



Taxonomic Revision and Clinical Importance of *Phlomis* Genus: A Comprehensive Review

Samin Mohammadi^{1,2,3}, Behzad Jafari⁴, Samira Pourtaghi Anvarian^{1,5}, Hossein Nazemiyeh^{2,6}, Solmaz Asnaashari⁷, Abbas Delazar^{2,3}, Parina Asgharian^{2,3}

¹Student Research Committee, Tabriz University of Medical Sciences, Tabriz, Iran.

²Department of Pharmacognosy, Faculty of Pharmacy, Tabriz University of Medical Sciences, Tabriz, Iran.

³Drug Applied Research Center, Tabriz University of Medical Sciences, Tabriz, Iran.

⁴Department of Medicinal Chemistry, Faculty of Pharmacy, Urmia University of Medical Sciences, Urmia, Iran.

⁵Department of Clinical Pharmacy, Faculty of Pharmacy, Tabriz University of Medical Sciences, Tabriz, Iran.

⁶Research Center for Pharmaceutical Nanotechnology, Tabriz University of Medical Sciences, Tabriz, Iran.

⁷Biotechnology Research Center, Tabriz University of Medical Sciences, Tabriz, Iran.

Article Info

Article History:

Received: 20 Nov 2022

Accepted: 5 Jan 2023

ePublished: 13 Aug 2023

Keywords:

- Anti-inflammatory Agent
- Bone Development
- Classification
- Iridoid
- Phlomis*
- Phytochemical

Abstract

Phlomis (L.) Moench belongs to the Lamiaceae family. It has recently undergone significant changes in taxonomy, with many species from *Eremostachys* and *Phlomis* added to the genus. The aforementioned species were studied in terms of morphological and phytochemical systematics. Species of *Phlomis* are distinguished from *Phlomis* by their densely bearded upper corolla lip and nutlet. However, *Eremostachys* and *Phlomis* have a lot in common morphologically. Plant chemosystematics present iridoids, phenylethanoids, and furanolanthenes as dominant constituents of *Phlomis* species. Long-term traditional uses, such as bone fracture therapy, local analgesic, and wound healing actions, pique researchers' interest in these plants. The species and their secondary metabolites have been implicated in drug discovery by their anti-inflammatory and bone-development properties *in vitro*, *in vivo*, and clinically. A review of the taxonomic status based on phytochemical and morphological characteristics, as well as the clinical importance of the *Phlomis* genus, is presented in the current study to provide a basis for further investigations.

Introduction

For millenniums, plants have been used as food and medicine. Today, plants are substantial sources of drugs as well as traditional medicine. Plant secondary metabolites are used originally or as lead compounds in pharmaceutical research and industries. Hundreds of examples indicate the importance of natural sources in drug development. For instance, artemisinin, which plays a fundamental role in malaria treatment, is derived from the *Artemisia* genus. Paclitaxel, obtained from *Taxus brevifolia*, is a crucial anti-cancer agent.¹ This valuable, ages-long experience of medicinal plant application is an opportunity that leads to a low incidence of side effects. However, using herbal materials could be expensive and harmful to the environment. Indiscriminate and unprincipled harvesting endangers natural resources. Following the identification and isolation of active ingredients, synthetic and semisynthetic methods are studied to develop an efficient industrial production of desired components. Therefore, screening flora for novel compounds remains an attractive

topic in pharmaceutical research to discover effective medications against various diseases.²

The Lamiaceae or Mint family of angiosperms is a prominent example of a medicinal and nutritional plant source. For example, peppermint (*Mentha × Piperita* L.), a representative of this family, is usually prescribed for gastrointestinal disorders such as irritable bowel syndrome (IBS) and is also used as a food ingredient.³ The mint family contains 245 genera and 7886 species. *Phlomis* (L.) Moench, comprising 168 accepted species, is one of the largest genera of the Lamiaceae family. This genus was recently revised by adding some *Phlomis* and *Eremostachys* species.^{4,5} From a phytochemical viewpoint, iridoids, phenylethanoids, flavonoids, and several other compounds have been isolated from this genus. *Phlomis* species have a relatively low essential oil yield, from 0.02% to 0.9%. Non-terpene hydrocarbons such as alcohols and aldehydes are the major components of volatile oil.^{6,7}

The reported pharmacological effects of various *Phlomis* species include menopausal symptoms relief,

*Corresponding Author: Parina Asgharian, E-mail: parina.asgharian@gmail.com

©2024 The Author(s). This is an open access article and applies the Creative Commons Attribution Non-Commercial License (<http://creativecommons.org/licenses/by-nc/4.0/>). Non-commercial uses of the work are permitted, provided the original work is properly cited.

anti-inflammatory activity, and anti-osteoporosis activity. Concerning the anti-osteoporosis activity, loganin and morroniside (iridoid glycosides) can stimulate the differentiation of osteoblasts through diverse mechanisms such as increasing collagen type 1 and inhibiting their apoptosis through anti-inflammatory effects. An *in vivo* study on ovariectomized female mice revealed the osteogenic effect of *P. umbrosa*. Increasing mineralization ratio and bone mineral density are two of the attributed mechanisms of this action.⁸⁻¹⁰ In addition to modern usages, these species have several traditional applications. For example, in Turkey, *P. tuberosa* was used for its wound-healing effects and in Iran, it is still used as a food ingredient. Traditional uses could be inspiring for novel drug innovation.¹¹ Plants of this genus are distributed throughout Asia and some European countries such as Turkey, Armenia, Georgia, and Azerbaijan. Forty-two various *Phlomis* species are found in China, and 59 diverse species grow in central Asian countries.¹²

Methodology

The evaluated papers were screened based on the inclusion criteria: (1) articles written in English and (2) articles containing data on species classified within the *Phlomis* genus. In addition, the following exclusion criteria were applied: (1) papers that did not discuss systematics, morphology, extracts' phytochemistry, or biological activities of *Phlomis* species and (2) Articles with unavailable full text.

Up until 2022, search engines and databases such as PubMed/Medline, Scopus, Google Scholar, Google Books, Google Patents, and Cochrane Library were combed through. The search terms were "*Phlomis*," "*Phlomis*," and "*Eremostachys*" in the article titles, abstracts, and keyword sections. We initially extracted the papers on the confirmed species currently classified under the genus *Phlomis* by looking through all the retrieved publications and using the World Flora Online plant list (<http://www.worldfloraonline.org/>). To proceed, we selected articles from the remaining 200 that were relevant to the outline of the present study and did not include duplicate information. Subsequently, articles were placed in the defined categories comprising taxonomy, morphology, phytochemistry, ethnomedicine, *in vitro*, *in vivo*, and clinical studies. EndNote software was used to manage and organize the references. Finally, About 120 scientific papers were incorporated in the text.¹³

Taxonomy

Taxonomy reflects the evolutionary progress of plants and determines their ancestors. It is also highly beneficial to study medicinal plants and authenticate herbals. Incorrect or inaccurate nomenclature of herbs may lead to either the omission or misuse of articles while searching the database. The use of synonyms or common names instead of confirmed ones in manuscripts is a frequent mistake. Various parameters such as morphological characters,

chromosome number, DNA sequences in chloroplast or nuclear regions, and phytochemical features are involved in the categorization of plants. Moreover, parsimony, maximum likelihood, and Bayesian analysis are some methods applied to draw phylogeny trees. Nevertheless, ongoing updates to our information may improve the current classifications.^{14,15}

The Lamiaceae family is divided into seven subfamilies (Table 1). Some genera, however, cannot be classified into any of these sections for several reasons. For example, a plant may possess the morphological characteristics of one group but the DNA sequence specifications of another. The plants mentioned herein belong to the incertae sedis class, which means uncertain position. Nepetoideae is the most populous subfamily of Lamiaceae and is genetically related to Symphorematoideae and Viticoideae in its evolutionary progress, but Lamioideae seems to be genetically independent of the other groups. In molecular data, Scutellarioideae is the closest subfamily to Lamioideae. The style is commonly terminal or subterminal within the Lamiaceae family, whereas it is gynobasic in Nepetoideae and Lamioideae. Despite similarities in style type, these two subfamilies have some differences. Lamioideae genera are mostly nonaromatic; their pollen is usually tricolpate; and Iridoids are their frequent phytochemicals. Hexacolpate pollen, however, is an essential characteristic of the Nepetoideae genera. They are commonly aromatic and comprise rosmarinic acid.¹⁶

Further investigations were accomplished to classify the Lamioideae subfamily at the tribal level. Scheen *et al.*¹⁷ reported Lamioideae as a monophyletic subfamily, but morphological features cannot singularly substantiate this. Based on molecular phylogenetic studies, Lamioideae consists of Pogostemoneae, Gomphostemmatae, Synandreae, Stachydeae, Phlomideae, Leonureae, Lamieae, Marrubieae, and Leucadeae tribes. Some genera, such as *Colquhounia*, also remain unclassified within the subfamily. *Ajugoides* and *Matsumurella*, which were not included in the aforementioned work, were classified by Bendiksby *et al.*¹⁸ based on molecular data and DNA sequence within a newly introduced tribe, Paraphlomideae.

The Phlomideae tribe used to have six members;

Table 1. Classification of Lamiaceae family according to "The families and genera of vascular plants" book by Kubitzki K.

| Subfamily | Tribe | Genus example |
|-------------------|--------------|--------------------|
| Symphorematoideae | - | <i>Sphenodesme</i> |
| Viticoideae | - | <i>Vitex</i> |
| Ajugoidae | - | <i>Karomia</i> |
| Prostantheroideae | Chloantheae | <i>Physopsis</i> |
| | Westringieae | <i>Hemigenia</i> |
| Scutellarioideae | - | <i>Scutellaria</i> |
| Lamioideae | - | <i>Phlomis</i> |
| | Elsholtzieae | <i>Collinsonia</i> |
| Nepetoideae | Mentheae | <i>Mentha</i> |
| | Ocimeae | <i>Lavandula</i> |
| Incertae sedis | - | <i>Tectona</i> |

however, according to the “world checklist of selected plant families” database, the species of the *Eremostachys*, *Pseuderemostachys*, *Notochaete*, and *Lamiophlomis* genera are now classified within the *Phlomoidea* genus. Therefore, *Phlomis* and *Phlomoidea* genera are the principal members of the Phlomoidea tribe.^{17,18} Despite the close relationship between *Eremostachys* and *Phlomoidea*, however, some *Eremostachys* species studied by Mathiesen *et al.*¹⁹ retained their last systematic positions. Moreover, Ryding introduced *Eremostachys* as a monophyletic and independent genus to make *Phlomoidea* less heterogeneous.²⁰ Salmaki *et al.*²¹ conducted a molecular phylogeny study in this regard and identified several intermediate species with both *Phlomoidea* and *Eremostachys* characteristics within the referred genera.

Furthermore, the *Phlomoidea* genus cannot be considered monophyletic by eliminating the *Eremostachys* species. Thus, *Eremostachys* is treated as a subclass of *Phlomoidea*, which confirms the hypothesis proposed by the “world checklist of selected plant families” database.²¹ The two principal genera of this tribe have an essential difference in their chromosome number. *Phlomis* and *Phlomoidea* have $x=10$ and $x=11$ chromosomes, respectively.¹⁹ Moreover, the existence of the indumentum is a common characteristic of Lamiaceae plants. Therefore, trichomes morphology may affect the plants phylogeny. For example, branched and multinodal trichomes can be considered as a synapomorphic feature of *Phlomis* species. This characteristic is also observed in some *Phlomoidea* species. The appearance of branched and multinodal trichomes is a plesiomorphic feature of the sister group *Phlomoidea*.²²

Phlomoidea is considered a non-monophyletic genus. It was introduced in 1794 by Moench when he transferred *Phlomis tuberosa* to a distinct and new category based on variations of corolla and fruit characteristics.²³ Five sections have been established within the genus *P. sect. Filipendula* and *P. sect. Phlomoidea*, two populous sections introduced based on morphological characteristics identified by Kamelin and Makhmedov. For example, *Filipendula* species, such as *P. tofetisowii*, are commonly identified by bicolor corolla and pinnatisect leaves.²² According to Kamelin and Makhmedov's classification, *P. sect. Phlomoidea* chiefly consists of the former *Phlomis* species, and *P. sect. Filipendula* mainly comprises the *ex-Eremostachys* species.¹⁹ Subsequently, Sennikov and Lazkov presented *Eremostachys*, *Paraeremostachys*, and *Moluccelloidea* sections, which were circumscribed using DNA sequence data and information about the recently transferred species.^{18,24} For example, Ranjbar and Mahmoudi²³ evaluated the morphological and taxonomic characteristics of *P. sect. Thyrsiflorae*, while Rechinger presented this section in 1982 as a subclass of *Eremostachys* genus.

Morphology

In terms of morphology, a determinant parameter of plant systematics, the variation rate is high among the family but

it becomes more limited in the subclasses. For example, the Lamiaceae family comprises trees, shrubs, and herbaceous plants, while the *Phlomoidea* genus includes herbaceous species.¹⁶ In the following paragraph, some of the most outstanding general characteristics of the Lamiaceae family are discussed.

Essential oil-secreting glandular trichomes other than covering hairs are present on the surface of the calyx (e.g., *Lavandula angustifolia* Mill.), leaves (e.g., *Rosmarinus officinalis* L.), and other aerial parts of most Lamiaceae members. Therefore, these plants are chiefly aromatic. They typically bear condensed flowers (verticillaster), opposite leaves, and quadrangular stems. The flowers possess corollas with two upper and lower lips and more than two segments. *Lavandula* L. corollas, for example, consist of two divisions on the upper lip and three on the lower one. There are usually two pairs of stamens within the corollas, and the pairs differ in size.^{25,26} In addition, the Lamioideae subfamily can be discerned by gynobasic style, tricolpate pollen, and being less aromatic compared to the other Lamiaceae subfamilies. The lower lip of the corolla usually has three smaller divisions with a larger central one. The upper lip bears two parts and infrequently a single part due to the segments merging. Small bracteoles and dry schizocarpic fruits are common in Lamioideae plants.¹⁶

Phlomoidea species are perennial herbaceous plants. The lateral roots are occasionally tuberiform. As with other Lamioideae species, they bear gynobasic style and schizocarpic fruits. There are four fertile stamens with the anterior higher pair within the corollas. Stellate and simple hairs cover the calyx surface. *Phlomoidea* species own arch-shaped upper corolla lips. Moreover, some principal features are needed to discriminate between *Phlomoidea* and its sister group *Phlomis*. *Phlomis* species are distinguished with their laterally compressed upper corolla lip. Unlike *Phlomoidea*, however, the margin of the upper corolla lip and nutlet of *Phlomis* species are not densely bearded. According to a study on the pericarp structure, the sclerenchyma region in *Phlomis* species is another distinguishing characteristic.^{20,21,27}

As previously discussed, the systematic position of *Eremostachys* is relatively controversial. The distinctive features of *Eremostachys* and *Phlomoidea* have been addressed by various studies. Leaves of the *Eremostachys* species are mostly pinnatisect, while they are often simple in *Phlomoidea* with a cordiform to triangular and oval shape. The nutlet pericarp of the *Eremostachys* class is thicker than most Lamioideae genera. Compared to the *Phlomoidea*, *Eremostachys* have huge flowers. However, similar characteristics are also discussed. For example, *Eremostachys* represents the same morphological aspects, referred to in the previous paragraph, as *Phlomoidea*. In regards to their cytology, the chromosome number is $X = 11$ for both groups.¹⁹⁻²¹ Figure 1 indicates the morphological features of *Phlomoidea labiosa*, a former *Eremostachys* member.

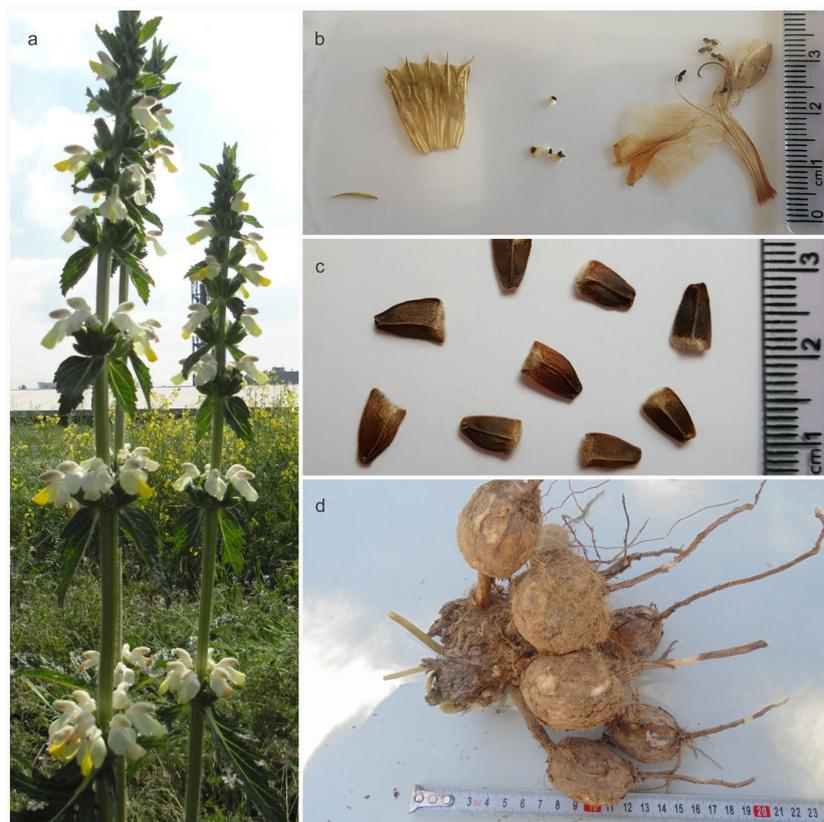


Figure 1. *Phlomoides labiosa*, a member of the *Filipendula* section, was formerly recognized as *Eremostachys labiosa*, and it was a subset of the *E. sect. Phlomoides* Bunge. a) Floral whorls are arranged with some intervals on verticillasters. b) *P. labiosa* has needle-like bracteoles, calyxes with five lobes, and typical characteristics of *Phlomoides* corollas. Also, hairs on four ovary lobes are apparent. c) Nutlets are almost hairy on top. Nevertheless, in contrast to most *Phlomoides* nutlets, they are not densely bearded. d) Tuberiform roots of *P. labiosa* were collected at Khalaj Mountains, Khorasan province, Iran, in May 2020.

Phytochemistry

Investigating structural similarities is a critical tool for classifying plants. Structural resemblances of secondary metabolites show the presence of similar metabolic pathways. To make inferences about phylogenetic links, it may be helpful to obtain networks of these compounds. Given the importance of plant chemicals from the perspective of biological activities, the aforementioned classifying system, plant chemosystematics, could be beneficial and practical in the discovery of drugs based on herbal metabolites.²⁸ In this regard, the isolated compounds of various *Phlomoides* species have been compiled and are listed in Table S1 in the supplementary data. Several secondary metabolites belonging to distinct phytochemical groups were evaluated for structural similarities based on the Tanimoto value using Open Babel and Cytoscape software programs as well as the ChEMBL website (Figure 2). The network of links with Tanimoto values greater than 0.4 was provided, and the top nodes were determined employing degree analysis (Figure 3). It is worth noting that Stachydrine (Alkaloid) had the least structural resemblance to the other components, and the associated Tanimoto values were all less than 0.4. Following that, the chemicals were categorized through clustering coefficient analysis (Table 2), and the correlations of clusters and species were assessed. As shown in Figure 4,

furanolabdanes, iridoids, and phenylethanoids (top node: cluster 15) were the dominant secondary metabolites of the studied species (Figure 5).²⁹

Ethnomedicine

Ethnomedicine refers to traditional practices orally passed through generations based on long-time observations. Traditional medicine depends mainly on well-known herbals which are valued in a particular location. Herbal products have always played an essential role in human life. Their application as medicines and foods predates written history. Herbals were the primary therapeutic agents used in earlier centuries. Today, plants also play a vital role in pharmacotherapy. A high proportion of the world's population still trusts and relies on traditional medicine for their healthcare, especially in developing countries.³⁰ Some *Phlomoides* species are also used in traditional medicine. An ethnobotanical survey in a village in Turkey showed that the leaves of *P. tuberosa* are used as a compress for wound healing.³¹ In addition, Rehman *et al.*³² revealed the application of a *P. laciniata* decoction for the treatment of headache and liver problems in Karak district, Pakistan. *P. umbrosa* is known in China as Xu Duan, which means "can reconnect broken bones." The roots of this plant are prescribed to accelerate bone fracture healing.³³ Reported folk uses are summarized in Table 3.

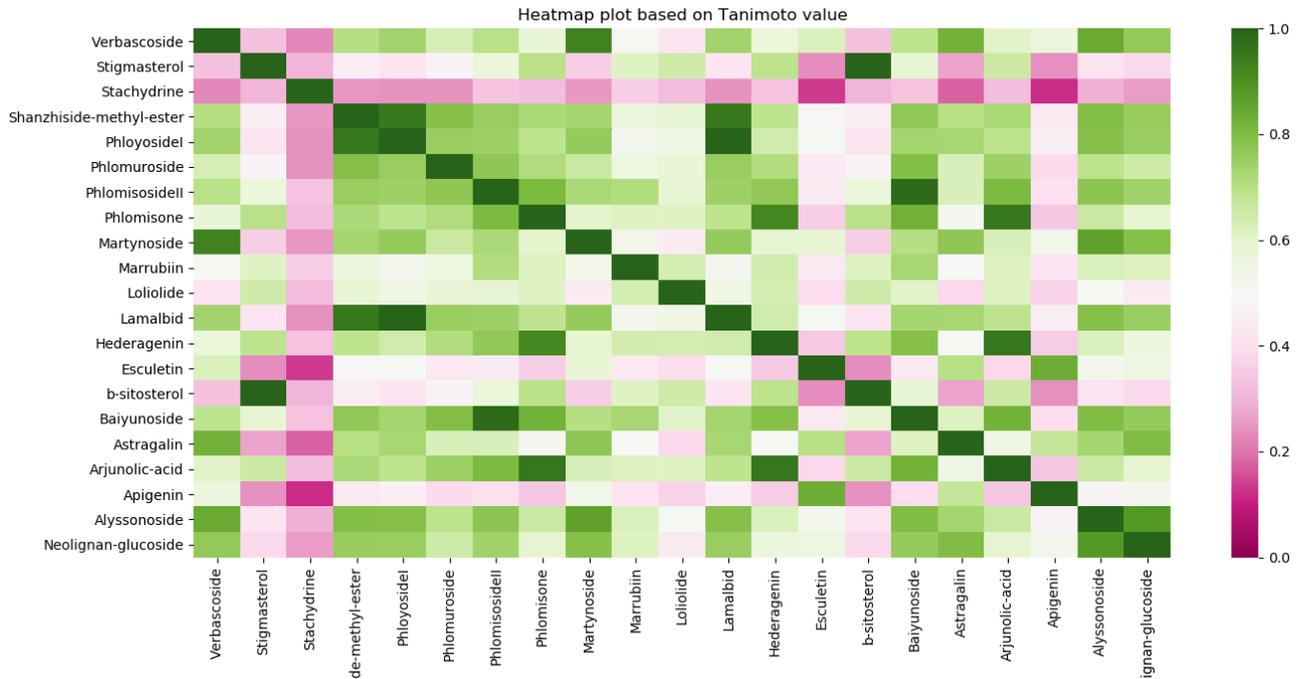


Figure 2. Heatmap of Secondary metabolites' Tanimoto value.

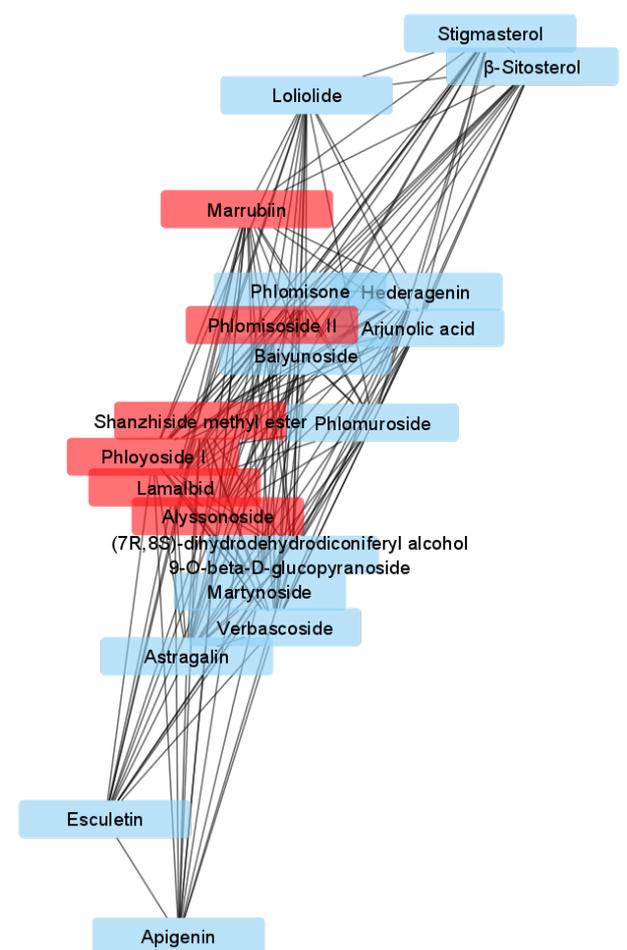


Figure 3. Structural similarity analysis based on Tanimoto value (Top nodes: Alyssonoside, Lamalbid, Phloyoside I, Shanzhiside methyl ester, Phlomisidell, and Marrubiin).

Table 2. Categories of clustering coefficient analysis.

| Rank | Name | Score |
|------|--|--------|
| 1 | Stigmasterol (Steroid) | 1.0000 |
| 1 | β-Sitosterol (Steroid) | 1.0000 |
| 3 | Apigenin (Flavonoid) | 0.9744 |
| 3 | Esculetin (Coumarin) | 0.9744 |
| 5 | Loliolide (Benzofuran) | 0.9500 |
| 6 | Arjunolic acid (Saponin) | 0.9338 |
| 6 | Hederagenin (Saponin) | 0.9338 |
| 6 | Phlomison (Triterpene) | 0.9338 |
| 9 | Astragalin (Flavonoid) | 0.9333 |
| 10 | (7R,8S)-dihydrodehydrodiconiferyl alcohol 9-O-beta-D-glucopyranoside (Neolignan glucoside) | 0.9191 |
| 10 | Verbascoside (Phenylethanoid) | 0.9191 |
| 10 | Martynoside (Phenylethanoid) | 0.9191 |
| 13 | Phlomuroside (Miscellaneous) | 0.9020 |
| 13 | Baiyunoside (Furanolabdane) | 0.9020 |
| 15 | Phlomisidell (Furanolabdane) | 0.8655 |
| 15 | Shanzhiside methyl ester (Iridoid) | 0.8655 |
| 15 | Lamalbid (Iridoid) | 0.8655 |
| 15 | Phloyoside I (Iridoid) | 0.8655 |
| 15 | Alyssonoside (Phenylethanoid) | 0.8655 |
| 15 | Marrubiin (Furanolabdane) | 0.8655 |

In Vitro Studies

Antioxidant effect

DPPH (2,2-diphenyl-1-picrylhydrazyl), suppression of PMNs' oxidative burst (polymorphonuclear leukocytes),

Table 3. Applications of *Phlomoides* species in ethnomedicine.

| Species | Region | Uses | Plant part used | Administration | Ref |
|---------------------|----------|--|-------------------|---|-----|
| <i>P. tuberosa</i> | Turkey | Wound healing | Leaves | Topical-Compress | 31 |
| | Iran | Culinary uses | Leaves | Oral-Grilled | 34 |
| <i>P. laciniata</i> | Pakistan | Headache Liver problems | Whole plant | Oral-Decoction | 32 |
| | Iran | local analgesic anti-inflammatory | Roots and flowers | Topical-Compress Oral-Decoction | 35 |
| <i>P. umbrosa</i> | Korea | Brain function enhancement Immunomodulation | Roots | Oral-Decoction (A combination of 18 dried herbs) | 36 |
| | China | Bone fractures | Roots | Oral | 33 |
| <i>P. bracteosa</i> | India | Stomach disorders | Aerial parts | Oral-Taken in warm water (A combination of 5 dried herbs) | 37 |

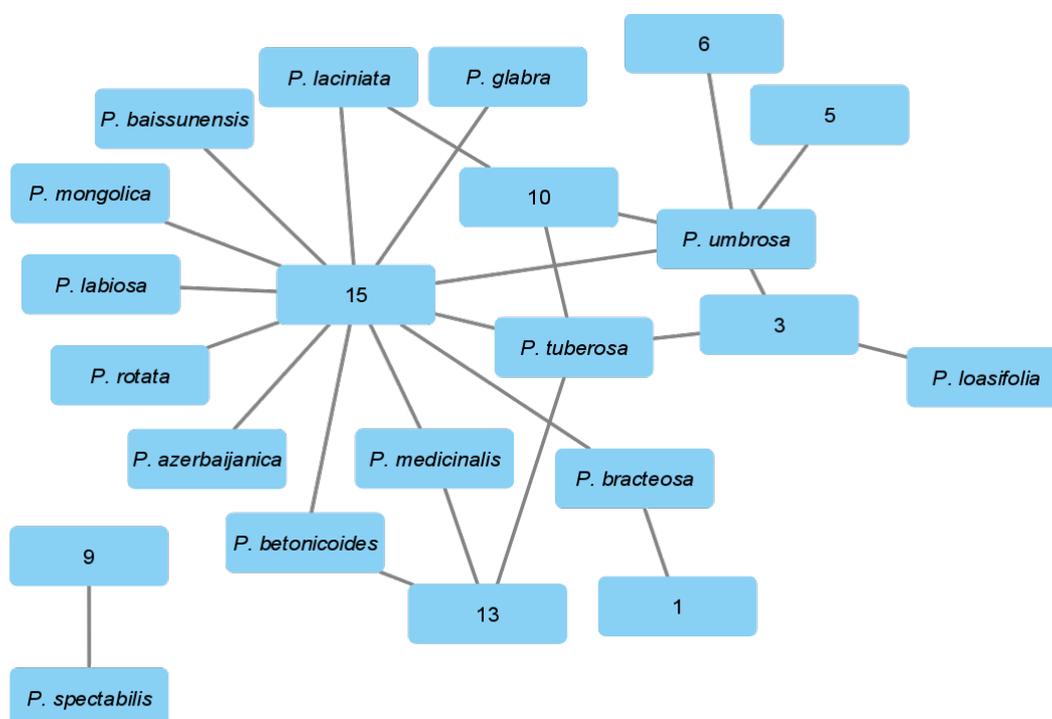


Figure 4. Correlations of clustering coefficient categories and *Phlomoides* species.

ABTS (2,2'-azino-bis(3-ethylbenzothiazoline-6-sulfonic acid)), FRAP (ferric reducing antioxidant power), β-Carotene-linoleic acid, DNA damage protection potential, and H₂O₂-luminol chemiluminescence assays have indicated the significant antioxidant capacity of several *Phlomoides* species such as *P. bracteosa*, *P. maximowiczii*, *P. megalantha*, and *P. laciniata*.³⁸⁻⁴¹ The radical scavenging activity of the genus is attributed to some secondary metabolites. Some identified active compounds include a flavonoid (luteolin-7-O-rutinoside) and a phenylethanoid (verbascoside) from the methanolic extract of *P. azerbaijanica*, phenylethanoid derivatives from *P. umbrosa* roots, phlolside G, (an iridoid from *P. likiangensis*), protocatechic and rosmarinic acids, all major phenolic components of *P. megalantha* and *P. umbrosa*.⁴¹⁻⁴⁴

Former studies have also shown a positive correlation between antioxidant activity and total phenolic content.⁴⁵ The volatile components of *Phlomoides* species, in contrast, do not bear antioxidant activity.⁴⁶ In addition, *P. labiosiformis* demonstrated *in vitro* neuroprotective effects in an Alzheimer's disease model by reducing Amyloid-peptide-induced ROS (reactive oxygen species) level.⁴⁷

Cytotoxicity

MTT and BSLT (Brine Shrimp Lethality Test) are the two most frequently used cytotoxicity assays.⁴⁸ Cancerous cell proliferation may also be studied using cell counting kits. Employing cell counting kit-8, the phenylethanoid verbascoside, isolated from *P. nissoli*, showed anti-cancer effects on the cell lines MCF7 and MDA-MB-231.⁴⁹

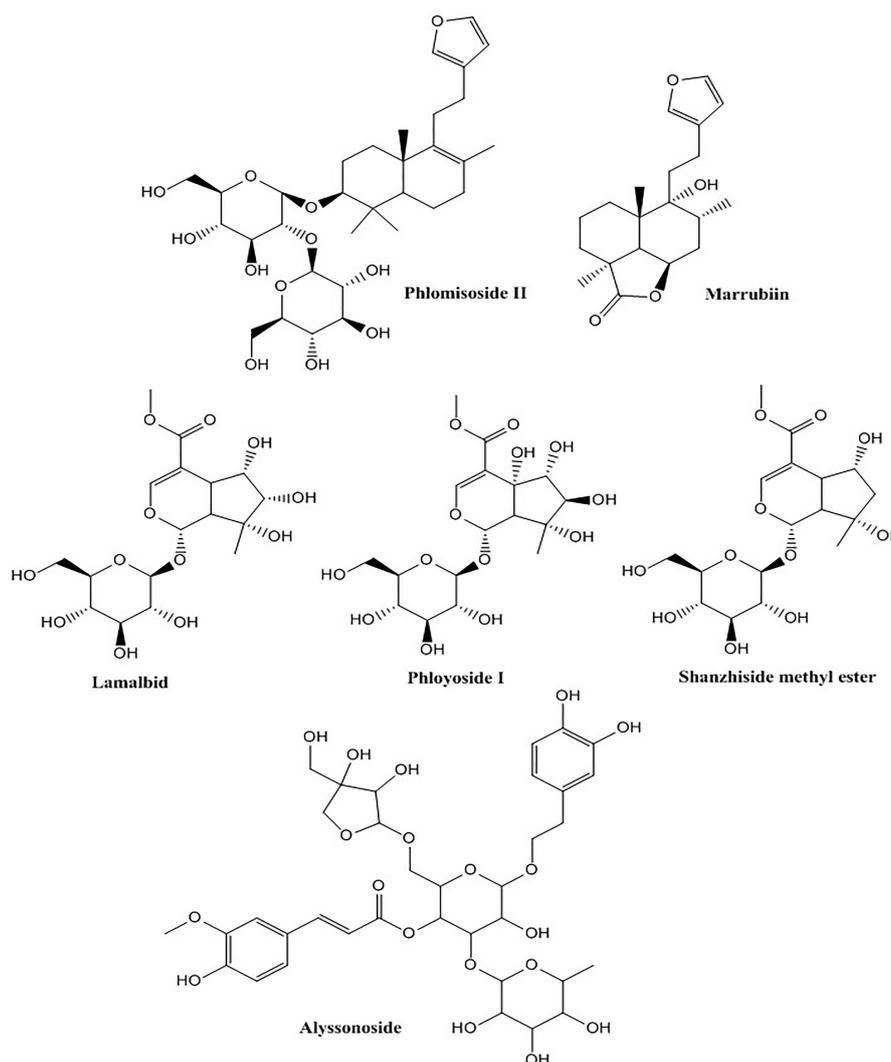


Figure 5. The chemical structures of the dominant secondary metabolites (Cluster 15).

However, iridoid glycosides purified from the underground parts of *P. laciniata* did not exhibit significant cytotoxic activity in the BSLT test.⁵⁰ Asgharian *et al.*⁴⁸ revealed the cytotoxicity of *P. azerbaijanica* dichloromethane and n-hexane extracts comprising sterols, terpenoids, and cardiac glycosides.⁴⁸ In addition, nanoencapsulation does not improve the cytotoxicity of *P. labiosa* dichloromethane extract, which might be a result of limited extract release through liposome lamella.⁴⁵

Antimicrobial effect

Various *Phlomoideis* species have been investigated for antimicrobial properties. *P. azerbaijanica* lacks antibacterial action, while *P. macrophylla* and *P. tuberosa* show significant activity against *Staphylococcus aureus*.^{48,51} Pulchelloside I, an iridoid glycoside purified from *P. laciniata* rhizomes, was indicated to have a MIC value of 0.05 mg/mL for the strains of *Staphylococcus aureus*, *Bacillus cereus*, *Escherichia coli*, and *Proteus mirabilis*.⁵⁰ Anti-rotavirus activity was reported from iridoids and isobenzofuranone derivatives of *P. betonicoides*.⁵² Moreover, sesquiterpene, coumarin, and steroid derivatives are assumed to be responsible for the

antimalarial effects of *P. azerbaijanica*.⁵³ The consequences of nanoencapsulation are inconsistent. Silver nanoparticles of *P. bracteosa* demonstrate antibacterial activity equivalent to the standard antibiotic agent; however, *P. labiosa* liposomes are less effective than the free extract.^{45,54}

Miscellaneous effects

The genus has also been reported to engage in other notable activities. By lowering macrophage nitric oxide generation, *P. labiosa* methanol extract has an anti-inflammatory effect.⁵⁵ In a combination of twelve medicinal herbs, *P. umbrosa* inhibits inflammatory mediators.⁵⁶ Catabolic and anabolic indicators of cartilage maintenance differ significantly using an herbal formulation including *P. umbrosa* and two additional species, which have been traditionally used to attenuate menopausal symptoms.¹⁰ The formulation also demonstrates no rise in estrogenic carcinogenesis; hence, it may be safe for the aforementioned applications.⁵⁷ To evaluate the efficacy of *P. umbrosa* in osteoporosis and enhancing bone development, the Saos-2 osteoblast cell line was treated with increasing concentrations of the plant extract, and the production of

osteoblast differentiation factors was measured. Following treatment, the mineralization ratio in differentiated osteoblasts rose significantly, as did the Runx2 level.⁸ Iridoid glycosides derived from *P. likiangensis* exhibited nitric oxide-dependent vasodilation in rat aortic rings.⁵⁸ The glucosidase inhibitory activity of the ethyl acetate fraction of the *P. tuberosa* extract, including diterpenoids, iridoids, and flavonoids, is comparable to that of the positive control acarbose.⁵⁹

In Vivo Studies

Anti-inflammatory effects

Mice with collagen-induced arthritis were given a mixture of 12 herbs, including *P. umbrosa*, determine any possible therapeutic benefits. Macroscopic analysis of paws and ankle TNF- α and IL-1 β levels demonstrated that the mixture has a significant cartilage-protecting effect.⁶⁰ The aqueous extract of *P. umbrosa* was tested for its anti-inflammatory and anti-nociceptive properties utilizing several techniques, including the acetic acid-induced writhing test (anti-nociceptive activity) and the carrageenan-induced paw edema test (anti-inflammatory activity). In the majority of these experiments, significant dose-dependent effects were seen. The above-mentioned benefits are attributed to the extract's predominant component, iridoid glucosides.⁶¹ The species was also evaluated in combination with *Angelica gigas* and *Cynanchum wilfordii*. Consequently, the mixture (200 mg/Kg) and celecoxib (60 mg/Kg) demonstrated equivalent inhibition percentages employing the carrageenan-induced paw edema assay.¹⁰ The monosodium iodoacetate-induced osteoarthritis model was employed to assess the protective properties of *P. umbrosa* extract. The extract reduced serum cytokine production, improved weight-bearing distribution, and ameliorated histological characteristics of osteoarthritic knee tissue.⁶² Regulating several genes, including those associated with the osteoarthritis pathway, was predicted as the molecular mechanism. By modulating the transcription factors, *P. umbrosa* extract reduces cartilage damage factors such as matrix metalloproteinases and increases chondrogenesis.⁶³ A systemic allergic response test was used to study the aqueous extract of *P. umbrosa* root; as a result, plasma histamine level and death rate were reduced in dose-dependent and time-dependent manners.⁶⁴ In addition, *P. umbrosa* was tested in an ovalbumin-induced asthma model using a 70% ethanol extract. Many parameters were affected, such as the production of inflammatory cytokines, airway inflammation, and eosinophilia. The findings introduced *P. umbrosa* as a potential agent for asthma management.⁶⁵ The carrageenan-induced paw edema test revealed an anti-inflammatory effect comparable to the positive control (aspirin) from the *P. laciniata* ethyl acetate fraction.⁶⁶ Moreover, the Tail Flick assay showed its analgesic and sedative properties in the crude extract and several fractions, particularly the chloroform fraction.⁶⁷ Analgesic effect's LD50/ED50 ratio of iridoids isolated from *P. labiosa*

exceeds diclofenac in the acetic acid-induced writhing assay and the hot plate test.⁶⁸

Bone effects

Oral administration of *P. umbrosa* to the ovariectomized mice for six weeks increased serum calcium concentration, bone mineral content, bone mineral density, and hyperplasia of the femoral growth plate.⁸ Unlike previous investigations, *P. umbrosa* in a mixture of 14 herbs showed a negligible impact on the protection of ovariectomized and calcium shortage-caused bone loss.⁶⁹ The ability of a standardized *P. umbrosa* root extract containing 6.62 mg/g shanzhiside methyl ester to improve bone development in adolescent female rats was investigated. The findings imply that the extract stimulates chondrocyte proliferation and hypertrophy with an increase in circulating insulin-like growth factor binding protein-3, which in turn enhances longitudinal bone growth.³³ Additionally, the combination of *Eleutherococcus senticosus*, *Astragalus membranaceus*, and *P. umbrosa* found in HT042 was examined in this respect. Consequently, enhancement was seen in trabecular bone mass, longitudinal bone growth, and bone microarchitecture during growth.⁷⁰ Some other studies with comparable results are also reported. In this context, the application of *P. umbrosa* mixture as a milk additive revealed a significant impact.⁷¹ However, Kim *et al.*⁷² showed that HT042 was ineffective for longitudinal bone growth in spontaneous dwarf rats.

Miscellaneous effects

In a carbon tetrachloride-induced liver damage model, the hepatoprotective activity of *P. maximowiczii* buthanolic extract was examined because of its substantial antioxidant action *in vitro* and the fact that oxidative stress is one of the mechanisms responsible for liver injury. In lowering the amount of malondialdehyde (a sign of oxidative stress) and raising the level of superoxide dismutase, the effects of this plant were comparable to those of the positive control (Bifendate).³⁹

Moreover, the antidepressant effect of *P. laciniata* was evaluated using a forced-swim test. Apigenin and luteolin were assumed responsible for the initial anti-depressive effect and the sedative properties at higher doses, respectively.⁷³

Safety studies

HT042 oral toxicity was studied in Sprague-Dawley rats by Song *et al.*⁷⁴ The fatal dosage was more than 5000 mg/Kg orally for the mixture and every species, including *P. umbrosa*. Moreover, sub-chronic toxicity tests up to 4000 mg/kg/day did not demonstrate any adverse effects.

Clinical Trials

Generally, the effects of *Phlomis* species have been investigated in seven different clinical trials to date (Table 4). Two clinical trials in Iran evaluated the analgesic effects of *P. laciniata*. A study by Gharabaghi *et al.*⁷⁵ examined

Table 4. Clinical studies on Phlomoides species.

| Status | Design | Country | Herbal extract | Population (n=patients) | Intervention group(s) | Comparison group(s) | Primary outcomes | Ref |
|----------------------------------|--|---------|---|--|--|--|---|-----|
| Analgesic Effects | | | | | | | | |
| Completed | A Double-blind randomized, placebo-controlled parallel study | Iran | Extract of <i>P. laciniata</i> (As suppository) | (N=90) females older than 35 years and candidate for selective hysterectomy | (N=30) Suppository of <i>P. laciniata</i> administered (35mg) 24h before to 24h after surgery every 12h | Case I(N=30): Suppository of placebo (contained starch and cacao butter) administered 24h before surgery and Suppository of <i>P. laciniata</i> administered 24h after surgery every 12h Case II(N=30): Suppository of placebo administered 24h before to 24h after surgery every 12h | A higher pain mitigation rate was observed in the patients who were given herbal extract before and after surgery compared to the placebo group. Results of pain severity evaluation after surgery showed that the herbal extract group was less painful than the placebo group. Prescribed sedative doses were also less. | 75 |
| Completed | A triple-blind controlled clinical trial | Iran | Rectal <i>P. laciniata</i> suppository was prepared from a 3.5% total rhizome extract | (N=86) women who gave childbirth by cesarean section | 35 mg <i>P. laciniata</i> suppository administered every 8 h up to three doses (8, 16, and 24 h after surgery) | 50 mg rectal diclofenac suppository administered at the same time | According to Friedman's test, a significant difference was found between the control group's distress score and the intervention group's score 24 hours after cesarean section. Wound healing scores also differed significantly between the two groups, indicating that rectal diclofenac suppository was more effective than <i>P. laciniata</i> . In both groups, no side effects were observed. | 76 |
| Anti-Inflammatory Effects | | | | | | | | |
| Completed | A Double-blind, Randomized Clinical Trial | Iran | Methanol extract of <i>P. laciniata</i> (As Ointment) | (N=40) patients between 15 and 65 years with mild and moderate CTS and without any other neurological problems | <i>P. laciniata</i> ointment was applied topically to the palmar wrist area twice a day for a period of 4 weeks | Eucerin ointment (as placebo) was applied topically twice a day for a period of 4 weeks | After four weeks, the herbal extract did not affect hand grip power. Also, this herbal extract had no significant effect on nerve conduction parameters. | 77 |
| Completed | A single-blinded randomized clinical trial | Iran | Methanol extract of <i>P. laciniata</i> (As Ointment) | (N=137) patients between 18-80 years with inflammatory diseases, e.g., osteoarthritis, rheumatoid arthritis and Reiter's syndrome. | The 5% <i>P. laciniata</i> ointment were gently massaged around the affected joint two times a day, for a consecutive 14 days. | The 5% piroxicam ointment were gently massaged around the affected joint two times a day, for a consecutive 14 days. | After 14 days of treatment with the <i>P. laciniata</i> and piroxicam ointments, all groups showed significant improvement compared to the control groups. <i>P. laciniata</i> (5%) ointment induced better initial therapeutic response than piroxicam (5%) ointment. | 78 |

Table 4. Continued.

| Bone Effects | | | | | | | | |
|------------------------|---|-------------|--|---|---|--|---|----|
| Completed | A double-blinded, randomized, placebo-controlled study | South Korea | Multi-herbal mixture consisting of <i>Eleutherococcus senticosus</i> , <i>Astragalus membranaceus</i> and <i>P. umbrosa</i> (HT042) | (N=99) children aged from 7 to 12 years with initial height below the 25th percentile for age and sex | 750 mg of HT042 administered twice daily for 12 weeks | Placebo administered twice daily for 12 weeks | In the HT042 group, fat-free mass, height percentile, and SDS increased, and fat mass notably decreased from baseline to week 12. A significant difference in the level of IGFBP-3, but not GH or IGF-1, was shown between the placebo and HT042 groups. There is no significant difference between the two groups in the nutrient intake or the amount of time spent exercising at the end of the study. | 79 |
| Active, not recruiting | A 6-month, multicenter, randomized, double-blind, placebo-controlled parallel study | South Korea | HT042 consists of the lyophilized 70% ethanolic extracts of three medicinal plants: the roots of <i>A. membranaceus</i> , the stems of <i>E. senticosus</i> , and the roots of <i>P. umbrosa</i> . | (N=140) children aged 6–8 years with height ranked below the 25th percentile | 750 mg of HT042 administered twice daily for 24 weeks | Placebo administered twice daily for 24 weeks | After 24 weeks, the HT042 group had a significantly greater height gain than the placebo group. Compared with baseline, serum IGF-1 and IGFBP-3 levels were significantly higher in the HT042 group. However, the group difference was not significant. | 80 |
| Estrogenic Effects | | | | | | | | |
| Completed | A randomized double-blind, Placebo-controlled Study | USA | Mixture of standardized extracts of <i>Cynanchum wilfordii</i> , <i>P. umbrosa</i> and <i>Angelica gigas</i> (EstroG-100) | (N=64) pre-, peri- and postmenopausal White Hispanic, White non-Hispanic and African American women | (N=31) one tablet (EstroG-100) administered twice a day orally for 12 weeks | (N=33) one placebo tablet administered twice a day orally for 12 weeks | A significant reduction in mean Kupperman menopause index (KMI), a significant improvement in constituting symptoms of vasomotor, paresthesia, insomnia, nervousness, melancholia, vertigo, fatigue, and rheumatic pain, as well as a significant amelioration in vaginal dryness in the intervention group compared to the placebo group, were reported. | 81 |

the analgesic effect of *P. laciniata* suppositories on women undergoing a hysterectomy. The results demonstrated that the rate of postoperative pain mitigation was higher in the intervention group than in the placebo group. After surgery, an assessment of pain intensity at different time intervals revealed that the group receiving herbal extract experienced less pain than the placebo group. The prescription of a total dose of sedatives was also declined. Mohammad Pour *et al.*⁷⁶ evaluated the analgesic effects of *P. laciniata* suppositories on pain and distress after cesarean delivery. According to their results, a rectal suppository of *P. laciniata* could effectively manage pain and distress following cesarean delivery with low side effects and acceptable performance. However, rectal diclofenac suppository as control was significantly more effective than the extract. Anti-inflammatory effects were evaluated in two different clinical trials in Iran. Eftekharsadat *et al.*⁷⁷ investigated the effects of *P. laciniata* ointment compared with a placebo group on patients suffering from carpal tunnel syndrome (CTS). The herbal extract did not affect handgrip strength. Besides, it did not significantly alter nerve conduction parameters over four weeks. Additional studies with larger sample sizes are needed to assess the long-term efficacy of this therapy. A clinical trial conducted by Delazar *et al.*⁷⁸ revealed that topical *P. laciniata* extract could be a safe and effective complementary treatment for patients with arthritis, rheumatoid arthritis, and Reiter's syndrome. Two clinical trials in South Korea investigated the effects of HT402 extract on bones. Studies were performed on children aged 7 to 12 years for three months,⁷⁹ and children aged 6 to 8 years for six months,⁸⁰ and the results were compared with those of placebo groups. These studies indicated that HT042 treatment for short children increased height and changed body composition positively. HT042 supplementation contributed to height growth in children without skeletal maturation, mainly for those who were much shorter, by increasing serum levels of IGF-1 and IGFBP-3.^{79,80} The estrogenic effects of *P. umbrosa* were investigated in a randomized, double-blind, placebo-controlled trial by Chang *et al.*⁸¹ A twelve-week therapy with EstroG-100 (a combination of *Angelica gigas*, *Cynanchum wilfordii*, and *P. umbrosa*) improved many menopausal ailments and vaginal dryness. The use of EstroG-100 was not associated with any adverse effects. Unlike hormone replacement therapy, no significant alterations in serum FSH, serum estrogen, BMI, body weight, and liver enzymes were reported. According to this study, EstroG-100 is a safe and effective supplement for women in the various menopausal phases.

Conclusion

Plant systematics undergo permanent modifications through ongoing studies. Common categorization approaches include morphology, genetics, and phytochemistry. Chemotaxonomy is of great importance because of its association with drug development. Recently, some species formerly classified in the genera *Eremostachys*

and *Phlomis* were moved to the *Phlomoidea* genus. It appears that evaluating all species under the latest classification might prevent future misunderstandings and biases in the area of position changes and names. Ethnomedical applications of these species, including wound healing effects, treatment for stomach disorders, and bone fracture therapy, have provided the basis for further evaluations. Their main identified secondary metabolites, iridoids, furanolanthenes, and phenylethanoids, are well known for various biological activities, such as anti-inflammatory effects. A United States patent (US8790727B2) introduced some iridoid-rich plant mixtures, including *Phlomoidea glabra*, as nutraceuticals having potent anti-inflammatory characteristics. Clinical trials, *in vivo*, and *in vitro* studies have also suggested potential therapeutic effects of the species. *P. umbrosa* mixtures significantly improved children's height and alleviated menopausal symptoms. *P. laciniata* was clinically effective as an analgesic and anti-inflammatory agent. As societies age, the prevalence of arthritis, inflammatory joint issues, bone fractures, and menopausal problems rises. In addition, due to the identification of plants as trustworthy sources of medicine, further studies on these plants could be beneficial and of interest.

Author Contributions

Samin Mohammadi: Formal Analysis, Investigation, Writing - Original Draft. Behzad Jafari: Formal analysis, Writing - Original Draft. Samira Pourtaghi Anvarian: Investigation, Writing - Original Draft. Hossein Nazemiyeh: Writing - Review & Editing. Solmaz Asnaashari: Writing - Review & Editing. Abbas Delazar: Supervision. Parina Asgharian: Conceptualization, Methodology, Funding acquisition, Writing - Review & Editing.

Acknowledgements

This is a report of database from PhD thesis registered in Faculty of Pharmacy, Tabriz University of Medical Sciences with the grant number of 64124.

Conflict of Interest

The authors report no conflicts of interest.

Supplementary Data

Supplementary data, Table S1, are available at <https://doi.org/10.34172/PS.2023.1>.

References

1. Fowler MW. Plants, medicines and man. *J Sci Food Agric.* 2006;86(12):1797-804. doi:10.1002/jsfa.2598
2. Katiyar C, Gupta A, Kanjilal S, Katiyar S. Drug discovery from plant sources: An integrated approach. *AYU.* 2012;33(1):10-9. doi: 10.4103/0974-8520.100295
3. Raja RR. Medicinally potential plants of labiateae (lamiaceae) family: An overview. *J Med Plant Res.* 2012;6(3):203-13. doi:10.3923/rjmp.2012.203.213
4. Ranjbar M, Mahmoudi C. A taxonomic note on the

- phlomoides sect. *Filipendula* (phlomisaceae, lamioideae, lamiaceae) from Iran. Feddes Repert. 2017;128(1-2):36-41. doi:10.1002/fedr.201600025
5. WFO. World flora online. 2022 [July 2022]; Available from: <http://www.worldfloraonline.org/>.
 6. Li MX, Shang XF, Jia ZP, Zhang RX. Phytochemical and biological studies of plants from the genus *phlomis*. Chem Biodivers. 2010;7(2):283-301. doi: 10.1002/cbdv.200800136
 7. Mohammadhosseini M, Frezza C, Venditti A, Akbarzadeh A. Ethnobotany and phytochemistry of the genus *Eremostachys bunge*. Curr Org Chem. 2019;23(17):1828-42. doi:10.2174/1385272823666191007161550
 8. Lee JE, Lee H, Kim MH, Yang WM. Osteogenic effects of *phlomis umbrosa* via up-regulation of *runx2* in osteoporosis. Biomed Rep. 2019;10(1):17-22. doi:10.3892/br.2018.1172
 9. Dinda B, Debnath S, Banik R. Naturally occurring iridoids and secoiridoids. An updated review, part 4. Chem Pharm Bull. 2011;59(7):803-33. doi:10.1248/cpb.59.803
 10. Kim J, Yang S, Choi CY. The evaluation of the effect of herbal extract on osteoarthritis: In vitro and in vivo study. Prev Nutr Food Sci. 2016;21(4):310-6. doi:10.3746/pnf.2016.21.4.310
 11. Amor ILB, Boubaker J, Sgaier MB, Skandrani I, Bhourri W, Neffati A, et al. Phytochemistry and biological activities of *phlomis* species. J Ethnopharmacol. 2009;125(2):183-202. doi:10.1016/j.jep.2009.06.022
 12. Salmaki Y, Zarre S, Heubl G. The genus *phlomis* *moench* (lamiaceae; lamioideae; phlomisaceae) in Iran: An updated synopsis. Iran J Bot. 2012;18(2):207-19. doi:10.22092/IJB.2012.101424
 13. Page MJ, McKenzie JE, Bossuyt PM, Boutron I, Hoffmann TC, Mulrow CD, et al. The PRISMA 2020 statement: An updated guideline for reporting systematic reviews. BMJ. 2021;372:n71. doi:10.1136/bmj.n71
 14. Bennett BC, Balick MJ. Does the name really matter? The importance of botanical nomenclature and plant taxonomy in biomedical research. J Ethnopharmacol. 2014;152(3):387-92. doi:10.1016/j.jep.2013.11.042
 15. Govindaraghavan S, Hennell J, Sucher N. From classical taxonomy to genome and metabolome: Towards comprehensive quality standards for medicinal herb raw materials and extracts. Fitoterapia. 2012;83:979-88. doi:10.1016/j.fitote.2012.05.001
 16. Harley RM, Atkins S, Budantsev AL, Cantino PD, Conn BJ, Grayer R, et al. Labiatae. In: Kadereit, JW. editor, Flowering Plants Dicotyledons. The Families and Genera of Vascular Plants, vol 7. Heidelberg, Berlin: Springer; 2004.
 17. Scheen A-C, Bendiksby M, Ryding O, Mathiesen C, Albert VA, Lindqvist C. Molecular phylogenetics, character evolution and suprageneric classification of Lamioideae (Lamiaceae). Ann Missouri Bot Gard. 2010;97:191-219. doi:10.3417/2007174
 18. Bendiksby M, Thorbek L, Scheen A-C, Lindqvist C, Ryding O. An updated phylogeny and classification of lamiaceae subfamily lamioideae. Taxon. 2011;60(2):471-84. doi:10.1002/tax.602015
 19. Mathiesen C, Scheen AC, Lindqvist C. Phylogeny and biogeography of the lamioideae genus *phlomis* (lamiaceae). Kew Bull. 2011;66(1):83-99. doi:10.1007/s12225-011-9257-0
 20. Ryding O. Pericarp structure and phylogeny of the *phlomis* group (lamiaceae subfam. Lamioideae). Bot Jahrb Syst Pflanzengesch. 2008;127:299-316. doi:10.1127/0006-8152/2008/0127-0002
 21. Salmaki Y, Zarre S, Ryding O, Lindqvist C, Scheunert A, Bräuchler C, et al. Phylogeny of the tribe phlomisaceae (lamioideae: Lamiaceae) with special focus on *eremostachys* and *phlomis*: New insights from nuclear and chloroplast sequences. Taxon. 2012;61(1):161-79. doi:10.1002/tax.611012
 22. Seyedi Z, Salmaki Y. Trichome morphology and its significance in the systematics of *phlomis* (lamiaceae; lamioideae; phlomisaceae). Flora. 2015;213:40-8. doi:10.1016/j.flora.2015.04.003
 23. Ranjbar M, Mahmoudi C. A revision of *phlomis* sect. *Thyrsiflorae* (lamiaceae). Webbia. 2015;70(2):237-45. doi:10.1080/00837792.2015.1041794
 24. Eyvazadeh Khosroshahi E, Salmaki Y. Evolution of trichome types and its systematic significance in the genus *Phlomis* (lamioideae-lamiaceae). Nord J Bot. 2019;37:5. doi:10.1111/njb.02132
 25. Evans WC. Trease and Evans' pharmacognosy. New York: Saunders/Elsevier; 2009.
 26. Kokkini S, Karousou R, Hanlidou E. Herbs | herbs of the Labiatae. Caballero B, Trugo LC, Finglas MP, editors. Encyclopedia of Food Sciences and Nutrition; 2003. p. 3082-90.
 27. Jamzad Z. Flora of Iran vol. 76 lamiaceae. Tehran: Research Institute of Forests & Rangelands; 2012.
 28. Liu K, Abdullah AA, Huang M, Nishioka T, Altaf-Ul-Amin M, Kanaya S. Novel approach to classify plants based on metabolite-content similarity. Biomed Res Int. 2017;2017:5296729. doi:10.1155/2017/5296729
 29. Chin CH, Chen SH, Wu HH, Ho CW, Ko MT, Lin CY. Cytohubba: Identifying hub objects and sub-networks from complex interactome. BMC Syst Biol. 2014;8(Suppl 4):S11. doi:10.1186/1752-0509-8-s4-s11
 30. Heinrich M. Ethnobotany and its role in drug development. Phytother Res. 2000;14(7):479-88. doi:10.1002/1099-1573(200011)14:7<479::aid-ptr958>3.0.co;2-2
 31. Mükemre M, Behçet L, Çakılcıoğlu U. Ethnobotanical study on medicinal plants in villages of Çatak (Van-turkey). J Ethnopharmacol. 2015;166:361-74. doi:10.1016/j.jep.2015.03.040
 32. Rehman K, Mashwani ZUR, Khan MA, Ullah Z, Chaudhary HJ. An ethnobotanical perspective of traditional medicinal plants from the Khattak tribe

- of chonthra karak, pakistan. J Ethnopharmacol. 2015;165:251-9. doi:10.1016/j.jep.2015.02.035
33. Lee D, Kim YS, Song J, Kim HS, Lee HJ, Guo H, et al. Effects of *Phlomis umbrosa* root on longitudinal bone growth rate in adolescent female rats. Molecules. 2016;21(4):461. doi:10.3390/molecules21040461
 34. Naghibi F, Mosadegh M, Mohammadi MS, Ghorbani A. Labiatae family in folk medicine in iran: From ethnobotany to pharmacology. Iran J Pharm Res. 2005;4(2):e128228. doi:10.22037/ijpr.2010.619
 35. Delazar A, Asl BH, Mohammadi O, Afshar FH, Nahar L, Modarresi M, et al. Evaluation of analgesic activity of *Eremostachys laciniata* in mice. J Nat Remedies. 2009;9(1):1-7. doi:10.18311/jnr/2009/213
 36. Adams M, Gmünder F, Hamburger M. Plants traditionally used in age related brain disorders--a survey of ethnobotanical literature. J Ethnopharmacol. 2007;113(3):363-81. doi:10.1016/j.jep.2007.07.016
 37. Singh KN, Lal B. Ethnomedicines used against four common ailments by the tribal communities of lahaul-spiti in western himalaya. J Ethnopharmacol. 2008;115(1):147-59. doi:10.1016/j.jep.2007.09.017
 38. Ullah R, Al-Zeghayer YS, Haider S. Immunomodulatory potential of *Phlomis bracteosa*. Afr J Pharm Pharmacol. 2011;5(15):1811-2. doi:10.5897/AJPP11.549
 39. Gu H, Gu X, Xu Q, Kang W. Antioxidant activity in vitro and hepatoprotective effect of *Phlomis maximowiczii* in vivo. Afr J Tradit Complement Altern Med. 2014;11(3):46-52. doi:10.4314/ajtcam.v11i3.8
 40. Erdemoglu N, Turan NN, Cakici I, Sener B, Aydin A. Antioxidant activities of some lamiaceae plant extracts. Phytother Res. 2006;20(1):9-13. doi:10.1002/ptr.1816
 41. Zhang Y, Wang Zz. Phenolic composition and antioxidant activities of two phlomis species: A correlation study. C R Biol. 2009;332(9):816-26. doi:10.1016/j.crvi.2009.05.006
 42. Asnaashari S, Afshar FH, Ebrahimi A, Moghadam SB, Delazar A. Chemical composition and radical scavenging activity of essential oil and methanolic extract of *Eremostachys azerbaijanica* rech.F. From iran. Res Pharm Sci. 2016;11(2):113-9.
 43. Le DD, Nguyen DH, Zhao BT, Min BS, Song SW, Woo MH. Quantitation and radical scavenging activity evaluation of iridoids and phenylethanoids from the roots of *Phlomis umbrosa* (turcz.) using DPPH free radical and DPPH-hplc methods, and their cytotoxicity. Nat Prod Sci. 2019;25(2):122-9. doi:10.20307/nps.2019.25.2.122
 44. Li XH, Lu LH, Li XH, Li YB, Zha YF, Fu DH, et al. Iridoid glycosides from phlomis likiangensis with free-radical scavenging activity. Tetrahedron Lett. 2017;58(46):4395-9. doi:10.1016/j.tetlet.2017.10.020
 45. Mohammadi S, Valizadeh H, Khalesh F, Bastani S, Delazar A, Asgharian P. Biological activities of extract-loaded nanocarriers: A comparison of aerial part, seed, and rhizome of phlomoïdes labiosa. Eur J Integr Med. 2022;52:102135. doi:10.1016/j.eujim.2022.102135
 46. Suleimen EM, Gorovoi PG, Dudkin RV, Drozdov KA, Tashenov EO, Iskakova ZB. Constituent composition and biological activity of essential oil from *Phlomis maximowiczii*. Chem Nat Compd. 2017;53(6):1186-8. doi:10.1007/s10600-017-2235-5
 47. Samandari-Bahraseman MR, Jahanshahi M, Asadi Barbariha S, Elyasi L. Altered micro-RNA regulation and neuroprotection activity of eremostachys labiosiformis in alzheimer's disease model. Ann Neurosci. 2019;25(3):160-5. doi:10.1159/000489551
 48. Asgharian P, Delazar A, Lotfipour F, Asnaashari S. Bioactive properties of *Eremostachys macrophylla* montbr. & auch. Rhizomes growing in iran. Pharm Sci. 2017;23(3):238-43. doi:10.15171/PS.2017.35
 49. Şenol H, Tulay P, Ergören M, Hanoğlu A, Çalış İ, Mocan G. Cytotoxic effects of verbascoside on MCF-7 and MDA-MB-231. Turk J Pharm Sci. 2021;18(5):637-44. doi:10.4274/tjps.galenos.2021.36599
 50. Modaressi M, Delazar A, Nazemiyeh H, Fathi-Azad F, Smith E, Rahman MM, et al. Antibacterial iridoid glucosides from *Eremostachys laciniata*. Phytother Res. 2009;23(1):99-103. doi:10.1002/ptr.2568
 51. Gonchig E, Erdenebat S, Togtoo O, Bataa S, Gendaram O, Young SK, et al. Antimicrobial activity of mongolian medicinal plants. Nat Prod Sci. 2008;14(1):32-6.
 52. Hou Y-T, Wu F, Yao J-H, Zhu Z-H, Mi Q-L, Gao Q, et al. Chemical constituents of the roots of phlomis betonicoides and their anti-rotavirus activity. Chem Nat Compd. 2021;57(5):864-8. doi:10.1007/s10600-021-03499-4
 53. Asnaashari S, Afshar FH, Moghadam SB, Delazar A. Evaluation of in vitro antimalarial activity of different extracts of *Eremostachys azerbaijanica* rech.F. Iran J Pharm Res. 2016;15(3):523-9.
 54. Anjum S, Abbasi BH. Biomimetic synthesis of antimicrobial silver nanoparticles using in vitro-propagated plantlets of a medicinally important endangered species: *Phlomis bracteosa*. Int J Nanomedicine. 2016;11:1663-75. doi:10.2147/IJN.S105532
 55. Zamani Taghizadeh Rabe S, Mahmoudi M, Ahmadsimab H, Zamani Taghizadeh Rabe SS, Emami A. Investigation of the biological activity of methanol extract from *Eremostachys labiosa* bunge. Food Agric Immunol. 2014;25(4):578-85. doi:10.1080/09540105.2013.858311
 56. Choi J, Kim SH, Kim S. Suppressive effects of PG201, an antiarthritic botanical formulation, on lipopolysaccharide-induced inflammatory mediators in Raw264.7 cells. Exp Biol Med. 2012;237(5):499-508. doi:10.1258/ebm.2011.011203
 57. Kim SJ, Jin SW, Lee GH, Kim YA, Jeong HG. Evaluation of Estrogenic Activity of Extract from the Herbal Mixture *Cynanchum wilfordii* Hemsley, *Phlomis umbrosa* Turczaninow, and *Angelica gigas* Nakai. Toxicol Res. 2017;33(1):71-7. doi:10.5487/TR.2017.33.1.071
 58. Hu LJ, Qi YY, Chen KJ, Yang C, Wu HY, Li

- GP. Vasodilatory effect and structural-activity relationship of a group of iridoid glucosides from *Phlomis likiangensis*. *Fitoterapia*. 2019;139: 104365. doi:10.1016/j.fitote.2019.104365
59. Yang Y, Gu L, Xiao Y, Liu Q, Hu H, Wang Z, et al. Rapid identification of α -glucosidase inhibitors from *phlomis tuberosa* by sepbox chromatography and thin-layer chromatography bioautography. *PLoS One*. 2015;10(2):e0116922. doi:10.1371/journal.pone.0116922
60. Shin SS, Jin M, Jung HJ, Kim B, Jeon H, Choi JJ, et al. Suppressive effects of PG201, an ethanol extract from herbs, on collagen-induced arthritis in mice. *Rheumatology* 2003;42(5):665-72. doi:10.1093/rheumatology/keg209
61. Shang X, Wang J, Li M, Miao X, Pan H, Yang Y, et al. Antinociceptive and anti-inflammatory activities of *Phlomis umbrosa* turcz extract. *Fitoterapia*. 2011;82(4):716-21. doi:10.1016/j.fitote.2011.03.001
62. Chun JM, Lee AY, Moon BC, Choi G, Kim JS. Effects of *Dipsacus asperoides* and *Phlomis umbrosa* extracts in a rat model of osteoarthritis. *Plants*. 2021;10(10):2030. doi: 10.3390/plants10102030
63. Chun JM, Lee AY, Nam JY, Lee MY, Choe MS, Lim KS, et al. Protective effects of *Phlomis umbrosa* extract on a monosodium iodoacetate-induced osteoarthritis model and prediction of molecular mechanisms using transcriptomics. *Phytomedicine*. 2021;81:153429. doi:10.1016/j.phymed.2020.153429
64. Shin TY, Kim SH, Kim DK, Leem KH, Park JS. *Phlomis umbrosa* root inhibits mast cell-dependent allergic reactions and inflammatory cytokine secretion. *Phytother Res*. 2008;22(2):153-8. doi:10.1002/ptr.2164
65. Pak S-W, Lee AY, Seo Y-S, Lee S-J, Kim W-I, Shin D-H, et al. Anti-asthmatic effects of *phlomis umbrosa* turczaninow using ovalbumin induced asthma murine model and network pharmacology analysis. *Biomed Pharmacother*. 2022;145:112410. doi:10.1016/j.biopha.2021.112410
66. Khan S, Nisar M, Rehman W, Khan R, Nasir F. Anti-inflammatory study on crude methanol extract and different fractions of *Eremostachys laciniata*. *Pharm Biol*. 2010;48(10):1115-8. doi:10.3109/13880200903517950
67. Khan S, Nisar M, Simjee SU, Rehman W, Khan R, Jan I, et al. Evaluation of micronutrients level and antinociceptive property of *Eremostachys laciniata* (L) bunge. *Afr J Biotechnol*. 2010;9(5):775-7. doi:10.5897/AJB09.1367
68. Usmanov D, Azamatov A, Baykuziyev T, Yusupova U, Rasulev B. Chemical constituents, anti-inflammatory and analgesic activities of iridoids preparation from *Phlomooides labiosa* bunge. *Nat Prod Res*. 2022;37(10):1709-13. doi:10.1080/14786419.2022.2104274
69. Lee JW, Jhee O, Yuan H, Kim T, Kim D, Lee M, et al. Effect of korean oriental medicine extract on bone mass as compared with alendronate in ovariectomized rats. *J Med Food*. 2005;8(3):369-76. doi:10.1089/jmf.2005.8.369
70. Song J, Lee SH, Lee D, Kim H. Astragalus extract mixture ht042 improves bone growth, mass, and microarchitecture in prepubertal female rats: A microcomputed tomographic study. *Evid Based Complement Altern Med*. 2017;2017:5219418. doi:10.1155/2017/5219418
71. Lee JK, Lee DG, Kwak BY, Kim JS, Yi K. Effect of combined extract of *hansogdan* (*Phlomis umbrosa*) and *dalgaebi* (*Commelina communis*) as a milk additive for enhancing the growth of physical height in vivo. *Food Sci Biotechnol*. 2012;21(3):875-9. doi:10.1007/s10068-012-0113-2
72. Kim JY, Song M, Lee D, Song J, Park SW, Park J, et al. Effect of HT042, herbal formula, on longitudinal bone growth in spontaneous dwarf rats. *Molecules*. 2013;18(11):13271-82. doi:10.3390/molecules181113271
73. Nisar M, Khan S, Dar A, Rehman W, Khan R, Jan I. Antidepressant screening and flavonoids isolation from *Eremostachys laciniata* (L) bunge. *Afr J Biotechnol*. 2011;10(9):1696-9. doi:10.5897/AJB10.1254
74. Song J, Lee D, Min B, Bae JS, Chang GT, Kim H. Safety evaluation of astragalus extract mixture ht042 and its constituent herbs in sprague-dawley rats. *Phytomedicine*. 2017;32:59-67. doi:10.1016/j.phymed.2017.03.005
75. Gharabagy PM, Zamany P, Delazar A, Ghojzadeh M, Goldust M. Efficacy of *Eremostachys laciniata* herbal extract on mitigation of pain after hysterectomy surgery. *Pak J Biol Sci*. 2013;16(17):891-4. doi:10.3923/pjbs.2013.891.894
76. Mohammad Pour S, Hakimi S, Delazar A, Javad Zadeh Y, Mallah F. *Eremostachys laciniata* as effective as rectal diclofenac suppository in cesarean section pain relief: A triple-blind controlled clinical trial. *J Endometr Pelvic Pain Disord*. 2020;12(1):26-34. doi:10.1177/2284026519899010
77. Eftekharsadat B, Kazem Shakouri S, Shimia M, Rahbar M, Ghojzadeh M, Rashidi MR, et al. Effect of *E. Laciniata* (L) ointment on mild and moderate carpal tunnel syndrome: A double-blind, randomized clinical trial. *Phytother Res*. 2011;25(2):290-5. doi:10.1002/ptr.3248
78. Delazar A, Sarker SD, Nahar L, Jalali SB, Modaresi M, Hamedeyazdan S, et al. Rhizomes of *Eremostachys laciniata*: Isolation and structure elucidation of chemical constituents and a clinical trial on inflammatory diseases. *Adv Pharm Bull*. 2013;3(2):385. doi:10.5681/apb.2013.062
79. Song J, Lee D, Kim HS, Lee HJ, Park J, Kim H. The effects of a multi-herbal mixture HT042 on height gain in short children: A double-blind, randomized, placebo-controlled trial (LB322). *FASEB J*. 2014;28(S1):LB322. doi:10.1096/fasebj.28.1_supplement.lb322
80. Lee D, Lee SH, Song J, Jee HJ, Cha SH, Chang GT.

Effects of *Astragalus extract* mixture HT042 on height growth in children with mild short stature: A multicenter randomized controlled trial. *Phytother Res.* 2018;32(1):49-57. doi:10.1002/ptr.5886

extract (estrog-100) on pre-, peri-and post-menopausal women: A randomized double-blind, placebo-controlled study. *Phytother Res.* 2012;26(4):510-6. doi:10.1002/ptr.3597

81. Chang A, Kwak BY, Yi K, Kim JS. The effect of herbal