9]; (3) World Gastroenterology, Russia, 2016[11]; (5) National Institute for Health and Care Excellence (NICE), 2015 [12]; (6) British Society of Gastroenterology (BSG), 2014[13]; and (7) America College of Gastroenterology (ACG), 2013[14].

The recommendations provided by each selected guideline were systematically explored and classified into five categories: patients to be tested for CD, diagnostic tests (serology, duodenal biopsy, genetic test, no-biopsy diagnosis), potential/ silent/seronegative CD, refractory/complicated CD, and follow-up. These categories represent the most discussed topics of CD.

The results are reported in different paragraphs, containing both a brief introduction to the specific topic (with references derived from the supporting evidence used by the guidelines and other relevant papers according to a narrative approach) and a comparative analysis of the guidelines' recommendations (collected using a strictly systematic approach).

RESULTS

Clinical presentation and risk factors: who should be tested for CD?

CD is a diagnostic challenge as it may develop at any age (even in older adults) and with a polymorphic clinical presentation[15]. The clinical spectrum of CD includes both symptomatic and silent forms revealed only by serological screening[16,17]. CD-related symptoms can be both intestinal and extraintestinal, reflecting the systemic nature of the disease. These manifestations are classified as 'classical' and 'non-classical' according to the historical presentation of first described cases. Table 1 reports the main manifestations of CD according to their categorization[1,17-26].

Some guidelines draw specific attention to some extraintestinal symptoms (Figure 2). In particular, the ESsCD 2019 guidelines focus on oral-dental and neuropsychiatric manifestations[9]. CD testing is advised in cases of dental enamel defects and recurrent oral aphthae. Special attention to neurological manifestations has also been drawn by the Russian Central Research Institute of Gastroenterology[11]. These guidelines also focus on reproductive disorders, such as delayed sexual development, amenorrhea, infertility, and miscarriage[11].

Despite these premises, all the guidelines agree on testing for CD in children, adolescents, and adults showing classical and non-classical symptoms of CD[7-13]. There is also a consensus on considering iron-deficiency anaemia and hypertransaminasemia as the most common laboratory abnormalities[8-14].

Zaishidene® WJG | https://www.wjgnet.com

Table 1 Most frequent clinical manifestions of celiac disease		
	Intestinal	Extraintestinal
Classical	Diarroea	Iron deficiency anaemia
	Failure to thrive	Muscle waisting
	Weight loss	Oedema
	Bloating	
Non classical	Chronic abdominal pain	Short stature
	Abdominal distension	Delayed puberty
	Constipation	Amenorrhea
	Vomiting	Irritability, unhappiness
		Chronic fatigue
		Epilepsy
		Peripheral neuropathy
		Joint/muscle pain
		Elevated aminotransferases
		Aphtous stomatitis
		Recurrent miscarriages
		Reduced bone mineral density

The high-risk group of patients did not change over time. These groups include first-degree relatives of patients with CD, patients with autoimmune conditions (such as type 1 diabetes mellitus and thyroid diseases) or genetic disorders such as IgA deficiency, Down syndrome, Turner syndrome, and Williams-Beuren syndrome[8-14].

Diagnosis.

There is no 'gold standard' for the diagnosis of CD. Clinical features, serology, or histology alone cannot provide a definitive diagnosis. Instead, the final diagnosis of CD relies on a combination of these elements. All the guidelines agree on a sequential approach to diagnosis, consisting of serology as a first-line test in high-risk patients, followed by duodenal biopsy in cases of positive serology or persistent suspicion of malabsorption (Figure 3). A positive serology paired with evidence of duodenal villous atrophy indicate a definite CD diagnosis, whereas cases with discordant findings should undergo HLA testing. All the guidelines also agree that patients with dis-cordance between serology, histology, and HLA DQ2/DQ8 positivity should be evaluated on a patient-by-patient basis in expert centres. The so-called 'four-out-offive rule' has long been advocated as a standard of care [27]. According to this rule, four of the following criteria are sufficient to establish CD diagnosis: (1) Typical signs and symptoms (diarrhoea and malabsorption), (2) Antibody positivity, (3) HLA-DQ2 or HLA-DQ8 positivity, (4) Intestinal damage (*i.e.*, villous atrophy and minor lesions); and (5) Clinical response to GFD. This rule also helps physicians to identify various subtypes of CD, that is, non-classic CD (absence of point 1), seronegative CD (absence of point 2), potential CD (absence of point 4), and non-responsive CD (absence of point 5). However, the 'four-out-of-five rule' is yet to be recognised by any guideline.

We will report the guidelines' detailed suggestions for obtaining key diagnostic elements from serology, histology, and genetic testing in the following paragraphs.

Serology

All diagnostic serological testing should be performed in patients on a glutencontaining diet[28]. Serum immunoglobulin A(IgA) anti-tissue transglutaminase antibody (anti-tTG-IgA) is widely accepted as the most sensitive test for CD diagnosis, although it suffers from low specificity, especially at low titres [29-33]. In contrast, IgA anti-endomysial antibodies (EMA-IgA) are nearly 100% specific for CD but are less sensitive, more expensive, and more operator-dependent than anti-tTG-IgA. Therefore, these characteristics make EMA-IgA an ideal second-line test[34]. The diagnostic performance of both anti-tTG-IgA and EMA-IgA is limited in patients with concurrent

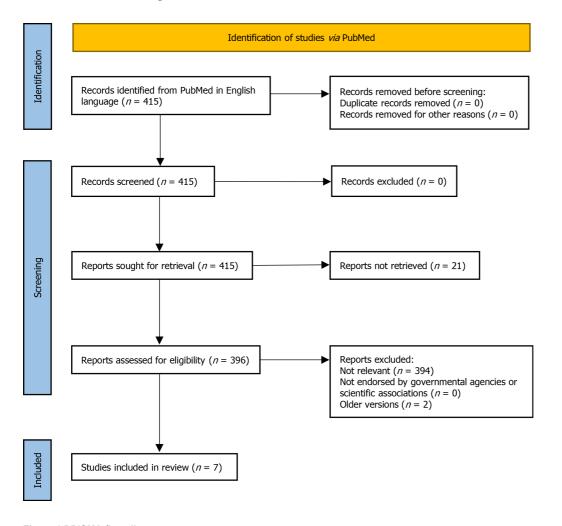


Figure 1 PRISMA flow diagram.

IgA deficiency. Antibodies to deamidated gliadin peptides (DGP) of the IgG class are advantageous in this setting and for younger children[35,36]. Even with the most recent advancements in CD serology, up to 2% of patients with CD have no circulating markers of gluten sensitivity, defining a condition of seronegative CD[37].

Currently, the guidelines are concordant and suggest anti-tTG-IgA as the initial serological test, complemented by a determination of total IgA levels to rule out concurrent IgA deficiency (Figure 4)[8-14]. This initial approach was suggested for both children and adults.TheACG2013 guidelines suggest a combination of different IgA and IgG antibodies in children younger than two years of age (for instance, anti-tTG IgA and DGP-IgG)[14]. This approach is still accepted only by the WGO2017 guidelines[10]. The remaining guidelines advise against this strategy, as a combination of antibodies implies a higher sensitivity at the expense of a reduced specificity, often leading to the necessity of histological confirmation. This scenario represents an obstacle in the pursuit of a no-biopsy approach in children, for whom the anti-tTG-IgA + total IgA strategy fits better[8]. Alternatively, DGP-IgG (together with anti-tTG-IgG) maintained the unanimous recommendation as the test of choice in patients with IgA deficiency[8-14].

Further, EMA-IgA is considered a confirmatory test, particularly when TG2 has a low titre, *i.e.*, < 2x the upper normal limit (UNL)[9,10,12]. A positive result is also required for a no-biopsy CD diagnosis in children with anti-tTG IgA > 10x[8]. However, the use of paired anti-tTG and EMA-IgA as the first diagnostic test is not supported by any guideline.

Currently, all of the guidelines strongly discourage urine, stool, and saliva tests in clinical practice due to their low-performances[8-14] and the consequent risk of initiating a GFD without a firm diagnosis, impacting the final diagnosis[13].

Biopsy

For a long time considered the 'gold standard' for diagnosing CD (ambiguously suggesting that other tests were of lesser importance), duodenal biopsies remain the



	Change over time
<u>*** *** *** *** ***</u>	No major changes over time
<u>*** *** *** ***</u> ***	No major changes over time
<u>***</u> *** *** ***	No major changes over time
Hepatology and Nutrition (ESPGHAN) 2020 (8).	
	می م

1	European Society for the Study of Coeliac Disease (ECD) 2019 (9).
<u></u>	World Gastroenterology Organization (WGO) 2017 (10).
1 %*	Central Research Institute of Gastroenterology, Russia, 2016 (11).
y	National Institute for Health and Care Excellence (NICE), 2015 (12).
yy'e	British Society of Gastroenterology (BSG), 2014 (13).
	America College of Gastroenterology (ACG), 2013 (14).

Figure 2 Recommendations about case finding.

mainstay of CD diagnosis, and all guidelines unanimously recognise this role. The presence of positive histology, however, was not considered CD-specific. Thus, clinical, and serological correlations are mandatory (Figure 5) [8-14].

Duodenal biopsies should be obtained from all patients with suspected CD. In highrisk symptomatic patients, duodenal biopsies should be performed irrespective of serology results for CD[9,13,14]. Some authors also suggested that duodenal biopsies should be considered in any individual undergoing endoscopy because of the relatively high prevalence of CD in the general population and its polymorphic presentation^[13].

Histology samples should be collected from multiple sites, given the possible patchy distribution of CD lesions. Current evidence suggests collecting four biopsies from the second duodenal portion and two biopsies from the bulb[38]. Biopsy sample orientation using cellulose acetate Millipore filters is of paramount importance to avoid artefacts, potentially leading to a false diagnosis of villous atrophy[39].

The histological findings are currently categorised according to the classification proposed by Marsh and subsequently modified by Oberhuber[40]. Pathology findings are reported as Marsh-Oberhuber 0 (normal histology), 1, 2, or 3 (subdivided into 3a, 3b, and 3c).

An increase in intraepithelial lymphocytes (IELs) without villous atrophy defines Marsh 1 Lesion. In most cases, Marsh 1 Lesions (also called minimal lesions) are attributable to other causes, including lymphocytic colitis, bacterial and parasitic intestinal infections (especially Helicobacter pylori and Giardia lamblia), small intestinal bacterial overgrowth, Crohn's disease, common variable immunodeficiency, and non-steroidal anti-inflammatory drugs[41]. While a Marsh 1 Lesion is not considered sufficient to diagnose CD, the BSG 2014 guidelines state that minimal lesions combined with positive serology could represent a probable CD. A trial with a GFD could be considered to support the diagnosis of CD[13]. When the increase in IELs is paired with hyperplasia of the duodenal crypts, the lesion is classified as Marsh 2.



Raiteri A et al. Celiac disease guidelines

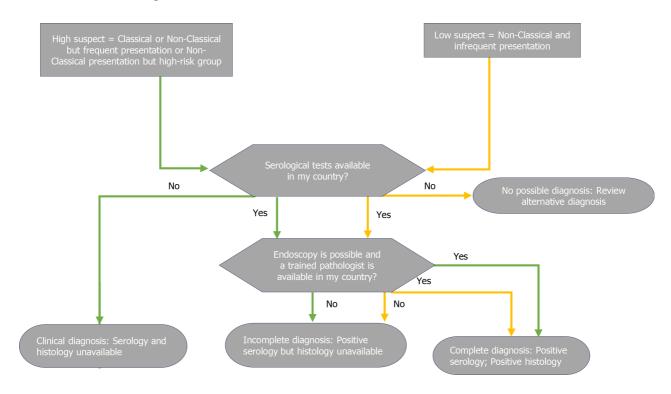


Figure 3 Worldwide adapted decision-making process for diagnosing celiac disease. Highly suspicious celiac disease (CD) comprises "classical presentation" (*i.e.*, classical symptoms in children include failure to thrive, weight loss, growth failure, vomiting, chronic diarrhea, bloating, Iron-deficiency anemia, muscle wasting, oedema due to hypoproteinemia, irritability and unhappiness; in adults, classical symptoms include chronic diarrhea, weight loss, iron-deficiency anemia, malaise and fatigue, oedema due to hypoproteinemia, and osteoporosis), frequent "non-classical presentation" (*i.e.*, iron deficiency and hypertransaminasemia) and "non-classical presentation" but high risk group (*i.e.*, CD first-degree relatives, autoimmune conditions such as type 1 Diabetes Mellitus, and thyroid disease, genetic conditions such as IgA deficiency, Down syndrome, Turner syndrome and Williams-Beuren syndrome).

Conversely, increased IELs in combination with villous atrophy define the typical CD lesion (Marsh 3), subclassified as mild (3a), moderate (3b), or subtotal (3c)[40]. Some authors proposed a simplified histopathological grading, reducing the possible grades from five to three, thus reducing the possible inter-operator variability in the histological interpretation[42]. This simplified classification is yet to be adopted by the international guidelines, which currently recommend the Marsh-Oberhuber classification[8-14].

At present, there is no alternative to duodenal biopsy for examining mucosal damage[8-14]. For instance, in children, video-capsule endoscopy (VCE) gives no indications[8], although in adults, VCE could support the diagnosis in cases of discordance between serology and biopsy^[13] or if the patient is unwilling or unable to undergo traditional endoscopy[14]. VCE could also play a role in detecting CD complications (*i.e.*, lymphoma, adenocarcinoma, ulcerative jejunitis)[9] and in helping to differentiate extended diseases (e.g., CD vs proximal Crohn's disease)[11]. Anti-actin IgA antibodies have been shown to be predictive of severe villous atrophy in CD patients at the time of diagnosis[43]. Theoretically, they may also provide indirect information about villous recovery following the introduction of the GFD; however, data are still lacking in this setting. The available information about faecal and salivary microbiome, at present, is not sufficient to allow a reliable conclusion for the diagnosis of CD[44,45]. Intestinal fatty-acid binding protein (I-FABP) are higher in dietary nonadherence and unintentional gluten intake and could be used as a sensible blood marker of mucosal damage[46,47]. This exam was first mentioned in the ESsCD guidelines[9].

A repeated small intestinal biopsy, including biopsies from the jejunum, could be considered in adults with discordance between histopathology and anti-tTG-IgA results[13]. In children, re-cutting biopsies and/or a second opinion from an experienced pathologist is preferred over endoscopic repetition[8].

In adults, a gluten challenge should be proposed for patients with uncertain CD diagnosis, who have been started on a GFD[9-14]. In children, gluten challenge is discouraged before the age of 5 years and during puberty, and in general, it should be reserved for unusual cases[8].

Recon	nmendation			Change over time
be used	sue Transglutaminase 2 IgA (TGA-IgA) should d as the initial serological test, complemented l IgA value in children of any age and adults	و 💥 💥 💥 💥	· · · · · · · · · · · · · · · · · · ·	Changes over time
based t	ents with low total IgA concentrations, an IgG- test, preferably TGA-IgG or DGP-IgG, should be ned as a second step	<u> ** ** ** **</u>	× ** **	No major changes over time
address	egy based on a combination of antibodies sing the same target (<i>i.e.</i> , TGA-IgA and EMA- a first approach is not recommended	%* %* %* %* %		No major changes over time
**	European Society Paediatric Gastroenterology, I	Hepatology and Nutrition (ESPGHAN) 20	20 (8).	
	European Society for the Study of Coeliac Disea	se (ECD) 2019 (9).		
<u>*</u> *	World Gastroenterology Organization (WGO) 20	17 (10).		
<u>}</u>	Central Research Institute of Gastroenterology,	Russia, 2016 (11).		
<u>,</u>	National Institute for Health and Care Excellence	e (NICE), 2015 (12).		
**	British Society of Gastroenterology (BSG), 2014	(13).		
<u>}</u>	America College of Gastroenterology (ACG), 20	13 (14).		

Figure 4 Recommendations about serology. IgA: Immunoglobulin A; IgG: Immunoglobulin G; DGP: Deamidated gliadin peptides; EMA: Anti-endomysium antibodies.

> Gluten challenge protocols are not homogeneous. A diet containing at least 10 g of gluten per day for 6-8 wk seems to be the most effective way to achieve disease relapse; however, the evidence is weak[28]. In shorter protocols, a diet containing at least 3 g of gluten per day for at least 2 wk seems to be sufficient for most patients[10, 13,14]. Certainly, a shorter and lighter approach would fit better for highly symptomatic patients. A strategy for optimising the result would be to undergo a serology test after two weeks and, if negative, to extend the challenge to 8 wk[13].

> After reintroducing gluten, physical symptoms should not be used for diagnosis in the absence of other supportive evidence[8,9,11-14]. A diagnosis based only on the disappearance of symptoms on GFD and relapse during gluten re-introduction can be relevant in geographic areas where serology tests are not available, as the only way to confirm the diagnosis and treat the disease[10].

Human Leukocyte Antigen testing

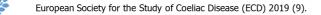
The strong genetic component of CD is testified by its high familial recurrence and high disease concordance among monozygotic twins (75%-80%)[48]. The presence of human leukocyte antigen (HLA) -DQ2/DQ8 is a pathogenic requisite for the development of the typical immune alterations found in CD. Simultaneously, HLA DQ2/DQ8 can be found in up to 30%-40% of the general population, so its specificity is remarkably poor^[49]. In contrast, the absence of HLA DQ2/DQ8 virtually excludes CD diagnosis[48,49].Restricting this observation to the sole HLA DQ2 alleles, a recent systematic review of the literature confirmed that only 5.06% of patients with CD were completely lacking the HLA-DQB1*02 allelic variant[50].

Consequently, all the guidelines advise against using HLA testing as a first-line tool for the diagnosis of CD (Figure 6)[8-14]. They are also concordant in allocating this resource for: (1) Patients with uncertain diagnosis of CD, already on a GFD; (2) Patients with a flat intestinal mucosa but negative serology; and (3) In patients already on a GFD, serology and histology can be inconclusive. In this context, before embarking on a so-called 'gluten-challenge', it is advisable to verify the presence of HLA-DQ2/DQ8[8-14].



Recommendation		Changes over time
Adult patients with a positive serology must undergo endoscopy with duodenal biopsies to achieve a final diagnosis	<u>منار منار منار منار منار منار منار منار</u>	With exceptions
In children, in precise conditions, diagnosis can be achieved without a duodenal biopsy	<u>***</u> *** *** ***	Major changes over time
Duodenal biopsy should be performed, irrespective of positive serology for CD, in case of high clinical suspicion of CD		No major changes over time
At least 4 biopsies from the distal duodenum and at least 1 from the duodenal bulb should be taken for histology assessment during a gluten-containing diet	<u>***</u> *** *** *** ***	No major changes over time
The diagnosis is confirmed in the presence of Marsh \geq 2 lesions. Marsh 1 is not sufficient to diagnose CD	<u>***</u> * <u>**</u> * <u>**</u> * <u>**</u>	No major changes over time. Minor exceptions
A gluten challenge should be proposed to patients who have been started on a GFD but have a doubtful diagnosis	<u>** ** ** **</u> **	Minor changes over time and guidelines
A diagnosis based only on the disappearance of symptoms on GFD and relapse during gluten re- introduction is absolutely discouraged.	<u>میں میں میں میں میں میں میں میں میں میں </u>	With exceptions
No exams can surrogate mucosal damage without biopsy	<u>**</u> ** ** ** **	No major changes over time. Minor exceptions

European Society Paediatric Gastroenterology, Hepatology and Nutrition (ESPGHAN) 2020 (8).



World Gastroenterology Organization (WGO) 2017 (10).



National Institute for Health and Care Excellence (NICE), 2015 (12).

British Society of Gastroenterology (BSG), 2014 (13).

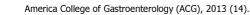


Figure 5 Recommendations about serology.

HLA tests would be useless for patients with positive serology before a glutenchallenge because virtually 100% of those patients would be positive. Therefore, HLA typing is no longer a criterion for the 'no-biopsy' approach of diagnosis in children with a TGA-IgA > 10x UNL[8]. In patients with positive histology (i.e., villous atrophy, though occasionally detected on esophagogastroduodenoscopy), and negative or questionable serology, HLA testing can exclude the diagnosis of CD[9]. In contrast, a positive result cannot confirm the diagnosis, which should be carefully evaluated on a patient-by-patient basis in expert centres.

The use of HLA typing in high-risk populations is controversial. HLA-DQ2/DQ8 can be found in more than 50% of first-degree relatives of patients with CD and in patients with other autoimmune or genetic disorders related to CD[14,49]. Most



1/2

Recommendation			Changes over time
HLA -DQ2/DQ8 testing has only a high negative predicting value and is recommended in selected patients to rule out coeliac disease in patients: Already on a gluten-free diet		<u>** ** ** ** ** **</u>	No major changes over time
histology	egative or questionable serology but positive ,		
	2/DQ8 testing is not recommended alone or d with serology tests to confirm the diagnosis	<u>** ** ** ** ** **</u>	No major changes over time
HLA) -DQ2/DQ8 testing in high-risk populations can indefinitely exclude these patients from a periodic screening		<u>اللہ میں میں بڑ بڑ بڑ</u>	Minor changes over time
<u></u>	European Society Paediatric Gastroenterology	Hepatology and Nutrition (ESPGHAN) 2020 (8).	
	European Society for the Study of Coeliac Disease (ECD) 2019 (9).		
<u>, , , , , , , , , , , , , , , , , , , </u>	World Gastroenterology Organization (WGO) 2017 (10).		
,,,,,	Central Research Institute of Gastroenterology, Russia, 2016 (11).		
,,,, ,,	National Institute for Health and Care Excellence (NICE), 2015 (12).		
<u>, , , , , , , , , , , , , , , , , , , </u>	British Society of Gastroenterology (BSG), 2014 (13).		

1/2 America College of Gastroenterology (ACG), 2013 (14).

Figure 6 Recommendations about Human Leukocyte Antigen testing.

guidelines suggest excluding HLA-DQ2/DQ8 in CD first-degree relatives and highrisk patients, even if asymptomatic, to avoid periodic monitoring[9,10,13,14]. This strategy can be questioned in terms of resources and costs[10,11,14]. Some authors hav esuggested screening high-risk patients only if they complain of gastrointestinal or extraintestinal symptoms or have laboratory abnormalities[11]. In addition, a two-step genetic screening procedure starting with HLA-DQ β chains has been proposed[51]. Thus, the choice of screening for symptomatic or asymptomatic first-degree relatives or high-risk patients, with or without a preliminary determination of HLA-type, remains debated, needing to take local resources and cost-benefit rates into account.

No-biopsy diagnosis

While most guidelines allow a no-biopsy diagnosis in children under strict conditions, endoscopy with duodenal biopsies is still mandatory to achieve a final diagnosis of CD in adults[9-14]. As the only exception, the WGO guidelines allow a diagnosis based on serology and clinical response to the GFD (Figure 7) in developing countries where endoscopy may not possible or trained pathologists may not be available[10].

The ESPGHAN2012 guidelines endorsed the possibility of a no-biopsy approach in children for the first time. This possibility was limited to certain conditions, which included the presence of classic symptoms, with tTG-IgA > 10x UNL, EMA-IgA positivity, and presence of permissive HLA[8].

This approach was subsequently adopted by a plurality of international guidelines [9-12]. although, the ACG2013 and BSG 2014 guidelines did not include this approach [13,14].

The 2020 update of the ESPGHAN guidelines removed classic symptoms, EMA-IgA positivity, and HLA DQ-2 or DQ-8 as crucial criteria for a diagnosis not based on biopsy[7]. However, EMA-IgA positivity is not discouraged[8,10]. The increasing confidence in diagnosing CD without biopsy in children has increased so rapidly that many recent studies consider tTGA > 10x as a new possible cut-off to further reduce the need for biopsies[52].

Recommendation		Changes over time
In children with classic symptoms, TGA-IgA titre > 10x, EMA-IgA positivity, and HLA DQ2/DQ8, the diagnosis can be achieved without a duodenal biopsy	<u>** ** **</u>	Major changes over time
In children, classic symptoms, EMA-IgA positivity and HLA DQ2 /DQ-8 are not mandatory to diagnose CD if TGA-IgA titre is > 10x	<u>,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,</u>	Major changes over time
In adults, a diagnosis of CD without a positive biopsy is still discouraged	<u>***</u> *** **** ****	Exception Probable future changes over time

	European Society Paediatric Gastroenterology, Hepatology and Nutrition (ESPGHAN) 2020 (8).
11/2	European Society for the Study of Coeliac Disease (ECD) 2019 (9).
<u>y</u> **	World Gastroenterology Organization (WGO) 2017 (10).
<u>,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,</u>	Central Research Institute of Gastroenterology, Russia, 2016 (11).
,,,,,	National Institute for Health and Care Excellence (NICE), 2015 (12).
<u>}</u>	British Society of Gastroenterology (BSG), 2014 (13).
	America College of Gastroenterology (ACG), 2013 (14).

Figure 7 Recommendations about the possibility of a no-biopsy diagnosis. TGA: Anti-transglutaminase antibodies; IgA: Immunoglobulin A; EMA: Antiendomysium antibodies; HLA: Human leukocytes antigen; CD: Celiac disease.

> CD diagnosis without a positive duodenal biopsy has always been discouraged in adults[9-14]. This choice was not dictated by the reduced reliability of the serological tests in adults. In fact, large population studies concluded that tTG-IgA>10x could accurately predict villous atrophy[53]. Rather, other considerations currently prevent the extension of paediatric criteria into the adult population. First, CD at onset can be associated with complications. In the case of primary or secondary resistance, or slow response to the GFD, the absence of baseline histology may make the diagnosis of complications difficult[9]. Index histology may also predict the risk of future complications, such as lymphoma[54]. Moreover, endoscopy may help diagnose other treatable disorders associated with CD, such as eosinophilic esophagitis, autoimmune gastritis, and lymphocytic gastritis[9].

> Both complicated CD and possible differential diagnoses of CD are virtually absent in children. However, they represent a serious concern in adults, thus justifying different diagnostic algorithms according to the age of presentation of the first symptoms.

Potential, silent and seronegative CD

Potential CD is characterised by a positive serology for CD in the absence of mucosal damage at biopsy[1]. As stated above, Marsh 1 Lesions (*i.e.*, an increased IELs count) are not suggestive of an active CD but may increase the risk of developing villous atrophy[41].

It is widely accepted that symptomatic potential CD may benefit from a GFD, and a direct challenge would be run[8-14]. In adult patients with both positive TGA-IgA and EMA-IgA CD is likely, and a GFD may be initiated irrespective of symptoms[9]. A serological response after a period of approximately 12 mo confirms the diagnosis of CD[9]. In EMA-IgA negativity, HLA-typing may exclude the diagnosis before embarking on follow-up[9]. If a follow-up is started, potential CD patients should be



retested after consuming a gluten-containing diet for 3-6 mo to confirm persistent seropositivity before referral for a new endoscopy (Figure 8)[9,10].

Silent CD is characterised by the presence of both positive serology and histology for CD in the absence of classical or non-classical symptoms^[1]. It is widely recommended to start a GFD in patients with silent CD because it is considered an active form of the disease[8-14].

Seronegative CD is characterised by the presence of active enteropathy and negative serology for CD, with no other causes, and with clinical and histological responses to a GFD[1,37]. In these cases, other causes of enteropathy should be excluded before embarking on the direct challenge of a GFD[37,55]. HLA-typing can also rule out the diagnosis of CD in seronegative enteropathies [9,14,37]. Finally, the direct challenge of a GFD is advised only in patients with seronegative enteropathy, positive HLA typing with no other causes. A documented histological response after 1-3 years of GFD is needed to confirm the diagnosis[9,14,37]. No major changes occurred over time in the management of seronegative CD[9,14].

Refractory and complicated CD

CD can be complicated by a persistent active form of the disease, independent of gluten intake, known as refractory CD (RCD)[1]. Other rare complications of CD can be neoplastic. Primarily, enteropathy-associated T-cell lymphoma (EATL) is a rare Tcell lymphoma associated with untreated CD. EATL has an abysmal prognosis and can occur primarily at diagnosis or as an evolution of RCD type 2[56]. Duodenal adenocarcinoma is possible, albeit less frequent in the CD population[57].

Refractory CD (RCD) is characterised by the persistence or recurrence of symptoms and signs of malabsorption, with documented villous atrophy, despite a strict GFD for more than 12 mo and in the absence of other causes [9-14]. No major changes occurred in this definition over time (Figure 9).

RCD can be primary (refractory at the time of the first diagnosis), or secondary (occurring after a period of response to the GFD)[1]. The first step in evaluating suspected RCD is to re-evaluate the initial diagnosis of CD by reviewing biopsies and serology tests obtained at the time of diagnosis[58]. The most common cause of GFD failure is inadvertent gluten ingestion [59]. Therefore, evaluation by an expert dietitian should always be included [9,10,13,14]. Other associated or concomitant pathological conditions should be excluded before RCD diagnosis. These include lactose and fructose intolerance, small intestinal bacterial overgrowth, microscopic colitis, pancreatic insufficiency, and inflammatory bowel diseases[59,60]. All guidelines recommend this strategy[9,10,13,14].

RCD is further classified into type 1 (RCD-1) and type 2 (RCD-2)[1]. T-cell flow cytometry is the most reliable method for classifying RCDs. Aberrant T cells lose the normal surface markers CD3 and CD8 with preserved expression of intracytoplasmic CD3. In RCD-1, the percentage of aberrant T cells is below 20%, whereas in RCD-2, they represent more than 20% of the total IELs [58]. RCD-2 can be considered a prelymphoma or low-grade lymphoma^[54]. T-cell receptor (TCR) g chain clonality analysis lacks sensitivity and specificity, and is of limited value in separating RCD-1 from RCD-2[54]. TCR analysis has been formerly indicated as a criterion for differentiating RCD-1 from RCD-2[11,13,14]. The latest ESsCD guidelines exclude TCR analysis in the RCD classification[9].

RCD-1 has an extremely high 5-year survival rate (> 90%)[54,59,60]. In RCD-1, the first-line therapy should be 'open-capsule' budesonide (OCB), 3 mg, 3 times a day [61]. Budesonide (open capsule or not) has been progressively accepted as the first-line therapy for RCD-1[9,11,13,14]. In the ACG 2013 guidelines, systemic steroids are considered the first-line therapy for RCD-1[14]. Second-line treatment for RCD-1 includes immunosuppressive drugs such as steroids (prednisone 0.5-1 mg/kg/day) and azathioprine (2-2.5 mg/kg/day)[60]. Most guidelines agree with this strategy [9,11, 12]. Systemic steroids can also be considered as first-line treatment while waiting for a specialist's advice[12]. Infliximab may be the preferred biological therapy for secondline treatment of RCD-1[62]. Evidence is still weak, and only one guideline includes infliximab as an RCD-1 treatment[9].Withdrawing of immunosuppressive therapy after 2-3 years of complete response may be considered[9,54].

RCD-2 is rarer than RCD-1, has a much higher mortality rate, and treatment is less well defined. Systemic steroids or open-capsule budesonide should be the first choice for milder presentations. In severe cases, cytoreductive therapies such as cladribine and fludarabine or autologous hematopoietic stem cell transplantation should be chosen[59,60]. Guidelines are mostly aligned with this strategy[9,13,14]. Some guidelines also report azathioprine, 6-mercaptopurine, methotrexate, cyclosporine, and anti-TNF antibodies as possible therapies, but the data are weaker[11,13,14]. Not every

Recommendation		Changes over time
Children and adults with symptomatic potential CD responding to GFD may be considered CD patients, despite the absence of villous atrophy	<u>**</u> ** ** ** **	Minor changes over time. Extensions
Children and adults with silent CD are considered CD patients and must be treated	<u>**</u> ** ** ** **	No major changes over time
In adults, a seronegative CD diagnosis can be achieved with a direct challenge of a GFD in patients with villous atrophy with no other causes, negative serology tests and positive HLA typing. A follow-up biopsy of confirmation is required	<u>** ** ** **</u>	No major changes over time

	European Society for the Study of Coeliac Disease (ECD) 2019 (9).
<u></u>	World Gastroenterology Organization (WGO) 2017 (10).
<u></u>	Central Research Institute of Gastroenterology, Russia, 2016 (11).
	National Institute for Health and Care Excellence (NICE), 2015 (12).
!	British Society of Gastroenterology (BSG), 2014 (13).
	America College of Gastroenterology (ACG), 2013 (14).

Figure 8 Recommendations about potential, silent, and seronegative celiac disease. GFD: Gluten-free diet; HLA: Human leukocytes antigen.

guideline has raised the topic of RCD-2 treatment[10-14].

Transformation to enteropathy-associated T-cell lymphoma (EATL) is likely in RCD-2[59]. VCE, positron-emission tomography (PET), and magnetic resonance (MR) enterography can be useful in cases of suspected progression to EATL to assess the extent of the disease^[63]. All guidelines advise the use of these tools in RCD-2 staging [9-14]. Severe RCD-2 and EATL may require surgery, chemotherapy, or bone marrow transplantation[64]. The former therapeutic strategies are mostly based on case reports, and only one guideline extensively discusses them[9].

Follow-up

Since CD is the only autoimmune disease with a known environmental trigger (*i.e.*, gluten), a periodical assessment of compliance to a GFD is essential [65]. Poor GFD compliance is not infrequent, and mucosal damage can persist despite negative serology and the absence of symptoms [66]. Follow-up is also essential for evaluating possible complications[54]. Osteoporosis and metabolic complications of GFD should also be evaluated during follow-up[67-69]. Suggested follow-up schedules are based on the frequency of complications, risk of GFD non-compliance, and reported quality of life[70].

Therefore, there is universal agreement on the necessity of long-term monitoring of patients with CD to assess the compliance and responsiveness to the GFD and allow early detection of complicated CD (Figure 10)[8-14]. Follow-up evaluations should be scheduled every 3-6 mo during the first year and then every 1-2 years[9-14]. In children, follow-up should continue until they reach their final height[9-11,14], focusing on normal growth and development[9,10,14].

There is disagreement about who should oversee follow-up. While most guidelines show no preference between primary care physicians, specialists, or dietitians[9-11,13, 14], the NICE 2015 guidelines suggest that dietitians with expertise in CD may be best suited to carry out an annual follow-up[12]. However, on a general principle, all guidelines agree that newly diagnosed patients should be referred to a dietitian[9-14]. Some guidelines suggest that nutritionist counselling should coincide with medical visits during follow-up[10,13]. The inclusion of a dietitian assessment at diagnosis and



Recommendation		Change over time
Slow-responder CD is defined as the persistence of symptoms, signs and laboratory abnormalities despite at least 6–12 months of GFD. This term replaces the former "non-responsive CD"		Major change over time
Refractory CD is defined as the persistence/recurrence of malabsorption, with documented villous atrophy, despite a strict GFD for > 12 mo and absence of other causes	<u>** ** ** ***</u> ***	No major changes over time
T-cell flow cytometry is the most reliable method for classification refractory CD	<u></u>	Major changes over time
TCR-gamma chain clonality analysis lacks sensitivity and specificity, and it is of limited value	1 %*	Major changes over time
Budesonide is recommended as first-line therapy for refractory CD type 1		Major changes over time
Second-line treatments for refractory CD type 1 includes steroids, azathioprine and infliximab	<u>***</u> * <u>**</u>	Major changes over time
Therapy for refractory CD type 2 is not supported by strong clinical data	<u>اللہ اللہ اللہ اللہ اللہ اللہ اللہ اللہ</u>	Major changes over time

1	European Society for the Study of Coeliac Disease (ECD) 2019 (9).
<u></u>	World Gastroenterology Organization (WGO) 2017 (10).
11/2	Central Research Institute of Gastroenterology, Russia, 2016 (11).
11/2	National Institute for Health and Care Excellence (NICE), 2015 (12).
<u>,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,</u>	British Society of Gastroenterology (BSG), 2014 (13).
)	America College of Gastroenterology (ACG), 2013 (14).

Figure 9 Recommendations about refractory and complicated celiac disease. GFD: Gluten-free diet; TCR: T-cell receptor.

during follow-up was supported by clinical data^[71]. Indeed, nutritional counselling could also help manage metabolic alterations, which frequently appear during the first years of the GFD[67].

All guidelines also provide information about the essential information that should be collected during follow-up evaluations. These evaluations should include a dietary interview, serology (TTG-IgA if normal IgA), and laboratory tests[9-14]. Laboratory tests should evaluate the presence of micronutrients malabsorption, including complete blood count, iron status, folate, vitamin B12, calcium, phosphate, vitamin D, and should monitor associated autoimmune conditions (thyroid-stimulating hormone and serum glucose) and liver disorders (aspartate aminotransferase/alanine aminotransferase)[9-11,13,14]. Normalisation of tTG-IgA levels do not predict full recovery of villous atrophy. In contrast, persistently positive serology 12 mo after GFD initiation is a strong indicator of gluten ingestion[72]. All guidelines were aligned with the interpretation of tTG-IgA levels during follow-up[8-14].

The inability of serology alone to predict mucosal healing automatically leads to consider the opportunity of repeating duodenal biopsies after the start of the GFD. While the general agreement is that follow-up biopsies are not mandatory in



Recommendation		Changes over time
In adults, follow-up should be scheduled every 3-6 month during the first year and then every 1-2 yr	** ** ** ** **	No major changes over time
A normal TGA level at the follow-up does not predict recovery of villous atrophy	<u>** ** ** ** **</u>	No major changes over time
On the contrary, persistently positive serology 12 mo after starting a GFD strongly suggests gluten contamination	%* <u>%</u> * %* %* %*	No major changes over time
The follow-up should include at least a dietary interview, serology, and laboratory tests evaluating absorption.	*** *** **** **** ****	No major changes over time
Follow-up biopsy is not universally recommended but may be reasonable after 2 yr of GFD in high-risk patients	*** *** *** *** ***	Minor changes over time
In children, follow-up should be scheduled every 3-6 mo during the first year and then every year until the end of development	<u>***</u> *** ***	No major changes over time
Newly diagnosed patients should be referred to a dietitian for management	****	Minor differences
Primary care physicians or dietitians with experience in dealing with CD may take responsibility for the follow- up	*** ***	Some differences
Follow-up should also include periodical bone densitometry, vaccinations and psychological support	تنیکو تنیکو تنیکو تنیکو تنیکو	Some differences

European Society Paediatric Gastroenterology, Hepatology and Nutrition (ESPGHAN) 2020 (8).



World Gastroenterology Organization (WGO) 2017 (10).

- Central Research Institute of Gastroenterology, Russia, 2016 (11).
 - National Institute for Health and Care Excellence (NICE), 2015 (12).
 - British Society of Gastroenterology (BSG), 2014 (13).

America College of Gastroenterology (ACG), 2013 (14).

Figure 10 Recommendations about follow-up of celiac disease. TGA: Anti-transglutaminase antibodies; GFD: Gluten-free diet.

asymptomatic patients on a GFD and without an increased risk of complications[9-14], the guidelines diverge regarding other points. Many guidelines consider it reasonable to repeat biopsy after 2 years of GFD to assess mucosal healing[9,11,14]. Other guidelines suggest repeating biopsies only for persistent symptoms or serological abnormalities after 12 mo of GFD[10,12,13]. A growing body of literature suggests that the risk of a complicated CD is higher in patients >40 years of age at the time of diagnosis or those with a classical presentation[54]. Some guidelines agree that repeating biopsies should be of interest in these selected populations[13,14].

Some guidelines also provide suggestions for further examinations to be performed during follow-up. According to the ECD and Russian guidelines, bone densitometry should be offered to every patient at the time of diagnosis and should be repeated after 3 years if abnormal, or 5 years if normal[9,11]. Other guidelines suggest performing



<u>,</u>

bone densitometry only in patients with a high risk of osteoporosis or those older than 55 years[12,13].

While there is a general agreement in recommending a pneumococcal vaccine[8-10, 12], the WGO2017 guidelines also recommend vaccinations against *Haemophilus influenzae typeB*, and *Meningococcus*, while other guidelines state that these vaccines have a less clear indication to be given to every patient with CD[9,11-13].

Mood disorders are another common problem in patients with dietary restrictions. Anxiety, depression, and fatigue may be associated with CD before and after diagnosis and can affect the quality of life^[73]. In this context, most guidelines agree on advising patients to join CD support groups and associations[9,10,12,13]. Some of them also suggest that psychological support provided by a specialist may be offered [12,13].

Gluten-free diet

Gluten is a protein with high proline and glutamine content, primarily found in wheat. Rye and barley belong to the same tribe as wheat and are known to contain gluten. In contrast, oats are derived from a different tribe and do not contain pure gluten[1].

Uncontaminated oats are safe for almost all patients with CD, but a small percentage of patients may be sensitive to some oat cultivars^[74] and should be monitored[9,10,12-14]. Some guidelines advise the initiation of a Gluten-free diet (GFD), excluding oats, and recently introduced them [10,13,14]. The Russian guidelines (2016) are against oat consumption in patients with CD because of the high risk of contamination[11]. Even if not stated, oat consumption would be safe in many countries, though it may be discouraged in developing countries where contamination could be widespread (Figure 11).

WHO guidelines on 'Standard for Foods for Special Dietary Use for Persons Intolerant to Gluten' state that foods labelled as 'gluten free' should contain ≤ 20 parts per million (ppm) of gluten[75].

Patients should be instructed to avoid contaminating their gluten-free food by using separate cooking utensils and cooking surfaces[9,10]. At present, shared items can be safely used if thoroughly cleaned with soap and water between use[9,76].

The duration of breastfeeding and the timing of gluten introduction to the infant seem to have no impact on the risk of developing CD, even in those at high risk[77]. Therefore, there are no strict indications for gluten introduction in infant diets[9]. Formerly, it was advised to avoid either early or late gluten introduction in children at risk of CD[13].

Dermatitis herpetiformis (DH) is a bullous cutaneous disease triggered by gluten consumption like CD[1]. DH and CD often coexist and share the same treatment, GFD [9,10,13,14]. Interestingly, the ESsCD guidelines suggest that psoriasis could also benefit from GFD in the case of documented CD serology, even in the absence of mucosal damage[9].

DISCUSSION

Our comparative analysis of the currently adopted CD guidelines underlined differences in diagnostic aspects and the management of the follow-up. These differences mirror some relevant clinical points in both developing and developed countries.

First, the differences in the diagnostic process of CD are important. The possibility of a no-biopsy diagnosis has relevant repercussions in developing countries. Most guidelines are still cautious in this regard, with the WGO2017 guidelines being the only ones contemplating this possibility in geographical areas with a paucity of resources. As correctly underlined by these guidelines, some absolute recommendations may not be valid for developing countries where the availability of serology or endoscopy may be lacking[10]. CD seems to have a non-negligible prevalence in Asia and sub-Saharan Africa 77,78]. Especially in Russia and Central Asia, the prevalence of CD is very likely to be underestimated due to poor disease awareness among physicians and/or patients, limited access to diagnostic resources, inappropriate use or interpretation of the serological tests, absence of standardised diagnostic and endoscopic protocols, and insufficient expertise in histopathological interpretation^[3]. Specific guidelines are lacking in these geographical areas [79]. In addition, the incidence of undiagnosed CD in children can be extremely high[80]. Knowing the high mortality and disability related to untreated CD in childhood, it would be advisable to develop specific protocols for specific geographical areas.

Recommendation		Change over time
The mainstay for treatment of CD is a strict GFD. which usually resolves both classical and non-classical manifestations	<u>**</u> **********************************	No major changes over time
Uncontaminated oat is safe for almost every patient. A small percentage of patients may be sensitive to oats and should be monitored	<u>**</u> ** ** ** **	Minor changes over time
GFD should be initiated also in psoriatic patients with positive CD serology		Major changes over time
The duration of breastfeeding and the timing of gluten introduction have no impact on the risk of developing CD		Major changes over time

945a	European Society Paediatric Gastroenterology, Hepatology and Nutrition (ESPGHAN) 2020 (8).
111 a	European Society for the Study of Coeliac Disease (ECD) 2019 (9).
<u></u>	World Gastroenterology Organization (WGO) 2017 (10).
94%e	Central Research Institute of Gastroenterology, Russia, 2016 (11).
945 C	National Institute for Health and Care Excellence (NICE), 2015 (12).
11/2	British Society of Gastroenterology (BSG), 2014 (13).
111 m	America College of Gastroenterology (ACG), 2013 (14).

Figure 11 Recommendations about the gluten-free diet for celiac disease.

The no-biopsy approach has been discouraged for a long time, especially in adults [13,14]. In contrast, most recent guidelines have incorporated the ESPGHAN 2012 recommendations for a no-biopsy approach in children[9,10]. The possibility of an outright extension of these criteria into the adult population still meets key obstacles. However, in an era during which the COVID-19 pandemic has caused a staggering drop in new CD diagnoses even in industrialised countries[81], ESPGHAN released the advice to lower the TGA-IgA threshold for diagnosing CD without biopsy[52]. Moreover, retrospective data on a possible no-biopsy approach in adults are increasing [53]. Prospective data will probably lead to the integration of such an approach to future guidelines over the next decade.

Second, the differences in follow-up recommendations reflect a relatively low interest in this topic in the past. Arguably, the search for more reliable diagnostic tools was the right priority in an era characterised by a severe under-diagnosis of CD .Nowadays, significant diagnostic delays can still occur in a minority of Central European children[82], with socioeconomically deprived children being more likely to be underdiagnosed despite improved and easily available serological testing[4].

Nonetheless, the current physicians' awareness of CD has reached fairly high levels, and the case-detection strategy has significantly contributed to the increased number of diagnoses. Consequently, the correct management of follow-up is crucial. This topic is of special interest in developed countries, in which metabolic problems possibly caused by an unbalanced GFD are particularly prevalent. Uncontrolled weight gain, metabolic syndrome, and non-alcoholic fatty liver disease are epidemic in these countries and can also be facilitated by the GFD[67,69,83-85]. In addition, quick detection of associated autoimmune conditions can prove highly beneficial, especially in autoimmune liver diseases[86]. Finally, early detection of complicated CD requires particular attention, as both neoplastic and non-neoplastic complications may arise years after the diagnosis^[6].

CONCLUSION

We found a relatively high concordance between CD guidelines. Important modifications have occurred in recent years, especially regarding the possibility of a no-biopsy diagnosis in children. Other modifications are expected in the future and will probably involve the extension of the non-invasive diagnosis to the adult population and the follow-up modalities.

ARTICLE HIGHLIGHTS

Research background

Celiac disease (CD) has risen from obscurity to global prominence in a few decades. These modifications have prompted experts from all over the world to identify effective strategies for the diagnosis and follow-up of CD. Different scientific societies, mainly from Europe and America regions, have proposed different guidelines.

Research motivation

CD guidelines are consistent when they deal key points in the diagnosis and follow-up of this condition. However, they differ in a number of other points.

Research objectives

To identify all of the existing guidelines across the globe and perform a comparative analysis to verify similarities and differences and, thus, discuss the most debated topics and the possible innovations in the next future.

Research methods

We searched PubMed for a complex string containing the terms "celiac disease", "management", and "guidelines". The results were subsequently explored to identify the most recent versions of existing guidelines of governmental agencies and scientific societies. The recommendations provided by each selected guideline were systematically explored and classified under five categories: Patients to be tested for CD, diagnostic tests (serology, duodenal biopsy, genetic test, no-biopsy diagnosis), potential/silent/seronegative CD, refractory/complicated CD, follow-up.

Research results

We identified 7 different guidelines [European Society Paediatric Gastroenterology, Hepatology and Nutrition (ESPGHAN) 2020; European Society for the Study of Coeliac Disease (ECD) 2019; World Gastroenterology Organization (WGO) 2017; Central Research Institute of Gastroenterology, Russia, 2016; National Institute for Health and Care Excellence (NICE), 2015; British Society of Gastroenterology (BSG), 2014; and America College of Gastroenterology (ACG), 2013]. These guidelines were mostly concordant but differed under certain recommendation for no-biopsy diagnosis, refractory CD, and follow-up.

Research conclusions

We found a relatively high concordance between the guidelines for CD. Important modifications have occurred in the last years, especially about the possibility of a nobiopsy diagnosis in children.

Research perspectives

Modifications of the current guidelines are expected in the near future. These modification will probably regard the possibility of a no-biopsy diagnosis (especially in developing countries) and the modalities of follow-up.

REFERENCES

- Ludvigsson JF, Leffler DA, Bai JC, Biagi F, Fasano A, Green PHR, Hadjivassiliou M, Kaukinen K, 1 Kelly CP, Leonard JN, Lundin KEA, Murray JA, Sanders DS, Walker MM, Zingone F, Ciacci C. The Oslo definitions for coeliac disease and related terms. Gut 2013; 62: 43-52 [DOI: 10.1136/gutjnl-2011-301346
- 2 Fasano A, Catassi C. Clinical practice. Celiac disease. N Engl J Med 2012; 367: 2419-2426 [PMID:



23252527 DOI: 10.1056/NEJMcp1113994]

- Poddighe D, Abdukhakimova D. Celiac Disease in Asia beyond the Middle East and Indian 3 subcontinent: Epidemiological burden and diagnostic barriers. World J Gastroenterol 2021; 27: 2251-2256 [PMID: 34040319 DOI: 10.3748/wjg.v27.i19.2251]
- 4 Whitburn J, Rao SR, Paul SP, Sandhu BK. Diagnosis of celiac disease is being missed in over 80% of children particularly in those from socioeconomically deprived backgrounds. Eur J Pediatr 2021; 180: 1941-1946 [PMID: 33569662 DOI: 10.1007/s00431-021-03974-8]
- 5 Ianiro G, Bibbò S, Bruno G, Ricci R, Arena V, Gasbarrini A, Cammarota G. Prior Misdiagnosis of Celiac Disease Is Common Among Patients Referred to a Tertiary Care Center: A Prospective Cohort Study. Clin Transl Gastroenterol 2016; 7: e139 [PMID: 26821194 DOI: 10.1038/ctg.2015.48]
- Biagi F, Schiepatti A, Maiorano G, Fraternale G, Agazzi S, Zingone F, Ciacci C, Volta U, Caio G, 6 Tortora R, Klersy C, Corazza GR. Risk of complications in coeliac patients depends on age at diagnosis and type of clinical presentation. Dig Liver Dis 2018; 50: 549-552 [PMID: 29277481]
- 7 D'Avino P, Serena G, Kenyon V, Fasano A. An updated overview on celiac disease: from immunopathogenesis and immuno-genetics to therapeutic implications. Expert Rev Clin Immunol 2021; 17: 269-284 [PMID: 33472447 DOI: 10.1080/1744666X.2021.1880320]
- Husby S, Koletzko S, Korponay-Szabó I, Kurppa K, Mearin ML, Ribes-Koninckx C, Shamir R, 8 Troncone R, Auricchio R, Castillejo G, Christensen R, Dolinsek J, Gillett P, Hróbjartsson A, Koltai T, Maki M, Nielsen SM, Popp A, Størdal K, Werkstetter K, Wessels M. European Society Paediatric Gastroenterology, Hepatology and Nutrition Guidelines for Diagnosing Coeliac Disease 2020. J Pediatr Gastroenterol Nutr 2020; 70: 141-156 [PMID: 31568151 DOI: 10.1097/MPG.00000000002497]
- Al-Toma A, Volta U, Auricchio R, Castillejo G, Sanders DS, Cellier C, Mulder CJ, Lundin KEA. European Society for the Study of Coeliac Disease (ESsCD) guideline for coeliac disease and other gluten-related disorders. United European Gastroenterol J 2019; 7: 583-613 [PMID: 31210940 DOI: 10.1177/2050640619844125
- 10 Bai JC, Ciacci C. World Gastroenterology Organisation Global Guidelines: Celiac Disease February 2017. J Clin Gastroenterol 2017; 51: 755-768 [PMID: 28877080 DOI: 10.1097/MCG.000000000000919]
- 11 Parfenov AI, Bykova SV, Sabel'nikova EA, Maev IV, Baranov AA, Bakulin IG, Krums LM, Bel'mer SV, Borovik TE, Zakharova IN, Dmitrieva YA, Roslavtseva EA, Kornienko EA, Khavkin AI, Potapov AS, Revnova MO, Mukhina YG, Shcherbakov PL, Fedorov ED, Belousova EA, Khalif IL, Khomeriki SG, Rotin DL, Vorob'eva NG, Pivnik AV, Gudkova RB, Chernin VV, Vokhmyanina NV, Pukhlikova TV, Degtyarev DA, Damulin IV, Mkrtumyan AM, Dzhulai GS, Tetruashvili NK, Baranovsky AY, Nazarenko LI, Kharitonov AG, Loranskaya ID, Saifutdinov RG, Livzan MA, Abramov DA, Osipenko MF, Oreshko LV, Tkachenko EI, Sitkin SI, Efremov LI. All-Russian Consensus on Diagnosis and Treatment of Celiac Disease in Children and Adults. Ter Arkh 2017; 89: 94-107 [PMID: 28378737 DOI: 10.17116/terarkh201789394-107]
- Downey L, Houten R, Murch S, Longson D; Guideline Development Group. Recognition, 12 assessment, and management of coeliac disease: summary of updated NICE guidance. BMJ 2015; 351: h4513 [PMID: 26333593 DOI: 10.1136/bmj.h4513]
- 13 Ludvigsson JF, Bai JC, Biagi F, Card TR, Ciacci C, Ciclitira PJ, Green PH, Hadjivassiliou M, Holdoway A, van Heel DA, Kaukinen K, Leffler DA, Leonard JN, Lundin KE, McGough N, Davidson M, Murray JA, Swift GL, Walker MM, Zingone F, Sanders DS; BSG Coeliac Disease Guidelines Development Group; British Society of Gastroenterology. Diagnosis and management of adult coeliac disease: guidelines from the British Society of Gastroenterology. Gut 2014; 63: 1210-1228 [PMID: 24917550 DOI: 10.1136/gutjnl-2013-306578]
- Rubio-Tapia A, Hill ID, Kelly CP, Calderwood AH, Murray JA; American College of 14 Gastroenterology. ACG clinical guidelines: diagnosis and management of celiac disease. Am J Gastroenterol 2013; 108: 656-676; quiz 677 [PMID: 23609613 DOI: 10.1038/ajg.2013.79]
- 15 Vilppula A, Kaukinen K, Luostarinen L, Krekelä I, Patrikainen H, Valve R, Mäki M, Collin P. Increasing prevalence and high incidence of celiac disease in elderly people: a population-based study. BMC Gastroenterol 2009; 9: 49 [PMID: 19558729 DOI: 10.1186/1471-230X-9-49]
- 16 Vivas S, Ruiz de Morales JM, Fernandez M, Hernando M, Herrero B, Casqueiro J, Gutierrez S. Agerelated clinical, serological, and histopathological features of celiac disease. Am J Gastroenterol 2008; 103: 2360-2365; quiz 2366 [PMID: 18702652 DOI: 10.1111/j.1572-0241.2008.01977.x]
- 17 Fasano A. Celiac disease--how to handle a clinical chameleon. N Engl J Med 2003; 348: 2568-2570 [PMID: 12815143 DOI: 10.1056/NEJMe030050]
- 18 Baydoun A, Maakaron JE, Halawi H, Abou Rahal J, Taher AT. Hematological manifestations of celiac disease. Scand J Gastroenterol 2012; 47: 1401-1411 [PMID: 22861356 DOI: 10.3109/00365521.2012.706828
- Kamycheva E, Goto T, Camargo CA Jr. Celiac disease is associated with reduced bone mineral 19 density and increased FRAX scores in the US National Health and Nutrition Examination Survey. Osteoporos Int 2017; 28: 781-790 [PMID: 27714440 DOI: 10.1007/s00198-016-3791-4]
- 20 Volta U, De Franceschi L, Lari F, Molinaro N, Zoli M, Bianchi FB. Coeliac disease hidden by cryptogenic hypertransaminasaemia. Lancet 1998; 352: 26-29 [PMID: 9800742 DOI: 10.1016/s0140-6736(97)11222-31
- Luostarinen L, Pirttilä T, Collin P. Coeliac disease presenting with neurological disorders. Eur 21 Neurol 1999; 42: 132-135 [PMID: 10529537 DOI: 10.1159/000008086]



- 22 Schiepatti A, Sprio E, Sanders DS, Lovati E, Biagi F. Coeliac disease and obstetric and gynaecological disorders: where are we now? Eur J Gastroenterol Hepatol 2019; 31: 425-433 [PMID: 30676472 DOI: 10.1097/MEG.00000000001361]
- 23 Agardh D, Lee HS, Kurppa K, Simell V, Aronsson CA, Jörneus O, Hummel M, Liu E, Koletzko S; TEDDY Study Group. Clinical features of celiac disease: a prospective birth cohort. Pediatrics 2015; 135: 627-634 [PMID: 25733751 DOI: 10.1542/peds.2014-3675]
- Imanzadeh F, Sayyari AA, Yaghoobi M, Akbari MR, Shafagh H, Farsar AR. Celiac disease in 24 children with diarrhea is more frequent than previously suspected. J Pediatr Gastroenterol Nutr 2005; 40: 309-311 [PMID: 15735484 DOI: 10.1097/01.mpg.0000154012.10420.08]
- 25 Reilly NR, Fasano A, Green PHR. Presentation of Celiac Disease. Gastrointest Endosc Clin N Am 2012; 22: 613-621 [DOI: 10.1016/j.giec.2012.07.008]
- 26 Leffler DA, Green PHR, Fasano A. Extraintestinal manifestations of coeliac disease. Nat Rev Gastroenterol Hepatol 2015; 12: 561-571 [DOI: 10.1038/nrgastro.2015.131]
- 27 Catassi C, Fasano A. Celiac disease diagnosis: simple rules are better than complicated algorithms. Am J Med 2010; 123: 691-693 [PMID: 20670718 DOI: 10.1016/j.amjmed.2010.02.019]
- Hischenhuber C, Crevel R, Jarry B, Maki M, Moneret-Vautrin DA, Romano A, Troncone R, Ward 28 R. Review article: safe amounts of gluten for patients with wheat allergy or coeliac disease. Aliment Pharmacol Ther 2006; 23: 559-575 [DOI: 10.1111/j.1365-2036.2006.02768.x]
- Reeves GEM, Squance ML, Duggan AE, Murugasu RR, Wilson RJ, Wong RC, Gibson RA, Steele RH, Pollock WK. Diagnostic accuracy of coeliac serological tests: a prospective study. Eur J Gastroenterol Hepatol 2006; 18: 493-501 [DOI: 10.1097/00042737-200605000-00006]
- 30 Volta U, Fabbri A, Parisi C, Piscaglia M, Caio G, Tovoli F, Fiorini E. Old and new serological tests for celiac disease screening. Expert Rev Gastroenterol Hepatol 2010; 4: 31-35 [PMID: 20136587 DOI: 10.1586/egh.09.66]
- 31 Stern M; Working Group on Serologic Screening for Celiac Disease. Comparative evaluation of serologic tests for celiac disease: a European initiative toward standardization. J Pediatr Gastroenterol Nutr 2000; 31: 513-519 [PMID: 11144436 DOI: 10.1097/00005176-200011000-00012
- Sood A, Khurana MS, Mahajan R, Midha V, Puri S, Kaur A, Gupta N, Sharma S. Prevalence and 32 clinical significance of IgA anti-tissue transglutaminase antibodies in patients with chronic liver disease. J Gastroenterol Hepatol 2017; 32: 446-450 [PMID: 27346589 DOI: 10.1111/jgh.13474]
- Granito A, Muratori L, Muratori P, Petrolini N, Bianchi FB, Volta U. Antitransglutaminase 33 antibodies and giardiasis. Am J Gastroenterol 2004; 99: 2505-2506 [PMID: 15571608 DOI: 10.1111/j.1572-0241.2004.41389 9.x
- 34 Carroccio A, Vitale G, Di Prima L, Chifari N, Napoli S, La Russa C, Gulotta G, Averna MR, Montalto G, Mansueto S, Notarbartolo A. Comparison of anti-transglutaminase ELISAs and an antiendomysial antibody assay in the diagnosis of celiac disease: a prospective study. Clin Chem 2002; 48: 1546-1550 [PMID: 12194932]
- 35 Korponay-Szabó IR, Dahlbom I, Laurila K, Koskinen S, Woolley N, Partanen J, Kovács JB, Mäki M, Hansson T. Elevation of IgG antibodies against tissue transglutaminase as a diagnostic tool for coeliac disease in selective IgA deficiency. Gut 2003; 52: 1567-1571 [PMID: 14570724 DOI: 10.1136/gut.52.11.1567]
- Mozo L, Gómez J, Escanlar E, Bousoño C, Gutiérrez C. Diagnostic value of anti-deamidated gliadin 36 peptide IgG antibodies for celiac disease in children and IgA-deficient patients. J Pediatr Gastroenterol Nutr 2012; 55: 50-55 [PMID: 22197936 DOI: 10.1097/MPG.0b013e31824703c7]
- 37 Leonard MM, Lebwohl B, Rubio-Tapia A, Biagi F. AGA Clinical Practice Update on the Evaluation and Management of Seronegative Enteropathies: Expert Review. Gastroenterology 2021; 160: 437-444 [PMID: 33010252 DOI: 10.1053/j.gastro.2020.08.061]
- 38 Pais WP, Duerksen DR, Pettigrew NM, Bernstein CN. How many duodenal biopsy specimens are required to make a diagnosis of celiac disease? Gastrointest Endosc 2008; 67: 1082-1087 [PMID: 18308317 DOI: 10.1016/j.gie.2007.10.015]
- Corazza GR, Villanacci V. Coeliac disease. J Clin Pathol 2005; 58: 573-574 [PMID: 15917404 DOI: 39 10.1136/jcp.2004.023978]
- 40 Oberhuber G, Granditsch G, Vogelsang H. The histopathology of coeliac disease: time for a standardized report scheme for pathologists. Eur J Gastroenterol Hepatol 1999; 11: 1185-1194 [PMID: 10524652 DOI: 10.1097/00042737-199910000-00019]
- Hammer ST, Greenson JK. The clinical significance of duodenal lymphocytosis with normal villus 41 architecture. Arch Pathol Lab Med 2013; 137: 1216-1219 [PMID: 23991733 DOI: 10.5858/arpa.2013-0261-RA]
- 42 Corazza GR, Villanacci V, Zambelli C, Milione M, Luinetti O, Vindigni C, Chioda C, Albarello L, Bartolini D, Donato F. Comparison of the interobserver reproducibility with different histologic criteria used in celiac disease. Clin Gastroenterol Hepatol 2007; 5: 838-843 [PMID: 17544877 DOI: 10.1016/j.cgh.2007.03.019
- Granito A, Muratori P, Cassani F, Pappas G, Muratori L, Agostinelli D, Veronesi L, Bortolotti R, 43 Petrolini N, Bianchi FB, Volta U. Anti-actin IgA antibodies in severe coeliac disease. Clin Exp Immunol 2004; 137: 386-392 [PMID: 15270857 DOI: 10.1111/j.1365-2249.2004.02541.x]
- Poddighe D, Kushugulova A. Salivary Microbiome in Pediatric and Adult Celiac Disease. Front Cell 44 Infect Microbiol 2021; 11: 625162 [PMID: 33680992 DOI: 10.3389/fcimb.2021.625162]
- 45 Abdukhakimova D, Dossybayeva K, Poddighe D. Fecal and Duodenal Microbiota in Pediatric Celiac



Disease. Front Pediatr 2021; 9: 652208 [PMID: 33968854 DOI: 10.3389/fped.2021.652208]

- Oldenburger IB, Wolters VM, Kardol-Hoefnagel T, Houwen RHJ, Otten HG. Serum intestinal fatty 46 acid-binding protein in the noninvasive diagnosis of celiac disease. APMIS 2018; 126: 186-190 [PMID: 29383769 DOI: 10.1111/apm.12800]
- 47 Vreugdenhil AC, Wolters VM, Adriaanse MP, Van den Neucker AM, van Bijnen AA, Houwen R, Buurman WA. Additional value of serum I-FABP levels for evaluating celiac disease activity in children. Scand J Gastroenterol 2011; 46: 1435-1441 [PMID: 22029621 DOI: 10.3109/00365521.2011.627447
- 48 Lundin KE, Wijmenga C. Coeliac disease and autoimmune disease-genetic overlap and screening. *Nat Rev Gastroenterol Hepatol* 2015; **12**: 507-515 [PMID: 26303674 DOI: 10.1038/nrgastro.2015.136
- Megiorni F, Pizzuti A. HLA-DQA1 and HLA-DQB1 in Celiac disease predisposition: practical 49 implications of the HLA molecular typing. J Biomed Sci 2012; 19: 88 [PMID: 23050549 DOI: 10.1186/1423-0127-19-88
- Poddighe D, Rebuffi C, De Silvestri A, Capittini C. Carrier frequency of HLA-DQB1*02 allele in patients affected with celiac disease: A systematic review assessing the potential rationale of a targeted allelic genotyping as a first-line screening. World J Gastroenterol 2020; 26: 1365-1381 [PMID: 32256023 DOI: 10.3748/wjg.v26.i12.1365]
- De Silvestri A, Capittini C, Poddighe D, Valsecchi C, Marseglia G, Tagliacarne SC, Scotti V, Rebuffi 51 C, Pasi A, Martinetti M, Tinelli C. HLA-DQ genetics in children with celiac disease: a meta-analysis suggesting a two-step genetic screening procedure starting with HLA-DQ β chains. *Pediatr Res* 2018; 83: 564-572 [PMID: 29244800 DOI: 10.1038/pr.2017.307]
- 52 Trovato CM, Montuori M, Cucchiara S, Oliva S. ESPGHAN 'biopsy-sparing' guidelines for celiac disease in children with low antitransglutaminase during COVID-19. Eur J Gastroenterol Hepatol 2020; 32: 1523-1526 [PMID: 32956181 DOI: 10.1097/MEG.000000000001924]
- 53 Penny HA, Raju SA, Lau MS, Marks LJ, Baggus EM, Bai JC, Bassotti G, Bontkes HJ, Carroccio A, Danciu M, Derakhshan MH, Ensari A, Ganji A, Green PHR, Johnson MW, Ishaq S, Lebwohl B, Levene A, Maxim R, Mohaghegh Shalmani H, Rostami-Nejad M, Rowlands D, Spiridon IA, Srivastava A, Volta U, Villanacci V, Wild G, Cross SS, Rostami K, Sanders DS. Accuracy of a nobiopsy approach for the diagnosis of coeliac disease across different adult cohorts. Gut 2021; 70: 876-883 [PMID: 33139268 DOI: 10.1136/gutjnl-2020-320913]
- 54 Malamut G, Afchain P, Verkarre V, Lecomte T, Amiot A, Damotte D, Bouhnik Y, Colombel JF, Delchier JC, Allez M, Cosnes J, Lavergne-Slove A, Meresse B, Trinquart L, Macintyre E, Radford-Weiss I, Hermine O, Brousse N, Cerf-Bensussan N, Cellier C. Presentation and long-term follow-up of refractory celiac disease: comparison of type I with type II. Gastroenterology 2009; 136: 81-90 [PMID: 19014942 DOI: 10.1053/j.gastro.2008.09.069]
- Schiepatti A, Biagi F, Fraternale G, Vattiato C, Balduzzi D, Agazzi S, Alpini C, Klersy C, Corazza GR. Short article: Mortality and differential diagnoses of villous atrophy without coeliac antibodies. Eur J Gastroenterol Hepatol 2017; 29: 572-576 [PMID: 28350748 DOI: 10.1097/MEG.00000000000836
- Delabie J, Holte H, Vose JM, Ullrich F, Jaffe ES, Savage KJ, Connors JM, Rimsza L, Harris NL, 56 Müller-Hermelink K, Rüdiger T, Coiffier B, Gascoyne RD, Berger F, Tobinai K, Au WY, Liang R, Montserrat E, Hochberg EP, Pileri S, Federico M, Nathwani B, Armitage JO, Weisenburger DD. Enteropathy-associated T-cell lymphoma: clinical and histological findings from the international peripheral T-cell lymphoma project. Blood 2011; 118: 148-155 [PMID: 21566094 DOI: 10.1182/blood-2011-02-335216]
- Biagi F, Gobbi P, Marchese A, Borsotti E, Zingone F, Ciacci C, Volta U, Caio G, Carroccio A, 57 Ambrosiano G, Mansueto P, Corazza GR. Low incidence but poor prognosis of complicated coeliac disease: a retrospective multicentre study. Dig Liver Dis 2014; 46: 227-230 [PMID: 24268568 DOI: 10.1016/j.dld.2013.10.010]
- 58 van Wanrooij RL, Schreurs MW, Bouma G, von Blomberg BM, Tack GJ, Verbeek WH, Mulder CJ. Accurate classification of RCD requires flow cytometry. Gut 2010; 59: 1732 [PMID: 20805314 DOI: 10.1136/gut.2010.223438]
- Al-Toma A, Verbeek WH, Hadithi M, von Blomberg BM, Mulder CJ. Survival in refractory coeliac 59 disease and enteropathy-associated T-cell lymphoma: retrospective evaluation of single-centre experience. Gut 2007; 56: 1373-1378 [PMID: 17470479 DOI: 10.1136/gut.2006.114512]
- Rubio-Tapia A, Kelly DG, Lahr BD, Dogan A, Wu TT, Murray JA. Clinical staging and survival in 60 refractory celiac disease: a single center experience. Gastroenterology 2009; 136: 99-107; quiz 352-353 [PMID: 18996383 DOI: 10.1053/j.gastro.2008.10.013]
- Mukewar SS, Sharma A, Rubio-Tapia A, Wu TT, Jabri B, Murray JA. Open-Capsule Budesonide for 61 Refractory Celiac Disease. Am J Gastroenterol 2017; 112: 959-967 [PMID: 28323276 DOI: 10.1038/ajg.2017.71]
- Chaudhary R, Ghosh S. Infliximab in refractory coeliac disease. Eur J Gastroenterol Hepatol 2005; 62 17: 603-604 [PMID: 15879720 DOI: 10.1097/00042737-200506000-00002]
- 63 Daum S, Wahnschaffe U, Glasenapp R, Borchert M, Ullrich R, Zeitz M, Faiss S. Capsule endoscopy in refractory celiac disease. Endoscopy 2007; 39: 455-458 [PMID: 17516353 DOI: 10.1055/s-2007-966239
- Al-toma A, Visser OJ, van Roessel HM, von Blomberg BM, Verbeek WH, Scholten PE, 64 Ossenkoppele GJ, Huijgens PC, Mulder CJ. Autologous hematopoietic stem cell transplantation in



refractory celiac disease with aberrant T cells. Blood 2007; 109: 2243-2249 [PMID: 17068146 DOI: 10.1182/blood-2006-08-042820]

- Pietzak MM. Follow-up of patients with celiac disease: achieving compliance with treatment. 65 Gastroenterology 2005; 128: S135-S141 [PMID: 15825121 DOI: 10.1053/j.gastro.2005.02.025]
- Ciacci C, Cirillo M, Cavallaro R, Mazzacca G. Long-term follow-up of celiac adults on gluten-free 66 diet: prevalence and correlates of intestinal damage. Digestion 2002; 66: 178-185 [PMID: 12481164 DOI: 10.1159/000066757]
- Tovoli F, Negrini G, Farì R, Guidetti E, Faggiano C, Napoli L, Bolondi L, Granito A. Increased risk 67 of nonalcoholic fatty liver disease in patients with coeliac disease on a gluten-free diet: beyond traditional metabolic factors. Aliment Pharmacol Ther 2018; 48: 538-546 [PMID: 29984415 DOI: 10.1111/apt.14910
- Kemppainen T, Kröger H, Janatuinen E, Arnala I, Kosma VM, Pikkarainen P, Julkunen R, Jurvelin 68 J, Alhava E, Uusitupa M. Osteoporosis in adult patients with celiac disease. Bone 1999; 24: 249-255 [PMID: 10071918 DOI: 10.1016/s8756-3282(98)00178-1]
- Valletta E, Fornaro M, Cipolli M, Conte S, Bissolo F, Danchielli C. Celiac disease and obesity: need 69 for nutritional follow-up after diagnosis. Eur J Clin Nutr 2010; 64: 1371-1372 [PMID: 20717130 DOI: 10.1038/ejcn.2010.161]
- 70 Hughey JJ, Ray BK, Lee AR, Voorhees KN, Kelly CP, Schuppan D. Self-reported dietary adherence, disease-specific symptoms, and quality of life are associated with healthcare provider follow-up in celiac disease. BMC Gastroenterol 2017; 17: 156 [PMID: 29228908 DOI: 10.1186/s12876-017-0713-7]
- 71 Johansson K, Malmberg Hård Af Segerstad E, Mårtensson H, Agardh D. Dietitian visits were a safe and cost-effective form of follow-up care for children with celiac disease. Acta Paediatr 2019; 108: 676-680 [PMID: 29782665 DOI: 10.1111/apa.14411]
- van Wanrooij RL, Bouma G, Bontkes HJ, Neefjes-Borst A, van Grieken NC, von Blomberg BM, 72 Mulder CJ. Outcome of Referrals for Non-Responsive Celiac Disease in a Tertiary Center: Low Incidence of Refractory Celiac Disease in the Netherlands. Clin Transl Gastroenterol 2017; 8: e218 [PMID: 28125074 DOI: 10.1038/ctg.2016.70]
- Lee A, Newman JM. Celiac diet: its impact on quality of life. J Am Diet Assoc 2003; 103: 1533-1535 73 [PMID: 14576723 DOI: 10.1016/j.jada.2003.08.027]
- 74 Comino I, Moreno Mde L, Sousa C. Role of oats in celiac disease. World J Gastroenterol 2015; 21: 11825-11831 [PMID: 26557006 DOI: 10.3748/wjg.v21.i41.11825]
- 75 Standard for Foods for Special Dietary Use for Persons Intolerant to Gluten, CXS 118-1979, Adopted in 1979. [cited 22 February 2021]. Available from: http://www.fao.org/fao-who-codexalimentarius
- Studerus D, Hampe EI, Fahrer D, Wilhelmi M, Vavricka SR. Cross-Contamination with Gluten by 76 Using Kitchen Utensils: Fact or Fiction? J Food Prot 2018; 81: 1679-1684 [PMID: 30230372 DOI: 10.4315/0362-028X.JFP-17-383]
- 77 Aronsson CA, Lee H-S, Liu E, Uusitalo U, Hummel S, Yang J, Hummel M, Rewers M, She J-X, Simell O, Toppari J, Ziegler A-G, Krischer J, Virtanen SM, Norris JM, Agardh D, for the TEDDY STUDY GROUP. Age at Gluten Introduction and Risk of Celiac Disease. Pediatrics 2015; 135: 239-245 [DOI: 10.1542/peds.2014-1787]
- Singh P, Arora A, Strand TA, Leffler DA, Catassi C, Green PH, Kelly CP, Ahuja V, Makharia GK. Global Prevalence of Celiac Disease: Systematic Review and Meta-analysis. Clin Gastroenterol Hepatol 2018; 16: 823-836.e2 [PMID: 29551598 DOI: 10.1016/j.cgh.2017.06.037]
- 79 Dhawan A, Agarwal A, Mulder CJ, Makharia GK. Celiac disease in the East and the West: Bridging the gaps between the guidelines and their implementation in daily practice is mandatory. Indian J Gastroenterol 2019; 38: 185-189 [PMID: 31313236 DOI: 10.1007/s12664-019-00970-7]
- Biagi F, Raiteri A, Schiepatti A, Klersy C, Corazza GR. The Relationship Between Child Mortality 80 Rates and Prevalence of Celiac Disease. J Pediatr Gastroenterol Nutr 2018; 66: 289-294 [PMID: 28753188 DOI: 10.1097/MPG.000000000001696]
- Valitutti F, Troncone R, Pisano P, Ciacci C; Campania Coeliac Disease Network. Where have all the 81 other coeliacs gone in 2020? Dig Liver Dis 2021; 53: 504-505 [PMID: 33541798 DOI: 10.1016/j.dld.2021.01.008
- 82 Riznik P, De Leo L, Dolinsek J, Gyimesi J, Klemenak M, Koletzko B, Koletzko S, Korponay-Szabó IR, Krencnik T, Not T, Palcevski G, Sblattero D, Werkstetter KJ. Clinical Presentation in Children With Coeliac Disease in Central. Europe J Pediatr Gastroenterol Nutr 2021; 72: 546-551 [DOI: 10.1097/MPG.000000000003015]
- 83 Reilly NR, Lebwohl B, Hultcrantz R, Green PH, Ludvigsson JF. Increased risk of non-alcoholic fatty liver disease after diagnosis of celiac disease. J Hepatol 2015; 62: 1405-1411 [PMID: 25617505 DOI: 10.1016/j.jhep.2015.01.013]
- Reilly NR, Aguilar K, Hassid BG, Cheng J, Defelice AR, Kazlow P, Bhagat G, Green PH. Celiac 84 disease in normal-weight and overweight children: clinical features and growth outcomes following a gluten-free diet. J Pediatr Gastroenterol Nutr 2011; 53: 528-531 [PMID: 21670710 DOI: 10.1097/MPG.0b013e3182276d5e
- 85 Tortora R, Capone P, De Stefano G, Imperatore N, Gerbino N, Donetto S, Monaco V, Caporaso N, Rispo A. Metabolic syndrome in patients with coeliac disease on a gluten-free diet. Aliment Pharmacol Ther 2015; 41: 352-359 [PMID: 25581084 DOI: 10.1111/apt.13062]
- Rubio-Tapia A, Murray JA. The Liver and Celiac Disease. Clin Liver Dis 2019; 23: 167-176 [PMID: 86 30947869 DOI: 10.1016/j.cld.2018.12.001]





Published by Baishideng Publishing Group Inc 7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA Telephone: +1-925-3991568 E-mail: bpgoffice@wjgnet.com Help Desk: https://www.f6publishing.com/helpdesk https://www.wjgnet.com

