

CORRECTED VERSION

(19) World Intellectual Property Organization  
International Bureau



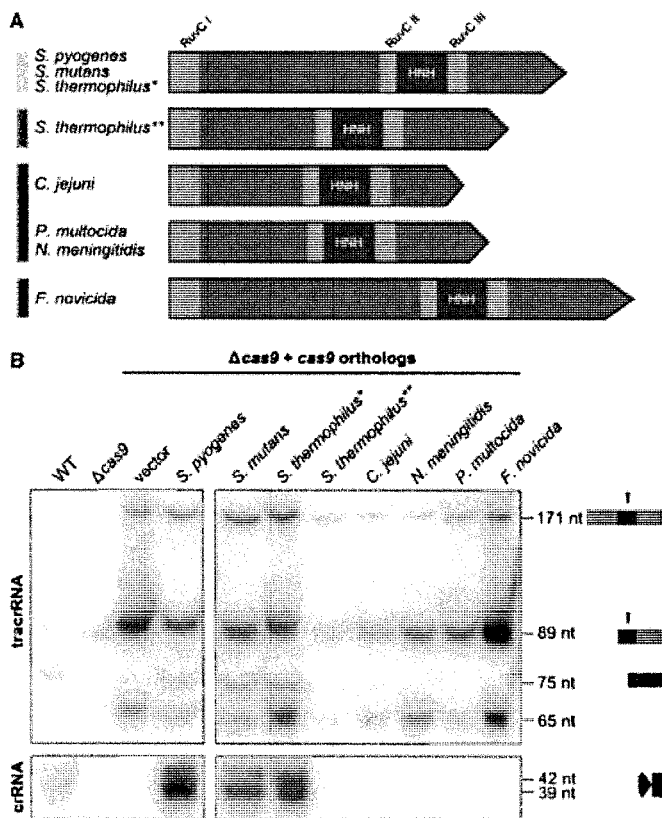
(10) International Publication Number  
**WO 2015/071474 A9**

(43) International Publication Date  
21 May 2015 (21.05.2015)

- (51) International Patent Classification:  
*C12N 15/11* (2006.01)    *C12N 15/63* (2006.01)  
*C07K 14/205* (2006.01)    *C12N 15/90* (2006.01)  
*C12N 9/22* (2006.01)
- (21) International Application Number:  
PCT/EP2014/074813
- (22) International Filing Date:  
17 November 2014 (17.11.2014)
- (25) Filing Language: English
- (26) Publication Language: English
- (30) Priority Data:  
61/905,835 18 November 2013 (18.11.2013) US
- (71) Applicant: **CRISPR THERAPEUTICS AG** [CH/CH];  
Aeschenvorstadt 36, CH-4051 Basel (CH).
- (72) Inventors: **CHARPENTIER, Emmanuelle**; Boecklerstrasse 18, 38102 Braunschweig (DE). **CHYLINSKI, Krzysztof**; Simmeringer Hauptstrasse 45/8, A-1110 Vienna (AT). **FONFARA, Ines**; Helmstedter Strasse 144, 38102 Braunschweig (DE).
- (74) Agent: **PILKINGTON, Stephanie Joan**; Potter Clarkson LLP, The Belgrave Centre, Talbot Street, Nottingham NG1 5GG (GB).
- (81) Designated States (*unless otherwise indicated, for every kind of national protection available*): AE, AG, AL, AM, AO, AT, AU, AZ, BA, BB, BG, BH, BN, BR, BW, BY, BZ, CA, CH, CL, CN, CO, CR, CU, CZ, DE, DK, DM,

[Continued on next page]

(54) Title: CRISPR-CAS SYSTEM MATERIALS AND METHODS



(57) Abstract: The invention relates to Type II CRISPR-Cas systems of Cas9 enzymes, guide RNAs and associated specific PAMs.

Figure 4

WO 2015/071474 A9



DO, DZ, EC, EE, EG, ES, FI, GB, GD, GE, GH, GM, GT, HN, HR, HU, ID, IL, IN, IR, IS, JP, KE, KG, KN, KP, KR, KZ, LA, LC, LK, LR, LS, LU, LY, MA, MD, ME, MG, MK, MN, MW, MX, MY, MZ, NA, NG, NI, NO, NZ, OM, PA, PE, PG, PH, PL, PT, QA, RO, RS, RU, RW, SA, SC, SD, SE, SG, SK, SL, SM, ST, SV, SY, TH, TJ, TM, TN, TR, TT, TZ, UA, UG, US, UZ, VC, VN, ZA, ZM, ZW.

**(84) Designated States** (unless otherwise indicated, for every kind of regional protection available): ARIPO (BW, GH, GM, KE, LR, LS, MW, MZ, NA, RW, SD, SL, ST, SZ, TZ, UG, ZM, ZW), Eurasian (AM, AZ, BY, KG, KZ, RU, TJ, TM), European (AL, AT, BE, BG, CH, CY, CZ, DE, DK, EE, ES, FI, FR, GB, GR, HR, HU, IE, IS, IT, LT, LU, LV, MC, MK, MT, NL, NO, PL, PT, RO, RS, SE, SI, SK, SM, TR), OAPI (BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, KM, ML, MR, NE, SN, TD, TG).

**Published:**

— with international search report (Art. 21(3))

- with sequence listing part of description (Rule 5.2(a))
- with information concerning authorization of rectification of an obvious mistake under Rule 91.3 (b) (Rule 48.2(i))

**(88) Date of publication of the international search report:**  
27 August 2015

**Date of publication of the revised international search report:**  
3 December 2015

**(48) Date of publication of this corrected version:**  
21 January 2016

**(15) Information about Corrections:**  
see Notice of 21 January 2016

**Previous Correction:**  
see Notice of 3 December 2015

## **CRISPR-CAS SYSTEM MATERIALS AND METHODS**

### **Field of the Invention**

**[0001]** The invention relates to type II CRISPR-Cas systems of Cas9 enzymes, guide RNAs and associated specific PAMs. This application claims the benefit of the filing date of U.S. Provisional Patent Application No. 61/905,835 filed November 18, 2013, which is incorporated by reference herein in its entirety.

### **Incorporation by Reference of the Sequence Listing**

**[0002]** This application contains, as a separate part of disclosure, a Sequence Listing in computer-readable form (filename: 48128\_SeqListing.txt; 7,869,256 bytes – ASCII text file; created November 14, 2014) which is incorporated by reference herein in its entirety.

### **Background**

**[0003]** Editing genomes using the RNA-guided DNA targeting principle of CRISPR-Cas (Clustered Regularly Interspaced Short Palindromic Repeats-CRISPR associated proteins) immunity has been exploited widely over the past few months (1-13). The main advantage provided by the bacterial type II CRISPR-Cas system lies in the minimal requirement for programmable DNA interference: an endonuclease, Cas9, guided by a customizable dual-RNA structure (14). As initially demonstrated in the original type II system of *Streptococcus pyogenes*, trans-activating CRISPR RNA (tracrRNA) (15,16) binds to the invariable repeats of precursor CRISPR RNA (pre-crRNA) forming a dual-RNA (14-17) that is essential for both RNA co-maturation by RNase III in the presence of Cas9 (15-17), and invading DNA cleavage by Cas9 (14,15,17-19). As demonstrated in *Streptococcus*, Cas9 guided by the duplex formed between mature activating tracrRNA and targeting crRNA (14-16) introduces site-specific double-stranded DNA (dsDNA) breaks in the invading cognate DNA (14,17-19). Cas9 is a multi-domain enzyme (14,20,21) that uses an HNH nuclease domain to cleave the target strand (defined as complementary to the spacer sequence of crRNA) and a RuvC-like domain to cleave the non-target strand (14,22,23), enabling the conversion of the dsDNA cleaving Cas9 into a nickase by selective motif inactivation (2,8,14,24,25). DNA cleavage specificity is determined by two parameters: the variable, spacer-derived sequence of crRNA targeting the protospacer sequence (a protospacer is defined as the sequence on the DNA target that is complementary to the spacer of crRNA) and a short sequence, the Protospacer Adjacent Motif (PAM), located immediately downstream of the protospacer on the non-target DNA strand (14,18,23,26-28).

**[0004]** Recent studies have demonstrated that RNA-guided Cas9 can be employed as an efficient genome editing tool in human cells (1,2,8,11), mice (9,10), zebrafish (6), drosophila (5), worms (4), plants (12,13), yeast (3) and bacteria (7). The system is versatile, enabling multiplex genome engineering by

programming Cas9 to edit several sites in a genome simultaneously by simply using multiple guide RNAs (2,7,8,10). The easy conversion of Cas9 into a nickase was shown to facilitate homology-directed repair in mammalian genomes with reduced mutagenic activity (2,8,24,25). In addition, the DNA-binding activity of a Cas9 catalytic inactive mutant has been exploited to engineer RNA-programmable transcriptional silencing and activating devices (29,30).

**[0005]** To date, RNA-guided Cas9 from *S. pyogenes*, *Streptococcus thermophilus*, *Neisseria meningitidis* and *Treponema denticola* have been described as tools for genome manipulation (1-13,24,25,31-34 and Esvelt et al. PMID: 24076762).

### Summary

**[0006]** The present invention expands the RNA-programmable Cas9 toolbox to additional orthologous systems. The diversity and interchangeability of dual-RNA:Cas9 in eight representatives of phylogenetically defined type II CRISPR-Cas groups was examined herein. The results of this work not only introduce a wider range of Cas9 enzymes, guide RNA structures and associated specific PAMs but also enlighten the evolutionary aspects of type II CRISPR-Cas systems, including coevolution and horizontal transfer of the system components.

**[0007]** In an aspect, the present disclosure provides guide RNAs, both single-molecule and double-molecule guide RNAs, as well as methods for manipulating DNA in a cell using the guide RNAs and/or DNAs (including vectors) encoding the guide RNAs. Complexes comprising the guide RNAs and Cas9 endonucleases are also provided

**[0008]** In some embodiments, the single-molecule guide RNAs comprise a DNA-targeting segment and a protein-binding segment, wherein the protein-binding segment comprises a tracrRNA set out in Supplementary Table S5 or wherein the protein-binding segment comprises a tracrRNA at least 80% identical over at least 20 nucleotides to a tracrRNA set out in Supplementary Table S5. In some embodiments, the protein-binding segment comprises a CRISPR repeat set out in Supplementary Table S5 that is the CRISPR repeat cognate to the tracrRNA of the protein-binding segment. In some embodiments, the DNA-targeting segment comprises RNA complementary to a protospacer-like sequence in a target DNA 5' to a PAM sequence. In some embodiments, the tracrRNA and CRISPR repeat are respectively the *C. jejuni* tracrRNA and its cognate CRISPR repeat set out in Supplementary Table S5 and the PAM sequence is NNNNACA. In some embodiments, the tracrRNA and CRISPR repeat are respectively at least 80% identical to the *C. jejuni* tracrRNA and its cognate CRISPR repeat set out in Supplementary Table S5 and the PAM sequence is NNNNACA.

**[0009]** In some embodiments, the single-molecule guide RNA comprises a sequence that hybridizes to a protospacer-like sequence set out in one of SEQ ID NOs: 801-2701.

**[0010]** In another aspect, the disclosure provides a DNA encoding a single-molecule guide RNA of the invention.



**[0011]** In yet another aspect, the disclosure provides a vector comprising a DNA encoding a single-molecule guide RNA of the invention.

**[0012]** In still another aspect, the disclosure provides a cell comprising a DNA encoding a single-molecule guide RNA of the invention.

**[0013]** In an aspect, the disclosure provides a double-molecule guide RNA comprising: a targeter-RNA and an activator-RNA complementary thereto, wherein the activator-RNA comprises a tracrRNA set out in Supplementary Table S5 or wherein the activator-RNA comprises a tracrRNA at least 80% identical over at least 20 nucleotides to a tracrRNA set out in Supplementary Table S5. In some embodiments, the double-molecule guide RNA comprises a modified backbone, a non-natural internucleoside linkage, a nucleic acid mimetic, a modified sugar moiety, a base modification, a modification or sequence that provides for modified or regulated stability, a modification or sequence that provides for subcellular tracking, a modification or sequence that provides for tracking, or a modification or sequence that provides for a binding site for a protein or protein complex. In some embodiments, the targeter-RNA comprises a CRISPR repeat set out in Supplementary Table S5. In some embodiments, the targeter-RNA comprises a CRISPR repeat set out in Supplementary Table S5 that is the cognate CRISPR repeat of the tracrRNA of the activator-RNA. In some embodiments, the targeter-RNA further comprises RNA complementary to a protospacer-like sequence in a target DNA 5' to a PAM sequence. In some embodiments, the tracrRNA and CRISPR repeat are respectively the *C. jejuni* tracrRNA and its cognate CRISPR repeat set out in Supplementary Table S5 and the PAM sequence is NNNNACA. In some embodiments, the tracrRNA and CRISPR repeat are at least 80% identical to respectively the *C. jejuni* tracrRNA and its cognate CRISPR repeat set out in Supplementary Table S5 and the PAM sequence is NNNNACA.

**[0014]** In some embodiments, the double-molecule guide RNA comprises a sequence that hybridizes to a protospacer-like sequence set out in one of SEQ ID NOs: 801-2701.

**[0015]** In another aspect, the disclosure provides a DNA encoding a double-molecule guide RNA of the invention.

**[0016]** In yet another aspect, the disclosure provides a vector comprising a DNA encoding a double-molecule guide RNA of the invention.

**[0017]** In still another aspect, the disclosure provides a cell comprising a DNA encoding a double-molecule guide RNA of the invention.

**[0018]** In an aspect, the disclosure provides methods for manipulating DNA in a cell, comprising contacting the DNA with a Cas9 ortholog-guideRNA complex, wherein the complex comprises: (a) a *C. jejuni* Cas9 endonuclease heterologous to the cell or an endonuclease with an activity portion at least 90% identical to the activity portion of the *C. jejuni* Cas9 endonuclease, and a guide RNA targeting the complex to a protospacer-like sequence in the DNA 5' to the PAM sequence NNNNACA; (b) a *P. multocida* Cas9 endonuclease heterologous to the cell or an endonuclease with an activity portion at least

90% identical to the activity portion of the *P. multocida* Cas9 endonuclease, and a guide RNA targeting the complex to a protospacer-like sequence in the DNA 5' to the PAM sequence GNNNCNNA or NNNNC; (c) an *F. novicida* Cas9 endonuclease heterologous to the cell or an endonuclease with an activity portion at least 90% identical to the activity portion of the *F. novicida* Cas9 endonuclease, and a guide RNA targeting the complex to a protospacer-like sequence in the DNA 5' to the PAM sequence NG; (d) an *S. thermophilus*\*\* Cas9 endonuclease heterologous to the cell or an endonuclease with an activity portion at least 90% identical to the activity portion of the *S. thermophilus*\*\* Cas9 endonuclease, and a guide RNA targeting the complex to a protospacer-like sequence in the DNA 5' to the PAM sequence NNAAAAW; (e) an *L. innocua* Cas9 endonuclease heterologous to the cell or an endonuclease with an activity portion at least 90% identical to the activity portion of the *L. innocua* Cas9 endonuclease, and a guide RNA targeting the complex to a protospacer-like sequence in the DNA 5' to the PAM sequence NGG; or (f) an *S. dysgalactiae* Cas9 endonuclease heterologous to the cell or an endonuclease with an activity portion at least 90% identical to the activity portion of the *S. dysgalactiae* Cas9 endonuclease, and a guide RNA targeting the complex to a protospacer-like sequence in the DNA 5' to the PAM sequence NGG. In some embodiments, the guide is a single-molecule guide RNA. In some embodiments, the guide RNA is a double-molecule guide RNA. The complexes used in the methods are also provided.

**[0019]** In some embodiments of the methods, the protospacer-like sequence targeted is in a CCR5, CXCR4, KRT5, KRT14, PLEC or COL7A1 gene. In some embodiments, the protospacer-like sequence is in a chronic granulomatous disease (CGD)-related gene CYBA, CYBB, NCF1, NCF2 or NCF4. In some embodiments, the protospacer-like sequence targeted is in a gene encoding B-cell lymphoma/leukemia IIA (BCL11A) protein, an erythroid enhancer of BCL11A or a BCL11A binding site. In some embodiments, the protospacer-like sequence targeted is up to 1000 nucleotides upstream of the above mentioned genes. In some embodiments of the methods, the guide RNA comprises a sequence complementary to a protospacer-like sequence set out in one of SEQ ID NOs: 801-2701.

**[0020]** In an aspect, the disclosure provides a recombinant vector encoding: (a) a guide RNA, wherein the guide RNA comprises a DNA-targeting segment complementary to a protospacer-like sequence in the DNA 5' to the PAM sequence NNNNACA; and (b) a *C. jejuni* Cas9 endonuclease (for example, set out in SEQ ID NO: 50) or an endonuclease with an activity portion at least 90% identical to the activity portion of the *C. jejuni* Cas9 endonuclease. In some embodiments, the DNA-targeting segment complementary to the protospacer-like sequence is RNA complementary to the target sequences set out in one of SEQ ID NOs: 801-973, 1079-1222, 1313-1348, 1372-1415, 1444-1900, 2163-2482 or 2667-2686. Methods of using the vectors to manipulate DNA in a cell are also provided.

**[0021]** In another aspect, the disclosure provides a recombinant vector encoding: (a) a guide RNA, wherein the guide RNA comprises a DNA-targeting segment complementary to a protospacer-like sequence in the DNA 5' to the PAM sequence GNNNCNNA or NNNNC; and (b) a *P. multocida* Cas9 endonuclease (for example, set out in SEQ ID NO: 1) or an endonuclease with an activity portion at least 90% identical to the activity portion of the *P. multocida* Cas9 endonuclease. In some embodiments, the

DNA-targeting segment complementary to the protospacer-like sequence is RNA complementary to the target sequences set out in one of SEQ ID NOs: 974-1078, 1223-1312, 1349-1371, 1416-1443, 1901-2162, 2483-2666 or 2687-2701. Methods of using the vectors to manipulate DNA in a cell are also provided.

**[0022]** In yet another aspect, the disclosure provides a recombinant vector encoding: (a) a guide RNA, wherein the guide RNA comprises a DNA-targeting segment complementary to a protospacer-like sequence in the DNA 5' to the PAM sequence NG; and (b) a *F. novicida* Cas9 endonuclease (for example, set out in SEQ ID NO: 43) or an endonuclease with an activity portion at least 90% identical to the activity portion of the *F. novicida* Cas9 endonuclease. Methods of using the vectors to manipulate DNA in a cell are also provided.

**[0023]** In still another aspect, the disclosure provides a recombinant vector encoding: (a) a guide RNA, wherein the guide RNA comprises a DNA-targeting segment complementary to a protospacer-like sequence in the DNA 5' to the PAM sequence NNAAAAW; and (b) a *S. thermophilus*\*\* Cas9 endonuclease or an endonuclease with an activity portion at least 90% identical to the activity portion of the *S. thermophilus*\*\* Cas9 endonuclease. Methods of using the vectors to manipulate DNA in a cell are also provided.

**[0024]** In yet another aspect, the disclosure provides a recombinant vector encoding: (a) a guide RNA, wherein the guide RNA comprises a DNA-targeting segment complementary to a protospacer-like sequence in the DNA 5' to the PAM sequence NGG; and (b) a *L. innocua* Cas9 endonuclease (for example, set out in SEQ ID NO: 3) or an endonuclease with an activity portion at least 90% identical to the activity portion of the *L. innocua* Cas9 endonuclease. Methods of using the vectors to manipulate DNA in a cell are also provided.

**[0025]** In still another aspect, the disclosure provides a recombinant vector encoding: (a) a guide RNA, wherein the guide RNA comprises a DNA-targeting segment complementary to a protospacer-like sequence in the DNA 5' to the PAM sequence NGG; and (b) a *S. dysgalactiae* Cas9 endonuclease (for example, set out in SEQ ID NO: 105) or an endonuclease with an activity portion at least 90% identical to the activity portion of the *S. dysgalactiae* Cas9 endonuclease.

**[0026]** In some embodiments of the vectors, the guide RNA comprises a sequence complementary to a protospacer-like sequence set out in one of SEQ ID NOs: 801-2701.

**[0027]** In a related aspect, the disclosure provides a method comprising (a) identifying at least 7-20 bases of mammalian genomic DNA adjacent to any of the preceding protospacer-like sequences, and (b) manipulating the mammalian genomic DNA sequence by contacting a mammalian cell with, or administering to a mammal, (i) a DNA-targeting segment complementary to the DNA sequence identified in step (a) and (ii) a protein-binding segment, or nucleic acid(s) encoding (i) and (ii), and (iii) a cas9 endonuclease or a nucleic acid encoding said cas9 endonuclease; and (c) detecting cleavage of the mammalian genomic DNA.

**[0028]** In an aspect, the disclosure provides a modified Cas9 endonuclease, modified from any of the Cas9 orthologs disclosed herein, comprising one or more mutations corresponding to *S. pyogenes* Cas9 mutation E762A, HH983AA or D986A. In some embodiments, the modified Cas 9 endonuclease further comprises one or more mutations corresponding to *S. pyogenes* Cas9 mutation D10A, H840A, G12A, G17A, N854A, N863A, N982A or A984A.

**[0029]** In an aspect, the disclosure provides a method for manipulating DNA in a cell, comprising contacting the DNA with a Cas9 ortholog-guide RNA complex, wherein the complex comprises: (a) a Cas9 endonuclease heterologous to the cell and (b) a cognate guide RNA of the Cas9 endonuclease comprising a tracrRNA set out in Supplementary Table S5 or a guide RNA comprising a tracrRNA at least 80% identical to a cognate tracrRNA set out in Supplementary Table S5 over at least 20 nucleotides. In some embodiments, the guide is a single-molecule guide RNA. In some embodiments, the guide RNA is a double-molecule guide RNA. In some embodiments of the methods, the guide RNA comprises a sequence complementary to a protospacer-like sequence set out in one of SEQ ID NOs: 801-2701. Complexes used in the methods are also provided.

**[0030]** In an aspect, the disclosure provides a method for manipulating DNA in a cell, comprising contacting the DNA with a Cas9 ortholog-guide RNA complex, wherein the complex comprises: (a) a cognate guide RNA for a first Cas9 endonuclease from a cluster in Supplementary Table S2 and (b) a second Cas9 endonuclease from the same cluster that is exchangeable with preserved high cleavage efficiency with the first endonuclease and shares at least 80% identity with the first endonuclease over 80% of their length. In some embodiments, the guide is a single-molecule guide RNA. In some embodiments, the guide RNA is a double-molecule guide RNA. In some embodiments, the first Cas9 endonuclease is from *S. pyogenes* and the second Cas9 endonuclease is from *S. mutans*. In some embodiments, the first Cas9 endonuclease is from *S. thermophilus*\* and the second Cas9 endonuclease is from *S. mutans*. In some embodiments, the first Cas9 endonuclease is from *N. meningitidis* and the second Cas9 endonuclease is from *P. multocida*. Complexes used in the methods are also provided.

**[0031]** In an aspect, the disclosure provides a method for manipulating DNA in a cell, comprising contacting the DNA with a Cas9 ortholog-guide RNA complex, wherein the complex comprises: (a) a cognate guide RNA of a first Cas9 endonuclease from a cluster in Supplementary Table S6 and (b) a Cas9 endonuclease from the same cluster in Supplementary Table S6 that is exchangeable with the same or lowered cleavage efficiency with the first endonuclease and shares at least 50% amino acid sequence identity with the first endonuclease over 70% of their length. In some embodiments, the guide is a single-molecule guide RNA. In some embodiments, the guide RNA is a double-molecule guide RNA. In some embodiments, the first Cas9 endonuclease is from *C. Jejuni* and the second Cas9 endonuclease is from *P. multocida*. In some embodiments, the first Cas9 endonuclease is from *N. meningitidis* and the second Cas9 endonuclease is from *P. multocida*. Complexes used in the methods are also provided.

**[0032]** In an aspect, the disclosure provides a method for manipulating DNA in a cell, comprising contacting the DNA with two or more Cas9-guide RNA complexes, wherein each Cas9-guideRNA complex comprises: (a) a Cas9 endonuclease from a different cluster in Supplementary Table S6 exhibiting less than 50% amino acid sequence identity with the other endonucleases of the method over 70% of their length, and (b) a guide RNA specifically complexed with each Cas9 endonuclease. In some embodiments, the guide is a single-molecule guide RNA. In some embodiments, the guide RNA is a double-molecule guide RNA. In some embodiments, the Cas9 endonucleases are from *F. novicida* and *S. pyogenes*. In some embodiments, the Cas9 endonucleases are from *N. meningitidis* and *S. mutans*. In some embodiments, the Cas9 endonucleases are the *S. thermophilus*\* Cas9 and the *S. thermophilus*\*\* Cas9. Complexes used in the methods are also provided.

**[0033]** In some embodiments of the manipulation methods, the DNA targeted in the cell is a CCR5, CXCR4, KRT5, KRT14, PLEC or COL7A1 gene. In some embodiments, the DNA targeted in the cell is a chronic granulomatous disease (CGD)-related gene CYBA, CYBB, NCF1, NCF2 or NCF4. In some embodiments, the protospacer-like sequence targeted is in a gene encoding B-cell lymphoma/leukemia IIA (BCL11A) protein, an erythroid enhancer of BCL11A or a BCL11A binding site. In some embodiments, the protospacer-like sequence targeted is up to 1000 nucleotides upstream of the above mentioned genes. In some embodiments of the methods, the guide RNA comprises a sequence complementary to a protospacer-like sequence set out in one of SEQ ID NOs: 801-2701.

**[0034]** It is contemplated that any of the methods provided herein may ex vivo or in vivo.

#### **Brief Description of the Drawings**

**[0035]** **Figure 1.** Phylogeny of representative Cas9 orthologs and schematic representation of selected bacterial type II CRISPR-Cas systems. **(A)** Phylogenetic tree of Cas9 reconstructed from selected, informative positions of representative Cas9 orthologs multiple sequence alignment is shown (see Supplementary Figure S2 and Supplementary Table S2). The Cas9 orthologs of the subtypes classified as II-A, II-B and II-C are highlighted with shaded boxes. The colored branches group distinct proteins of closely related loci with similar locus architecture (15). Each protein is represented by the GenInfo (GI) identifier followed by the bacterial strain name. The bootstrap values are given for each node (see Materials and Methods). Note that the monophyletic clusters of subtypes II-A and II-B are supported by high bootstrap values. The scale bar for the branch length is given as the estimated number of amino acid substitution per site. **(B)** Genetic loci of type II (Nmeni/CASS4) CRISPR-Cas in *Streptococcus pyogenes* SF370, *Streptococcus mutans* UA159, *Streptococcus thermophilus* LMD-9 \*(CRISPR3), \*\*(CRISPR1), *Campylobacter jejuni* NCTC 11168, *Neisseria meningitidis* Z2491, *Pasteurella multocida* Pm70 and *Francisella novicida* U112. Red arrow, transcription direction of tracrRNA; blue arrows, *cas* genes; black rectangles, CRISPR repeats; green diamonds, spacers; thick black line, leader sequence; black arrow, putative pre-crRNA promoter; HP, Hypothetical Protein. The colored bars

represented on the left correspond to Cas9 tree branches colors. The transcription direction and putative leader position of *C. jejuni* and *N. meningitidis* pre-crRNAs were derived from previously published RNA sequencing data (15). The CRISPR-Cas locus architecture of *P. multocida* was predicted based on its close similarity to that of *N. meningitidis* and further confirmed by bioinformatics prediction of tracrRNA based on a strongly predicted promoter and a transcriptional terminator as described in (15). Type II CRISPR-Cas loci can differ in the *cas* gene composition, mostly with *cas9*, *cas1* and *cas2* being the minimal set of genes (type II-C, blue), sometimes accompanied with a fourth gene *csn2a/b* (type II-A, yellow and orange) or *cas4* (type II-B, green). The CRISPR array can be transcribed in the same (type II-A, yellow and orange) or in the opposite (types II-B and C, blue and green) direction of the *cas* operon. The location of tracrRNA and the direction of its transcription differ within the groups (compare type II-A of *S. thermophilus*\*\* with type II-A from the other species indicated here (yellow) and compare type II-C of *C. jejuni* with type II-C of *N. meningitidis* and *P. multocida* (blue)).

**[0036] Figure 2.** RNase III is a general executioner of tracrRNA:pre-crRNA processing in type II CRISPR-Cas. Northern blot analysis of total RNA from *S. pyogenes* WT,  $\Delta rnc$  and  $\Delta rnc$  complemented with *rnc* orthologs or mutants (truncated *rnc* and inactivated (dead) (D51A) *mc*) probed for tracrRNA (top) and crRNA repeat (bottom). RNA sizes in nt and schematic representations of tracrRNA (red-black) and crRNA (green-black) are indicated on the right (16). The vertical black arrows indicate the processing sites. tracrRNA-171 nt and tracrRNA-89 nt forms correspond to primary tracrRNA transcripts. The presence of tracrRNA-75 nt and crRNA 39-42 nt forms indicates tracrRNA and pre-crRNA co-processing. *S. pyogenes* tracrRNA and pre-crRNA are co-processed by all analyzed RNase III orthologs. The truncated version and catalytic inactive mutant of *S. pyogenes* RNase III are both deficient in tracrRNA:pre-crRNA processing.

**[0037] Figure 3.** Conserved motifs of Cas9 are required for DNA interference but not for dual-RNA processing by RNase III. (A) Schematic representation of *S. pyogenes* Cas9. The conserved HNH and splitted RuvC motifs and analyzed amino acids are indicated. (B) Northern blot analysis of total RNA from *S. pyogenes* WT,  $\Delta cas9$  and  $\Delta cas9$  complemented with pEC342 or pEC342 containing *cas9* WT or mutant genes, probed for tracrRNA and crRNA repeat. Maturation of tracrRNA and pre-crRNA generating tracrRNA-75 nt and crRNA-39-42 nt forms is observed in all  $\Delta cas9$  strains complemented with the *cas9* mutants. (C) *In vivo* protospacer targeting. Transformation assays of *S. pyogenes* WT and  $\Delta cas9$  with pEC85 (vector), pEC85 $\Omega cas9$  (*cas9*), pEC85 $\Omega speM$  (*speM*), and pEC85 $\Omega tracrRNA-171$  nt plasmids containing *speM* and *cas9* mutants. The CFUs (colony forming units) per  $\mu\text{g}$  of plasmid DNA were determined in at least three independent experiments. The results +/- SD of technical triplicates of one representative experiment are shown. Cas9 N854A is the only mutant that did not tolerate the protospacer plasmid as observed for WT Cas9, indicating that this residue is not involved in DNA interference. (D) *In vitro* plasmid cleavage. Agarose gel electrophoresis of plasmid DNA (5 nM) containing *speM* protospacer (pEC287) incubated with 25 nM Cas9 WT or mutants in the presence of equimolar

amounts of dual-RNA-*speM* (see Materials and Methods). Cas9 WT and N854A generated linear cleavage products while the other Cas9 mutants created only nicked products. M, 1 kb DNA ladder (Fermentas); oc: open circular, li: linear; sc: supercoiled.

**[0038] Figure 4.** Cas9 from closely related CRISPR-Cas systems can substitute the role of *S. pyogenes* Cas9 in RNA processing by RNase III. **(A)** Schematic representation of Cas9 from selected bacterial species. The protein sizes and distances between conserved motifs (RuvC and HNH) are drawn in scale. See Supplementary Figure S1. **(B)** Northern blot analysis of total RNA extracted from *S. pyogenes* WT,  $\Delta cas9$  and  $\Delta cas9$  complemented with pEC342 (backbone vector containing *tracrRNA*-171 nt and the *cas* operon promoter from *S. pyogenes*) or pEC342-based plasmids containing *cas9* orthologous genes, probed for *tracrRNA* and crRNA repeat. Mature forms of *S. pyogenes* *tracrRNA* and pre-crRNA are observed only in the presence of *S. pyogenes* Cas9 WT or closely related Cas9 orthologs from *S. mutans* and *S. thermophilus*\*

**[0039] Figure 5.** Cas9 orthologs cleave DNA in the presence of their cognate dual-RNA and specific PAM *in vitro*. **(A)** Logo plot of protospacer adjacent sequences derived from BLAST analysis of spacer sequences for selected bacterial species. The logo plot gives graphical representation of most abundant nucleotides downstream of the protospacer sequence. The numbers in brackets correspond to the number of analyzed protospacers. **(B)** DNA substrates designed for specific PAM verification. Based on the logo plot for each species, plasmid DNA substrates were designed to contain the *speM* protospacer and the indicated sequence downstream, either comprising (PAM+) or not (PAM-) the proposed PAM. The predicted PAMs were verified by cleavage assays narrowing down the necessary nucleotides for activity (data not shown); therefore the sequence used differs slightly from the logoplot shown in **(A)**. The high abundance of other nucleotides not being part of the PAM can be explained by redundancy of the coding sequences containing the protospacers, and by the limited number of found protospacer targets. The last column shows the PAM sequence for each species, which was already published (no symbol) or derived from this work (#). **(C)** *In vitro* plasmid cleavage assays by dual-RNA:Cas9 orthologs on plasmid DNA with the 10 bp protospacer adjacent sequence (summarized in **(B)**). Each Cas9 ortholog in complex with its cognate dual-RNA cleaves plasmids containing the corresponding species-specific PAM (PAM+). No cleavage is observed with plasmids that did not contain the specific PAM (PAM-). li: linear cleavage product, sc: supercoiled plasmid DNA.

**[0040] Figure 6.** Cas9 and dual-RNA co-evolved. **(A)** *In vitro* plasmid cleavage assays using *S. pyogenes* Cas9 in complex with orthologous dual-RNA (upper panel) and orthologous Cas9 enzymes in complex with *S. pyogenes* dual-RNA (lower panel). Plasmid DNA containing protospacer *speM* and *S. pyogenes* PAM (NGG) was incubated with different dual-RNAs in complex with *S. pyogenes* Cas9. *tracrRNA* and crRNA-repeat sequences of the dual-RNAs are from the indicated bacterial species, with crRNA spacer targeting *speM*. In the lower panel, plasmid DNA containing *speM* protospacer and the specific PAM was incubated with Cas9 orthologs in complex with *S. pyogenes* dual-RNA. *S. pyogenes*

Cas9 can cleave plasmid DNA only in the presence of dual-RNA from *S. pyogenes*, *S. mutans* and *S. thermophilus*\* (yellow). Dual-RNA from *S. pyogenes* can mediate DNA cleavage only with Cas9 from *S. pyogenes*, *S. mutans* and *S. thermophilus*\* (yellow). li: linear cleavage product; sc: supercoiled plasmid DNA. (B) Summary of Cas9 and dual-RNA orthologs exchangeability. Specific PAM sequences were used according to Figure 5. The color code reflects the type II CRISPR-Cas subgroups (Figure 1). +++: 100 - 75% cleavage activity; ++: 75 - 50% cleavage activity; +: 50 - 25% cleavage activity; -: 25 - 0% cleavage activity observed under the conditions tested. Cas9 and dual-RNA duplexes from the same type II group can be interchanged and still mediate plasmid cleavage providing that the PAM sequence is specific for Cas9. See also Supplementary Figure S10.

**[0041] Supplementary Figure S1.** Biochemical characteristics and SDS-PAGE analysis of Cas9 proteins purified in this study. (A) Overview of characteristics of Cas9 orthologous proteins <sup>a</sup>Note that the biochemical characteristics of *S. pyogenes* Cas9 WT and mutants are identical; <sup>b</sup>GenInfo (GI) Identifier; <sup>c</sup> $\epsilon$ , Extinction coefficient. (B) SDS PAGE analysis of purified mutants of Cas9 from *S. pyogenes*. (C) SDS PAGE analysis of purified Cas9 orthologs. M: PageRuler™ Unstained Protein Ladder (Thermo Scientific).

**[0042] Supplementary Figure S2.** Multiple sequence alignment of representative Cas9 sequences (see Supplementary Table S2 and Material and Methods). The rows described as Jnet with following GI identifier of a selected Cas9 sequence provide the predicted secondary structure of Cas9 within the corresponding subgroups (sequences indicated below each Jnet). Conserved motifs are marked below the alignment and the mutated amino acid residues are highlighted. Asterisks indicate informative positions chosen for the Cas9 tree reconstruction.

**[0043] Supplementary Figure S3.** Multiple sequence alignment of representative Cas1 sequences (see Supplementary Table S2 and Materials and Methods). Informative positions chosen for the Cas1 tree reconstruction are marked with asterisks at the bottom of the alignment.

**[0044] Supplementary Figure S4.** Phylogenetic analysis of representative Cas9 and Cas1 sequences. Phylogenetic trees of Cas1 (left) and Cas9 (right) reconstructed from selected, informative positions of Cas1 and Cas9 multiple sequence alignments are shown (see Figure 1 and Supplementary Figures S2 and S3). The Cas1 tree is rooted to the outgroup of selected Cas1 orthologs of type I CRISPR-Cas systems. The Cas1 and Cas9 orthologs of the types classified as II-A, II-B and II-C are highlighted with shaded boxes. The same branch colors were used for each bacterial strain on both trees. Each protein is represented by the GenInfo (GI) identifier followed by the bacterial strain name. The bootstrap values are given for each node (see Materials and Methods). The scale bars for the branch length are given as the estimated number of amino acid substitution per site. Note the similarity of the trees topology and monophyletic clusters of subtypes II-A and II-B on both trees supported by high bootstrap values.



**[0045] Supplementary Figure S5.** RNase III is a general executioner of tracrRNA:pre-crRNA processing in type II CRISPR-Cas. Northern blot analysis of total RNA from *S. pyogenes* WT,  $\Delta rnc$  and  $\Delta rnc$  complemented with *rnc* orthologs or *rnc* mutants probed with (A) tracrRNA and (B) crRNA repeat (Supplementary Table S1). The dashed-line boxes represented below the Northern blots in (B) show the area of the blots with enhanced exposure. All RNase III orthologs can co-process *S. pyogenes* tracrRNA and pre-crRNA. No mature forms of tracrRNA and crRNAs could be observed in  $\Delta rnc$  complemented with the truncated version or catalytically inactive (dead) mutant of RNase III.

**[0046] Supplementary Figure S6.** Multiple sequence alignment of bacterial endoribonucleases III used in the study. Domains indicated below the alignment are according to the domains identified in RNase III from *E. coli* (58, 59). The conserved catalytic aspartate residue mutated in the catalytically inactive “*rnc* dead” mutant and the last amino acid of the truncated *rnc* mutant are indicated above the alignment with an asterisk and an arrow, respectively.

**[0047] Supplementary Figure S7.** Conserved catalytic amino acid residues of Cas9 are not involved in dual-RNA processing by RNase III. Northern blot analysis of total RNA extracted from *S. pyogenes* WT,  $\Delta cas9$  and  $\Delta cas9$  complemented with pEC342 (backbone vector containing tracrRNA-171 nt and the native *cas* operon promoter from *S. pyogenes*) or pEC342-derived plasmids encoding Cas9 WT or mutants, hybridized with (A) tracrRNA or (B) crRNA repeat probe (Supplementary Table S1). tracrRNA:crRNA co-processing is observed in all strains encoding Cas9 point mutants. Note that in a previous study, we observed low abundance of tracrRNA in the *cas9* deletion mutant (16). For this reason, plasmids used in *cas9* complementation studies were designed to encode tracrRNA in addition to *cas9*.

**[0048] Supplementary Figure S8.** Cas9 and tracrRNA:crRNA co-evolved. Northern blot analysis of total RNA extracted from *S. pyogenes* WT,  $\Delta cas9$  and  $\Delta cas9$  complemented with pEC342 or pEC342-derived plasmids encoding Cas9 WT or mutants - hybridized with (A) tracrRNA or (B) crRNA repeat probe (Supplementary Table S1). Only *S. pyogenes* Cas9 WT and closely related Cas9 orthologs from *S. mutans* and *S. thermophilus*\* (CRISPR3) can contribute to coprocessing of *S. pyogenes* tracrRNA:pre-crRNA.

**[0049] Supplementary Figure S9.** Cas9 orthologs cleave plasmid DNA in the presence of their cognate dual-RNA and specific PAM. Agarose gel electrophoresis analysis of dual-RNA:Cas9 titration (0-100 nM dual-RNA-Cas9 complex) on plasmid DNA (5 nM) containing *speM* protospacer and adjacent WT PAM (PAM+), imperfect PAM (PAM $\pm$ ) or no PAM (PAM-). For *S. pyogenes*, *S. mutans*, *S. thermophilus*\*, *S. thermophilus*\*\* and *N. meningitidis*, the PAM sequence has already been published (27,28,53,54). For the other bacterial species, PAMs were predicted based on the downstream sequence of protospacer identified in the investigated or related strains (see Supplementary Table S2 and Materials and Methods). The 10 bp sequence located directly downstream of the crRNA-targeted *speM* protospacer is shown. The

nucleotide(s) predicted to belong to the PAM sequence are shaded in grey. li: linear cleavage product, sc: supercoiled plasmid DNA, M: 1 kb DNA ladder.

**[0050] Supplementary Figure S10.** Summary of *in vitro* plasmid cleavage assays of Cas9 orthologs in combination with dual-RNAs. Agarose gel electrophoresis of cleavage assays. (A) *S. mutans* Cas9 (50 nM), (B) *S. thermophilus*\* Cas9 (25 nM), (C) *S. thermophilus*\*\* Cas9 (100 nM), (D) *C. jejuni* Cas9 (100 nM), (E) *N. meningitidis* Cas9 (100 nM), (F) *P. multocida* Cas9 (25 nM), (G) *F. novicida* Cas9 (100 nM) in complex with equimolar concentrations of each of the dual-RNA orthologs were incubated with plasmid DNA (5 nM) containing *speM* protospacer sequence and the PAM sequence specific to the Cas9 ortholog analyzed. li: linear cleavage product, sc: supercoiled plasmid DNA, M: 1 kb DNA ladder.

**[0051] Supplementary Figure S11.** Cas9 tree topology suggests both horizontal and vertical transfer of type II CRISPR-Cas systems. See Figure 1, Supplementary Figure S4 and Supplementary Table S4. The codes for taxonomy (phyla in color) and habitat (symbols) of the bacterial strains harbouring representative Cas9 orthologs are indicated (right panel). The clusters grouping evolutionary distant bacteria (1 and 3) but isolated mainly from similar sources (human for cluster 1 and mostly environmental samples for cluster 3) suggest horizontal transfer of type II systems. Clusters 2, 4 and 5 group closely related bacteria isolated from diverse habitats indicating vertical transfer of the systems.

**[0052] Supplementary Figure S12.** tracrRNA:crRNA repeat duplexes form similar secondary structures in loci with closely related Cas9 orthologs. Antirepeat sequence of processed tracrRNA (red) and repeat-derived sequence of mature crRNA (grey) were co-folded for each type II CRISPR-Cas locus studied (see Materials and Methods). Color bars indicated on the left group dual-RNAs from loci with closely related Cas9 (see Figure 1 and Supplementary Figure S4). RNA duplexes belonging to the same groups display structural similarities, suggesting a role of the structure in dual-RNA recognition by Cas9.

### Detailed Description

**[0053]** Terminology

**[0054]** All technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this invention belongs, unless the technical or scientific term is defined differently herein.

**[0055]** The terms "polynucleotide" and "nucleic acid," used interchangeably herein, refer to a polymeric form of nucleotides of any length, either ribonucleotides or deoxyribonucleotides. Thus, this term includes, but is not limited to, single-, double-, or multi-stranded DNA or RNA, genomic DNA, cDNA, DNA-RNA hybrids, or a polymer comprising purine and pyrimidine bases or other natural, chemically or biochemically modified, non-natural, or derivatized nucleotide bases. "Oligonucleotide" generally refers to

polynucleotides of between about 5 and about 100 nucleotides of single- or double-stranded DNA. However, for the purposes of this disclosure, there is no upper limit to the length of an oligonucleotide. Oligonucleotides are also known as "oligomers" or "oligos" and may be isolated from genes, or chemically synthesized by methods known in the art. The terms "polynucleotide" and "nucleic acid" should be understood to include, as applicable to the embodiments being described, single-stranded (such as sense or antisense) and double-stranded polynucleotides.

**[0056]** "Genomic DNA" refers to the DNA of a genome of an organism including, but not limited to, the DNA of the genome of a bacterium, fungus, archea, plant or animal.

**[0057]** "Manipulating" DNA encompasses binding, nicking one strand, or cleaving (i.e., cutting) both strands of the DNA, or encompasses modifying the DNA or a polypeptide associated with the DNA (e.g., the modifications of paragraphs [00161] or [00162]). Manipulating DNA can silence, activate, or modulate (either increase or decrease) the expression of an RNA or polypeptide encoded by the DNA.

**[0058]** A "stem-loop structure" refers to a nucleic acid having a secondary structure that includes a region of nucleotides which are known or predicted to form a double strand (stem portion) that is linked on one side by a region of predominantly single-stranded nucleotides (loop portion). The terms "hairpin" and "fold-back" structures are also used herein to refer to stem-loop structures. Such structures are well known in the art and these terms are used consistently with their known meanings in the art. As is known in the art, a stem-loop structure does not require exact base-pairing. Thus, the stem may include one or more base mismatches. Alternatively, the base-pairing may be exact, i.e. not include any mismatches.

**[0059]** By "hybridizable" or "complementary" or "substantially complementary" it is meant that a nucleic acid (e.g. RNA) comprises a sequence of nucleotides that enables it to non-covalently bind, i.e. form Watson-Crick base pairs and/or G/U base pairs, "anneal", or "hybridize," to another nucleic acid in a sequence-specific, antiparallel, manner (i.e., a nucleic acid specifically binds to a complementary nucleic acid) under the appropriate in vitro and/or in vivo conditions of temperature and solution ionic strength. As is known in the art, standard Watson-Crick base-pairing includes: adenine (A) pairing with thymidine (T), adenine (A) pairing with uracil (U), and guanine (G) pairing with cytosine (C) [DNA, RNA]. In addition, it is also known in the art that for hybridization between two RNA molecules (e.g., dsRNA), guanine (G) base pairs with uracil (U). For example, G/U base-pairing is partially responsible for the degeneracy (i.e., redundancy) of the genetic code in the context of tRNA anti-codon base-pairing with codons in mRNA. In the context of this disclosure, a guanine (G) of a protein-binding segment (dsRNA duplex) of a guide RNA molecule is considered complementary to a uracil (U), and vice versa. As such, when a G/U base-pair can be made at a given nucleotide position a protein-binding segment (dsRNA duplex) of a guide RNA molecule, the position is not considered to be non-complementary, but is instead considered to be complementary.

**[0060]** Hybridization and washing conditions are well known and exemplified in Sambrook, J., Fritsch, E. F. and Maniatis, T. *Molecular Cloning: A Laboratory Manual*, Second Edition, Cold Spring Harbor Laboratory Press, Cold Spring Harbor (1989), particularly Chapter 11 and Table 11.1 therein; and Sambrook, J. and Russell, W., *Molecular Cloning: A Laboratory Manual*, Third Edition, Cold Spring Harbor Laboratory Press, Cold Spring Harbor (2001). The conditions of temperature and ionic strength determine the "stringency" of the hybridization.

**[0061]** Hybridization requires that the two nucleic acids contain complementary sequences, although mismatches between bases are possible. The conditions appropriate for hybridization between two nucleic acids depend on the length of the nucleic acids and the degree of complementation, variables well known in the art. The greater the degree of complementation between two nucleotide sequences, the greater the value of the melting temperature ( $T_m$ ) for hybrids of nucleic acids having those sequences. For hybridizations between nucleic acids with short stretches of complementarity (e.g. complementarity over 35 or less, 30 or less, 25 or less, 22 or less, 20 or less, or 18 or less nucleotides) the position of mismatches becomes important (see Sambrook et al., *supra*, 11.7-11.8). Typically, the length for a hybridizable nucleic acid is at least about 10 nucleotides. Illustrative minimum lengths for a hybridizable nucleic acid are: at least about 15 nucleotides; at least about 20 nucleotides; at least about 22 nucleotides; at least about 25 nucleotides; and at least about 30 nucleotides). Furthermore, the skilled artisan will recognize that the temperature and wash solution salt concentration may be adjusted as necessary according to factors such as length of the region of complementation and the degree of complementation.

**[0062]** It is understood in the art that the sequence of polynucleotide need not be 100% complementary to that of its target nucleic acid to be specifically hybridizable or hybridizable. Moreover, a polynucleotide may hybridize over one or more segments such that intervening or adjacent segments are not involved in the hybridization event (e.g., a loop structure or hairpin structure). A polynucleotide can comprise at least 70%, at least 80%, at least 90%, at least 95%, at least 99%, or 100% sequence complementarity to a target region within the target nucleic acid sequence to which they are targeted. For example, an antisense nucleic acid in which 18 of 20 nucleotides of the antisense compound are complementary to a target region, and would therefore specifically hybridize, would represent 90 percent complementarity. In this example, the remaining noncomplementary nucleotides may be clustered or interspersed with complementary nucleotides and need not be contiguous to each other or to complementary nucleotides. Percent complementarity between particular stretches of nucleic acid sequences within nucleic acids can be determined routinely using BLAST programs (basic local alignment search tools) and PowerBLAST programs known in the art (Altschul et al., *J. Mol. Biol.*, 1990, 215, 403-410; Zhang and Madden, *Genome Res.*, 1997, 7, 649-656) or by using the Gap program (Wisconsin Sequence Analysis Package, Version 8 for Unix, Genetics Computer Group, University

Research Park, Madison Wis.), using default settings, which uses the algorithm of Smith and Waterman (Adv. Appl. Math., 1981, 2, 482-489).

**[0063]** The terms "peptide," "polypeptide," and "protein" are used interchangeably herein, and refer to a polymeric form of amino acids of any length, which can include coded and non-coded amino acids, chemically or biochemically modified or derivatized amino acids, and polypeptides having modified peptide backbones.

**[0064]** "Binding" as used herein (e.g. with reference to an RNA-binding domain of a polypeptide) refers to a non-covalent interaction between macromolecules (e.g., between a protein and a nucleic acid). While in a state of non-covalent interaction, the macromolecules are said to be "associated" or "interacting" or "binding" (e.g., when a molecule X is said to interact with a molecule Y, it is meant the molecule X binds to molecule Y in a non-covalent manner). Not all components of a binding interaction need be sequence-specific (e.g., contacts with phosphate residues in a DNA backbone), but some portions of a binding interaction may be sequence-specific. Binding interactions are generally characterized by a dissociation constant ( $K_d$ ) of less than  $10^{-6}$  M, less than  $10^{-7}$  M, less than  $10^{-8}$  M, less than  $10^{-9}$  M, less than  $10^{-10}$  M, less than  $10^{-11}$  M, less than  $10^{-12}$  M, less than  $10^{-13}$  M, less than  $10^{-14}$  M, or less than  $10^{-15}$  M. "Affinity" refers to the strength of binding, increased binding affinity being correlated with a lower  $K_d$ . By "binding domain" it is meant a protein domain that is able to bind non-covalently to another molecule. A binding domain can bind to, for example, a DNA molecule (a DNA-binding protein), an RNA molecule (an RNA-binding protein) and/or a protein molecule (a protein-binding protein). In the case of a protein domain-binding protein, it can bind to itself (to form homodimers, homotrimers, etc.) and/or it can bind to one or more molecules of a different protein or proteins.

**[0065]** The term "conservative amino acid substitution" refers to the interchangeability in proteins of amino acid residues having similar side chains. For example, a group of amino acids having aliphatic side chains consists of glycine, alanine, valine, leucine, and isoleucine; a group of amino acids having aliphatic-hydroxyl side chains consists of serine and threonine; a group of amino acids having amide containing side chains consisting of asparagine and glutamine; a group of amino acids having aromatic side chains consists of phenylalanine, tyrosine, and tryptophan; a group of amino acids having basic side chains consists of lysine, arginine, and histidine; a group of amino acids having acidic side chains consists of glutamate and aspartate; and a group of amino acids having sulfur containing side chains consists of cysteine and methionine. Exemplary conservative amino acid substitution groups are: valine-leucine-isoleucine, phenylalanine-tyrosine, lysine-arginine, alanine-valine, and asparagine-glutamine.

**[0066]** A polynucleotide or polypeptide has a certain percent "sequence identity" to another polynucleotide or polypeptide, meaning that, when aligned, that percentage of bases or amino acids are the same, and in the same relative position, when comparing the two sequences. Sequence identity can be determined in a number of different manners. To determine sequence identity, sequences can be aligned

using various methods and computer programs (e.g., BLAST, T-COFFEE, MUSCLE, MAFFT, etc.), available over the world wide web at sites including [ncbi.nlm.nih.gov/BLAST](http://ncbi.nlm.nih.gov/BLAST), [ebi.ac.uk/Tools/msa/tcoffee/](http://ebi.ac.uk/Tools/msa/tcoffee/), [ebi.ac.uk/Tools/msa/muscle/](http://ebi.ac.uk/Tools/msa/muscle/), [mafft.cbrc.jp/alignment/software/](http://mafft.cbrc.jp/alignment/software/). See, e.g., Altschul et al. (1990), J. Mol. Biol. 215:403-10. Sequence alignments standard in the art are used according to the invention to determine amino acid residues in a Cas9 ortholog that "correspond to" amino acid residues in another Cas9 ortholog. The amino acid residues of Cas9 orthologs that correspond to amino acid residues of other Cas9 orthologs appear at the same position in alignments of the sequences.

**[0067]** A DNA sequence that "encodes" a particular RNA is a DNA nucleic acid sequence that is transcribed into RNA. A DNA polynucleotide may encode an RNA (mRNA) that is translated into protein, or a DNA polynucleotide may encode an RNA that is not translated into protein (e.g. tRNA, rRNA, or a guide RNA; also called "non-coding" RNA or "ncRNA"). A "protein coding sequence" or a sequence that encodes a particular protein or polypeptide, is a nucleic acid sequence that is transcribed into mRNA (in the case of DNA) and is translated (in the case of mRNA) into a polypeptide in vitro or in vivo when placed under the control of appropriate regulatory sequences. The boundaries of the coding sequence are determined by a start codon at the 5' terminus (N-terminus) and a translation stop nonsense codon at the 3' terminus (C-terminus). A coding sequence can include, but is not limited to, cDNA from prokaryotic or eukaryotic mRNA, genomic DNA sequences from prokaryotic or eukaryotic DNA, and synthetic nucleic acids. A transcription termination sequence will usually be located 3' to the coding sequence.

**[0068]** As used herein, a "promoter sequence" is a DNA regulatory region capable of binding RNA polymerase and initiating transcription of a downstream (3' direction) coding or non-coding sequence. For purposes of defining the present invention, the promoter sequence is bounded at its 3' terminus by the transcription initiation site and extends upstream (5' direction) to include the minimum number of bases or elements necessary to initiate transcription at levels detectable above background. Within the promoter sequence will be found a transcription initiation site, as well as protein binding domains responsible for the binding of RNA polymerase. Eukaryotic promoters will often, but not always, contain "TATA" boxes and "CAT" boxes. Various promoters, including inducible promoters, may be used to drive the various vectors of the present invention.

**[0069]** A promoter can be a constitutively active promoter (i.e., a promoter that is constitutively in an active/"ON" state), it may be an inducible promoter (i.e., a promoter whose state, active/"ON" or inactive/"OFF", is controlled by an external stimulus, e.g., the presence of a particular temperature, compound, or protein.), it may be a spatially restricted promoter (i.e., transcriptional control element, enhancer, etc.)(e.g., tissue specific promoter, cell type specific promoter, etc.), and it may be a temporally restricted promoter (i.e., the promoter is in the "ON" state or "OFF" state during specific stages of embryonic development or during specific stages of a biological process, e.g., hair follicle cycle in mice).

**[0070]** Suitable promoters can be derived from viruses and can therefore be referred to as viral promoters, or they can be derived from any organism, including prokaryotic or eukaryotic organisms. Suitable promoters can be used to drive expression by any RNA polymerase (e.g., pol I, pol II, pol III). Exemplary promoters include, but are not limited to the SV40 early promoter, mouse mammary tumor virus long terminal repeat (LTR) promoter; adenovirus major late promoter (Ad MLP); a herpes simplex virus (HSV) promoter, a cytomegalovirus (CMV) promoter such as the CMV immediate early promoter region (CMVIE), a rous sarcoma virus (RSV) promoter, a human U6 small nuclear promoter (U6) (Miyagishi et al. , Nature Biotechnology 20, 497 - 500 (2002)), an enhanced U6 promoter (e.g., Xia et al., Nucleic Acids Res. 2003 Sep 1;31(17)), a human H1 promoter (H1), and the like.

**[0071]** Examples of inducible promoters include, but are not limited to T7 RNA polymerase promoter, T3 RNA polymerase promoter, Isopropyl-beta-D-thiogalactopyranoside (IPTG)-regulated promoter, lactose induced promoter, heat shock promoter, Tetracycline-regulated promoter, Steroid-regulated promoter, Metal-regulated promoter, estrogen receptor-regulated promoter, etc. Inducible promoters can therefore be regulated by molecules including, but not limited to, doxycycline; RNA polymerase, e.g., T7 RNA polymerase; an estrogen receptor; an estrogen receptor fusion; etc.

**[0072]** In some embodiments, the promoter is a spatially restricted promoter (i.e., cell type specific promoter, tissue specific promoter, etc.) such that in a multi-cellular organism, the promoter is active (i.e., "ON") in a subset of specific cells. Spatially restricted promoters may also be referred to as enhancers, transcriptional control elements, control sequences, etc. Any convenient spatially restricted promoter may be used and the choice of suitable promoter (e.g., a brain specific promoter, a promoter that drives expression in a subset of neurons, a promoter that drives expression in the germline, a promoter that drives expression in the lungs, a promoter that drives expression in muscles, a promoter that drives expression in islet cells of the pancreas, etc.) will depend on the organism. For example, various spatially restricted promoters are known for plants, flies, worms, mammals, mice, etc. Thus, a spatially restricted promoter can be used to regulate the expression of a nucleic acid encoding a site-directed modifying polypeptide in a wide variety of different tissues and cell types, depending on the organism. Some spatially restricted promoters are also temporally restricted such that the promoter is in the "ON" state or "OFF" state during specific stages of embryonic development or during specific stages of a biological process (e.g., hair follicle cycle in mice).

**[0073]** For illustration purposes, examples of spatially restricted promoters include, but are not limited to, neuron-specific promoters, adipocyte-specific promoters, cardiomyocyte-specific promoters, smooth muscle-specific promoters, photoreceptor-specific promoters, etc. Neuron-specific spatially restricted promoters include, but are not limited to, a neuron-specific enolase (NSE) promoter (see, e.g., EMBL HSENO2, X51956); an aromatic amino acid decarboxylase (AADC) promoter; a neurofilament promoter (see, e.g., GenBank HUMNFL, L04147); a synapsin promoter (see, e.g., GenBank HUMSYNIB, M55301); a thy-1 promoter (see, e.g., Chen et al. (1987) Cell 51:7-19; and Llewellyn, et al. (2010) Nat. Med.

16(10):1161-1166); a serotonin receptor promoter (see, e.g., GenBank S62283); a tyrosine hydroxylase promoter (TH) (see, e.g., Oh et al. (2009) *Gene Ther* 16:437; Sasaoka et al. (1992) *Mol. Brain Res.* 16:274; Boundy et al. (1998) *J. Neurosci.* 18:9989; and Kaneda et al. (1991) *Neuron* 6:583-594); a GnRH promoter (see, e.g., Radovick et al. (1991) *Proc. Natl. Acad. Sci. USA* 88:3402-3406); an L7 promoter (see, e.g., Oberdick et al. (1990) *Science* 248:223-226); a DNMT promoter (see, e.g., Bartge et al. (1988) *Proc. Natl. Acad. Sci. USA* 85:3648-3652); an enkephalin promoter (see, e.g., Comb et al. (1988) *EMBO J.* 17:3793-3805); a myelin basic protein (MBP) promoter; a Ca<sup>2+</sup>-calmodulin-dependent protein kinase II-alpha (CamKIM) promoter (see, e.g., Mayford et al. (1996) *Proc. Natl. Acad. Sci. USA* 93:13250; and Casanova et al. (2001) *Genesis* 31:37); a CMV enhancer/platelet-derived growth factor-p promoter (see, e.g., Liu et al. (2004) *Gene Therapy* 11:52-60); and the like.

**[0074]** Adipocyte-specific spatially restricted promoters include, but are not limited to aP2 gene promoter/enhancer, e.g., a region from -5.4 kb to +21 bp of a human aP2 gene (see, e.g., Tozzo et al. (1997) *Endocrinol.* 138:1604; Ross et al. (1990) *Proc. Natl. Acad. Sci. USA* 87:9590; and Pavjani et al. (2005) *Nat. Med.* 11:797); a glucose transporter-4 (GLUT4) promoter (see, e.g., Knight et al. (2003) *Proc. Natl. Acad. Sci. USA* 100:14725); a fatty acid translocase (FAT/CD36) promoter (see, e.g., Kuriki et al. (2002) *Biol. Pharm. Bull.* 25:1476; and Sato et al. (2002) *J. Biol. Chem.* 277:15703); a stearoyl-CoA desaturase-1 (SCD1) promoter (Tabor et al. (1999) *J. Biol. Chem.* 274:20603); a leptin promoter (see, e.g., Mason et al. (1998) *Endocrinol.* 139:1013; and Chen et al. (1999) *Biochem. Biophys. Res. Comm.* 262:187); an adiponectin promoter (see, e.g., Kita et al. (2005) *Biochem. Biophys. Res. Comm.* 331:484; and Chakrabarti (2010) *Endocrinol.* 151:2408); an adipsin promoter (see, e.g., Platt et al. (1989) *Proc. Natl. Acad. Sci. USA* 86:7490); a resistin promoter (see, e.g., Seo et al. (2003) *Molec. Endocrinol.* 17:1522); and the like.

**[0075]** Cardiomyocyte-specific spatially restricted promoters include, but are not limited to control sequences derived from the following genes: myosin light chain-2, a-myosin heavy chain, AE3, cardiac troponin C, cardiac actin, and the like. Franz et al. (1997) *Cardiovasc. Res.* 35:560-566; Robbins et al. (1995) *Ann. N.Y. Acad. Sci.* 752:492-505; Linn et al. (1995) *Circ. Res.* 76:584591; Parmacek et al. (1994) *Mol. Cell. Biol.* 14:1870-1885; Hunter et al. (1993) *Hypertension* 22:608-617; and Sartorelli et al. (1992) *Proc. Natl. Acad. Sci. USA* 89:4047-4051.

**[0076]** Smooth muscle-specific spatially restricted promoters include, but are not limited to an SM22a promoter (see, e.g., Akyiirek et al. (2000) *Mol. Med.* 6:983; and U.S. Patent No. 7,169,874); a smoothelin promoter (see, e.g., WO 2001/018048); an a-smooth muscle actin promoter; and the like. For example, a 0.4 kb region of the SM22a promoter, within which lie two CArG elements, has been shown to mediate vascular smooth muscle cell-specific expression (see, e.g., Kim, et al. (1997) *Mol. Cell. Biol.* 17, 2266-2278; Li, et al., (1996) *J. Cell Biol.* 132, 849-859; and Moessler, et al. (1996) *Development* 122, 2415-2425).



**[0077]** Photoreceptor-specific spatially restricted promoters include, but are not limited to, a rhodopsin promoter; a rhodopsin kinase promoter (Young et al. (2003) *Ophthalmol. Vis. Sci.* 44:4076); a beta phosphodiesterase gene promoter (Nicoud et al. (2007) *J. Gene Med.* 9:1015); a retinitis pigmentosa gene promoter (Nicoud et al. (2007) *supra*); an interphotoreceptor retinoid-binding protein (IRBP) gene enhancer (Nicoud et al. (2007) *supra*); an IRBP gene promoter (Yokoyama et al. (1992) *Exp Eye Res.* 55:225); and the like.

**[0078]** The terms "DNA regulatory sequences," "control elements," and "regulatory elements," used interchangeably herein, refer to transcriptional and translational control sequences, such as promoters, enhancers, polyadenylation signals, terminators, protein degradation signals, and the like, that provide for and/or regulate transcription of a non-coding sequence (e.g., guide RNA) or a coding sequence (e.g., site-directed modifying polypeptide, or Cas9 polypeptide) and/or regulate translation of an encoded polypeptide.

**[0079]** The term "naturally-occurring" or "unmodified" as used herein as applied to a nucleic acid, a polypeptide, a cell, or an organism, refers to a nucleic acid, polypeptide, cell, or organism that is found in nature. For example, a polypeptide or polynucleotide sequence that is present in an organism (including viruses) that can be isolated from a source in nature and which has not been intentionally modified by a human in the laboratory is naturally occurring.

**[0080]** The term "chimeric" as used herein as applied to a nucleic acid or polypeptide refers to two components that are defined by structures derived from different sources. For example, where "chimeric" is used in the context of a chimeric polypeptide (e.g., a chimeric Cas9 protein), the chimeric polypeptide includes amino acid sequences that are derived from different polypeptides. A chimeric polypeptide may comprise either modified or naturally-occurring polypeptide sequences (e.g., a first amino acid sequence from a modified or unmodified Cas9 protein; and a second amino acid sequence other than the Cas9 protein). Similarly, "chimeric" in the context of a polynucleotide encoding a chimeric polypeptide includes nucleotide sequences derived from different coding regions (e.g., a first nucleotide sequence encoding a modified or unmodified Cas9 protein; and a second nucleotide sequence encoding a polypeptide other than a Cas9 protein).

**[0081]** The term "chimeric polypeptide" refers to a polypeptide which is not naturally occurring, e.g., is made by the artificial combination (i.e., "fusion") of two otherwise separated segments of amino sequence through human intervention. A polypeptide that comprises a chimeric amino acid sequence is a chimeric polypeptide. Some chimeric polypeptides can be referred to as "fusion variants."

**[0082]** "Heterologous," as used herein, means a nucleotide or peptide that is not found in the native nucleic acid or protein, respectively. For example, in a chimeric Cas9 protein, the RNA-binding domain of a naturally-occurring bacterial Cas9 polypeptide (or a variant thereof) may be fused to a heterologous

polypeptide sequence (i.e. a polypeptide sequence from a protein other than Cas9 or a polypeptide sequence from another organism). The heterologous polypeptide may exhibit an activity (e.g., enzymatic activity) that will also be exhibited by the chimeric Cas9 protein (e.g., methyltransferase activity, acetyltransferase activity, kinase activity, ubiquitinating activity, etc.). A heterologous nucleic acid may be linked to a naturally-occurring nucleic acid (or a variant thereof) (e.g., by genetic engineering) to generate a chimeric polynucleotide encoding a chimeric polypeptide. As another example, in a fusion variant Cas9 site-directed polypeptide, a variant Cas9 site-directed polypeptide may be fused to a heterologous polypeptide (i.e. a polypeptide other than Cas9), which exhibits an activity that will also be exhibited by the fusion variant Cas9 site-directed polypeptide. A heterologous nucleic acid may be linked to a variant Cas9 site-directed polypeptide (e.g., by genetic engineering) to generate a polynucleotide encoding a fusion variant Cas9 site-directed polypeptide. "Heterologous," as used herein, additionally means a nucleotide or polypeptide in a cell that is not its native cell.

**[0083]** The term "cognate" refers to two biomolecules that normally interact or co-exist in nature.

**[0084]** "Recombinant," as used herein, means that a particular nucleic acid (DNA or RNA) or vector is the product of various combinations of cloning, restriction, polymerase chain reaction (PCR) and/or ligation steps resulting in a construct having a structural coding or non-coding sequence distinguishable from endogenous nucleic acids found in natural systems. DNA sequences encoding polypeptides can be assembled from cDNA fragments or from a series of synthetic oligonucleotides, to provide a synthetic nucleic acid which is capable of being expressed from a recombinant transcriptional unit contained in a cell or in a cell-free transcription and translation system. Genomic DNA comprising the relevant sequences can also be used in the formation of a recombinant gene or transcriptional unit. Sequences of non-translated DNA may be present 5' or 3' from the open reading frame, where such sequences do not interfere with manipulation or expression of the coding regions, and may indeed act to modulate production of a desired product by various mechanisms (see "DNA regulatory sequences", below). Alternatively, DNA sequences encoding RNA (e.g., guide RNA) that is not translated may also be considered recombinant. Thus, e.g., the term "recombinant" nucleic acid refers to one which is not naturally occurring, e.g., is made by the artificial combination of two otherwise separated segments of sequence through human intervention. This artificial combination is often accomplished by either chemical synthesis means, or by the artificial manipulation of isolated segments of nucleic acids, e.g., by genetic engineering techniques. Such is usually done to replace a codon with a codon encoding the same amino acid, a conservative amino acid, or a non-conservative amino acid. Alternatively, it is performed to join together nucleic acid segments of desired functions to generate a desired combination of functions. This artificial combination is often accomplished by either chemical synthesis means, or by the artificial manipulation of isolated segments of nucleic acids, e.g., by genetic engineering techniques. When a recombinant polynucleotide encodes a polypeptide, the sequence of the encoded polypeptide can be naturally occurring ("wild type") or can be a variant (e.g., a mutant) of the naturally occurring sequence.

Thus, the term "recombinant" polypeptide does not necessarily refer to a polypeptide whose sequence does not naturally occur. Instead, a "recombinant" polypeptide is encoded by a recombinant DNA sequence, but the sequence of the polypeptide can be naturally occurring ("wild type") or non-naturally occurring (e.g., a variant, a mutant, etc.). Thus, a "recombinant" polypeptide is the result of human intervention, but may be a naturally occurring amino acid sequence.

**[0085]** A "vector" or "expression vector" is a replicon, such as plasmid, phage, virus, or cosmid, to which another DNA segment, i.e. an "insert", may be attached so as to bring about the replication of the attached segment in a cell.

**[0086]** An "expression cassette" comprises a DNA coding sequence operably linked to a promoter. "Operably linked" refers to a juxtaposition wherein the components so described are in a relationship permitting them to function in their intended manner. For instance, a promoter is operably linked to a coding sequence if the promoter affects its transcription or expression. The terms "recombinant expression vector," or "DNA construct" are used interchangeably herein to refer to a DNA molecule comprising a vector and at least one insert. Recombinant expression vectors are usually generated for the purpose of expressing and/or propagating the insert(s), or for the construction of other recombinant nucleotide sequences. The nucleic acid(s) may or may not be operably linked to a promoter sequence and may or may not be operably linked to DNA regulatory sequences.

**[0087]** A cell has been "genetically modified" or "transformed" or "transfected" by exogenous DNA, e.g. a recombinant expression vector, when such DNA has been introduced inside the cell. The presence of the exogenous DNA results in permanent or transient genetic change. The transforming DNA may or may not be integrated (covalently linked) into the genome of the cell.

**[0088]** In prokaryotes, yeast, and mammalian cells for example, the transforming DNA may be maintained on an episomal element such as a plasmid. With respect to eukaryotic cells, a stably transformed cell is one in which the transforming DNA has become integrated into a chromosome so that it is inherited by daughter cells through chromosome replication. This stability is demonstrated by the ability of the eukaryotic cell to establish cell lines or clones that comprise a population of daughter cells containing the transforming DNA. A "clone" is a population of cells derived from a single cell or common ancestor by mitosis. A "cell line" is a clone of a primary cell that is capable of stable growth *in vitro* for many generations.

**[0089]** Suitable methods of genetic modification (also referred to as "transformation") include e.g., viral or bacteriophage infection, transfection, conjugation, protoplast fusion, lipofection, electroporation, calcium phosphate precipitation, polyethyleneimine (PEI)-mediated transfection, DEAE-dextran mediated transfection, liposome-mediated transfection, particle gun technology, calcium phosphate precipitation, direct micro injection, nanoparticle-mediated nucleic acid delivery (see, e.g., Panyam et. al Adv Drug Deliv Rev. 2012 Sep 13. pii: S0169-409X(12)00283-9. doi: 10.1016/j.addr.2012.09.023 ), and the like.

**[0090]** The choice of method of genetic modification is generally dependent on the type of cell being transformed and the circumstances under which the transformation is taking place (e.g., in vitro, ex vivo, or in vivo). A general discussion of these methods can be found in Ausubel, et al., Short Protocols in Molecular Biology, 3rd ed., Wiley & Sons, 1995.

**[0091]** A "host cell," as used herein, denotes an in vivo or in vitro eukaryotic cell, a prokaryotic cell (e.g., bacterial or archaeal cell), or a cell from a multicellular organism (e.g., a cell line) cultured as a unicellular entity, which eukaryotic or prokaryotic cells can be, or have been, used as recipients for a nucleic acid, and include the progeny of the original cell which has been transformed by the nucleic acid. It is understood that the progeny of a single cell may not necessarily be completely identical in morphology or in genomic or total DNA complement as the original parent, due to natural, accidental, or deliberate mutation. A "recombinant host cell" (also referred to as a "genetically modified host cell") is a host cell into which has been introduced a heterologous nucleic acid, e.g., an expression vector. For example, a bacterial host cell is a genetically modified bacterial host cell by virtue of introduction into a suitable bacterial host cell of an exogenous nucleic acid (e.g., a plasmid or recombinant expression vector) and a eukaryotic host cell is a genetically modified eukaryotic host cell (e.g., a mammalian germ cell), by virtue of introduction into a suitable eukaryotic host cell of an exogenous nucleic acid.

**[0092]** A "target DNA" as used herein is a DNA polynucleotide that comprises a "target site" or "target sequence." The terms "target site," "target sequence," "target protospacer DNA," or "protospacer-like sequence" are used interchangeably herein to refer to a nucleic acid sequence present in a target DNA to which a DNA-targeting segment of a guide RNA will bind, provided sufficient conditions for binding exist. For example, the target site (or target sequence) 5'-GAGCATATC-3' within a target DNA is targeted by (or is bound by, or hybridizes with, or is complementary to) the RNA sequence 5'-GAUAUGCUC-3'. Suitable DNA/RNA binding conditions include physiological conditions normally present in a cell. Other suitable DNA/RNA binding conditions (e.g., conditions in a cell-free system) are known in the art; see, e.g., Sambrook, *supra*. The strand of the target DNA that is complementary to and hybridizes with the guide RNA is referred to as the "complementary strand" and the strand of the target DNA that is complementary to the "complementary strand" (and is therefore not complementary to the guide RNA) is referred to as the "noncomplementary strand" or "non-complementary strand." By "site-directed modifying polypeptide" or "RNA-binding site-directed polypeptide" or "RNA-binding site-directed modifying polypeptide" or "site-directed polypeptide" it is meant a polypeptide that binds RNA and is targeted to a specific DNA sequence. A site-directed modifying polypeptide as described herein is targeted to a specific DNA sequence by the RNA molecule to which it is bound. The RNA molecule comprises a sequence that binds, hybridizes to, or is complementary to a target sequence within the target DNA, thus targeting the bound polypeptide to a specific location within the target DNA (the target sequence). Exemplary target sequences of the invention are set out in SEQ ID NOs: 801-2701. SEQ ID NOs: 801-973 are protospacer-like target sequences 5' to the PAM sequence NNNNACA in the human CCR5 gene. SEQ

ID NOs: 974-1078 are protospacer-like sequences 5' to the PAM sequence GNNNCNNA in the human CCR5 gene. SEQ ID NOs: 1079-1222 are protospacer-like target sequences 5' to the PAM sequence NNNNACA in the exons of the human CCR5 gene. SEQ ID NOs: 1223-1312 are protospacer-like sequences 5' to the PAM sequence GNNNCNNA in the exons of the human CCR5 gene. SEQ ID NOs: 1313-1348 are protospacer-like target sequences 5' to the PAM sequence NNNNACA around the 5' end of the human CCR5 gene. SEQ ID NOs: 1349-1371 are protospacer-like sequences 5' to the PAM sequence GNNNCNNA around the 5' end of the human CCR5 gene. SEQ ID NOs: 1372-1415 are protospacer-like target sequences 5' to the PAM sequence NNNNACA around the delta 32 locus in the human CCR5 gene. SEQ ID NOs: 1416-1443 are protospacer-like sequences 5' to the PAM sequence GNNNCNNA around the delta 32 locus in the human CCR5 gene. SEQ ID NOs: 1444-1900 are protospacer-like target sequences 5' to the PAM sequence NNNNACA in the human BCL11A gene. SEQ ID NOs: 1901-2162 are protospacer-like sequences 5' to the PAM sequence GNNNCNNA in the human BCL11A gene. SEQ ID NOs: 2163-2482 are protospacer-like target sequences 5' to the PAM sequence NNNNACA in the exons of the human BCL11A gene. SEQ ID NOs: 2483-2666 are protospacer-like sequences 5' to the PAM sequence GNNNCNNA in the exons of the human BCL11A gene. SEQ ID NOs: 2667-2686 are protospacer-like target sequences 5' to the PAM sequence NNNNACA around the 5' end of the human BCL11A gene. SEQ ID NOs: 2687-2701 are protospacer-like sequences 5' to the PAM sequence GNNNCNNA around the 5' end of the human BCL11A gene. Target sequences at least 80% identical to the sequences set out in SEQ ID NOs: 801-2701 are also contemplated.

**[0093]** By "cleavage" it is meant the breakage of the covalent backbone of a DNA molecule. Cleavage can be initiated by a variety of methods including, but not limited to, enzymatic or chemical hydrolysis of a phosphodiester bond. Both single-stranded cleavage and double-stranded cleavage are possible, and double-stranded cleavage can occur as a result of two distinct single-stranded cleavage events. DNA cleavage can result in the production of either blunt ends or staggered ends. In certain embodiments, a complex comprising a guide RNA and a site-directed modifying polypeptide is used for targeted double-stranded DNA cleavage.

**[0094]** "Nuclease" and "endonuclease" are used interchangeably herein to mean an enzyme which possesses endonucleolytic catalytic activity for DNA cleavage.

**[0095]** By "cleavage domain" or "active domain" or "nuclease domain" of a nuclease it is meant the polypeptide sequence or domain within the nuclease which possesses the catalytic activity for DNA cleavage. A cleavage domain can be contained in a single polypeptide chain or cleavage activity can result from the association of two (or more) polypeptides. A single nuclease domain may consist of more than one isolated stretch of amino acids within a given polypeptide.

**[0096]** By "site-directed polypeptide" or "RNA-binding site-directed polypeptide" or "RNA-binding site-directed polypeptide" it is meant a polypeptide that binds RNA and is targeted to a specific DNA

sequence. A site-directed polypeptide as described herein is targeted to a specific DNA sequence by the RNA molecule to which it is bound. The RNA molecule comprises a sequence that is complementary to a target sequence within the target DNA, thus targeting the bound polypeptide to a specific location within the target DNA (the target sequence).

**[0097]** The RNA molecule that binds to the site-directed modifying polypeptide and targets the polypeptide to a specific location within the target DNA is referred to herein as the "guide RNA" or "guide RNA polynucleotide" (also referred to herein as a "guide RNA" or "gRNA"). A guide RNA comprises two segments, a "DNA-targeting segment" and a "protein-binding segment." By "segment" it is meant a segment/section/region of a molecule, e.g., a contiguous stretch of nucleotides in an RNA. A segment can also mean a region/section of a complex such that a segment may comprise regions of more than one molecule. For example, in some cases the protein-binding segment (described below) of a guide RNA is one RNA molecule and the protein-binding segment therefore comprises a region of that RNA molecule. In other cases, the protein-binding segment (described below) of a guide RNA comprises two separate molecules that are hybridized along a region of complementarity. As an illustrative, non-limiting example, a protein-binding segment of a guide RNA that comprises two separate molecules can comprise (i) base pairs 40-75 of a first RNA molecule that is 100 base pairs in length; and (ii) base pairs 10-25 of a second RNA molecule that is 50 base pairs in length. The definition of "segment," unless otherwise specifically defined in a particular context, is not limited to a specific number of total base pairs, is not limited to any particular number of base pairs from a given RNA molecule, is not limited to a particular number of separate molecules within a complex, and may include regions of RNA molecules that are of any total length and may or may not include regions with complementarity to other molecules.

**[0098]** The DNA-targeting segment (or "DNA-targeting sequence") comprises a nucleotide sequence that is complementary to a specific sequence within a target DNA (the complementary strand of the target DNA) designated the "protospacer-like" sequence herein. The protein-binding segment (or "protein-binding sequence") interacts with a site-directed modifying polypeptide. When the site-directed modifying polypeptide is a Cas9 or Cas9 related polypeptide (described in more detail below), site-specific cleavage of the target DNA occurs at locations determined by both (i) base-pairing complementarity between the guide RNA and the target DNA; and (ii) a short motif (referred to as the protospacer adjacent motif (PAM)) in the target DNA.

**[0099]** The protein-binding segment of a guide RNA comprises, in part, two complementary stretches of nucleotides that hybridize to one another to form a double stranded RNA duplex (dsRNA duplex).

**[00100]** In some embodiments, a nucleic acid (e.g., a guide RNA, a nucleic acid comprising a nucleotide sequence encoding a guide RNA; a nucleic acid encoding a site-directed polypeptide; etc.) comprises a modification or sequence that provides for an additional desirable feature (e.g., modified or regulated stability; subcellular targeting; tracking, e.g., a fluorescent label; a binding site for a protein or

protein complex; etc.). Non-limiting examples include: a 5' cap (e.g., a 7-methylguanylate cap (m7G)); a 3' polyadenylated tail (i.e., a 3' poly(A) tail); a riboswitch sequence (e.g., to allow for regulated stability and/or regulated accessibility by proteins and/or protein complexes); a stability control sequence; a sequence that forms a dsRNA duplex (i.e., a hairpin); a modification or sequence that targets the RNA to a subcellular location (e.g., nucleus, mitochondria, chloroplasts, and the like); a modification or sequence that provides for tracking (e.g., direct conjugation to a fluorescent molecule, conjugation to a moiety that facilitates fluorescent detection, a sequence that allows for fluorescent detection, etc.); a modification or sequence that provides a binding site for proteins (e.g., proteins that act on DNA, including transcriptional activators, transcriptional repressors, DNA methyltransferases, DNA demethylases, histone acetyltransferases, histone deacetylases, and the like); and combinations thereof.

**[00101]** In some embodiments, a guide RNA comprises an additional segment at either the 5' or 3' end that provides for any of the features described above. For example, a suitable third segment can comprise a 5' cap (e.g., a 7-methylguanylate cap (m7G)); a 3' polyadenylated tail (i.e., a 3' poly(A) tail); a riboswitch sequence (e.g., to allow for regulated stability and/or regulated accessibility by proteins and protein complexes); a stability control sequence; a sequence that forms a dsRNA duplex (i.e., a hairpin); a sequence that targets the RNA to a subcellular location (e.g., nucleus, mitochondria, chloroplasts, and the like); a modification or sequence that provides for tracking (e.g., direct conjugation to a fluorescent molecule, conjugation to a moiety that facilitates fluorescent detection, a sequence that allows for fluorescent detection, etc.); a modification or sequence that provides a binding site for proteins (e.g., proteins that act on DNA, including transcriptional activators, transcriptional repressors, DNA methyltransferases, DNA demethylases, histone acetyltransferases, histone deacetylases, and the like); and combinations thereof.

**[00102]** A guide RNA and a site-directed modifying polypeptide (i.e., site-directed polypeptide) form a complex (i.e., bind via non-covalent interactions). The guide RNA provides target specificity to the complex by comprising a nucleotide sequence that is complementary to a sequence of a target DNA. The site-directed modifying polypeptide of the complex provides the site-specific activity. In other words, the site-directed modifying polypeptide is guided to a target DNA sequence (e.g. a target sequence in a chromosomal nucleic acid; a target sequence in an extrachromosomal nucleic acid, e.g. an episomal nucleic acid, a minicircle, etc.; a target sequence in a mitochondrial nucleic acid; a target sequence in a chloroplast nucleic acid; a target sequence in a plasmid; etc.) by virtue of its association with the protein-binding segment of the guide RNA.

**[00103]** In some embodiments, a guide RNA comprises two separate RNA molecules (RNA polynucleotides: an "activator-RNA" and a "targeter-RNA", see below) and is referred to herein as a "double-molecule guide RNA" or a "two-molecule guide RNA." In other embodiments, the guide RNA is a single RNA molecule (single RNA polynucleotide) and is referred to herein as a "single-molecule guide

RNA," a "single-guide RNA," or an "sgRNA." The term "guide RNA" or "gRNA" is inclusive, referring both to double-molecule guide RNAs and to single-molecule guide RNAs (i.e., sgRNAs).

**[00104]** A two-molecule guide RNA comprises two separate RNA molecules (a "targeter-RNA" and an "activator-RNA"). Each of the two RNA molecules of a two-molecule guide RNA comprises a stretch of nucleotides that are complementary to one another such that the complementary nucleotides of the two RNA molecules hybridize to form the double stranded RNA duplex of the protein-binding segment.

**[00105]** An exemplary two-molecule guide RNA comprises a crRNA-like ("CRISPR RNA" or "targeter-RNA") molecule (which includes a CRISPR repeat or CRISPR repeat-like sequence) and a corresponding tracrRNA-like ("trans-activating CRISPR RNA" or "activator-RNA" or "tracrRNA") molecule. A crRNA-like molecule (targeter-RNA) comprises both the DNA-targeting segment (single stranded) of the guide RNA and a stretch ("duplex-forming segment") of nucleotides that forms one half of the dsRNA duplex of the protein-binding segment of the guide RNA. A corresponding tracrRNA-like molecule (activator-RNA) comprises a stretch of nucleotides (duplex-forming segment) that forms the other half of the dsRNA duplex of the protein-binding segment of the guide RNA. In other words, a stretch of nucleotides of a crRNA-like molecule are complementary to and hybridize with a stretch of nucleotides of a tracrRNA-like molecule to form the dsRNA duplex of the protein-binding domain of the guide RNA. As such, each crRNA-like molecule can be said to have a corresponding tracrRNA-like molecule. The crRNA-like molecule additionally provides the single stranded DNA-targeting segment. Thus, a crRNA-like and a tracrRNA-like molecule (as a corresponding pair) hybridize to form a guide RNA. A double-molecule guide RNA can comprise any corresponding crRNA and tracrRNA pair.

**[00106]** A two-molecule guide RNA can be designed to allow for controlled (i.e., conditional) binding of a targeter-RNA with an activator-RNA. Because a two-molecule guide RNA is not functional unless both the activator-RNA and the targeter-RNA are bound in a functional complex with Cas9, a two-molecule guide RNA can be inducible (e.g., drug inducible) by rendering the binding between the activator-RNA and the targeter-RNA to be inducible. As one non-limiting example, RNA aptamers can be used to regulate (i.e., control) the binding of the activator-RNA with the targeter-RNA. Accordingly, the activator-RNA and/or the targeter-RNA can comprise an RNA aptamer sequence.

**[00107]** A single-molecule guide RNA comprises two stretches of nucleotides (a targeter-RNA and an activator-RNA) that are complementary to one another, are covalently linked (directly, or by intervening nucleotides), and hybridize to form the double stranded RNA duplex (dsRNA duplex) of the protein-binding segment, thus resulting in a stem-loop structure. The targeter-RNA and the activator-RNA can be covalently linked via the 3' end of the targeter-RNA and the 5' end of the activator-RNA. Alternatively, targeter-RNA and the activator-RNA can be covalently linked via the 5' end of the targeter-RNA and the 3' end of the activator-RNA.



**[00108]** An exemplary single-molecule guide RNA comprises two complementary stretches of nucleotides that hybridize to form a dsRNA duplex. In some embodiments, one of the two complementary stretches of nucleotides of the single-molecule guide RNA (or the DNA encoding the stretch) is at least about 60% identical to one of the activator-RNA (tracrRNA) sequences set forth in Supplementary Table S5 over a stretch of at least 8 contiguous nucleotides. For example, one of the two complementary stretches of nucleotides of the single-molecule guide RNA (or the DNA encoding the stretch) is at least about 65% identical, at least about 70% identical, at least about 75% identical, at least about 80% identical, at least about 85% identical, at least about 90% identical, at least about 95% identical, at least about 98% identical, at least about 99% identical or 100 % identical to one of the tracrRNA sequences set forth in Supplementary Table S5 over a stretch of at least 8 contiguous , at least 9 contiguous, at least 10 contiguous, at least 11 contiguous, at least 12 contiguous, at least 13 contiguous, at least 14 contiguous or at least 15 contiguous nucleotides. For example, the single-molecule guide RNA may comprise a nucleotide sequence that is at least 70% identical over at least 10 contiguous nucleotides, at least 80% identical over at least 10 contiguous nucleotides, at least 70% identical over at least 11 contiguous nucleotides, at least 80% identical over at least 11 contiguous nucleotides, at least 70% identical over at least 12 contiguous nucleotides, or at least 80% identical over at least 12 contiguous nucleotides of one of the tracrRNA sequences set forth in Supplementary Table S5. It is understood that where a series of percent identities and a series of lengths of nucleotides sequences are set out as options, each and every combination of a percent identity with a length (e.g. 8, 9, 10, 12, 13, 14, 15 nucleotides) of nucleotide sequence is contemplated.

**[00109]** In some embodiments, one of the two complementary stretches of nucleotides of the single-molecule guide RNA (or the DNA encoding the stretch) is at least about 60% identical to one of the targeter-RNA (crRNA/CRISPR repeat) sequences set forth in Supplementary Table S5 over a stretch of at least 8 contiguous nucleotides. For example, one of the two complementary stretches of nucleotides of the single-molecule guide RNA (or the DNA encoding the stretch) is at least about 65% identical, at least about 70% identical, at least about 75% identical, at least about 80% identical, at least about 85% identical, at least about 90% identical, at least about 95% identical, at least about 98% identical, at least about 99% identical or 100 % identical to one of the crRNA/CRISPR repeat sequences set forth in Supplementary Table S5 over a stretch of at least 8 contiguous, at least 9 contiguous, at least 10 contiguous, at least 11 contiguous, at least 12 contiguous, at least 13 contiguous, at least 14 contiguous or at least 15 contiguous nucleotides. For example, the single-molecule guide RNA may comprise a nucleotide sequence that is at least 70% identical over at least 10 contiguous nucleotides, at least 80% identical over at least 10 contiguous nucleotides, at least 70% identical over at least 11 contiguous nucleotides, at least 80% identical over at least 11 contiguous nucleotides, at least 70% identical over at least 12 contiguous nucleotides, or at least 80% identical over at least 12 contiguous nucleotides of one of the CRISPR repeat sequences set forth in Supplementary Table S5. It is understood that where a series of percent identities and a series of lengths of nucleotides sequences are set out as options, each

and every combination of a percent identity with a length (e.g. 8, 9, 10, 11, 12, 13, 14, 15 nucleotides) of nucleotide sequence is contemplated.

**[00110]** The term "activator-RNA" is used herein to mean a tracrRNA-like molecule of a double-molecule guide RNA. The term "targeter-RNA" is used herein to mean a crRNA-like molecule of a double-molecule guide RNA. The term "duplex-forming segment" is used herein to mean the stretch of nucleotides of an activator-RNA or a targeter-RNA that contributes to the formation of the dsRNA duplex by hybridizing to a stretch of nucleotides of a corresponding activator-RNA or targeter-RNA molecule. In other words, an activator-RNA comprises a duplex-forming segment that is complementary to the duplex-forming segment of the corresponding targeter-RNA. As such, an activator-RNA comprises a duplex-forming segment while a targeter-RNA comprises both a duplex-forming segment and the DNA-targeting segment of the guide RNA. Therefore, a double-molecule guide RNA can be comprised of any corresponding activator-RNA and targeter-RNA pair.

**[00111]** RNA aptamers are known in the art and are generally a synthetic version of a riboswitch. The terms "RNA aptamer" and "riboswitch" are used interchangeably herein to encompass both synthetic and natural nucleic acid sequences that provide for inducible regulation of the structure (and therefore the availability of specific sequences) of the RNA molecule of which they are part. RNA aptamers usually comprise a sequence that folds into a particular structure (e.g., a hairpin), which specifically binds a particular drug (e.g., a small molecule). Binding of the drug causes a structural change in the folding of the RNA, which changes a feature of the nucleic acid of which the aptamer is a part. As non-limiting examples: (i) an activator-RNA with an aptamer may not be able to bind to the cognate targeter-RNA unless the aptamer is bound by the appropriate drug; (ii) a targeter-RNA with an aptamer may not be able to bind to the cognate activator-RNA unless the aptamer is bound by the appropriate drug; and (iii) a targeter-RNA and an activator-RNA, each comprising a different aptamer that binds a different drug, may not be able to bind to each other unless both drugs are present. As illustrated by these examples, a two-molecule guide RNA can be designed to be inducible.

**[00112]** Examples of aptamers and riboswitches can be found, for example, in: Nakamura et al., *Genes Cells*. 2012 May;17(5):344-64; Vavalle et al., *Future Cardiol*. 2012 May;8(3):371-82; Citartan et al., *Biosens Bioelectron*. 2012 Apr 15;34(1):1-11; and Liberman et al., *Wiley Interdiscip Rev RNA*. 2012 May-Jun;3(3):369-84; all of which are herein incorporated by reference in their entirety.

**[00113]** The term "stem cell" is used herein to refer to a cell (e.g., plant stem cell, vertebrate stem cell) that has the ability both to self-renew and to generate a differentiated cell type (see Morrison et al. (1997) *Cell* 88:287-298). In the context of cell ontogeny, the adjective "differentiated", or "differentiating" is a relative term. A "differentiated cell" is a cell that has progressed further down the developmental pathway than the cell it is being compared with. Thus, pluripotent stem cells (described below) can differentiate into lineage-restricted progenitor cells (e.g., mesodermal stem cells), which in turn can differentiate into

cells that are further restricted (e.g., neuron progenitors), which can differentiate into end-stage cells (i.e., terminally differentiated cells, e.g., neurons, cardiomyocytes, etc.), which play a characteristic role in a certain tissue type, and may or may not retain the capacity to proliferate further. Stem cells may be characterized by both the presence of specific markers (e.g., proteins, RNAs, etc.) and the absence of specific markers. Stem cells may also be identified by functional assays both in vitro and in vivo, particularly assays relating to the ability of stem cells to give rise to multiple differentiated progeny.

**[00114]** Stem cells of interest include pluripotent stem cells (PSCs). The term "pluripotent stem cell" or "PSC" is used herein to mean a stem cell capable of producing all cell types of the organism. Therefore, a PSC can give rise to cells of all germ layers of the organism (e.g., the endoderm, mesoderm, and ectoderm of a vertebrate). Pluripotent cells are capable of forming teratomas and of contributing to ectoderm, mesoderm, or endoderm tissues in a living organism. Pluripotent stem cells of plants are capable of giving rise to all cell types of the plant (e.g., cells of the root, stem, leaves, etc.).

**[00115]** PSCs of animals can be derived in a number of different ways. For example, embryonic stem cells (ESCs) are derived from the inner cell mass of an embryo (Thomson et. al, Science. 1998 Nov 6;282(5391):1145-7) whereas induced pluripotent stem cells (iPSCs) are derived from somatic cells (Takahashi et. al, Cell. 2007 Nov 30;131(5):861-72; Takahashi et. al, Nat Protoc. 2007;2(12):3081-9; Yu et. al, Science. 2007 Dec 21;318(5858):1917-20. Epub 2007 Nov 20). Because the term PSC refers to pluripotent stem cells regardless of their derivation, the term PSC encompasses the terms ESC and iPSC, as well as the term embryonic germ stem cells (EGSC), which are another example of a PSC. PSCs may be in the form of an established cell line, they may be obtained directly from primary embryonic tissue, or they may be derived from a somatic cell. PSCs can be target cells of the methods described herein.

**[00116]** By "embryonic stem cell" (ESC) is meant a PSC that was isolated from an embryo, typically from the inner cell mass of the blastocyst. ESC lines are listed in the NIH Human Embryonic Stem Cell Registry, e.g. hESBGN-01, hESBGN-02, hESBGN-03, hESBGN-04 (BresaGen, Inc.); HES-1, HES-2, HES-3, HES-4, HES-5, HES-6 (ES Cell International); Miz-hES1 (MizMedi Hospital-Seoul National University); HSF-1, HSF-6 (University of California at San Francisco); and H1, H7, H9, H13, H14 (Wisconsin Alumni Research Foundation (WiCell Research Institute)). Stem cells of interest also include embryonic stem cells from other primates, such as Rhesus stem cells and marmoset stem cells. The stem cells may be obtained from any mammalian species, e.g. human, equine, bovine, porcine, canine, feline, rodent, e.g. mice, rats, hamster, primate, etc. (Thomson et al. (1998) Science 282:1145; Thomson et al. (1995) Proc. Natl. Acad. Sci USA 92:7844; Thomson et al. (1996) Biol. Reprod. 55:254; Shambloott et al., Proc. Natl. Acad. Sci. USA 95:13726, 1998). In culture, ESCs typically grow as flat colonies with large nucleo-cytoplasmic ratios, defined borders and prominent nucleoli. In addition, ESCs express SSEA-3, SSEA-4, TRA-1-60, TRA-1-81, and Alkaline Phosphatase, but not SSEA-1. Examples of methods of generating and characterizing ESCs may be found in, for example, US Patent No.

7,029,913, US Patent No. 5,843,780, and US Patent No. 6,200,806, the disclosures of which are incorporated herein by reference. Methods for proliferating hESCs in the undifferentiated form are described in WO 99/20741, WO 01/51616, and WO 03/020920. By "embryonic germ stem cell" (EGSC) or "embryonic germ cell" or "EG cell" is meant a PSC that is derived from germ cells and/or germ cell progenitors, e.g. primordial germ cells, i.e. those that would become sperm and eggs. Embryonic germ cells (EG cells) are thought to have properties similar to embryonic stem cells as described above. Examples of methods of generating and characterizing EG cells may be found in, for example, US Patent No. 7,153,684; Matsui, Y., et al., (1992) Cell 70:841; Shambloott, M., et al. (2001) Proc. Natl. Acad. Sci. USA 98: 113; Shambloott, M., et al. (1998) Proc. Natl. Acad. Sci. USA, 95:13726; and Koshimizu, U., et al. (1996) Development, 122:1235, the disclosures of which are incorporated herein by reference.

**[00117]** By "induced pluripotent stem cell" or "iPSC" it is meant a PSC that is derived from a cell that is not a PSC (i.e., from a cell this is differentiated relative to a PSC). iPSCs can be derived from multiple different cell types, including terminally differentiated cells. iPSCs have an ES cell-like morphology, growing as flat colonies with large nucleo-cytoplasmic ratios, defined borders and prominent nuclei. In addition, iPSCs express one or more key pluripotency markers known by one of ordinary skill in the art, including but not limited to Alkaline Phosphatase, SSEA3, SSEA4, Sox2, Oct3/4, Nanog, TRA160, TRA181, TDGF 1, Dnmt3b, FoxD3, GDF3, Cyp26a1, TERT, and zfp42. Examples of methods of generating and characterizing iPSCs may be found in, for example, U.S. Patent Publication Nos. US20090047263, US20090068742, US20090191159, US20090227032, US20090246875, and US20090304646, the disclosures of which are incorporated herein by reference. Generally, to generate iPSCs, somatic cells are provided with reprogramming factors (e.g. Oct4, SOX2, KLF4, MYC, Nanog, Lin28, etc.) known in the art to reprogram the somatic cells to become pluripotent stem cells.

**[00118]** By "somatic cell" it is meant any cell in an organism that, in the absence of experimental manipulation, does not ordinarily give rise to all types of cells in an organism. In other words, somatic cells are cells that have differentiated sufficiently that they will not naturally generate cells of all three germ layers of the body, i.e. ectoderm, mesoderm and endoderm. For example, somatic cells would include both neurons and neural progenitors, the latter of which may be able to naturally give rise to all or some cell types of the central nervous system but cannot give rise to cells of the mesoderm or endoderm lineages.

**[00119]** By "mitotic cell" it is meant a cell undergoing mitosis. Mitosis is the process by which a eukaryotic cell separates the chromosomes in its nucleus into two identical sets in two separate nuclei. It is generally followed immediately by cytokinesis, which divides the nuclei, cytoplasm, organelles and cell membrane into two cells containing roughly equal shares of these cellular components.

**[00120]** By "post-mitotic cell" it is meant a cell that has exited from mitosis, i.e., it is "quiescent", i.e. it is no longer undergoing divisions. This quiescent state may be temporary, i.e. reversible, or it may be permanent.

**[00121]** By "meiotic cell" it is meant a cell that is undergoing meiosis. Meiosis is the process by which a cell divides its nuclear material for the purpose of producing gametes or spores. Unlike mitosis, in meiosis, the chromosomes undergo a recombination step which shuffles genetic material between chromosomes. Additionally, the outcome of meiosis is four (genetically unique) haploid cells, as compared with the two (genetically identical) diploid cells produced from mitosis.

**[00122]** By "recombination" it is meant a process of exchange of genetic information between two polynucleotides. As used herein, "homology-directed repair (HDR)" refers to the specialized form DNA repair that takes place, for example, during repair of double-strand breaks in cells. This process requires nucleotide sequence homology, uses a "donor" molecule to template repair of a "target" molecule (i.e., the one that experienced the double-strand break), and leads to the transfer of genetic information from the donor to the target. Homology-directed repair may result in an alteration of the sequence of the target molecule (e.g., insertion, deletion, mutation), if the donor polynucleotide differs from the target molecule and part or all of the sequence of the donor polynucleotide is incorporated into the target DNA. In some embodiments, the donor polynucleotide, a portion of the donor polynucleotide, a copy of the donor polynucleotide, or a portion of a copy of the donor polynucleotide integrates into the target DNA.

**[00123]** By "non-homologous end joining (NHEJ)" it is meant the repair of double-strand breaks in DNA by direct ligation of the break ends to one another without the need for a homologous template (in contrast to homology-directed repair, which requires a homologous sequence to guide repair). NHEJ often results in the loss (deletion) of nucleotide sequence near the site of the double-strand break.

**[00124]** The terms "treatment", "treating" and the like are used herein to generally mean obtaining a desired pharmacologic and/or physiologic effect. The effect may be prophylactic in terms of completely or partially preventing a disease or symptom thereof and/or may be therapeutic in terms of a partial or complete cure for a disease and/or adverse effect attributable to the disease. "Treatment" as used herein covers any treatment of a disease or symptom in a mammal, and includes: (a) preventing the disease or symptom from occurring in a subject which may be predisposed to acquiring the disease or symptom but has not yet been diagnosed as having it; (b) inhibiting the disease or symptom, i.e., arresting its development; or (c) relieving the disease, i.e., causing regression of the disease. The therapeutic agent may be administered before, during or after the onset of disease or injury. The treatment of ongoing disease, where the treatment stabilizes or reduces the undesirable clinical symptoms of the patient, is of particular interest. Such treatment is desirably performed prior to complete loss of function in the affected tissues. The therapy will desirably be administered during the symptomatic stage of the disease, and in some cases after the symptomatic stage of the disease.

**[00125]** The terms "individual," "subject," "host," and "patient," are used interchangeably herein and refer to any mammalian subject for whom diagnosis, treatment, or therapy is desired, particularly humans.

**[00126]** General methods in molecular and cellular biochemistry can be found in such standard textbooks as *Molecular Cloning: A Laboratory Manual*, 3rd Ed. (Sambrook et al., HaRBor Laboratory Press 2001); *Short Protocols in Molecular Biology*, 4th Ed. (Ausubel et al. eds., John Wiley & Sons 1999); *Protein Methods* (Bollag et al., John Wiley & Sons 1996); *Nonviral Vectors for Gene Therapy* (Wagner et al. eds., Academic Press 1999); *Viral Vectors* (Kapliff & Loewy eds., Academic Press 1995); *Immunology Methods Manual* (I. Lefkovits ed., Academic Press 1997); and *Cell and Tissue Culture: Laboratory Procedures in Biotechnology* (Doyle & Griffiths, John Wiley & Sons 1998), the disclosures of which are incorporated herein by reference.

**[00127]** Where a range of values is provided, it is understood that each intervening value, to the tenth of the unit of the lower limit unless the context clearly dictates otherwise, between the upper and lower limit of that range and any other stated or intervening value in that stated range, is encompassed within the invention. The upper and lower limits of these smaller ranges may independently be included in the smaller ranges, and are also encompassed within the invention, subject to any specifically excluded limit in the stated range. Where the stated range includes one or both of the limits, ranges excluding either or both of those included limits are also included in the invention.

**[00128]** The phrase "consisting essentially of" is meant herein to exclude anything that is not the specified active component or components of a system, or that is not the specified active portion or portions of a molecule.

**[00129]** Certain ranges are presented herein with numerical values being preceded by the term "about." The term "about" is used herein to provide literal support for the exact number that it precedes, as well as a number that is near to or approximately the number that the term precedes. In determining whether a number is near to or approximately a specifically recited number, the near or approximating unrecited number may be a number which, in the context in which it is presented, provides the substantial equivalent of the specifically recited number.

**[00130]** It is appreciated that certain features of the invention, which are, for clarity, described in the context of separate embodiments, may also be provided in combination in a single embodiment. Conversely, various features of the invention, which are, for brevity, described in the context of a single embodiment, may also be provided separately or in any suitable sub-combination. All combinations of the embodiments pertaining to the invention are specifically embraced by the present invention and are disclosed herein just as if each and every combination was individually and explicitly disclosed. In addition, all sub-combinations of the various embodiments and elements thereof are also specifically embraced by

the present invention and are disclosed herein just as if each and every such sub-combination was individually and explicitly disclosed herein.

**[00131]** Aspects of the Disclosure - Part I

**[00132]** Nucleic Acids

**[00133]** Guide RNA

**[00134]** The present disclosure provides a guide RNA that directs the activities of an associated polypeptide (e.g., a site-directed modifying polypeptide) to a specific target sequence within a target DNA. A guide RNA comprises: a first segment (also referred to herein as a "DNA-targeting segment" or a "DNA-targeting sequence") and a second segment (also referred to herein as a "protein-binding segment" or a "protein-binding sequence").

**[00135]** DNA-targeting segment of a guide RNA

**[00136]** The DNA-targeting segment of a guide RNA comprises a nucleotide sequence that is complementary to a sequence in a target DNA. In other words, the DNA-targeting segment of a guide RNA interacts with a target DNA in a sequence-specific manner via hybridization (i.e., base pairing). As such, the nucleotide sequence of the DNA-targeting segment may vary and determines the location within the target DNA that the guide RNA and the target DNA will interact. The DNA-targeting segment of a guide RNA can be modified (e.g., by genetic engineering) to hybridize to any desired sequence within a target DNA.

**[00137]** The DNA-targeting segment can have a length of from about 12 nucleotides to about 100 nucleotides. For example, the DNA-targeting segment can have a length of from about 12 nucleotides (nt) to about 80 nt, from about 12 nt to about 50 nt, from about 12 nt to about 40 nt, from about 12 nt to about 30 nt, from about 12 nt to about 25 nt, from about 12 nt to about 20 nt, or from about 12 nt to about 19 nt. For example, the DNA-targeting segment can have a length of from about 19 nt to about 20 nt, from about 19 nt to about 25 nt, from about 19 nt to about 30 nt, from about 19 nt to about 35 nt, from about 19 nt to about 40 nt, from about 19 nt to about 45 nt, from about 19 nt to about 50 nt, from about 19 nt to about 60 nt, from about 19 nt to about 70 nt, from about 19 nt to about 80 nt, from about 19 nt to about 90 nt, from about 19 nt to about 100 nt, from about 20 nt to about 25 nt, from about 20 nt to about 30 nt, from about 20 nt to about 35 nt, from about 20 nt to about 40 nt, from about 20 nt to about 45 nt, from about 20 nt to about 50 nt, from about 20 nt to about 60 nt, from about 20 nt to about 70 nt, from about 20 nt to about 80 nt, from about 20 nt to about 90 nt, or from about 20 nt to about 100 nt. The nucleotide sequence (the DNA-targeting sequence) of the DNA-targeting segment that is complementary to a nucleotide sequence (target sequence) of the target DNA can have a length at least about 12 nt. For example, the DNA-targeting sequence of the DNA-targeting segment that is complementary to a target sequence of the target DNA can have a length at least about 12 nt, at least about 15 nt, at least about 18 nt, at least about 19 nt,

at least about 20 nt, at least about 25 nt, at least about 30 nt, at least about 35 nt or at least about 40 nt. For example, the DNA-targeting sequence of the DNA-targeting segment that is complementary to a target sequence of the target DNA can have a length of from about 12 nucleotides (nt) to about 80 nt, from about 12 nt to about 50 nt, from about 12 nt to about 45 nt, from about 12 nt to about 40 nt, from about 12 nt to about 35 nt, from about 12 nt to about 30 nt, from about 12 nt to about 25 nt, from about 12 nt to about 20 nt, from about 12 nt to about 19 nt, from about 19 nt to about 20 nt, from about 19 nt to about 25 nt, from about 19 nt to about 30 nt, from about 19 nt to about 35 nt, from about 19 nt to about 40 nt, from about 19 nt to about 45 nt, from about 19 nt to about 50 nt, from about 19 nt to about 60 nt, from about 20 nt to about 25 nt, from about 20 nt to about 30 nt, from about 20 nt to about 35 nt, from about 20 nt to about 40 nt, from about 20 nt to about 45 nt, from about 20 nt to about 50 nt, or from about 20 nt to about 60 nt. The nucleotide sequence (the DNA-targeting sequence) of the DNA-targeting segment that is complementary to a nucleotide sequence (target sequence) of the target DNA can have a length at least about 12 nt.

**[00138]** In some cases, the DNA-targeting sequence of the DNA-targeting segment that is complementary to a target sequence of the target DNA is 20 nucleotides in length. In some cases, the DNA-targeting sequence of the DNA-targeting segment that is complementary to a target sequence of the target DNA is 16 nucleotides, 17 nucleotides, 18 nucleotides or 19 nucleotides in length.

**[00139]** The percent complementarity between the DNA-targeting sequence of the DNA-targeting segment and the target sequence of the target DNA can be at least 60% (e.g., at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 97%, at least 98%, at least 99%, or 100%). For example, the DNA-targeting sequence may be at least about 80% identical to about 10 contiguous nucleotides, or at least about 80% identical to about 11 contiguous nucleotides, or at least about 80% identical to about 12 contiguous nucleotides, or at least about 80% identical to about 13 contiguous nucleotides, or at least about 80% identical to about 14 contiguous nucleotides, or at least about 80% identical to about 15 contiguous nucleotides, or at least about 80% identical to about 16 contiguous nucleotides, or at least about 80% identical to about 17 contiguous nucleotides of the target sequence. In some cases, the percent complementarity between the DNA-targeting sequence of the DNA-targeting segment and the target sequence of the target DNA is 100% over the seven contiguous 5'-most nucleotides of the target sequence of the complementary strand of the target DNA. In some cases, the percent complementarity between the DNA-targeting sequence of the DNA-targeting segment and the target sequence of the target DNA is at least 60% over about 20 contiguous nucleotides. In some cases, the percent complementarity between the DNA-targeting sequence of the DNA-targeting segment and the target sequence of the target DNA is 100% over the fourteen contiguous 5'-most nucleotides of the target sequence of the complementary strand of the target DNA and as low as 0% over the remainder. In such a case, the DNA-targeting sequence can be considered to be 14 nucleotides in length. In some cases, the percent complementarity between the DNA-targeting sequence of the DNA-targeting segment and the



target sequence of the target DNA is 100% over the seven contiguous 5'-most nucleotides of the target sequence of the complementary strand of the target DNA and as low as 0% over the remainder. In such a case, the DNA-targeting sequence can be considered to be 7 nucleotides in length.

**[00140]** Protein-binding segment of a guide RNA

**[00141]** The protein-binding segment of a guide RNA interacts with a site-directed modifying polypeptide. The guide RNA guides the bound polypeptide to a specific nucleotide sequence within target DNA via the above mentioned DNA-targeting segment. The protein-binding segment of a guide RNA comprises two stretches of nucleotides that are complementary to one another. The complementary nucleotides of the protein-binding segment hybridize to form a double stranded RNA duplex (dsRNA).

**[00142]** A double-molecule guide RNA comprises two separate RNA molecules. Each of the two RNA molecules of a double-molecule guide RNA comprises a stretch of nucleotides that are complementary to one another such that the complementary nucleotides of the two RNA molecules hybridize to form the double-stranded RNA duplex of the protein-binding segment.

**[00143]** In some embodiments, the duplex-forming segment of the activator-RNA is at least about 60% identical to one of the activator-RNA (tracrRNA) molecules set forth in Supplementary Table S5, or a complement thereof, over a stretch of at least 8 contiguous nucleotides. For example, the duplex-forming segment of the activator-RNA (or the DNA encoding the duplex-forming segment of the activator-RNA) is at least about 60% identical, at least about 65% identical, at least about 70% identical, at least about 75% identical, at least about 80% identical, at least about 85% identical, at least about 90% identical, at least about 95% identical, at least about 98% identical, at least about 99% identical, or 100% identical, to one of the tracrRNA sequences set forth in Supplementary Table S5, or a complement thereof, over a stretch of at least 8 contiguous, at least 9 contiguous, at least 10 contiguous, at least 11 contiguous, at least 12 contiguous, at least 13 contiguous, at least 14 contiguous or at least 15 contiguous nucleotides. For example, the activator-RNA may comprise a nucleotide sequence that is at least 70% identical over at least 10 contiguous nucleotides, at least 80% identical over at least 10 contiguous nucleotides, at least 70% identical over at least 11 contiguous nucleotides, at least 80% identical over at least 11 contiguous nucleotides, at least 70% identical over at least 12 contiguous nucleotides, or at least 80% identical over at least 12 contiguous nucleotides of one of the tracrRNA sequences set forth in Supplementary Table S5.

**[00144]** In some embodiments, the duplex-forming segment of the targeter-RNA is at least about 60% identical to one of the targeter-RNA (crRNA/CRISPR repeat) sequences set forth in Supplementary Table S5, or a complement thereof, over a stretch of at least 8 contiguous nucleotides. For example, the duplex-forming segment of the targeter-RNA (or the DNA encoding the duplex-forming segment of the targeter-RNA) is at least about 65% identical, at least about 70% identical, at least about 75% identical, at least about 80% identical, at least about 85% identical, at least about 90% identical, at least about 95%

identical, at least about 98% identical, at least about 99% identical or 100 % identical to one of the crRNA/CRISPR repeat sequences set forth in Supplementary Table S5, or a complement thereof, over a stretch of at least 8 contiguous, at least 9 contiguous, at least 10 contiguous, at least 11 contiguous, at least 12 contiguous, at least 13 contiguous, at least 14 contiguous or at least 15 contiguous nucleotides. For example, the targeter-RNA may comprise a nucleotide sequence that is at least 70% identical over at least 10 contiguous nucleotides, at least 80% identical over at least 10 contiguous nucleotides, at least 70% identical over at least 11 contiguous nucleotides, at least 80% identical over at least 11 contiguous nucleotides, at least 70% identical over at least 12 contiguous nucleotides, at least 80% identical over at least 12 contiguous nucleotides, at least 80% identical over at least 13 contiguous nucleotides, at least 80% identical over at least 14 contiguous nucleotides, at least 80% identical over at least 15 contiguous nucleotides, at least 80% identical over at least 16 contiguous nucleotides, or at least 80% identical over at least 17 contiguous nucleotides, to one of the CRISPR repeat sequences set forth in Supplementary Table S5.

**[00145]** A two-molecule guide RNA can be designed to allow for controlled (i.e., conditional) binding of a targeter-RNA with an activator-RNA. Because a two-molecule guide RNA is not functional unless both the activator-RNA and the targeter-RNA are bound in a functional complex with Cas9, a two-molecule guide RNA can be inducible (e.g., drug inducible) by rendering the binding between the activator-RNA and the targeter-RNA to be inducible. As one non-limiting example, RNA aptamers can be used to regulate (i.e., control) the binding of the activator-RNA with the targeter-RNA. Accordingly, the activator-RNA and/or the targeter-RNA can comprise an RNA aptamer sequence.

**[00146]** RNA aptamers are known in the art and are generally a synthetic version of a riboswitch. The terms "RNA aptamer" and "riboswitch" are used interchangeably herein to encompass both synthetic and natural nucleic acid sequences that provide for inducible regulation of the structure (and therefore the availability of specific sequences) of the RNA molecule of which they are part. RNA aptamers usually comprise a sequence that folds into a particular structure (e.g., a hairpin), which specifically binds a particular drug (e.g., a small molecule). Binding of the drug causes a structural change in the folding of the RNA, which changes a feature of the nucleic acid of which the aptamer is a part. As non-limiting examples: (i) an activator-RNA with an aptamer may not be able to bind to the cognate targeter-RNA unless the aptamer is bound by the appropriate drug; (ii) a targeter-RNA with an aptamer may not be able to bind to the cognate activator-RNA unless the aptamer is bound by the appropriate drug; and (iii) a targeter-RNA and an activator-RNA, each comprising a different aptamer that binds a different drug, may not be able to bind to each other unless both drugs are present. As illustrated by these examples, a two-molecule guide RNA can be designed to be inducible.

**[00147]** Examples of aptamers and riboswitches can be found, for example, in: Nakamura et al., *Genes Cells*. 2012 May;17(5):344-64; Vavalle et al., *Future Cardiol*. 2012 May;8(3):371-82; Citartan et al.,

Biosens Bioelectron. 2012 Apr 15;34(1):1-11; and Liberman et al., Wiley Interdiscip Rev RNA. 2012 May-Jun;3(3):369-84; all of which are herein incorporated by reference in their entirety.

**[00148]** Non-limiting examples of nucleotide sequences that can be included in a two-molecule guide RNA include either of the sequences set forth in Supplementary Table S5, or complements thereof pairing with any sequences set forth in Supplementary Table S5, or complements thereof that can hybridize to form a protein binding segment.

**[00149]** A single-molecule guide RNA comprises two stretches of nucleotides (a targeter-RNA and an activator-RNA) that are complementary to one another, are covalently linked (directly, or by intervening nucleotides referred to as "linkers" or "linker nucleotides"), and hybridize to form the double stranded RNA duplex (dsRNA duplex) of the protein-binding segment, thus resulting in a stem-loop structure. The targeter-RNA and the activator-RNA can be covalently linked via the 3' end of the targeter-RNA and the 5' end of the activator-RNA. Alternatively, targeter-RNA and the activator-RNA can be covalently linked via the 5' end of the targeter-RNA and the 3' end of the activator-RNA.

**[00150]** The linker of a single-molecule guide RNA can have a length of from about 3 nucleotides to about 100 nucleotides. For example, the linker can have a length of from about 3 nucleotides (nt) to about 90 nt, from about 3 nucleotides (nt) to about 80 nt, from about 3 nucleotides (nt) to about 70 nt, from about 3 nucleotides (nt) to about 60 nt, from about 3 nucleotides (nt) to about 50 nt, from about 3 nucleotides (nt) to about 40 nt, from about 3 nucleotides (nt) to about 30 nt, from about 3 nucleotides (nt) to about 20 nt or from about 3 nucleotides (nt) to about 10 nt. For example, the linker can have a length of from about 3 nt to about 5 nt, from about 5 nt to about 10 nt, from about 10 nt to about 15 nt, from about 15 nt to about 20 nt, from about 20 nt to about 25 nt, from about 25 nt to about 30 nt, from about 30 nt to about 35 nt, from about 35 nt to about 40 nt, from about 40 nt to about 50 nt, from about 50 nt to about 60 nt, from about 60 nt to about 70 nt, from about 70 nt to about 80 nt, from about 80 nt to about 90 nt, or from about 90 nt to about 100 nt. In some embodiments, the linker of a single-molecule guide RNA is 4 nt.

**[00151]** An exemplary single-molecule guide RNA comprises two complementary stretches of nucleotides that hybridize to form a dsRNA duplex. In some embodiments, one of the two complementary stretches of nucleotides of the single-molecule guide RNA (or the DNA encoding the stretch) is at least about 60% identical to one of the activator-RNA (tracrRNA) molecules set forth in Supplementary Table S5, or a complement thereof, over a stretch of at least 8 contiguous nucleotides. For example, one of the two complementary stretches of nucleotides of the single-molecule guide RNA (or the DNA encoding the stretch) is at least about 65% identical, at least about 70% identical, at least about 75% identical, at least about 80% identical, at least about 85% identical, at least about 90% identical, at least about 95% identical, at least about 98% identical, at least about 99% identical or 100% identical to one of the tracrRNA sequences set forth in Supplementary Table S5, or a complement thereof, over a stretch of at least 8 contiguous, at least 9 contiguous, at least 10 contiguous, at least 11 contiguous, at least 12

contiguous, at least 13 contiguous, at least 14 contiguous or at least 15 contiguous nucleotides. For example, the single-molecule guide RNA may comprise a nucleotide sequence that is at least 70% identical over at least 10 contiguous nucleotides, at least 80% identical over at least 10 contiguous nucleotides, at least 70% identical over at least 11 contiguous nucleotides, at least 80% identical over at least 11 contiguous nucleotides, at least 70% identical over at least 12 contiguous nucleotides, or at least 80% identical over at least 12 contiguous nucleotides of one of the tracrRNA sequences set forth in Supplementary Table S5.

**[00152]** In some embodiments, one of the two complementary stretches of nucleotides of the single-molecule guide RNA (or the DNA encoding the stretch) is at least about 60% identical to one of the targeter-RNA (crRNA/CRISPR repeat) sequences set forth in Supplementary Table S5, or a complement thereof, over a stretch of at least 8 contiguous nucleotides. For example, one of the two complementary stretches of nucleotides of the single-molecule guide RNA (or the DNA encoding the stretch) is at least about 65% identical, at least about 70% identical, at least about 75% identical, at least about 80% identical, at least about 85% identical, at least about 90% identical, at least about 95% identical, at least about 98% identical, at least about 99% identical or 100 % identical to one of the crRNA/CRISPR repeat sequences set forth in Supplementary Table S5, or a complement thereof, over a stretch of at least 8 contiguous , at least 9 contiguous, at least 10 contiguous, at least 11 contiguous, at least 12 contiguous, at least 13 contiguous, at least 14 contiguous or at least 15 contiguous nucleotides. For example, the single-molecule guide RNA may comprise a nucleotide sequence that is at least 70% identical over at least 10 contiguous nucleotides, at least 80% identical over at least 10 contiguous nucleotides, at least 70% identical over at least 11 contiguous nucleotides, at least 80% identical over at least 11 contiguous nucleotides, at least 70% identical over at least 12 contiguous nucleotides, or at least 80% identical over at least 12 contiguous nucleotides, or at least about 80% identical to about 13 contiguous nucleotides, or at least about 80% identical to about 14 contiguous nucleotides, or at least about 80% identical to about 15 contiguous nucleotides, or at least about 80% identical to about 16 contiguous nucleotides, or at least about 80% identical to about 17 contiguous nucleotides of one of the CRISPR repeat sequences set forth in Supplementary Table S5.

**[00153]** Appropriate naturally occurring cognate pairs of crRNAs and tracrRNAs can be routinely determined by taking into account the species name and base-pairing (for the dsRNA duplex of the protein-binding domain) when determining appropriate cognate pairs. Non-cognate pairs are also contemplated for use in the invention. In some embodiments of non-cognate pairs, each RNA is from a Cas9 cluster herein wherein the Cas9 endonucleases share 80% identity over 80% of their amino acid sequences.

**[00154]** Artificial sequences that share very little identity (roughly 50% identity, or alternatively about 70% identity over about 50% of the full length protein) with naturally occurring a tracrRNAs and crRNAs can function with Cas9 to cleave target DNA as long as the structure of the protein-binding domain of the

guide RNA is conserved. Thus, RNA folding structure of a naturally occurring protein-binding domain of a DNA-targeting RNA can be taken into account in order to design artificial protein-binding domains (either two-molecule or single-molecule versions). As structures can readily be produced by one of ordinary skill in the art for any naturally occurring crRNA:tracrRNA pair from any, an artificial DNA-targeting-RNA can be designed to mimic the natural structure for a given species when using the Cas9 (or a related Cas9) from that species. Thus, a suitable guide RNA can be an artificially designed RNA (non-naturally occurring) comprising a protein-binding domain that was designed to mimic the structure of a protein-binding domain of a naturally occurring guide RNA.

**[00155]** The protein-binding segment can have a length of from about 10 nucleotides to about 100 nucleotides. For example, the protein-binding segment can have a length of from about 15 nucleotides (nt) to about 80 nt, from about 15 nt to about 50 nt, from about 15 nt to about 40 nt, from about 15 nt to about 30 nt or from about 15 nt to about 25 nt.

**[00156]** Also with regard to both a single-molecule guide RNA and to a double-molecule guide RNA, the dsRNA duplex of the protein-binding segment can have a length from about 6 base pairs (bp) to about 50bp. For example, the dsRNA duplex of the protein-binding segment can have a length from about 6 bp to about 40 bp, from about 6 bp to about 30bp, from about 6 bp to about 25 bp, from about 6 bp to about 20 bp, from about 6 bp to about 15 bp, from about 8 bp to about 40 bp, from about 8 bp to about 30bp, from about 8 bp to about 25 bp, from about 8 bp to about 20 bp or from about 8 bp to about 15 bp. For example, the dsRNA duplex of the protein-binding segment can have a length from about from about 8 bp to about 10 bp, from about 10 bp to about 15 bp, from about 15 bp to about 18 bp, from about 18 bp to about 20 bp, from about 20 bp to about 25 bp, from about 25 bp to about 30 bp, from about 30 bp to about 35 bp, from about 35 bp to about 40 bp, or from about 40 bp to about 50 bp. In some embodiments, the dsRNA duplex of the protein-binding segment has a length of 36 base pairs. The percent complementarity between the nucleotide sequences that hybridize to form the dsRNA duplex of the protein-binding segment can be at least about 60%. For example, the percent complementarity between the nucleotide sequences that hybridize to form the dsRNA duplex of the protein-binding segment can be at least about 65%, at least about 70%, at least about 75%, at least about 80%, at least about 85%, at least about 90%, at least about 95%, at least about 98%, or at least about 99% . In some cases, the percent complementarity between the nucleotide sequences that hybridize to form the dsRNA duplex of the protein-binding segment is 100%.

**[00157]** Site-directed modifying polypeptide

**[00158]** A guide RNA and a site-directed modifying polypeptide form a complex. The guide RNA provides target specificity to the complex by comprising a nucleotide sequence that is complementary to a sequence of a target DNA (as noted above). The site-directed modifying polypeptide is guided to a DNA sequence (e.g. a chromosomal sequence or an extrachromosomal sequence, e.g. an episomal

sequence, a minicircle sequence, a mitochondrial sequence, a chloroplast sequence, etc.) by virtue of its association with at least the protein-binding segment of the guide RNA (described above).

**[00159]** A site-directed modifying polypeptide modifies target DNA (e.g., cleavage or methylation of target DNA) and/or a polypeptide associated with target DNA (e.g., methylation or acetylation of a histone tail). A site-directed modifying polypeptide is also referred to herein as a "site-directed polypeptide" or an "RNA binding site-directed modifying polypeptide." In some cases, the site-directed modifying polypeptide is a naturally-occurring modifying polypeptide. In other cases, the site-directed modifying polypeptide is not a naturally-occurring polypeptide (e.g., a chimeric polypeptide as discussed below or a naturally-occurring polypeptide that is modified, e.g., mutation, deletion, insertion).

**[00160]** Naturally-occurring site-directed modifying polypeptides bind a guide RNA, are thereby directed to a specific sequence within a target DNA, and cleave the target DNA to generate a double strand break. The amino acid sequences of exemplary naturally-occurring Cas9 site-directed modifying polypeptide orthologs are set out in SEQ ID NOs: 1-800. The amino acid sequence of the *S. pyogenes* Cas9 endonuclease is set out in SEQ ID NO: 8. A site-directed modifying polypeptide comprises two portions, an RNA-binding portion and an activity portion. In some embodiments, a site-directed modifying polypeptide comprises: (i) an RNA-binding portion that interacts with a guide RNA, wherein the guide RNA comprises a nucleotide sequence that is complementary to a sequence in a target DNA; and (ii) an activity portion that exhibits site-directed enzymatic activity (e.g., activity for DNA methylation, activity for DNA cleavage, activity for histone acetylation, activity for histone methylation, etc.), wherein the site of enzymatic activity is determined by the guide RNA.

**[00161]** In other embodiments, a site-directed modifying polypeptide comprises: (i) an RNA-binding portion that interacts with a guide RNA, wherein the guide RNA comprises a nucleotide sequence that is complementary to a sequence in a target DNA; and (ii) an activity portion that modulates transcription within the target DNA (e.g., to increase or decrease transcription), wherein the site of modulated transcription within the target DNA is determined by the guide RNA.

**[00162]** In some cases, a site-directed modifying polypeptide has enzymatic activity that modifies target DNA (e.g., nuclease activity, methyltransferase activity, demethylase activity, DNA repair activity, DNA damage activity, deamination activity, dismutase activity, alkylation activity, depurination activity, oxidation activity, pyrimidine dimer forming activity, integrase activity, transposase activity, recombinase activity, polymerase activity, ligase activity, helicase activity, photolyase activity or glycosylase activity).

**[00163]** In other cases, a site-directed modifying polypeptide has enzymatic activity that modifies a polypeptide (e.g., a histone) associated with target DNA (e.g., methyltransferase activity, demethylase activity, acetyltransferase activity, deacetylase activity, kinase activity, phosphatase activity, ubiquitin ligase activity, deubiquitinating activity, adenylation activity, deadenylation activity, SUMOylating activity,

deSUMOylating activity, ribosylation activity, deribosylation activity, myristoylation activity or demyristoylation activity).

**[00164]** Exemplary site-directed modifying polypeptides

**[00165]** In some cases, the site-directed modifying polypeptide comprises an amino acid sequence having at least about 75%, at least about 80%, at least about 85%, at least about 90%, at least about 95%, at least about 99%, or 100%, amino acid sequence identity to amino acids 7-166 and/or 731-1003 of SEQ ID NO: 8, or to the corresponding portions in any of the amino acid sequences set forth as SEQ ID NOs: 1-800.

**[00166]** Nucleic acid modifications

**[00167]** In some embodiments, a nucleic acid (e.g., a guide RNA) comprises one or more modifications, e.g., a base modification, a backbone modification, etc, to provide the nucleic acid with a new or enhanced feature (e.g., improved stability). As is known in the art, a nucleoside is a base-sugar combination. The base portion of the nucleoside is normally a heterocyclic base. The two most common classes of such heterocyclic bases are the purines and the pyrimidines. Nucleotides are nucleosides that further include a phosphate group covalently linked to the sugar portion of the nucleoside. For those nucleosides that include a pentofuranosyl sugar, the phosphate group can be linked to the 2', the 3', or the 5' hydroxyl moiety of the sugar. In forming oligonucleotides, the phosphate groups covalently link adjacent nucleosides to one another to form a linear polymeric compound. In turn, the respective ends of this linear polymeric compound can be further joined to form a circular compound, however, linear compounds are generally suitable. In addition, linear compounds may have internal nucleotide base complementarity and may therefore fold in a manner as to produce a fully or partially double-stranded compound. Within oligonucleotides, the phosphate groups are commonly referred to as forming the internucleoside backbone of the oligonucleotide. The normal linkage or backbone of RNA and DNA is a 3' to 5' phosphodiester linkage.

**[00168]** Modified backbones and modified internucleoside linkages

**[00169]** Examples of suitable nucleic acids containing modifications include nucleic acids containing modified backbones or non-natural internucleoside linkages. Nucleic acids having modified backbones include those that retain a phosphorus atom in the backbone and those that do not have a phosphorus atom in the backbone.

**[00170]** Suitable modified oligonucleotide backbones containing a phosphorus atom therein include, for example, phosphorothioates, chiral phosphorothioates, phosphorodithioates, phosphotriesters, aminoalkylphosphotriesters, methyl and other alkyl phosphonates including 3'-alkylene phosphonates, 5'-alkylene phosphonates and chiral phosphonates, phosphinates, phosphoramidates including 3<sup>1</sup>-amino phosphoramidate and aminoalkylphosphoramidates, phosphorodiamidates, thionophosphoramidates,

thionoalkylphosphonates, thionoalkylphosphotriesters, selenophosphates and boranophosphates having normal 3'-5' linkages, 2'-5' linked analogs of these, and those having inverted polarity wherein one or more internucleotide linkages is a 3' to 3', 5' to 5' or 2' to 2' linkage. Suitable oligonucleotides having inverted polarity comprise a single 3' to 3' linkage at the 3'-most internucleotide linkage i.e. a single inverted nucleoside residue which may be a basic (the nucleobase is missing or has a hydroxyl group in place thereof). Various salts (such as, for example, potassium or sodium), mixed salts and free acid forms are also included.

**[00171]** In some embodiments, a nucleic acid comprises one or more phosphorothioate and/or heteroatom internucleoside linkages, in particular  $-\text{CH}_2\text{-NH-O-CH}_2-$ ,  $-\text{CH}_2\text{-N(CH}_3\text{)-O-CH}_2-$  (known as a methylene (methylimino) or MMI backbone),  $-\text{CH}_2\text{-O-N(CH}_3\text{)-CH}_2-$ ,  $-\text{CH}_2\text{-N(CH}_3\text{)-N(CH}_3\text{)-CH}_2-$  and  $-\text{O-N(CH}_3\text{)-CH}_2\text{-CH}_2-$  (wherein the native phosphodiester internucleotide linkage is represented as  $-\text{O-P(=O)(OH)-O-CH}_2-$ ). MMI type internucleoside linkages are disclosed in the above referenced U.S. Pat. No. 5,489,677. Suitable amide internucleoside linkages are disclosed in U.S. Pat. No. 5,602,240.

**[00172]** Also suitable are nucleic acids having morpholino backbone structures as described in, e.g., U.S. Pat. No. 5,034,506. For example, in some embodiments, a nucleic acid comprises a 6-membered morpholino ring in place of a ribose ring. In some of these embodiments, a phosphorodiamidate or other non-phosphodiester internucleoside linkage replaces a phosphodiester linkage.

**[00173]** Suitable modified polynucleotide backbones that do not include a phosphorus atom therein have backbones that are formed by short chain alkyl or cycloalkyl internucleoside linkages, mixed heteroatom and alkyl or cycloalkyl internucleoside linkages, or one or more short chain heteroatomic or heterocyclic internucleoside linkages. These include those having morpholino linkages (formed in part from the sugar portion of a nucleoside); siloxane backbones; sulfide, sulfoxide and sulfone backbones; formacetyl and thioformacetyl backbones; methylene formacetyl and thioformacetyl backbones; riboacetyl backbones; alkene containing backbones; sulfamate backbones; methyleneimino and methylenehydrazino backbones; sulfonate and sulfonamide backbones; amide backbones; and others having mixed N, O, S and  $\text{CH}_2$  component parts.

**[00174]** Mimetics

**[00175]** A nucleic acid can be a nucleic acid mimetic. The term "mimetic" as it is applied to polynucleotides is intended to include polynucleotides wherein only the furanose ring or both the furanose ring and the internucleotide linkage are replaced with non-furanose groups, replacement of only the furanose ring is also referred to in the art as being a sugar surrogate. The heterocyclic base moiety or a modified heterocyclic base moiety is maintained for hybridization with an appropriate target nucleic acid. One such nucleic acid, a polynucleotide mimetic that has been shown to have excellent hybridization properties, is referred to as a peptide nucleic acid (PNA). In PNA, the sugar-backbone of a polynucleotide is



replaced with an amide containing backbone, in particular an aminoethylglycine backbone. The nucleotides are retained and are bound directly or indirectly to aza nitrogen atoms of the amide portion of the backbone.

**[00176]** One polynucleotide mimetic that has been reported to have excellent hybridization properties is a peptide nucleic acid (PNA). The backbone in PNA compounds is two or more linked aminoethylglycine units which gives PNA an amide containing backbone. The heterocyclic base moieties are bound directly or indirectly to aza nitrogen atoms of the amide portion of the backbone. Representative U.S. patents that describe the preparation of PNA compounds include, but are not limited to: U.S. Pat. Nos. 5,539,082; 5,714,331; and 5,719,262.

**[00177]** Another class of polynucleotide mimetic that has been studied is based on linked morpholino units (morpholino nucleic acid) having heterocyclic bases attached to the morpholino ring. A number of linking groups have been reported that link the morpholino monomeric units in a morpholino nucleic acid. One class of linking groups has been selected to give a non-ionic oligomeric compound. The non-ionic morpholino-based oligomeric compounds are less likely to have undesired interactions with cellular proteins. Morpholino-based polynucleotides are nonionic mimics of oligonucleotides which are less likely to form undesired interactions with cellular proteins (Dwaine A. Braasch and David R. Corey, *Biochemistry*, 2002, 41(14), 45034510). Morpholino-based polynucleotides are disclosed in U.S. Pat. No. 5,034,506. A variety of compounds within the morpholino class of polynucleotides have been prepared, having a variety of different linking groups joining the monomeric subunits.

**[00178]** A further class of polynucleotide mimetic is referred to as cyclohexenyl nucleic acids (CeNA). The furanose ring normally present in a DNA/RNA molecule is replaced with a cyclohexenyl ring. CeNA DMT protected phosphoramidite monomers have been prepared and used for oligomeric compound synthesis following classical phosphoramidite chemistry. Fully modified CeNA oligomeric compounds and oligonucleotides having specific positions modified with CeNA have been prepared and studied (see Wang et al., *J. Am. Chem. Soc.*, 2000, 122, 85958602). In general the incorporation of CeNA monomers into a DNA chain increases its stability of a DNA/RNA hybrid. CeNA oligoadenylates formed complexes with RNA and DNA complements with similar stability to the native complexes. The study of incorporating CeNA structures into natural nucleic acid structures was shown by NMR and circular dichroism to proceed with easy conformational adaptation.

**[00179]** A further modification includes Locked Nucleic Acids (LNAs) in which the 2'-hydroxyl group is linked to the 4' carbon atom of the sugar ring thereby forming a 2'-C,4'-C-oxymethylene linkage thereby forming a bicyclic sugar moiety. The linkage can be a methylene (-CH<sub>2</sub>-), group bridging the 2' oxygen atom and the 4' carbon atom wherein n is 1 or 2 (Singh et al., *Chem. Commun.*, 1998, 4, 455-456). LNA and LNA analogs display very high duplex thermal stabilities with complementary DNA and RNA (T<sub>m</sub>=+3 to +10° C), stability towards 3'-exonucleolytic degradation and good solubility properties. Potent and

nontoxic antisense oligonucleotides containing LNAs have been described (Wahlestedt et al., Proc. Natl. Acad. Sci. U.S.A., 2000, 97, 5633-5638).

**[00180]** The synthesis and preparation of the LNA monomers adenine, cytosine, guanine, 5-methyl-cytosine, thymine and uracil, along with their oligomerization, and nucleic acid recognition properties have been described (Koshkin et al., Tetrahedron, 1998, 54, 3607-3630). LNAs and preparation thereof are also described in WO 98/39352 and WO 99/14226.

**[00181]** Modified sugar moieties

**[00182]** A nucleic acid can also include one or more substituted sugar moieties. Suitable polynucleotides comprise a sugar substituent group selected from: OH; F; O-, S-, or N-alkyl; O-, S-, or N-alkenyl; O-, S- or N-alkynyl; or O-alkyl-O-alkyl, wherein the alkyl, alkenyl and alkynyl may be substituted or unsubstituted C.sub.1 to C<sub>10</sub> alkyl or C<sub>2</sub> to C<sub>10</sub> alkenyl and alkynyl. Particularly suitable are O((CH<sub>2</sub>)<sub>n</sub>O)<sub>m</sub>CH<sub>3</sub>, O(CH<sub>2</sub>)<sub>n</sub>OCH<sub>3</sub>, O(CH<sub>2</sub>)<sub>n</sub>NH<sub>2</sub>, O(CH<sub>2</sub>)CH<sub>3</sub>, O(CH<sub>2</sub>)<sub>n</sub>ONH<sub>2</sub>, and O(CH<sub>2</sub>)<sub>n</sub>ON((CH<sub>2</sub>)<sub>n</sub>CH<sub>3</sub>)<sub>2</sub>, where n and m are from 1 to about 10. Other suitable polynucleotides comprise a sugar substituent group selected from: C<sub>1</sub> to C<sub>10</sub> lower alkyl, substituted lower alkyl, alkenyl, alkynyl, alkaryl, aralkyl, O-alkaryl or O-aralkyl, SH, SCH<sub>3</sub>, OCN, Cl, Br, CN, CF<sub>3</sub>, OCF<sub>3</sub>, SOCH<sub>3</sub>, SO<sub>2</sub>CH<sub>3</sub>, ONO<sub>2</sub>, NO<sub>2</sub>, N<sub>3</sub>, NH<sub>2</sub>, heterocycloalkyl, heterocycloalkaryl, aminoalkylamino, polyalkylamino, substituted silyl, an RNA cleaving group, a reporter group, an intercalator, a group for improving the pharmacokinetic properties of an oligonucleotide, or a group for improving the pharmacodynamic properties of an oligonucleotide, and other substituents having similar properties. A suitable modification includes 2'-methoxyethoxy 2'-O-CH<sub>2</sub>CH<sub>2</sub>OCH<sub>3</sub>, also known as -2'-O-(2-methoxyethyl) or 2'-MOE) (Martin et al., Hely. Chim. Acta, 1995, 78, 486-504) i.e., an alkoxyalkoxy group. A further suitable modification includes 2'-dimethylaminoethoxy, i.e., a O(CH<sub>2</sub>)<sub>2</sub>ON(CH<sub>3</sub>)<sub>2</sub> group, also known as 2'-DMAOE, as described in examples hereinbelow, and 2'-dimethylaminoethoxyethoxy (also known in the art as 2'-O-dimethyl-amino-ethoxy-ethyl or 2'-DMAEOE), i.e., 2'-O-CH<sub>2</sub>-O-CH<sub>2</sub>-N(CH<sub>3</sub>)<sub>2</sub>.

**[00183]** Other suitable sugar substituent groups include methoxy (-O-CH<sub>3</sub>), aminopropoxy (--O--CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>NH<sub>2</sub>), allyl (-CH<sub>2</sub>-CH=CH<sub>2</sub>), -O-allyl (--O--CH<sub>2</sub>-CH=CH<sub>2</sub>) and fluoro (F). 2'-sugar substituent groups may be in the arabino (up) position or ribo (down) position. A suitable 2'-arabino modification is 2'-F. Similar modifications may also be made at other positions on the oligomeric compound, particularly the 3' position of the sugar on the 3' terminal nucleoside or in 2'-5' linked oligonucleotides and the 5' position of 5' terminal nucleotide. Oligomeric compounds may also have sugar mimetics such as cyclobutyl moieties in place of the pentofuranosyl sugar.

**[00184]** Base modifications and substitutions

**[00185]** A nucleic acid may also include nucleobase (often referred to in the art simply as "base") modifications or substitutions. As used herein, "unmodified" or "natural" nucleobases include the purine

bases adenine (A) and guanine (G), and the pyrimidine bases thymine (T), cytosine (C) and uracil (U). Modified nucleobases include other synthetic and natural nucleobases such as 5-methylcytosine (5-me-C), 5-hydroxymethyl cytosine, xanthine, hypoxanthine, 2-aminoadenine, 6-methyl and other alkyl derivatives of adenine and guanine, 2-propyl and other alkyl derivatives of adenine and guanine, 2-thiouracil, 2-thiothymine and 2-thiocytosine, 5-halouracil and cytosine, 5-propynyl (-C≡C-CH<sub>3</sub>) uracil and cytosine and other alkynyl derivatives of pyrimidine bases, 6-azo uracil, cytosine and thymine, 5-uracil (pseudouracil), 4-thiouracil, 8-halo, 8-amino, 8-thiol, 8-thioalkyl, 8-hydroxyl and other 8-substituted adenines and guanines, 5-halo particularly 5-bromo, 5-trifluoromethyl and other 5-substituted uracils and cytosines, 7-methylguanine and 7-methyladenine, 2-F-adenine, 2-amino-adenine, 8-azaguanine and 8-azaadenine, 7-deazaguanine and 7-deazaadenine and 3-deazaguanine and 3-deazaadenine. Further modified nucleobases include tricyclic pyrimidines such as phenoxazine cytidine(1H-pyrimido(5,4-b)(1,4)benzoxazin-2(3H)-one), phenothiazine cytidine (1H-pyrimido(5,4-b)(1,4)benzothiazin-2(3H)-one), G-clamps such as a substituted phenoxazine cytidine (e.g. 9-(2-aminoethoxy)-H-pyrimido(5,4-(b)(1,4)benzoxazin-2(3H)-one), carbazole cytidine (2H-pyrimido(4,5-b)indol-2-one), pyridoindole cytidine (H-pyrido(3',2':4,5)pyrrolo(2,3-d)pyrimidin-2-one).

**[00186]** Heterocyclic base moieties may also include those in which the purine or pyrimidine base is replaced with other heterocycles, for example 7-deaza-adenine, 7-deazaguanosine, 2-aminopyridine and 2-pyridone. Further nucleobases include those disclosed in U.S. Pat. No. 3,687,808, those disclosed in The Concise Encyclopedia Of Polymer Science And Engineering, pages 858-859, Kroschwitz, J. I., ed. John Wiley & Sons, 1990, those disclosed by Englisch et al., *Angewandte Chemie, International Edition*, 1991, 30, 613, and those disclosed by Sanghvi, Y. S., Chapter 15, *Antisense Research and Applications*, pages 289-302, Crooke, S. T. and Lebleu, B., ed., CRC Press, 1993. Certain of these nucleobases are useful for increasing the binding affinity of an oligomeric compound. These include 5-substituted pyrimidines, 6-azapyrimidines and N-2, N-6 and O-6 substituted purines, including 2-aminopropyladenine, 5-propynyluracil and 5-propynylcytosine. 5-methylcytosine substitutions have been shown to increase nucleic acid duplex stability by 0.6-1.2° C. (Sanghvi et al., eds., *Antisense Research and Applications*, CRC Press, Boca Raton, 1993, pp. 276-278) and are suitable base substitutions, e.g., when combined with 2'-O-methoxyethyl sugar modifications.

**[00187]** "Complementary" refers to the capacity for pairing, through base stacking and specific hydrogen bonding, between two sequences comprising naturally or non-naturally occurring (e.g., modified as described above) bases (nucleosides) or analogs thereof. For example, if a base at one position of a nucleic acid is capable of hydrogen bonding with a base at the corresponding position of a target, then the bases are considered to be complementary to each other at that position. Nucleic acids can comprise universal bases, or inert abasic spacers that provide no positive or negative contribution to hydrogen bonding. Base pairings may include both canonical Watson-Crick base pairing and non-Watson-Crick base pairing (e.g., Wobble base pairing and Hoogsteen base pairing). It is understood that for

complementary base pairings, adenosine-type bases (A) are complementary to thymidine-type bases (T) or uracil-type bases (U), that cytosine-type bases (C) are complementary to guanosine-type bases (G), and that universal bases such as such as 3-nitropyrrole or 5-nitroindole can hybridize to and are considered complementary to any A, C, U, or T. Nichols et al., *Nature*, 1994;369:492-493 and Loakes et al., *Nucleic Acids Res.*, 1994;22:4039-4043. Inosine (I) has also been considered in the art to be a universal base and is considered complementary to any A, C, U, or T. See Watkins and SantaLucia, *Nucl. Acids Research*, 2005; 33 (19): 6258-6267.

**[00188]** Conjugates

**[00189]** Another possible modification of a nucleic acid involves chemically linking to the polynucleotide one or more moieties or conjugates which enhance the activity, cellular distribution or cellular uptake of the oligonucleotide. These moieties or conjugates can include conjugate groups covalently bound to functional groups such as primary or secondary hydroxyl groups. Conjugate groups include, but are not limited to, intercalators, reporter molecules, polyamines, polyamides, polyethylene glycols, polyethers, groups that enhance the pharmacodynamic properties of oligomers, and groups that enhance the pharmacokinetic properties of oligomers. Suitable conjugate groups include, but are not limited to, cholesterol, lipids, phospholipids, biotin, phenazine, folate, phenanthridine, anthraquinone, acridine, fluoresceins, rhodamines, coumarins, and dyes. Groups that enhance the pharmacodynamic properties include groups that improve uptake, enhance resistance to degradation, and/or strengthen sequence-specific hybridization with the target nucleic acid. Groups that enhance the pharmacokinetic properties include groups that improve uptake, distribution, metabolism or excretion of a nucleic acid.

**[00190]** Conjugate moieties include but are not limited to lipid moieties such as a cholesterol moiety (Letsinger et al., *Proc. Natl. Acad. Sci. USA*, 1989, 86, 6553-6556), cholic acid (Manoharan et al., *Bioorg. Med. Chem. Lett.*, 1994, 4, 1053-1060), a thioether, e.g., hexyl-S-tritylthiol (Manoharan et al., *Ann. N.Y. Acad. Sci.*, 1992, 660, 306-309; Manoharan et al., *Bioorg. Med. Chem. Lett.*, 1993, 3, 2765-2770), a thiocholesterol (Oberhauser et al., *Nucl. Acids Res.*, 1992, 20, 533-538), an aliphatic chain, e.g., dodecandiol or undecyl residues (Saison-Behmoaras et al., *EMBO J.*, 1991, 10, 1111-1118; Kabanov et al., *FEBS Lett.*, 1990, 259, 327-330; Svinarchuk et al., *Biochimie*, 1993, 75, 49-54), a phospholipid, e.g., di-hexadecyl-rac-glycerol or triethylammonium 1,2-di-O-hexadecyl-rac-glycero-3-H-phosphonate (Manoharan et al., *Tetrahedron Lett.*, 1995, 36, 3651-3654; Shea et al., *Nucl. Acids Res.*, 1990, 18, 3777-3783), a polyamine or a polyethylene glycol chain (Manoharan et al., *Nucleosides & Nucleotides*, 1995, 14, 969-973), or adamantane acetic acid (Manoharan et al., *Tetrahedron Lett.*, 1995, 36, 3651-3654), a palmitoyl moiety (Mishra et al., *Biochim. Biophys. Acta*, 1995, 1264, 229-237), or an octadecylamine or hexylamino-carbonyl-oxcholesterol moiety (Crooke et al., *J. Pharmacol. Exp. Ther.*, 1996, 277, 923-937).

**[00191]** A conjugate may include a "Protein Transduction Domain" or PTD (also known as a CPP - cell penetrating peptide), which may refer to a polypeptide, polynucleotide, carbohydrate, or organic or inorganic compound that facilitates traversing a lipid bilayer, micelle, cell membrane, organelle membrane, or vesicle

membrane. A PTD attached to another molecule, which can range from a small polar molecule to a large macromolecule and/or a nanoparticle, facilitates the molecule traversing a membrane, for example going from extracellular space to intracellular space, or cytosol to within an organelle. In some embodiments, a PTD is covalently linked to the amino terminus of an exogenous polypeptide (e.g., a site-directed modifying polypeptide). In some embodiments, a PTD is covalently linked to the carboxyl terminus of an exogenous polypeptide (e.g., a site-directed modifying polypeptide). In some embodiments, a PTD is covalently linked to a nucleic acid (e.g., a guide RNA, a polynucleotide encoding a guide RNA, a polynucleotide encoding a site-directed modifying polypeptide, etc.). Exemplary PTDs include but are not limited to a minimal undecapeptide protein transduction domain (corresponding to residues 47-57 of HIV-1 TAT comprising YGRKKRRQRRR; a polyarginine sequence comprising a number of arginines sufficient to direct entry into a cell (e.g., 3, 4, 5, 6, 7, 8, 9, 10, or 10-50 arginines); a VP22 domain (Zender et al. (2002) *Cancer Gene Ther.* 9(6):489-96); an *Drosophila* Antennapedia protein transduction domain (Noguchi et al. (2003) *Diabetes* 52(7):1732-1737); a truncated human calcitonin peptide (Trehin et al. (2004) *Pharm. Research* 21:1248-1256); polylysine (Wender et al. (2000) *Proc. Natl. Acad. Sci. USA* 97:13003-13008); RRQRRTSKLMKR; Transport= GWTLNSAGYLLGKINLKALAALAKKIL; KALAWKAKLAKALAKALAKHLAKALAKALKCEA; and RQIKIWFQNRRMKWKK. Exemplary PTDs include but are not limited to, YGRKKRRQRRR; RKKRRQRRR; an arginine homopolymer of from 3 arginine residues to 50 arginine residues; Exemplary PTD domain amino acid sequences include, but are not limited to, any of the following: YGRKKRRQRRR; RKKRRQRRR; YARAAARQARA; THRLPRRRRRR; and GGRRARRRRRR. In some embodiments, the PTD is an activatable CPP (ACPP) (Aguilera et al. (2009) *Integr Biol (Camb)* June; 1(5-6): 371-381). ACPPs comprise a polycationic CPP (e.g., Arg9 or "R9") connected via a cleavable linker to a matching polyanion (e.g., Glu9 or "E9"), which reduces the net charge to nearly zero and thereby inhibits adhesion and uptake into cells. Upon cleavage of the linker, the polyanion is released, locally unmasking the polyarginine and its inherent adhesiveness, thus "activating" the ACPP to traverse the membrane.

**[00192]** Exemplary guide RNAs

**[00193]** In some embodiments, a guide RNA comprises two separate RNA polynucleotide molecules. The first of the two separate RNA polynucleotide molecules (the activator-RNA) comprises a nucleotide sequence having at least about 60%, at least about 65%, at least about 70%, at least about 75%, at least about 80%, at least about 85%, at least about 90%, at least about 95%, at least about 98%, at least about 99%, or 100% nucleotide sequence identity over a stretch of at least 8 contiguous, at least 9 contiguous, at least 10 contiguous, at least 11 contiguous, at least 12 contiguous, at least 13 contiguous, at least 14 contiguous or at least 15 contiguous nucleotides to any one of the tracrRNA nucleotide sequences set forth in Supplementary Table S5, or complements thereof. The second of the two separate RNA polynucleotide molecules (the targeter-RNA) comprises a nucleotide sequence having at least about 60%, at least about 65%, at least about 70%, at least about 75%, at least about 80%, at least about 85%,

at least about 90%, at least about 95%, at least about 98%, at least about 99%, or 100% nucleotide sequence identity over a stretch of at least 8 contiguous, at least 9 contiguous, at least 10 contiguous, at least 11 contiguous, at least 12 contiguous, at least 13 contiguous, at least 14 contiguous or at least 15 contiguous nucleotides to the cognate CRISPR repeat nucleotide sequence set forth in Supplementary Table S5, or complements thereof. In some embodiments, a suitable guide RNA is a single-molecule RNA polynucleotide and comprises a first nucleotide sequence having at least about 60%, at least about 65%, at least about 70%, at least about 75%, at least about 80%, at least about 85%, at least about 90%, at least about 95%, at least about 98%, at least about 99%, or 100% nucleotide sequence identity over a stretch of at least 8 contiguous, at least 9 contiguous, at least 10 contiguous, at least 11 contiguous, at least 12 contiguous, at least 13 contiguous, at least 14 contiguous or at least 15 contiguous nucleotides to any one of the tracrRNA nucleotide sequences set forth in Supplementary Table S5 and a second nucleotide sequence having at least about 60%, at least about 65%, at least about 70%, at least about 75%, at least about 80%, at least about 85%, at least about 90%, at least about 95%, at least about 98%, at least about 99%, or 100% nucleotide sequence identity over a stretch of at least 8 contiguous, at least 9 contiguous, at least 10 contiguous, at least 11 contiguous, at least 12 contiguous, at least 13 contiguous, at least 14 contiguous or at least 15 contiguous nucleotides to the cognate CRISPR repeat nucleotide sequence set forth in Supplementary Table S5, or complements thereof.

**[00194]** In some embodiments, the single-molecule guide RNAs comprise a DNA-targeting segment and a protein-binding segment complementary thereto, wherein the protein-binding segment comprises a tracrRNA set out in Supplementary Table S5 or wherein the protein-binding segment comprises a tracrRNA at least 80% identical over at least 20 nucleotides to a tracrRNA set out in Supplementary Table S5, or at least about 60%, at least about 65%, at least about 70%, at least about 75%, at least about 80%, at least about 85%, at least about 90%, at least about 95%, at least about 98%, at least about 99%, or 100% nucleotide sequence identity over a stretch of at least 8 contiguous, at least 9 contiguous, at least 10 contiguous, at least 11 contiguous, at least 12 contiguous, at least 13 contiguous, at least 14 contiguous or at least 15 contiguous nucleotides of any one of the tracrRNA nucleotide sequences set forth in Supplementary Table S5. For example, the protein-binding segment may comprise a tracrRNA at least 70% identical over at least 10 contiguous nucleotides, at least 80% identical over at least 10 contiguous nucleotides, at least 70% identical over at least 11 contiguous nucleotides, at least 80% identical over at least 11 contiguous nucleotides, at least 70% identical over at least 12 contiguous nucleotides, or at least 80% identical over at least 12 contiguous nucleotides.

**[00195]** In some embodiments, the single-molecule guide RNAs comprise a DNA-targeting segment and a protein-binding segment, wherein the protein-binding segment comprises a tracrRNA set out in Supplementary Table S5 or wherein the protein-binding segment comprises a tracrRNA at least 80% identical over at least 20 nucleotides to a tracrRNA set out in Supplementary Table S5. In some embodiments, the protein-binding segment comprises a CRISPR repeat set out in Supplementary Table S5 that is the CRISPR repeat cognate to the tracrRNA of the protein-binding segment. In some

embodiments, the DNA-targeting segment comprises RNA complementary to a protospacer-like sequence in a target DNA 5' to a PAM sequence. In some embodiments, the tracrRNA and CRISPR repeat are respectively the *C. jejuni* tracrRNA and its cognate CRISPR repeat set out in Supplementary Table S5 and the PAM sequence is NNNNACA. In some embodiments, the tracrRNA and CRISPR repeat are respectively at least 80% identical to the *C. jejuni* tracrRNA and its cognate CRISPR repeat set out in Supplementary Table S5 and the PAM sequence is NNNNACA. In some embodiments, the single-molecule guide RNA comprises a sequence that hybridizes to a protospacer-like sequence set out in one of SEQ ID NOs: 801-2701.

**[00196]** In some embodiments, the double-molecule guide RNAs comprise a targeter-RNA and an activator-RNA complementary thereto, wherein the activator-RNA comprises a tracrRNA set out in Supplementary Table S5 or wherein the activator-RNA comprises a tracrRNA at least 80% identical over at least 20 nucleotides to a tracrRNA set out in Supplementary Table S5. In some embodiments, the double-molecule guide RNA comprises a modified backbone, a non-natural internucleoside linkage, a nucleic acid mimetic, a modified sugar moiety, a base modification, a modification or sequence that provides for modified or regulated stability, a modification or sequence that provides for subcellular tracking, a modification or sequence that provides for tracking, or a modification or sequence that provides for a binding site for a protein or protein complex. In some embodiments, the targeter-RNA comprises a CRISPR repeat set out in Supplementary Table S5. In some embodiments, the targeter-RNA comprises a CRISPR repeat set out in Supplementary Table S5 that is the cognate CRISPR repeat of the tracrRNA of the activator-RNA. In some embodiments, the targeter-RNA further comprises RNA complementary to a protospacer-like sequence in a target DNA 5' to a PAM sequence. In some embodiments, the tracrRNA and CRISPR repeat are respectively the *C. jejuni* tracrRNA and its cognate CRISPR repeat set out in Supplementary Table S5 and the PAM sequence is NNNNACA. In some embodiments, the tracrRNA and CRISPR repeat are at least 80% identical to respectively the *C. jejuni* tracrRNA and its cognate CRISPR repeat set out in Supplementary Table S5 and the PAM sequence is NNNNACA. In some embodiments, the double-molecule guide RNA comprises a sequence that hybridizes to a protospacer-like sequence set out in one of SEQ ID NOs: 801-2701.

**[00197]** Nucleic acids encoding a guide RNA and/or a site-directed modifying polypeptide

**[00198]** The present disclosure provides a nucleic acid comprising a nucleotide sequence encoding a guide RNA and/or a site-directed modifying polypeptide. In some embodiments, a guide RNA-encoding nucleic acid is an expression vector, e.g., a recombinant expression vector.

**[00199]** In some embodiments, a method involves contacting a target DNA or introducing into a cell (or a population of cells) one or more nucleic acids comprising nucleotide sequences encoding a guide RNA and/or a site-directed modifying polypeptide. In some embodiments a cell comprising a target DNA is *in vitro*. In some embodiments a cell comprising a target DNA is *in vivo*. Suitable nucleic acids comprising nucleotide sequences encoding a guide RNA and/or a site-directed modifying polypeptide include

expression vectors, where an expression vector comprising a nucleotide sequence encoding a guide RNA and/or a site-directed modifying polypeptide is a "recombinant expression vector."

**[00200]** In some embodiments, the recombinant expression vector is a viral construct, e.g., a recombinant adeno-associated virus construct (see, e.g., U.S. Patent No. 7,078,387), a recombinant adenoviral construct, a recombinant lentiviral construct, a recombinant retroviral construct, etc.

**[00201]** Suitable expression vectors include, but are not limited to, viral vectors (e.g. viral vectors based on vaccinia virus; poliovirus; adenovirus (see, e.g., Li et al., *Invest Ophthalmol Vis Sci* 35:2543-2549, 1994; Borrás et al., *Gene Ther* 6:515-524, 1999; Li and Davidson, *PNAS* 92:7700-7704, 1995; Sakamoto et al., *H Gene Ther* 5:1088-1097, 1999; WO 94/12649, WO 93/03769; WO 93/19191; WO 94/28938; WO 95/11984 and WO 95/00655); adeno-associated virus (see, e.g., Ali et al., *Hum Gene Ther* 9:81-86, 1998, Flannery et al., *PNAS* 94:6916-6921, 1997; Bennett et al., *Invest Ophthalmol Vis Sci* 38:2857-2863, 1997; Jomary et al., *Gene Ther* 4:683-690, 1997, Rolling et al., *Hum Gene Ther* 10:641-648, 1999; Ali et al., *Hum Mol Genet* 5:591-594, 1996; Srivastava in WO 93/09239, Samulski et al., *J. Vir.* (1989) 63:3822-3828; Mendelson et al., *Virology* (1988) 166:154-165; and Flotte et al., *PNAS* (1993) 90:10613-10617); SV40; herpes simplex virus; human immunodeficiency virus (see, e.g., Miyoshi et al., *PNAS* 94:10319-23, 1997; Takahashi et al., *J Virol* 73:7812-7816, 1999); a retroviral vector (e.g., Murine Leukemia Virus, spleen necrosis virus, and vectors derived from retroviruses such as Rous Sarcoma Virus, Harvey Sarcoma Virus, avian leukosis virus, a lentivirus, human immunodeficiency virus, myeloproliferative sarcoma virus, and mammary tumor virus); and the like.

**[00202]** Numerous suitable expression vectors are known to those of skill in the art, and many are commercially available. The following vectors are provided by way of example; for eukaryotic host cells: pXT1, pSG5 (Stratagene), pSVK3, pBPV, pMSG, and pSVLSV40 (Pharmacia). However, any other vector may be used so long as it is compatible with the host cell. Depending on the host/vector system utilized, any of a number of suitable transcription and translation control elements, including constitutive and inducible promoters, transcription enhancer elements, transcription terminators, etc. may be used in the expression vector (see e.g., Bitter et al. (1987) *Methods in Enzymology*, 153:516-544).

**[00203]** In some embodiments, a nucleotide sequence encoding a guide RNA and/or a site-directed modifying polypeptide is operably linked to a control element, e.g., a transcriptional control element, such as a promoter. The transcriptional control element may be functional in either a eukaryotic cell, e.g., a mammalian cell; or a prokaryotic cell (e.g., bacterial or archaeal cell). In some embodiments, a nucleotide sequence encoding a guide RNA and/or a site-directed modifying polypeptide is operably linked to multiple control elements that allow expression of the nucleotide sequence encoding a guide RNA and/or a site-directed modifying polypeptide in both prokaryotic and eukaryotic cells.

**[00204]** Non-limiting examples of suitable eukaryotic promoters (promoters functional in a eukaryotic cell) include those from cytomegalovirus (CMV) immediate early, herpes simplex virus (HSV) thymidine kinase, early and late SV40, long terminal repeats (LTRs) from retrovirus, and mouse metallothionein-I.



Selection of the appropriate vector and promoter is well within the level of ordinary skill in the art. The expression vector may also contain a ribosome binding site for translation initiation and a transcription terminator. The expression vector may also include appropriate sequences for amplifying expression. The expression vector may also include nucleotide sequences encoding protein tags (e.g., 6xHis tag, hemagglutinin tag, green fluorescent protein, etc.) that are fused to the site-directed modifying polypeptide, thus resulting in a chimeric polypeptide.

**[00205]** In some embodiments, a nucleotide sequence encoding a guide RNA and/or a site-directed modifying polypeptide is operably linked to an inducible promoter. In some embodiments, a nucleotide sequence encoding a guide RNA and/or a site-directed modifying polypeptide is operably linked to a constitutive promoter.

**[00206]** Methods of introducing a nucleic acid into a host cell are known in the art, and any known method can be used to introduce a nucleic acid (e.g., an expression construct) into a cell. Suitable methods include e.g., viral or bacteriophage infection, transfection, conjugation, protoplast fusion, lipofection, electroporation, calcium phosphate precipitation, polyethyleneimine (PEI)-mediated transfection, DEAE-dextran mediated transfection, liposome-mediated transfection, particle gun technology, calcium phosphate precipitation, direct micro injection, nanoparticle-mediated nucleic acid delivery (see, e.g., Panyam et., al *Adv Drug Deliv Rev.* 2012 Sep 13. pii: S0169-409X(12)00283-9. doi: 10.1016/j.addr.2012.09.023 ), and the like.

**[00207]** Chimeric Polypeptides

**[00208]** The present disclosure provides a chimeric site-directed modifying polypeptide. A chimeric site-directed modifying polypeptide interacts with (e.g., binds to) a guide RNA (described above). The guide RNA guides the chimeric site-directed modifying polypeptide to a target sequence within target DNA (e.g. a chromosomal sequence or an extrachromosomal sequence, e.g. an episomal sequence, a minicircle sequence, a mitochondrial sequence, a chloroplast sequence, etc.). A chimeric site-directed modifying polypeptide modifies target DNA (e.g., cleavage or methylation of target DNA) and/or a polypeptide associated with target DNA (e.g., methylation or acetylation of a histone tail).

**[00209]** A chimeric site-directed modifying polypeptide modifies target DNA (e.g., cleavage or methylation of target DNA) and/or a polypeptide associated with target DNA (e.g., methylation or acetylation of a histone tail). A chimeric site-directed modifying polypeptide is also referred to herein as a "chimeric site-directed polypeptide" or a "chimeric RNA binding site-directed modifying polypeptide."

**[00210]** A chimeric site-directed modifying polypeptide comprises two portions, an RNA-binding portion and an activity portion. A chimeric site-directed modifying polypeptide comprises amino acid sequences that are derived from at least two different polypeptides. A chimeric site-directed modifying polypeptide can comprise modified and/or naturally-occurring polypeptide sequences (e.g., a first amino

acid sequence from a modified or unmodified Cas9 protein; and a second amino acid sequence other than the Cas9 protein).

**[00211]** RNA-binding portion

**[00212]** In some cases, the RNA-binding portion of a chimeric site-directed modifying polypeptide is a naturally-occurring polypeptide. In other cases, the RNA-binding portion of a chimeric site-directed modifying polypeptide is not a naturally-occurring molecule (modified, e.g., mutation, deletion, insertion). Naturally-occurring RNA-binding portions of interest are derived from site-directed modifying polypeptides known in the art. For example, SEQ ID NOs: 1-800 provide a non-limiting set of naturally occurring Cas9 endonucleases that can be used as site-directed modifying polypeptides. In some cases, the RNA-binding portion of a chimeric site-directed modifying polypeptide comprises an amino acid sequence having at least about 75%, at least about 80%, at least about 85%, at least about 90%, at least about 95%, at least about 98%, at least about 99%, or 100%, amino acid sequence identity to the RNA-binding portion of a polypeptide set forth in SEQ ID NOs: 1-800.

**[00213]** In some cases, the site-directed modifying polypeptide comprises an amino acid sequence having at least about 75%, at least about 80%, at least about 85%, at least about 90%, at least about 95%, at least about 99%, or 100%, amino acid sequence identity to amino acids 7-166 and/or 731-1003 of SEQ ID NO: 8, or to the corresponding portions in any of the amino acid sequences set forth as SEQ ID NOs: 1-800.

**[00214]** Activity portion

**[00215]** In addition to the RNA-binding portion, the chimeric site-directed modifying polypeptide comprises an "activity portion." In some embodiments, the activity portion of a chimeric site-directed modifying polypeptide comprises the naturally-occurring activity portion of a site-directed modifying polypeptide (e.g., Cas9 endonuclease). In other embodiments, the activity portion of a subject chimeric site-directed modifying polypeptide comprises a modified amino acid sequence (e.g., substitution, deletion, insertion) of a naturally-occurring activity portion of a site-directed modifying polypeptide. Naturally-occurring activity portions of interest are derived from site-directed modifying polypeptides known in the art. For example, SEQ ID NOs: 1-800 are a non-limiting set of naturally occurring Cas9 endonucleases that can be used as site-directed modifying polypeptides. The activity portion of a chimeric site-directed modifying polypeptide is variable and may comprise any heterologous polypeptide sequence that may be useful in the methods disclosed herein. In some embodiments, the activity portion of a site-directed modifying polypeptide comprises a portion of a Cas9 ortholog (including, but not limited to, the Cas9 orthologs set out in one of SEQ ID NOs: 1-800) that is at least 90% identical to amino acids 7-166 of SEQ ID NO: 8 and/or at least 90% identical to amino acids 731-1003 of SEQ ID NO: 8. In some embodiments, a chimeric site-directed modifying polypeptide comprises: (i) an RNA-binding portion that interacts with a guide RNA, wherein the guide RNA comprises a nucleotide sequence that is

complementary to a sequence in a target DNA; and (ii) an activity portion that exhibits site-directed enzymatic activity (e.g., activity for DNA methylation, activity for DNA cleavage, activity for histone acetylation, activity for histone methylation, etc.), wherein the site of enzymatic activity is determined by the guide RNA.

**[00216]** In other embodiments, a chimeric site-directed modifying polypeptide comprises: (i) an RNA-binding portion that interacts with a guide RNA, wherein the guide RNA comprises a nucleotide sequence that is complementary to a sequence in a target DNA; and (ii) an activity portion that modulates transcription within the target DNA (e.g., to increase or decrease transcription), wherein the site of modulated transcription within the target DNA is determined by the guide RNA.

**[00217]** In some cases, the activity portion of a chimeric site-directed modifying polypeptide has enzymatic activity that modifies target DNA (e.g., nuclease activity, methyltransferase activity, demethylase activity, DNA repair activity, DNA damage activity, deamination activity, dismutase activity, alkylation activity, depurination activity, oxidation activity, pyrimidine dimer forming activity, integrase activity, transposase activity, recombinase activity, polymerase activity, ligase activity, helicase activity, photolyase activity or glycosylase activity).

**[00218]** In other cases, the activity portion of a chimeric site-directed modifying polypeptide has enzymatic activity (e.g., methyltransferase activity, demethylase activity, acetyltransferase activity, deacetylase activity, kinase activity, phosphatase activity, ubiquitin ligase activity, deubiquitinating activity, adenylation activity, deadenylation activity, SUMOylating activity, deSUMOylating activity, ribosylation activity, deribosylation activity, myristoylation activity or demyristoylation activity) that modifies a polypeptide associated with target DNA (e.g., a histone).

**[00219]** In some cases, the activity portion of a chimeric site-directed modifying polypeptide exhibits enzymatic activity (described above). In other cases, the activity portion of a chimeric site-directed modifying polypeptide modulates transcription of the target DNA (described above). The activity portion of a chimeric site-directed modifying polypeptide is variable and may comprise any heterologous polypeptide sequence that may be useful in the methods disclosed herein.

**[00220]** Exemplary chimeric site-directed modifying polypeptides

**[00221]** In some embodiments, the activity portion of the chimeric site-directed modifying polypeptide comprises a modified form of the Cas9 protein, including modified forms of any of the Cas9 orthologs described herein, such as SEQ ID NOs: 1-800). In some instances, the modified form of the Cas9 protein comprises an amino acid change (e.g., deletion, insertion, or substitution) that reduces the naturally-occurring nuclease activity of the Cas9 protein. For example, in some instances, the modified form of the Cas9 protein has less than 50%, less than 40%, less than 30%, less than 20%, less than 10%, less than

5%, or less than 1% of the nuclease activity of the corresponding wild-type Cas9 polypeptide. In some cases, the modified form of the Cas9 polypeptide has no substantial nuclease activity.

**[00222]** In some embodiments, the modified form of the Cas9 polypeptide is a D10A (aspartate to alanine at amino acid position 10 of SEQ ID NO:8) mutation (or the corresponding mutation of any of the proteins presented in SEQ ID NOs: 1-800) that can cleave the complementary strand of the target DNA but has reduced ability to cleave the non-complementary strand of the target DNA. In some embodiments, the modified form of the SEQ ID NO: 8 Cas9 polypeptide is a H840A (histidine to alanine at amino acid position 840) mutation (or the corresponding mutation of any of the proteins set forth as SEQ ID NOs: 1-800) that can cleave the non-complementary strand of the target DNA but has reduced ability to cleave the complementary strand of the target DNA. In some embodiments, the modified form of the SEQ ID NO: 8 Cas9 polypeptide harbors both the D10A and the H840A mutations (or the corresponding mutations of any of the proteins set forth as SEQ ID NOs: 1-800) such that the polypeptide has a reduced ability to cleave both the complementary and the non-complementary strands of the target DNA. Other residues can be mutated to achieve the above effects (i.e. inactivate one or the other nuclease portions). As non-limiting examples, *S. pyogenes* Cas9 residues D10, G12, G17, E762, H840, N863, H982, H983, A984, D986, and/or A987 of SEQ ID NO: 8 (or the corresponding mutations of any of the proteins set forth as SEQ ID NOs: 1-800) can be altered (i.e., substituted). Also, mutations other than alanine substitutions are contemplated.

**[00223]** In some some embodiments, a modified Cas9 endonuclease comprises one or more mutations corresponding to *S. pyogenes* Cas9 mutation E762A, HH983AA or D986A in SEQ ID NO: 8. In some embodiments, the modified Cas 9 endonuclease further comprises one or more mutations corresponding to *S. pyogenes* Cas9 mutation D10A, H840A, G12A, G17A, N854A, N863A, N982A or A984A in SEQ ID NO: 8. For example, the modified Cas9 endonuclease may comprise a variant at least about 75% identical to any of SEQ ID NOs: 1-800 that comprises one or more mutations corresponding to a mutation E762A, HH983AA or D986A in SEQ ID NO: 8; and/or one or more mutations corresponding to a mutation D10A, H840A, G12A, G17A, N854A, N863A, N982A or A984A in SEQ ID NO: 8. In some embodiments, such a variant comprises a region at least about 75%, at least about 80%, at least about 85%, at least about 90%, at least about 95%, at least about 99% or 100% amino acid sequence identity to the regions corresponding to amino acids 7-166 and/or 731-1003 of SEQ ID NO: 8.

**[00224]** **Table 1.** Table 1 lists four motifs that are present in Cas9 sequences from various species. The amino acids listed here are from the Cas9 from *S. pyogenes* (SEQ ID NO:8).

	Motif	Amino acids (residue #s)	Highly conserved
	RuvC-like I	IGLDIGTNSVGVAVI (7-21)	D10, G12, G17
	RuvC-like II	IVIAMARE (759-766)	E762

	HNH-motif	DVDHIVPQSFLKDDSIDNKVLTRSDKN (837-863)	H840, N854, N863
	RuvC-like II	HHAHDAYL (982-989)	H982, H983, A984, D986, A987

**[00225]** In some cases, the chimeric site-directed modifying polypeptide comprises an amino acid sequence having at least about 75%, at least about 80%, at least about 85%, at least about 90%, at least about 95%, at least about 99% or 100% amino acid sequence identity to amino acids 7-166 and/or 731-1003 of SEQ ID NO: 8, or to the corresponding portions in any of the amino acid sequences set forth as SEQ ID NOs: 1-800. In some cases, the chimeric site-directed modifying polypeptide comprises 4 motifs (as listed in Table 1), each with amino acid sequences having at least about 75%, at least about 80%, at least about 85%, at least about 90%, at least about 95%, at least about 99% or 100% amino acid sequence identity to each of the 4 motifs listed in Table 1, or to the corresponding portions in any of the amino acid sequences set forth as SEQ ID NOs: 1-800. In some cases, the chimeric site-directed modifying polypeptide comprises amino acid sequences having at least about 75%, at least about 80%, at least about 85%, at least about 90%, at least about 95%, at least about 99% or 100% amino acid sequence identity to amino acids 7-166 and/or 731-1003 of SEQ ID NO: 8, or to the corresponding portions in any of the amino acid sequences set forth as SEQ ID NOs: 1-800.

**[00226]** In some embodiments, the activity portion of the site-directed modifying polypeptide comprises a heterologous polypeptide that has DNA-modifying activity and/or transcription factor activity and/or DNA-associated polypeptide-modifying activity. In some cases, a heterologous polypeptide replaces a portion of the Cas9 polypeptide that provides nuclease activity. In other embodiments, a site-directed modifying polypeptide comprises both a portion of the Cas9 polypeptide that normally provides nuclease activity (and that portion can be fully active or can instead be modified to have less than 100% of the corresponding wild-type activity) and a heterologous polypeptide. In other words, in some cases, a chimeric site-directed modifying polypeptide is a fusion polypeptide comprising both the portion of the Cas9 polypeptide that normally provides nuclease activity and the heterologous polypeptide. In other cases, a chimeric site-directed modifying polypeptide is a fusion polypeptide comprising a modified variant of the activity portion of the Cas9 polypeptide (e.g., amino acid change, deletion, insertion) and a heterologous polypeptide. In yet other cases, a chimeric site-directed modifying polypeptide is a fusion polypeptide comprising a heterologous polypeptide and the RNA-binding portion of a naturally-occurring or a modified site-directed modifying polypeptide.

**[00227]** For example, in a chimeric Cas9 protein, a naturally-occurring (or modified, e.g., mutation, deletion, insertion) bacterial Cas9 polypeptide may be fused to a heterologous polypeptide sequence (i.e. a polypeptide sequence from a protein other than Cas9 or a polypeptide sequence from another organism).

The heterologous polypeptide sequence may exhibit an activity (e.g., enzymatic activity) that will also be exhibited by the chimeric Cas9 protein (e.g., methyltransferase activity, acetyltransferase activity, kinase activity, ubiquitinating activity, etc.). A heterologous nucleic acid sequence may be linked to another nucleic acid sequence (e.g., by genetic engineering) to generate a chimeric nucleotide sequence encoding a chimeric polypeptide. In some embodiments, a chimeric Cas9 polypeptide is generated by fusing a Cas9 polypeptide (e.g., wild type Cas9 or a Cas9 variant, e.g., a Cas9 with reduced or inactivated nuclease activity) with a heterologous sequence that provides for subcellular localization (e.g., a nuclear localization signal (NLS) for targeting to the nucleus; a mitochondrial localization signal for targeting to the mitochondria; a chloroplast localization signal for targeting to a chloroplast; an ER retention signal; and the like). In some embodiments, the heterologous sequence can provide a tag for ease of tracking or purification (e.g., a fluorescent protein, e.g., green fluorescent protein (GFP), YFP, RFP, CFP, mCherry, tdTomato, and the like; a HIS tag, e.g., a 6XHis tag; a hemagglutinin (HA) tag; a FLAG tag; a Myc tag; and the like). In some embodiments, the heterologous sequence can provide for increased or decreased stability. In some embodiments, the heterologous sequence can provide a binding domain (e.g., to provide the ability of a chimeric Cas9 polypeptide to bind to another protein of interest, e.g., a DNA or histone modifying protein, a transcription factor or transcription repressor, a recruiting protein, etc.).

**[00228]** Nucleic acid encoding a chimeric site-directed modifying polypeptide

**[00229]** The present disclosure provides a nucleic acid comprising a nucleotide sequence encoding a chimeric site-directed modifying polypeptide. In some embodiments, the nucleic acid comprising a nucleotide sequence encoding a chimeric site-directed modifying polypeptide is an expression vector, e.g., a recombinant expression vector.

**[00230]** In some embodiments, a method involves contacting a target DNA or introducing into a cell (or a population of cells) one or more nucleic acids comprising a chimeric site-directed modifying polypeptide. Suitable nucleic acids comprising nucleotide sequences encoding a chimeric site-directed modifying polypeptide include expression vectors, where an expression vector comprising a nucleotide sequence encoding a chimeric site-directed modifying polypeptide is a "recombinant expression vector."

**[00231]** In some embodiments, the recombinant expression vector is a viral construct, e.g., a recombinant adeno-associated virus construct (see, e.g., U.S. Patent No. 7,078,387), a recombinant adenoviral construct, a recombinant lentiviral construct, etc.

**[00232]** Suitable expression vectors include, but are not limited to, viral vectors (e.g. viral vectors based on vaccinia virus; poliovirus; adenovirus (see, e.g., Li et al., Invest Ophthalmol Vis Sci 35:2543 2549, 1994; Borrás et al., Gene Ther 6:515 524, 1999; Li and Davidson, PNAS 92:7700 7704, 1995; Sakamoto et al., Hum Gene Ther 5:1088 1097, 1999; WO 94/12649, WO 93/03769; WO 93/19191; WO 94/28938; WO 95/11984 and WO 95/00655); adeno-associated virus (see, e.g., Ali et al., Hum Gene Ther 9:81 86, 1998, Flannery et al., PNAS 94:6916 6921, 1997; Bennett et al., Invest Ophthalmol Vis Sci

38:2857-2863, 1997; Jomary et al., *Gene Ther* 4:683-690, 1997; Rolling et al., *Hum Gene Ther* 10:641-648, 1999; Ali et al., *Hum Mol Genet* 5:591-594, 1996; Srivastava in WO 93/09239, Samulski et al., *J. Vir.* (1989) 63:3822-3828; Mendelson et al., *Viol.* (1988) 166:154-165; and Flotte et al., *PNAS* (1993) 90:10613-10617); SV40; herpes simplex virus; human immunodeficiency virus (see, e.g., Miyoshi et al., *PNAS* 94:10319-23, 1997; Takahashi et al., *J Virol* 73:7812-7816, 1999); a retroviral vector (e.g., Murine Leukemia Virus, spleen necrosis virus, and vectors derived from retroviruses such as Rous Sarcoma Virus, Harvey Sarcoma Virus, avian leukosis virus, a lentivirus, human immunodeficiency virus, myeloproliferative sarcoma virus, and mammary tumor virus); and the like.

**[00233]** Numerous suitable expression vectors are known to those of skill in the art, and many are commercially available. The following vectors are provided by way of example; for eukaryotic host cells: pXT1, pSG5 (Stratagene), pSVK3, pBPV, pMSG, and pSVLSV40 (Pharmacia). However, any other vector may be used so long as it is compatible with the host cell.

**[00234]** Depending on the host/vector system utilized, any of a number of suitable transcription and translation control elements, including constitutive and inducible promoters, transcription enhancer elements, transcription terminators, etc. may be used in the expression vector (see e.g., Bitter et al. (1987) *Methods in Enzymology*, 153:516-544).

**[00235]** In some embodiments, a nucleotide sequence encoding a chimeric site-directed modifying polypeptide is operably linked to a control element, e.g., a transcriptional control element, such as a promoter. The transcriptional control element may be functional in either a eukaryotic cell, e.g., a mammalian cell; or a prokaryotic cell (e.g., bacterial or archaeal cell). In some embodiments, a nucleotide sequence encoding a chimeric site-directed modifying polypeptide is operably linked to multiple control elements that allow expression of the nucleotide sequence encoding a chimeric site-directed modifying polypeptide in both prokaryotic and eukaryotic cells.

**[00236]** Non-limiting examples of suitable eukaryotic promoters (promoters functional in a eukaryotic cell) include those from cytomegalovirus (CMV) immediate early, herpes simplex virus (HSV) thymidine kinase, early and late SV40, long terminal repeats (LTRs) from retrovirus, and mouse metallothionein-I. Selection of the appropriate vector and promoter is well within the level of ordinary skill in the art. The expression vector may also contain a ribosome binding site for translation initiation and a transcription terminator. The expression vector may also include appropriate sequences for amplifying expression. The expression vector may also include nucleotide sequences encoding protein tags (e.g., 6xHis tag, hemagglutinin (HA) tag, a fluorescent protein (e.g., a green fluorescent protein; a yellow fluorescent protein, etc.), etc.) that are fused to the chimeric site-directed modifying polypeptide.

**[00237]** In some embodiments, a nucleotide sequence encoding a chimeric site-directed modifying polypeptide is operably linked to an inducible promoter (e.g., heat shock promoter, Tetracycline-regulated promoter, Steroid-regulated promoter, Metal-regulated promoter, estrogen receptor-regulated promoter,

etc.). In some embodiments, a nucleotide sequence encoding a chimeric site-directed modifying polypeptide is operably linked to a spatially restricted and/or temporally restricted promoter (e.g., a tissue specific promoter, a cell type specific promoter, etc.). In some embodiments, a nucleotide sequence encoding a chimeric site-directed modifying polypeptide is operably linked to a constitutive promoter.

**[00238]** Methods of introducing a nucleic acid into a host cell are known in the art, and any known method can be used to introduce a nucleic acid (e.g., an expression construct) into a stem cell or progenitor cell. Suitable methods include, include e.g., viral or bacteriophage infection, transfection, conjugation, protoplast fusion, lipofection, electroporation, calcium phosphate precipitation, polyethyleneimine (PEI)-mediated transfection, DEAE-dextran mediated transfection, liposome-mediated transfection, particle gun technology, calcium phosphate precipitation, direct micro injection, nanoparticle-mediated nucleic acid delivery (see, e.g., Panyam et., al *Adv Drug Deliv Rev.* 2012 Sep 13. pii: 50169-409X(12)00283-9. doi: 10.1016/j.addr.2012.09.023 ), and the like.

**[00239]** Methods

**[00240]** The present disclosure provides methods for modifying a target DNA and/or a target DNA-associated polypeptide. Generally, a method involves contacting a target DNA with a complex (a "targeting complex"), which complex comprises a guide RNA and a site-directed modifying polypeptide.

**[00241]** As discussed above, a guide RNA and a site-directed modifying polypeptide form a complex. The guide RNA provides target specificity to the complex by comprising a nucleotide sequence that is complementary to a sequence of a target DNA. The site-directed modifying polypeptide of the complex provides the site-specific activity. In some embodiments, a complex modifies a target DNA, leading to, for example, DNA cleavage, DNA methylation, DNA damage, DNA repair, etc. In other embodiments, a complex modifies a target polypeptide associated with target DNA (e.g., a histone, a DNA-binding protein, etc.), leading to, for example, histone methylation, histone acetylation, histone ubiquitination, and the like. The target DNA may be, for example, naked DNA *in vitro*, chromosomal DNA in cells *in vitro*, chromosomal DNA in cells *in vivo*, etc.

**[00242]** In some cases, the site-directed modifying polypeptide exhibits nuclease activity that cleaves target DNA at a target DNA sequence defined by the region of complementarity between the guide RNA and the target DNA. In some cases, when the site-directed modifying polypeptide is a Cas9 or Cas9 related polypeptide, site-specific cleavage of the target DNA occurs at locations determined by both (i) base-pairing complementarity between the guide RNA and the target DNA; and (ii) a short motif [referred to as the protospacer adjacent motif (PAM)] in the target DNA. In some embodiments (e.g., when Cas9 from *S. pyogenes* is used), the PAM sequence of the non-complementary strand is 5'-XGG-3', where X is any DNA nucleotide and X is immediately 3' of the target sequence of the non-complementary strand of the target DNA. As such, the PAM sequence of the complementary strand is 5'-CCY-3', where Y is any DNA nucleotide and Y is immediately 5' of the target sequence of the complementary strand of the target



DNA (where the PAM of the non-complementary strand is 5'-GGG-3' and the PAM of the complementary strand is 5'-CCC-3'). In some such embodiments, X and Y can be complementary and the X-Y base pair can be any basepair (e.g., X=C and Y=G; X=G and Y=C; X=A and Y=T, X=T and Y=A).

**[00243]** In some cases, different Cas9 proteins (i.e., Cas9 proteins from various species) may be advantageous to use in the various provided methods in order to capitalize on various enzymatic characteristics of the different Cas9 proteins (e.g., for different PAM sequence preferences; for increased or decreased enzymatic activity; for an increased or decreased level of cellular toxicity; to change the balance between NHEJ, homology-directed repair, single strand breaks, double strand breaks, etc.).

**[00244]** Cas9 proteins from various species (see SEQ ID NOs: 1-800) may require different PAM sequences in the target DNA. Thus, for a particular Cas9 protein of choice, the PAM sequence requirement may be different than the 5'-XGG-3' sequence described above. The present disclosure, for example, provides a *C. jejuni* PAM sequence NNNNACA; *P. multocida* PAM sequences GNNNCNNA or NNNNC; an *F. novicida* PAM sequence NG; an *S. thermophilus*\*\* PAM sequence NNAAAAW; an *L. innocua* PAM sequence NGG; and an *S. dysgalactiae* PAM sequence NGG.

**[00245]** Exemplary methods provided that take advantage of characteristics of Cas9 orthologs include the following.

**[00246]** A method for manipulating DNA in a cell, comprising contacting the DNA with a Cas9 ortholog-guideRNA complex, wherein the complex comprises: (a) a cognate guide RNA for a first Cas9 endonuclease from a cluster in Supplementary Table S2 and (b) a second Cas9 endonuclease from the cluster that is exchangeable with preserved high cleavage efficiency with the first endonuclease and shares at least 80% identity with the first endonuclease over 80% of their length. In some embodiments, the guide is a single-molecule guide RNA. In some embodiments, the guide RNA is a double-molecule guide RNA. In some embodiments, the first Cas9 endonuclease is from *S. pyogenes* and the second Cas9 endonuclease is from *S. mutans*. In some embodiments, the first Cas9 endonuclease is from *S. thermophilus*\* and the second Cas9 endonuclease is from *S. mutans*. In some embodiments, the first Cas9 endonuclease is from *N. meningitidis* and the second Cas9 endonuclease is from *P. multocida*.

**[00247]** A method for manipulating DNA in a cell, comprising contacting the DNA with a Cas9 ortholog-guideRNA complex, wherein the complex comprises: (a) a cognate guide RNA of a first Cas9 endonuclease from a cluster in Supplementary Table S6 and (b) an Cas9 endonuclease from a cluster in Supplementary Table S6 that is exchangeable with lowered cleavage efficiency with the first endonuclease and shares at least 50% amino acid sequence identity with the first endonuclease over 70% of their length. In some embodiments, the guide is a single-molecule guide RNA. In some embodiments, the guide RNA is a double-molecule guide RNA. In some embodiments, the first Cas9 endonuclease is from *C. Jejuni* and the second Cas9 endonuclease is from *P. multocida*. In some

embodiments, the first Cas9 endonuclease is from *N. meningitidis* and the second Cas9 endonuclease is from *P. multocida*.

**[00248]** A method for manipulating DNA in a cell, comprising contacting the DNA with two or more Cas9-guideRNA complexes, wherein each Cas9-guideRNA complex comprises: (a) a Cas9 endonuclease from a different cluster in Supplementary Table S6 exhibiting less than 50% amino acid sequence identity with the other endonucleases of the method over 70% of their length, and (b) a guide RNA specifically complexed with each Cas9 endonuclease. In some embodiments, the guide is a single-molecule guide RNA. In some embodiments, the guide RNA is a double-molecule guide RNA. In some embodiments, the Cas9 endonucleases are from *F. novicida* and *S. pyogenes*. In some embodiments, the Cas9 endonucleases are from *N. meningitidis* and *S. mutans*. In some embodiments, the *S. thermophilus*\* and *S. thermophilus*\*\* Cas9 endonucleases.

**[00249]** Many Cas9 orthologs from a wide variety of species have been identified herein. All identified Cas9 orthologs have the same domain architecture with a central HNH endonuclease domain and a split RuvC/RNaseH domain. Cas9 proteins share four key motifs with a conserved architecture. Motifs 1, 2, and 4 are RuvC like motifs while motif 3 is an HNH-motif. In some cases, a suitable site-directed modifying polypeptide comprises an amino acid sequence having four motifs, each of motifs 1-4 having at least about 75%, at least about 80%, at least about 85%, at least about 90%, at least about 95%, at least about 99% or 100% amino acid sequence identity to the motifs 1-4 of the Cas9 amino acid sequence depicted in Table 1), or to the corresponding portions in any of the amino acid sequences set forth in SEQ ID NOs: 1-800. In some cases, a suitable site-directed modifying polypeptide comprises an amino acid sequence having at least about 75%, at least about 80%, at least about 85%, at least about 90%, at least about 95%, at least about 99% or 100% amino acid sequence identity to amino acids 7-166 and/or 731-1003 of SEQ ID NO: 8, or to the corresponding portions in any of the amino acid sequences set forth as SEQ ID NOs: 1-800.

**[00250]** The nuclease activity cleaves target DNA to produce double strand breaks. These breaks are then repaired by the cell in one of two ways: non-homologous end joining, and homology-directed repair. In non-homologous end joining (NHEJ), the double-strand breaks are repaired by direct ligation of the break ends to one another. As such, no new nucleic acid material is inserted into the site, although some nucleic acid material may be lost, resulting in a deletion. In homology-directed repair, a donor polynucleotide with homology to the cleaved target DNA sequence is used as a template for repair of the cleaved target DNA sequence, resulting in the transfer of genetic information from the donor polynucleotide to the target DNA. As such, new nucleic acid material may be inserted/copied into the site. In some cases, a target DNA is contacted with a donor polynucleotide. In some cases, a donor polynucleotide is introduced into a cell. The modifications of the target DNA due to NHEJ and/or homology-directed repair lead to, for example, gene correction, gene replacement, gene tagging, transgene insertion, nucleotide deletion, gene disruption, gene mutation, sequence replacement, etc.

Accordingly, cleavage of DNA by a site-directed modifying polypeptide may be used to delete nucleic acid material from a target DNA sequence (e.g., to disrupt a gene that makes cells susceptible to infection (e.g. the CCR5 or CXCR4 gene, which makes T cells susceptible to HIV infection), to remove disease-causing trinucleotide repeat sequences in neurons, to create gene knockouts and mutations as disease models in research, etc.) by cleaving the target DNA sequence and allowing the cell to repair the sequence in the absence of an exogenously provided donor polynucleotide. Thus, the methods can be used to knock out a gene (resulting in complete lack of transcription or altered transcription) or to knock in genetic material into a locus of choice in the target DNA.

**[00251]** Alternatively, if a guide RNA and a site-directed modifying polypeptide are coadministered to cells with a donor polynucleotide sequence that includes at least a segment with homology to the target DNA sequence, the subject methods may be used to add, i.e. insert or replace, nucleic acid material to a target DNA sequence (e.g. to "knock in" a nucleic acid that encodes for a protein, an siRNA, an miRNA, etc.), to add a tag (e.g., 6xHis, a fluorescent protein (e.g., a green fluorescent protein; a yellow fluorescent protein, etc.), hemagglutinin (HA), FLAG, etc.), to add a regulatory sequence to a gene (e.g. promoter, polyadenylation signal, internal ribosome entry sequence (IRES), 2A peptide, start codon, stop codon, splice signal, localization signal, etc.), to modify a nucleic acid sequence (e.g., introduce a mutation), and the like. As such, a complex comprising a guide RNA and a site-directed modifying polypeptide is useful in any *in vitro* or *in vivo* application in which it is desirable to modify DNA in a site-specific, i.e. "targeted", way, for example gene knock-out, gene knock-in, gene editing, gene tagging, sequence replacement, etc., as used in, for example, gene therapy, e.g. to treat a disease or as an antiviral, antipathogenic, or anticancer therapeutic, the production of genetically modified organisms in agriculture, the large scale production of proteins by cells for therapeutic, diagnostic, or research purposes, the induction of iPS cells, biological research, the targeting of genes of pathogens for deletion or replacement, etc.

**[00252]** In some embodiments, the site-directed modifying polypeptide comprises a modified form of the Cas9 protein. In some instances, the modified form of the Cas9 protein comprises an amino acid change (e.g., deletion, insertion, or substitution) that reduces the naturally-occurring nuclease activity of the Cas9 protein. For example, in some instances, the modified form of the Cas9 protein has less than 50%, less than 40%, less than 30%, less than 20%, less than 10%, less than 5%, or less than 1% of the nuclease activity of the corresponding wild-type Cas9 polypeptide. In some cases, the modified form of the Cas9 polypeptide has no substantial nuclease activity. When a site-directed modifying polypeptide is a modified form of the Cas9 polypeptide that has no substantial nuclease activity, it can be referred to as "dCas9."

**[00253]** In some embodiments, the modified form of the Cas9 polypeptide is a D10A (aspartate to alanine at amino acid position 10 of SEQ ID NO:8) mutation (or the corresponding mutation of any of the proteins set forth as SEQ ID NOs: 1-800) that can cleave the complementary strand of the target DNA but has reduced ability to cleave the non-complementary strand of the target DNA (thus resulting in a single

strand break (SSB) instead of a DSB). In some embodiments, the modified form of the Cas9 polypeptide is a H840A (histidine to alanine at amino acid position 840 of SEQ ID NO:8) mutation (or the corresponding mutation of any of the proteins set forth as SEQ ID NOs: 1-800) that can cleave the non-complementary strand of the target DNA but has reduced ability to cleave the complementary strand of the target DNA (thus resulting in a single strand break (SSB) instead of a DSB). The use of the D10A or H840A variant of SEQ ID NO: 8 Cas9 (or the corresponding mutations in any of the proteins set forth as SEQ ID NOs: 1-800) can alter the expected biological outcome because the non-homologous end joining (NHEJ) is much more likely to occur when DSBs are present as opposed to SSBs. Thus, in some cases where one wishes to reduce the likelihood of DSB (and therefore reduce the likelihood of NHEJ), a D10A or H840A variant of Cas9 can be used. Other residues can be mutated to achieve the same effect (i.e. inactivate one or the other nuclease portions). As non-limiting examples, SEQ ID NO: 8 *S. pyogenes* Cas9 residues D10, G12, G17, E762, H840, N863, H982, H983, A984, D986, and/or A987 (or the corresponding mutations of any of the proteins set forth as SEQ ID NOs: 1-800) can be altered (i.e., substituted). Also, mutations other than alanine substitutions are contemplated. In some embodiments when a site-directed polypeptide (e.g., site-directed modifying polypeptide) has reduced catalytic activity (e.g., when a SEQ ID NO: 8 Cas9 protein has a D10, G12, G17, E762, H840, N863, H982, H983, A984, D986, and/or a A987 mutation, e.g., D 10A, G12A, G17A, E762A, H840A, N863A, H982A, H983A, A984A, and/or D986A), the polypeptide can still bind to target DNA in a site-specific manner (because it is still guided to a target DNA sequence by a guide RNA) as long as it retains the ability to interact with the guide RNA.

**[00254]** In some embodiments, the modified form of the SEQ ID NO: 8 Cas9 polypeptide harbors both the D10A and the H840A mutations (or the corresponding mutations of any of the proteins set forth as SEQ ID NOs: 1-800) such that the polypeptide has a reduced ability to cleave both the complementary and the non-complementary strands of the target DNA (i.e., the variant can have no substantial nuclease activity). Other residues can be mutated to achieve the same effect (i.e. inactivate one or the other nuclease portions). As non-limiting examples, SEQ ID NO: 8 residues D10, G12, G17, E762, H840, N863, H982, H983, A984, D986, and/or A987 (or the corresponding mutations of any of the proteins set forth as SEQ ID NOs: 1-800) can be altered (i.e., substituted). Also, mutations other than alanine substitutions are contemplated.

**[00255]** In some embodiments, the site-directed modifying polypeptide comprises a heterologous sequence (e.g., a fusion). In some embodiments, a heterologous sequence can provide for subcellular localization of the site-directed modifying polypeptide (e.g., a nuclear localization signal (NLS) for targeting to the nucleus; a mitochondrial localization signal for targeting to the mitochondria; a chloroplast localization signal for targeting to a chloroplast; a ER retention signal; and the like). In some embodiments, a heterologous sequence can provide a tag for ease of tracking or purification (e.g., a fluorescent protein, e.g., green fluorescent protein (GFP), YFP, RFP, CFP, mCherry, tdTomato, and the

like; a his tag, e.g., a 6XHis tag; a hemagglutinin (HA) tag; a FLAG tag; a Myc tag; and the like). In some embodiments, the heterologous sequence can provide for increased or decreased stability.

**[00256]** In some embodiments, a site-directed modifying polypeptide can be codon-optimized. This type of optimization is known in the art and entails the mutation of foreign-derived DNA to mimic the codon preferences of the intended host organism or cell while encoding the same protein. Thus, the codons are changed, but the encoded protein remains unchanged. For example, if the intended target cell was a human cell, a human codon-optimized Cas9 (or variant, e.g., enzymatically inactive variant) would be a suitable site-directed modifying polypeptide. Any suitable site-directed modifying polypeptide (e.g., any Cas9 such as any of the sequences set forth in SEQ ID NOs: 1-800) can be codon optimized. As another non-limiting example, if the intended host cell were a mouse cell, than a mouse codon-optimized Cas9 (or variant, e.g., enzymatically inactive variant) would be a suitable site-directed modifying polypeptide. While codon optimization is not required, it is acceptable and may be preferable in certain cases.

**[00257]** Polyadenylation signals can also be chosen to optimize expression in the intended host.

**[00258]** In some embodiments, a guide RNA and a site-directed modifying polypeptide are used as an inducible system for shutting off gene expression in bacterial cells. In some cases, nucleic acids encoding an appropriate guide RNA and/or an appropriate site-directed polypeptide are incorporated into the chromosome of a target cell and are under control of an inducible promoter. When the guide RNA and/or the site-directed polypeptide are induced, the target DNA is cleaved (or otherwise modified) at the location of interest (e.g., a target gene on a separate plasmid), when both the guide RNA and the site-directed modifying polypeptide are present and form a complex. As such, in some cases, bacterial expression strains are engineered to include nucleic acid sequences encoding an appropriate site-directed modifying polypeptide in the bacterial genome and/or an appropriate guide RNA on a plasmid (e.g., under control of an inducible promoter), allowing experiments in which the expression of any targeted gene (expressed from a separate plasmid introduced into the strain) could be controlled by inducing expression of the guide RNA and the site-directed polypeptide.

**[00259]** In some cases, the site-directed modifying polypeptide has enzymatic activity that modifies target DNA in ways other than introducing double strand breaks. Enzymatic activity of interest that may be used to modify target DNA (e.g., by fusing a heterologous polypeptide with enzymatic activity to a site-directed modifying polypeptide, thereby generating a chimeric site-directed modifying polypeptide) includes, but is not limited methyltransferase activity, demethylase activity, DNA repair activity, DNA damage activity, deamination activity, dismutase activity, alkylation activity, depurination activity, oxidation activity, pyrimidine dimer forming activity, integrase activity, transposase activity, recombinase activity, polymerase activity, ligase activity, helicase activity, photolyase activity or glycosylase activity). Methylation and demethylation is recognized in the art as an important mode of epigenetic gene

regulation while DNA damage and repair activity is essential for cell survival and for proper genome maintenance in response to environmental stresses.

**[00260]** As such, the methods herein find use in the epigenetic modification of target DNA and may be employed to control epigenetic modification of target DNA at any location in a target DNA by genetically engineering the desired complementary nucleic acid sequence into the DNA-targeting segment of a guide RNA. The methods herein also find use in the intentional and controlled damage of DNA at any desired location within the target DNA. The methods herein also find use in the sequence-specific and controlled repair of DNA at any desired location within the target DNA. Methods to target DNA-modifying enzymatic activities to specific locations in target DNA find use in both research and clinical applications.

**[00261]** In some cases, the site-directed modifying polypeptide has activity that modulates the transcription of target DNA (e.g., in the case of a chimeric site-directed modifying polypeptide, etc.). In some cases, a chimeric site-directed modifying polypeptides comprising a heterologous polypeptide that exhibits the ability to increase or decrease transcription (e.g., transcriptional activator or transcription repressor polypeptides) is used to increase or decrease the transcription of target DNA at a specific location in a target DNA, which is guided by the DNA-targeting segment of the guide RNA. Examples of source polypeptides for providing a chimeric site-directed modifying polypeptide with transcription modulatory activity include, but are not limited to light-inducible transcription regulators, small molecule/drug-responsive transcription regulators, transcription factors, transcription repressors, etc. In some cases, the method is used to control the expression of a targeted coding-RNA (protein-encoding gene) and/or a targeted non-coding RNA (e.g., tRNA, rRNA, snoRNA, siRNA, miRNA, long ncRNA, etc.). In some cases, the site-directed modifying polypeptide has enzymatic activity that modifies a polypeptide associated with DNA (e.g. histone). In some embodiments, the enzymatic activity is methyltransferase activity, demethylase activity, acetyltransferase activity, deacetylase activity, kinase activity, phosphatase activity, ubiquitin ligase activity (i.e., ubiquitination activity), deubiquitinating activity, adenylation activity, deadenylation activity, SUMOylating activity, deSUMOylating activity, ribosylation activity, deribosylation activity, myristoylation activity, demyristoylation activity glycosylation activity (e.g., from GlcNAc transferase) or deglycosylation activity. The enzymatic activities listed herein catalyze covalent modifications to proteins. Such modifications are known in the art to alter the stability or activity of the target protein (e.g., phosphorylation due to kinase activity can stimulate or silence protein activity depending on the target protein). Of particular interest as protein targets are histones. Histone proteins are known in the art to bind DNA and form complexes known as nucleosomes. Histones can be modified (e.g., by methylation, acetylation, ubiquitination, phosphorylation) to elicit structural changes in the surrounding DNA, thus controlling the accessibility of potentially large portions of DNA to interacting factors such as transcription factors, polymerases and the like. A single histone can be modified in many different ways and in many different combinations (e.g., trimethylation of lysine 27 of histone 3, H3K27, is associated with DNA regions of repressed transcription while trimethylation of lysine 4 of histone 3, H3K4, is associated with DNA regions of active transcription). Thus, a site-directed modifying polypeptide with

histone-modifying activity finds use in the site specific control of DNA structure and can be used to alter the histone modification pattern in a selected region of target DNA. Such methods find use in both research and clinical applications.

**[00262]** In some embodiments, multiple guide RNAs are used simultaneously to simultaneously modify different locations on the same target DNA or on different target DNAs. In some embodiments, two or more guide RNAs target the same gene or transcript or locus. In some embodiments, two or more guide RNAs target different unrelated loci. In some embodiments, two or more guide RNAs target different, but related loci.

**[00263]** In some cases, the site-directed modifying polypeptide is provided directly as a protein. As one non-limiting example, fungi (e.g., yeast) can be transformed with exogenous protein and/or nucleic acid using spheroplast transformation (see Kawai et al., *Bioeng Bugs*. 2010 Nov-Dec;1(6):395-403 : "Transformation of *Saccharomyces cerevisiae* and other fungi: methods and possible underlying mechanism"; and Tanka et al., *Nature*. 2004 Mar 18;428(6980):323-8: "Conformational variations in an infectious protein determine prion strain differences"; both of which are herein incorporated by reference in their entirety). Thus, a site-directed modifying polypeptide (e.g., Cas9) can be incorporated into a spheroplast (with or without nucleic acid encoding a guide RNA and with or without a donor polynucleotide) and the spheroplast can be used to introduce the content into a yeast cell. A site-directed modifying polypeptide can be introduced into a cell (provided to the cell) by any convenient method; such methods are known to those of ordinary skill in the art. As another non-limiting example, a site-directed modifying polypeptide can be injected directly into a cell (e.g., with or without nucleic acid encoding a guide RNA and with or without a donor polynucleotide), e.g., a cell of a zebrafish embryo, the pronucleus of a fertilized mouse oocyte, etc.

**[00264]** Target cells of interest

**[00265]** In some of the above applications, the methods may be employed to induce DNA cleavage, DNA modification, and/or transcriptional modulation in mitotic or post-mitotic cells *in vivo* and/or *ex vivo* and/or *in vitro* (e.g., to produce genetically modified cells that can be reintroduced into an individual). Because the guide RNA provide specificity by hybridizing to target DNA, a mitotic and/or post-mitotic cell of interest in the disclosed methods may include a cell from any organism (e.g. a bacterial cell, an archaeal cell, a cell of a single-cell eukaryotic organism, a plant cell, an algal cell, e.g., *Botryococcus braunii*, *Chlamydomonas reinhardtii*, *Nannochloropsis gaditana*, *Chlorella pyrenoidosa*, *Sargassum patens* C. *Agardh*, and the like, a fungal cell (e.g., a yeast cell), an animal cell, a cell from an invertebrate animal (e.g. fruit fly, cnidarian, echinoderm, nematode, etc.), a cell from a vertebrate animal (e.g., fish, amphibian, reptile, bird, mammal), a cell from a mammal, a cell from a rodent, a cell from a primate, a cell from a human, etc.).

**[00266]** Any type of cell may be of interest (e.g. a stem cell, e.g. an embryonic stem (ES) cell, an induced pluripotent stem (iPS) cell, a germ cell; a somatic cell, e.g. a fibroblast, a hematopoietic cell, a neuron, a muscle cell, a bone cell, a hepatocyte, a pancreatic cell; an in vitro or in vivo embryonic cell of an embryo at any stage, e.g., a 1-cell, 2-cell, 4-cell, 8-cell, etc. stage zebrafish embryo; etc.). Cells may be from established cell lines or they may be primary cells, where "primary cells", "primary cell lines", and "primary cultures" are used interchangeably herein to refer to cells and cells cultures that have been derived from a and allowed to grow in vitro for a limited number of passages, i.e. splittings, of the culture. For example, primary cultures are cultures that may have been passaged 0 times, 1 time, 2 times, 4 times, 5 times, 10 times, or 15 times, but not enough times go through the crisis stage. Typically, the primary cell lines of the present invention are maintained for fewer than 10 passages in vitro. Target cells are in many embodiments unicellular organisms, or are grown in culture.

**[00267]** If the cells are primary cells, they may be harvest from an individual by any convenient method. For example, leukocytes may be conveniently harvested by apheresis, leukocytapheresis, density gradient separation, etc., while cells from tissues such as skin, muscle, bone marrow, spleen, liver, pancreas, lung, intestine, stomach, etc. are most conveniently harvested by biopsy. An appropriate solution may be used for dispersion or suspension of the harvested cells. Such solution will generally be a balanced salt solution, e.g. normal saline, phosphate-buffered saline (PBS), Hank's balanced salt solution, etc., conveniently supplemented with fetal calf serum or other naturally occurring factors, in conjunction with an acceptable buffer at low concentration, generally from 5-25 mM. Convenient buffers include HEPES, phosphate buffers, lactate buffers, etc. The cells may be used immediately, or they may be stored, frozen, for long periods of time, being thawed and capable of being reused. In such cases, the cells will usually be frozen in 10% DMSO, 50% serum, 40% buffered medium, or some other such solution as is commonly used in the art to preserve cells at such freezing temperatures, and thawed in a manner as commonly known in the art for thawing frozen cultured cells.

**[00268]** Nucleic acids encoding a guide RNA and/or a site-directed modifying polypeptide

**[00269]** In some embodiments, a method involves contacting a target DNA or introducing into a cell (or a population of cells) one or more nucleic acids comprising nucleotide sequences encoding a guide RNA and/or a site-directed modifying polypeptide and/or a donor polynucleotide. Suitable nucleic acids comprising nucleotide sequences encoding a guide RNA and/or a site-directed modifying polypeptide include expression vectors, where an expression vector comprising a nucleotide sequence encoding a guide RNA and/or a site-directed modifying polypeptide is a "recombinant expression vector."

**[00270]** In some embodiments, the recombinant expression vector is a viral construct, e.g., a recombinant adeno-associated virus construct (see, e.g., U.S. Patent No. 7,078,387), a recombinant adenoviral construct, a recombinant lentiviral construct, etc.



**[00271]** Suitable expression vectors include, but are not limited to, viral vectors (e.g. viral vectors based on vaccinia virus; poliovirus; adenovirus (see, e.g., Li et al., *Invest Ophthalmol Vis Sci* 35:2543-2549, 1994; Borrás et al., *Gene Ther* 6:515-524, 1999; Li and Davidson, *PNAS* 92:7700-7704, 1995; Sakamoto et al., *Hum Gene Ther* 5:1088-1097, 1999; WO 94/12649, WO 93/03769; WO 93/19191; WO 94/28938; WO 95/11984 and WO 95/00655); adeno-associated virus (see, e.g., Ali et al., *Hum Gene Ther* 9:81-86, 1998; Flannery et al., *PNAS* 94:6916-6921, 1997; Bennett et al., *Invest Ophthalmol Vis Sci* 38:2857-2863, 1997; Jomary et al., *Gene Ther* 4:683-690, 1997; Rolling et al., *Hum Gene Ther* 10:641-648, 1999; Ali et al., *Hum Mol Genet* 5:591-594, 1996; Srivastava in WO 93/09239, Samulski et al., *J. Vir.* (1989) 63:3822-3828; Mendelson et al., *Virology* (1988) 166:154-165; and Flotte et al., *PNAS* (1993) 90:10613-10617); SV40; herpes simplex virus; human immunodeficiency virus (see, e.g., Miyoshi et al., *PNAS* 94:10319-23, 1997; Takahashi et al., *J Virol* 73:7812-7816, 1999); a retroviral vector (e.g., Murine Leukemia Virus, spleen necrosis virus, and vectors derived from retroviruses such as Rous Sarcoma Virus, Harvey Sarcoma Virus, avian leukosis virus, a lentivirus, human immunodeficiency virus, myeloproliferative sarcoma virus, and mammary tumor virus); and the like.

**[00272]** Numerous suitable expression vectors are known to those of skill in the art, and many are commercially available. The following vectors are provided by way of example; for eukaryotic host cells: pXT1, pSG5 (Stratagene), pSVK3, pBPV, pMSG, and pSVLSV40 (Pharmacia). However, any other vector may be used so long as it is compatible with the host cell.

**[00273]** In some embodiments, a nucleotide sequence encoding a guide RNA and/or a site-directed modifying polypeptide is operably linked to a control element, e.g., a transcriptional control element, such as a promoter. The transcriptional control element may be functional in either a eukaryotic cell, e.g., a mammalian cell, or a prokaryotic cell (e.g., bacterial or archaeal cell). In some embodiments, a nucleotide sequence encoding a guide RNA and/or a site-directed modifying polypeptide is operably linked to multiple control elements that allow expression of the nucleotide sequence encoding a guide RNA and/or a site-directed modifying polypeptide in both prokaryotic and eukaryotic cells.

**[00274]** Depending on the host/vector system utilized, any of a number of suitable transcription and translation control elements, including constitutive and inducible promoters, transcription enhancer elements, transcription terminators, etc. may be used in the expression vector (e.g., U6 promoter, H1 promoter, etc.; see above) (see e.g., Bitter et al. (1987) *Methods in Enzymology*, 153:516-544).

**[00275]** In some embodiments, a guide RNA and/or a site-directed modifying polypeptide can be provided as RNA. In such cases, the guide RNA and/or the RNA encoding the site-directed modifying polypeptide can be produced by direct chemical synthesis or may be transcribed *in vitro* from a DNA encoding the guide RNA. Methods of synthesizing RNA from a DNA template are well known in the art. In some cases, the guide RNA and/or the RNA encoding the site-directed modifying polypeptide will be synthesized *in vitro* using an RNA polymerase enzyme (e.g., T7 polymerase, T3 polymerase, SP6

polymerase, etc.). Once synthesized, the RNA may directly contact a target DNA or may be introduced into a cell by any of the well-known techniques for introducing nucleic acids into cells (e.g., microinjection, electroporation, transfection, etc).

**[00276]** Nucleotides encoding a guide RNA (introduced either as DNA or RNA) and/or a site-directed modifying polypeptide (introduced as DNA or RNA) and/or a donor polynucleotide may be provided to the cells using well-developed transfection techniques; see, e.g. Angel and Yanik (2010) PLoS ONE 5(7): e 11756, and the commercially available TransMessenger® reagents from Qiagen, Stemfect™ RNA Transfection Kit from Stemgent, and TransIT®-mRNA Transfection Kit from Mims Bio LLC. See also Beumer et al. (2008) Efficient gene targeting in *Drosophila* by direct embryo injection with zinc-finger nucleases. PNAS 105(50):19821-19826. Alternatively, nucleic acids encoding a guide RNA and/or a site-directed modifying polypeptide and/or a chimeric site-directed modifying polypeptide and/or a donor polynucleotide may be provided on DNA vectors. Many vectors, e.g. plasmids, cosmids, minicircles, phage, viruses, etc., useful for transferring nucleic acids into target cells are available. The vectors comprising the nucleic acid(s) may be maintained episomally, e.g. as plasmids, minicircle DNAs, viruses such as cytomegalovirus, adenovirus, etc., or they may be integrated into the target cell genome, through homologous recombination or random integration, e.g. retrovirus-derived vectors such as MMLV, HIV-1, ALV, etc.

**[00277]** Vectors may be provided directly to the cells. In other words, the cells are contacted with vectors comprising the nucleic acid encoding guide RNA and/or a site-directed modifying polypeptide and/or a chimeric site-directed modifying polypeptide and/or a donor polynucleotide such that the vectors are taken up by the cells. Methods for contacting cells with nucleic acid vectors that are plasmids, including electroporation, calcium chloride transfection, microinjection, and lipofection are well known in the art. For viral vector delivery, the cells are contacted with viral particles comprising the nucleic acid encoding a guide RNA and/or a site-directed modifying polypeptide and/or a chimeric site-directed modifying polypeptide and/or a donor polynucleotide. Retroviruses, for example, lentiviruses, are particularly suitable to the method of the invention. Commonly used retroviral vectors are "defective", i.e. unable to produce viral proteins required for productive infection. Rather, replication of the vector requires growth in a packaging cell line. To generate viral particles comprising nucleic acids of interest, the retroviral nucleic acids comprising the nucleic acid are packaged into viral capsids by a packaging cell line. Different packaging cell lines provide a different envelope protein (ecotropic, amphotropic or xenotropic) to be incorporated into the capsid, this envelope protein determining the specificity of the viral particle for the cells (ecotropic for murine and rat; amphotropic for most mammalian cell types including human, dog and mouse; and xenotropic for most mammalian cell types except murine cells). The appropriate packaging cell line may be used to ensure that the cells are targeted by the packaged viral particles. Methods of introducing the retroviral vectors comprising the nucleic acid encoding the reprogramming factors into packaging cell lines and of collecting the viral particles that are generated by

the packaging lines are well known in the art. Nucleic acids can also be introduced by direct micro-injection (e.g., injection of RNA into a zebrafish embryo).

**[00278]** Vectors used for providing the nucleic acids encoding guide RNA and/or a site-directed modifying polypeptide and/or a chimeric site-directed modifying polypeptide and/or a donor polynucleotide to the cells will typically comprise suitable promoters for driving the expression, that is, transcriptional activation, of the nucleic acid of interest. In other words, the nucleic acid of interest will be operably linked to a promoter. This may include ubiquitously acting promoters, for example, the CMV-13-actin promoter, or inducible promoters, such as promoters that are active in particular cell populations or that respond to the presence of drugs such as tetracycline. By transcriptional activation, it is intended that transcription will be increased above basal levels in the target cell by at least about 10 fold, by at least about 100 fold, more usually by at least about 1000 fold. In addition, vectors used for providing a guide RNA and/or a site-directed modifying polypeptide and/or a chimeric site-directed modifying polypeptide and/or a donor polynucleotide to the cells may include nucleic acid sequences that encode for selectable markers in the target cells, so as to identify cells that have taken up the guide RNA and/or a site-directed modifying polypeptide and/or a chimeric site-directed modifying polypeptide and/or a donor polynucleotide.

**[00279]** A guide RNA and/or a site-directed modifying polypeptide and/or a chimeric site-directed modifying polypeptide may instead be used to contact DNA or introduced into cells as RNA. Methods of introducing RNA into cells are known in the art and may include, for example, direct injection, transfection, or any other method used for the introduction of DNA. A site-directed modifying polypeptide may instead be provided to cells as a polypeptide. Such a polypeptide may optionally be fused to a polypeptide domain that increases solubility of the product. The domain may be linked to the polypeptide through a defined protease cleavage site, e.g. a TEV sequence, which is cleaved by TEV protease. The linker may also include one or more flexible sequences, e.g. from 1 to 10 glycine residues. In some embodiments, the cleavage of the fusion protein is performed in a buffer that maintains solubility of the product, e.g. in the presence of from 0.5 to 2 M urea, in the presence of polypeptides and/or polynucleotides that increase solubility, and the like. Domains of interest include endosomolytic domains, e.g. influenza HA domain; and other polypeptides that aid in production, e.g. IF2 domain, GST domain, GRPE domain, and the like. The polypeptide may be formulated for improved stability. For example, the peptides may be PEGylated, where the polyethyleneoxy group provides for enhanced lifetime in the blood stream.

**[00280]** Additionally or alternatively, the site-directed modifying polypeptide may be fused to a polypeptide permeant domain to promote uptake by the cell. A number of permeant domains are known in the art and may be used in the non-integrating polypeptides of the present invention, including peptides, peptidomimetics, and non-peptide carriers. For example, a permeant peptide may be derived from the third alpha helix of *Drosophila melanogaster* transcription factor Antennapedia, referred to as penetratin, which comprises the amino acid sequence RQIKIWFAQNRRMKWKK. As another example, the

permeant peptide comprises the HIV-1 tat basic region amino acid sequence, which may include, for example, amino acids 49-57 of naturally-occurring tat protein. Other permeant domains include poly-arginine motifs, for example, the region of amino acids 34-56 of HIV-1 rev protein, nona-arginine, octa-arginine, and the like. (See, for example, Futaki et al. (2003) *Curr Protein Pept Sci.* 2003 Apr; 4(2): 87-9 and 446; and Wender et al. (2000) *Proc. Natl. Acad. Sci. U.S.A* 2000 Nov. 21; 97(24):13003-8; published U.S. Patent applications 20030220334; 20030083256; 20030032593; and 20030022831, herein specifically incorporated by reference for the teachings of translocation peptides and peptoids). The nona-arginine (R9) sequence is one of the more efficient PTDs that have been characterized (Wender et al. 2000; Uemura et al. 2002). The site at which the fusion is made may be selected in order to optimize the biological activity, secretion or binding characteristics of the polypeptide. The optimal site will be determined by routine experimentation.

**[00281]** A site-directed modifying polypeptide may be produced *in vitro* or by eukaryotic cells or by prokaryotic cells, and it may be further processed by unfolding, e.g. heat denaturation, DTT reduction, etc. and may be further refolded, using methods known in the art.

**[00282]** Modifications of interest that do not alter primary sequence include chemical derivatization of polypeptides, e.g., acylation, acetylation, carboxylation, amidation, etc. Also included are modifications of glycosylation, e.g. those made by modifying the glycosylation patterns of a polypeptide during its synthesis and processing or in further processing steps; e.g. by exposing the polypeptide to enzymes which affect glycosylation, such as mammalian glycosylating or deglycosylating enzymes. Also embraced are sequences that have phosphorylated amino acid residues, e.g. phosphotyrosine, phosphoserine, or phosphothreonine.

**[00283]** Also included in the invention are guide RNAs and site-directed modifying polypeptides that have been modified using ordinary molecular biological techniques and synthetic chemistry so as to improve their resistance to proteolytic degradation, to change the target sequence specificity, to optimize solubility properties, to alter protein activity (e.g., transcription modulatory activity, enzymatic activity, etc) or to render them more suitable as a therapeutic agent. Analogs of such polypeptides include those containing residues other than naturally occurring L-amino acids, e.g. D-amino acids or non-naturally occurring synthetic amino acids. D-amino acids may be substituted for some or all of the amino acid residues. The site-directed modifying polypeptides may be prepared by *in vitro* synthesis, using conventional methods as known in the art. Various commercial synthetic apparatuses are available, for example, automated synthesizers by Applied Biosystems, Inc., Beckman, etc. By using synthesizers, naturally occurring amino acids may be substituted with unnatural amino acids. The particular sequence and the manner of preparation will be determined by convenience, economics, purity required, and the like.

**[00284]** If desired, various groups may be introduced into the peptide during synthesis or during expression, which allow for linking to other molecules or to a surface. Thus cysteines can be used to make thioethers, histidines for linking to a metal ion complex, carboxyl groups for forming amides or esters, amino groups for forming amides, and the like.

**[00285]** The site-directed modifying polypeptides may also be isolated and purified in accordance with conventional methods of recombinant synthesis. A lysate may be prepared of the expression host and the lysate purified using HPLC, exclusion chromatography, gel electrophoresis, affinity chromatography, or other purification technique. For the most part, the compositions which are used will comprise at least 20% by weight of the desired product, more usually at least about 75% by weight, preferably at least about 95% by weight, and for therapeutic purposes, usually at least about 99.5% by weight, in relation to contaminants related to the method of preparation of the product and its purification. Usually, the percentages will be based upon total protein. To induce DNA cleavage and recombination, or any desired modification to a target DNA, or any desired modification to a polypeptide associated with target DNA, the guide RNA and/or the site-directed modifying polypeptide and/or the donor polynucleotide, whether they be introduced as nucleic acids or polypeptides, are provided to the cells for about 30 minutes to about 24 hours, e.g., 1 hour, 1.5 hours, 2 hours, 2.5 hours, 3 hours, 3.5 hours, 4 hours, 5 hours, 6 hours, 7 hours, 8 hours, 12 hours, 16 hours, 18 hours, 20 hours, or any other period from about 30 minutes to about 24 hours, which may be repeated with a frequency of about every day to about every 4 days, e.g., every 1.5 days, every 2 days, every 3 days, or any other frequency from about every day to about every four days. The agent(s) may be provided to the cells one or more times, e.g. one time, twice, three times, or more than three times, and the cells allowed to incubate with the agent(s) for some amount of time following each contacting event e.g. 16-24 hours, after which time the media is replaced with fresh media and the cells are cultured further. In cases in which two or more different targeting complexes are provided to the cell (e.g., two different guide RNAs that are complementary to different sequences within the same or different target DNA), the complexes may be provided simultaneously (e.g. as two polypeptides and/or nucleic acids), or delivered simultaneously. Alternatively, they may be provided consecutively, e.g. the targeting complex being provided first, followed by the second targeting complex, etc. or vice versa.

**[00286]** Typically, an effective amount of the guide RNA and/or site-directed modifying polypeptide and/or donor polynucleotide is provided to the target DNA or cells to induce target modification. An effective amount of the guide RNA and/or site-directed modifying polypeptide and/or donor polynucleotide is the amount to induce a 2-fold increase or more in the amount of target modification observed between two homologous sequences relative to a negative control, e.g. a cell contacted with an empty vector or irrelevant polypeptide. That is to say, an effective amount or dose of the guide RNA and/or site-directed modifying polypeptide and/or donor polynucleotide will induce a 2-fold increase, a 3-fold increase, a 4-fold increase or more in the amount of target modification observed at a target DNA region, in some instances a 5-fold increase, a 6-fold increase or more, sometimes a 7-fold or 8-fold increase or more in the amount of

recombination observed, e.g. an increase of 10-fold, 50-fold, or 100-fold or more, in some instances, an increase of 200-fold, 500-fold, 700-fold, or 1000-fold or more, e.g. a 5000-fold, or 10,000-fold increase in the amount of recombination observed. The amount of target modification may be measured by any convenient method. For example, a silent reporter construct comprising complementary sequence to the targeting segment (targeting sequence) of the guide RNA flanked by repeat sequences that, when recombined, will reconstitute a nucleic acid encoding an active reporter may be cotransfected into the cells, and the amount of reporter protein assessed after contact with the guide RNA and/or site-directed modifying polypeptide and/or donor polynucleotide, e.g. 2 hours, 4 hours, 8 hours, 12 hours, 24 hours, 36 hours, 48 hours, 72 hours or more after contact with the guide RNA and/or site-directed modifying polypeptide and/or donor polynucleotide. As another, more sensitivity assay, for example, the extent of recombination at a genomic DNA region of interest comprising target DNA sequences may be assessed by PCR or Southern hybridization of the region after contact with a guide RNA and/or site-directed modifying polypeptide and/or donor polynucleotide, e.g. 2 hours, 4 hours, 8 hours, 12 hours, 24 hours, 36 hours, 48 hours, 72 hours or more after contact with the guide RNA and/or site-directed modifying polypeptide and/or donor polynucleotide.

**[00287]** Contacting the cells with a guide RNA and/or site-directed modifying polypeptide and/or donor polynucleotide may occur in any culture media and under any culture conditions that promote the survival of the cells. For example, cells may be suspended in any appropriate nutrient medium that is convenient, such as Iscove's modified DMEM or RPMI 1640, supplemented with fetal calf serum or heat inactivated goat serum (about 5-10%), L-glutamine, a thiol, particularly 2-mercaptoethanol, and antibiotics, e.g. penicillin and streptomycin. The culture may contain growth factors to which the cells are responsive. Growth factors, as defined herein, are molecules capable of promoting survival, growth and/or differentiation of cells, either in culture or in the intact tissue, through specific effects on a transmembrane receptor. Growth factors include polypeptides and non-polypeptide factors. Conditions that promote the survival of cells are typically permissive of nonhomologous end joining and homology-directed repair. In applications in which it is desirable to insert a polynucleotide sequence into a target DNA sequence, a polynucleotide comprising a donor sequence to be inserted is also provided to the cell. By a "donor sequence" or "donor polynucleotide" it is meant a nucleic acid sequence to be inserted at the cleavage site induced by a site-directed modifying polypeptide. The donor polynucleotide will contain sufficient homology to a genomic sequence at the cleavage site, e.g. 70%, 80%, 85%, 90%, 95%, or 100% homology with the nucleotide sequences flanking the cleavage site, e.g. within about 50 bases or less of the cleavage site, e.g. within about 30 bases, within about 15 bases, within about 10 bases, within about 5 bases, or immediately flanking the cleavage site, to support homology-directed repair between it and the genomic sequence to which it bears homology. Approximately 25, 50, 100, or 200 nucleotides, or more than 200 nucleotides, of sequence homology between a donor and a genomic sequence (or any integral value between 10 and 200 nucleotides, or more) will support homology-directed repair. Donor sequences can be

of any length, e.g. 10 nucleotides or more, 50 nucleotides or more, 100 nucleotides or more, 250 nucleotides or more, 500 nucleotides or more, 1000 nucleotides or more, 5000 nucleotides or more, etc.

**[00288]** The donor sequence is typically not identical to the genomic sequence that it replaces. Rather, the donor sequence may contain at least one or more single base changes, insertions, deletions, inversions or rearrangements with respect to the genomic sequence, so long as sufficient homology is present to support homology-directed repair. In some embodiments, the donor sequence comprises a non-homologous sequence flanked by two regions of homology, such that homology-directed repair between the target DNA region and the two flanking sequences results in insertion of the non-homologous sequence at the target region. Donor sequences may also comprise a vector backbone containing sequences that are not homologous to the DNA region of interest and that are not intended for insertion into the DNA region of interest. Generally, the homologous region(s) of a donor sequence will have at least 50% sequence identity to a genomic sequence with which recombination is desired. In certain embodiments, 60%, 70%, 80%, 90%, 95%, 98%, 99%, or 99.9% sequence identity is present. Any value between 1% and 100% sequence identity can be present, depending upon the length of the donor polynucleotide. The donor sequence may comprise certain sequence differences as compared to the genomic sequence, e.g. restriction sites, nucleotide polymorphisms, selectable markers (e.g., drug resistance genes, fluorescent proteins, enzymes etc.), etc., which may be used to assess for successful insertion of the donor sequence at the cleavage site or in some cases may be used for other purposes (e.g., to signify expression at the targeted genomic locus). In some cases, if located in a coding region, such nucleotide sequence differences will not change the amino acid sequence, or will make silent amino acid changes (i.e., changes which do not affect the structure or function of the protein). Alternatively, these sequences differences may include flanking recombination sequences such as FLPs, loxP sequences, or the like, that can be activated at a later time for removal of the marker sequence.

**[00289]** The donor sequence may be provided to the cell as single-stranded DNA, single-stranded RNA, double-stranded DNA, or double-stranded RNA. It may be introduced into a cell in linear or circular form. If introduced in linear form, the ends of the donor sequence may be protected (e.g., from exonucleolytic degradation) by methods known to those of skill in the art. For example, one or more dideoxynucleotide residues are added to the 3' terminus of a linear molecule and/or self-complementary oligonucleotides are ligated to one or both ends. See, for example, Chang et al. (1987) Proc. Natl. Acad. Sci USA 84:4959-4963; Nehls et al. (1996) Science 272:886-889. Additional methods for protecting exogenous polynucleotides from degradation include, but are not limited to, addition of terminal amino group(s) and the use of modified internucleotide linkages such as, for example, phosphorothioates, phosphoramidates, and 0-methyl ribose or deoxyribose residues. As an alternative to protecting the termini of a linear donor sequence, additional lengths of sequence may be included outside of the regions of homology that can be degraded without impacting recombination. A donor sequence can be introduced into a cell as part of a vector molecule having additional sequences such as, for example, replication

origins, promoters and genes encoding antibiotic resistance. Moreover, donor sequences can be introduced as naked nucleic acid, as nucleic acid complexed with an agent such as a liposome or poloxamer, or can be delivered by viruses (e.g., adenovirus, AAV), as described above for nucleic acids encoding a guide RNA and/or site-directed modifying polypeptide and/or donor polynucleotide.

**[00290]** Following the methods described above, a DNA region of interest may be cleaved and modified, i.e. "genetically modified", *ex vivo*. In some embodiments, as when a selectable marker has been inserted into the DNA region of interest, the population of cells may be enriched for those comprising the genetic modification by separating the genetically modified cells from the remaining population. Prior to enriching, the "genetically modified" cells may make up only about 1% or more (e.g., 2% or more, 3% or more, 4% or more, 5% or more, 6% or more, 7% or more, 8% or more, 9% or more, 10% or more, 15% or more, or 20% or more) of the cellular population. Separation of "genetically modified" cells may be achieved by any convenient separation technique appropriate for the selectable marker used. For example, if a fluorescent marker has been inserted, cells may be separated by fluorescence activated cell sorting, whereas if a cell surface marker has been inserted, cells may be separated from the heterogeneous population by affinity separation techniques, e.g. magnetic separation, affinity chromatography, "panning" with an affinity reagent attached to a solid matrix, or other convenient technique. Techniques providing accurate separation include fluorescence activated cell sorters, which can have varying degrees of sophistication, such as multiple color channels, low angle and obtuse light scattering detecting channels, impedance channels, etc. The cells may be selected against dead cells by employing dyes associated with dead cells (e.g. propidium iodide). Any technique may be employed which is not unduly detrimental to the viability of the genetically modified cells. Cell compositions that are highly enriched for cells comprising modified DNA are achieved in this manner. By "highly enriched", it is meant that the genetically modified cells will be 70% or more, 75% or more, 80% or more, 85% or more, 90% or more of the cell composition, for example, about 95% or more, or 98% or more of the cell composition. In other words, the composition may be a substantially pure composition of genetically modified cells.

**[00291]** Genetically modified cells produced by the methods described herein may be used immediately. Alternatively, the cells may be frozen at liquid nitrogen temperatures and stored for long periods of time, being thawed and capable of being reused. In such cases, the cells will usually be frozen in 10% dimethylsulfoxide (DMSO), 50% serum, 40% buffered medium, or some other such solution as is commonly used in the art to preserve cells at such freezing temperatures, and thawed in a manner as commonly known in the art for thawing frozen cultured cells.

**[00292]** The genetically modified cells may be cultured *in vitro* under various culture conditions. The cells may be expanded in culture, i.e. grown under conditions that promote their proliferation. Culture medium may be liquid or semi-solid, e.g. containing agar, methylcellulose, etc. The cell population may be suspended in an appropriate nutrient medium, such as Iscove's modified DMEM or RPMI 1640,



normally supplemented with fetal calf serum (about 5-10%), L-glutamine, a thiol, particularly 2-mercaptoethanol, and antibiotics, e.g. penicillin and streptomycin. The culture may contain growth factors to which the regulatory T cells are responsive. Growth factors, as defined herein, are molecules capable of promoting survival, growth and/or differentiation of cells, either in culture or in the intact tissue, through specific effects on a transmembrane receptor. Growth factors include polypeptides and non-polypeptide factors.

**[00293]** Cells that have been genetically modified in this way may be transplanted to a subject for purposes such as gene therapy, e.g. to treat a disease or as an antiviral, antipathogenic, or anticancer therapeutic, for the production of genetically modified organisms in agriculture, or for biological research. The subject may be a neonate, a juvenile, or an adult. Of particular interest are mammalian subjects. Mammalian species that may be treated with the present methods include canines and felines; equines; bovines; ovines; etc. and primates, particularly humans. Animal models, particularly small mammals (e.g. mouse, rat, guinea pig, hamster, lagomorpha (e.g., rabbit), etc.) may be used for experimental investigations.

**[00294]** Cells may be provided to the subject alone or with a suitable substrate or matrix, e.g. to support their growth and/or organization in the tissue to which they are being transplanted. Usually, at least  $1 \times 10^3$  cells will be administered, for example  $5 \times 10^3$  cells,  $1 \times 10^4$  cells,  $5 \times 10^4$  cells,  $1 \times 10^5$  cells,  $1 \times 10^6$  cells or more. The cells may be introduced to the subject via any of the following routes: parenteral, subcutaneous, intravenous, intracranial, intraspinal, intraocular, or into spinal fluid. The cells may be introduced by injection, catheter, or the like. Examples of methods for local delivery, that is, delivery to the site of injury, include, e.g. through an Ommaya reservoir, e.g. for intrathecal delivery (see e.g. US Patent Nos. 5,222,982 and 5,385,582, incorporated herein by reference); by bolus injection, e.g. by a syringe, e.g. into a joint; by continuous infusion, e.g. by cannulation, e.g. with convection (see e.g. US Application No. 20070254842, incorporated herein by reference); or by implanting a device upon which the cells have been reversibly affixed (see e.g. US Application Nos. 20080081064 and 20090196903, incorporated herein by reference). Cells may also be introduced into an embryo (e.g., a blastocyst) for the purpose of generating a transgenic animal (e.g., a transgenic mouse).

**[00295]** The number of administrations of treatment to a subject may vary. Introducing the genetically modified cells into the subject may be a one-time event; but in certain situations, such treatment may elicit improvement for a limited period of time and require an on-going series of repeated treatments. In other situations, multiple administrations of the genetically modified cells may be required before an effect is observed. The exact protocols depend upon the disease or condition, the stage of the disease and parameters of the individual subject being treated.

**[00296]** In other aspects of the disclosure, the guide RNA and/or site-directed modifying polypeptide and/or donor polynucleotide are employed to modify cellular DNA in vivo, again for purposes such as

gene therapy, e.g. to treat a disease or as an antiviral, antipathogenic, or anticancer therapeutic, for the production of genetically modified organisms in agriculture, or for biological research. In these in vivo embodiments, a guide RNA and/or site-directed modifying polypeptide and/or donor polynucleotide are administered directly to the individual. A guide RNA and/or site-directed modifying polypeptide and/or donor polynucleotide may be administered by any of a number of well-known methods in the art for the administration of peptides, small molecules and nucleic acids to a subject. A guide RNA and/or site-directed modifying polypeptide and/or donor polynucleotide can be incorporated into a variety of formulations. More particularly, a guide RNA and/or site-directed modifying polypeptide and/or donor polynucleotide of the present invention can be formulated into pharmaceutical compositions by combination with appropriate pharmaceutically acceptable carriers or diluents.

**[00297]** Pharmaceutical preparations are compositions that include one or more a guide RNA and/or site-directed modifying polypeptide and/or donor polynucleotide present in a pharmaceutically acceptable vehicle. "Pharmaceutically acceptable vehicles" may be vehicles approved by a regulatory agency of the Federal or a state government or listed in the U.S. Pharmacopeia or other generally recognized pharmacopeia for use in mammals, such as humans. The term "vehicle" refers to a diluent, adjuvant, excipient, or carrier with which a compound of the invention is formulated for administration to a mammal. Such pharmaceutical vehicles can be lipids, e.g. liposomes, e.g. liposome dendrimers; liquids, such as water and oils, including those of petroleum, animal, vegetable or synthetic origin, such as peanut oil, soybean oil, mineral oil, sesame oil and the like, saline; gum acacia, gelatin, starch paste, talc, keratin, colloidal silica, urea, and the like. In addition, auxiliary, stabilizing, thickening, lubricating and coloring agents may be used. Pharmaceutical compositions may be formulated into preparations in solid, semi-solid, liquid or gaseous forms, such as tablets, capsules, powders, granules, ointments, solutions, suppositories, injections, inhalants, gels, microspheres, and aerosols. As such, administration of the a guide RNA and/or site-directed modifying polypeptide and/or donor polynucleotide can be achieved in various ways, including oral, buccal, rectal, parenteral, intraperitoneal, intradermal, transdermal, intratracheal, intraocular, etc., administration. The active agent may be systemic after administration or may be localized by the use of regional administration, intramural administration, or use of an implant that acts to retain the active dose at the site of implantation. The active agent may be formulated for immediate activity or it may be formulated for sustained release.

**[00298]** For some conditions, particularly central nervous system conditions, it may be necessary to formulate agents to cross the blood-brain barrier (BBB). One strategy for drug delivery through the BBB entails disruption of the BBB, either by osmotic means such as mannitol or leukotrienes, or biochemically by the use of vasoactive substances such as bradykinin. The potential for using BBB opening to target specific agents to brain tumors is also an option. A BBB disrupting agent can be co-administered with the therapeutic compositions of the invention when the compositions are administered by intravascular injection. Other strategies to go through the BBB may entail the use of endogenous transport systems,

including Caveolin-1 mediated transcytosis, carrier-mediated transporters such as glucose and amino acid carriers, receptor-mediated transcytosis for insulin or transferrin, and active efflux transporters such as p-glycoprotein. Active transport moieties may also be conjugated to the therapeutic compounds for use in the invention to facilitate transport across the endothelial wall of the blood vessel. Alternatively, drug delivery of therapeutics agents behind the BBB may be by local delivery, for example by intrathecal delivery, e.g. through an Ommaya reservoir (see e.g. US Patent Nos. 5,222,982 and 5385582, incorporated herein by reference); by bolus injection, e.g. by a syringe, e.g. intravitreally or intracranially; by continuous infusion, e.g. by cannulation, e.g. with convection (see e.g. US Application No. 20070254842, incorporated here by reference); or by implanting a device upon which the agent has been reversibly affixed (see e.g. US Application Nos. 20080081064 and 20090196903, incorporated herein by reference).

**[00299]** Typically, an effective amount of a guide RNA and/or site-directed modifying polypeptide and/or donor polynucleotide are provided. As discussed above with regard to ex vivo methods, an effective amount or effective dose of a guide RNA and/or site-directed modifying polypeptide and/or donor polynucleotide in vivo is the amount to induce a 2 fold increase or more in the amount of recombination observed between two homologous sequences relative to a negative control, e.g. a cell contacted with an empty vector or irrelevant polypeptide. The amount of recombination may be measured by any convenient method, e.g. as described above and known in the art. The calculation of the effective amount or effective dose of a guide RNA and/or site-directed modifying polypeptide and/or donor polynucleotide to be administered is within the skill of one of ordinary skill in the art, and will be routine to those persons skilled in the art. The final amount to be administered will be dependent upon the route of administration and upon the nature of the disorder or condition that is to be treated.

**[00300]** The effective amount given to a particular patient will depend on a variety of factors, several of which will differ from patient to patient. A competent clinician will be able to determine an effective amount of a therapeutic agent to administer to a patient to halt or reverse the progression the disease condition as required. Utilizing LD50 animal data, and other information available for the agent, a clinician can determine the maximum safe dose for an individual, depending on the route of administration. For instance, an intravenously administered dose may be more than an intrathecally administered dose, given the greater body of fluid into which the therapeutic composition is being administered. Similarly, compositions which are rapidly cleared from the body may be administered at higher doses, or in repeated doses, in order to maintain a therapeutic concentration. Utilizing ordinary skill, the competent clinician will be able to optimize the dosage of a particular therapeutic in the course of routine clinical trials.

**[00301]** For inclusion in a medicament, a guide RNA and/or site-directed modifying polypeptide and/or donor polynucleotide may be obtained from a suitable commercial source. As a general proposition, the total pharmaceutically effective amount of the a guide RNA and/or site-directed modifying polypeptide and/or donor polynucleotide administered parenterally per dose will be in a range that can be measured by a dose response curve.

**[00302]** Therapies based on a guide RNA and/or site-directed modifying polypeptide and/or donor polynucleotides, i.e. preparations of a guide RNA and/or site-directed modifying polypeptide and/or donor polynucleotide to be used for therapeutic administration, must be sterile. Sterility is readily accomplished by filtration through sterile filtration membranes (e.g., 0.2 µm membranes). Therapeutic compositions generally are placed into a container having a sterile access port, for example, an intravenous solution bag or vial having a stopper pierceable by a hypodermic injection needle. The therapies based on a guide RNA and/or site-directed modifying polypeptide and/or donor polynucleotide may be stored in unit or multi-dose containers, for example, sealed ampules or vials, as an aqueous solution or as a lyophilized formulation for reconstitution. As an example of a lyophilized formulation, 10-ml vials are filled with 5 ml of sterile-filtered 1% (w/v) aqueous solution of compound, and the resulting mixture is lyophilized. The infusion solution is prepared by reconstituting the lyophilized compound using bacteriostatic Water-for-Injection.

**[00303]** Pharmaceutical compositions can include, depending on the formulation desired, pharmaceutically-acceptable, non-toxic carriers of diluents, which are defined as vehicles commonly used to formulate pharmaceutical compositions for animal or human administration. The diluent is selected so as not to affect the biological activity of the combination. Examples of such diluents are distilled water, buffered water, physiological saline, PBS, Ringer's solution, dextrose solution, and Hank's solution. In addition, the pharmaceutical composition or formulation can include other carriers, adjuvants, or non-toxic, nontherapeutic, nonimmunogenic stabilizers, excipients and the like. The compositions can also include additional substances to approximate physiological conditions, such as pH adjusting and buffering agents, toxicity adjusting agents, wetting agents and detergents.

**[00304]** The composition can also include any of a variety of stabilizing agents, such as an antioxidant for example. When the pharmaceutical composition includes a polypeptide, the polypeptide can be complexed with various well-known compounds that enhance the in vivo stability of the polypeptide, or otherwise enhance its pharmacological properties (e.g., increase the half-life of the polypeptide, reduce its toxicity, enhance solubility or uptake). Examples of such modifications or complexing agents include sulfate, gluconate, citrate and phosphate. The nucleic acids or polypeptides of a composition can also be complexed with molecules that enhance their in vivo attributes. Such molecules include, for example, carbohydrates, polyamines, amino acids, other peptides, ions (e.g., sodium, potassium, calcium, magnesium, manganese), and lipids.

**[00305]** Further guidance regarding formulations that are suitable for various types of administration can be found in Remington's Pharmaceutical Sciences, Mace Publishing Company, Philadelphia, Pa., 17th ed. (1985). For a brief review of methods for drug delivery, see, Langer, Science 249:1527-1533 (1990).

**[00306]** The pharmaceutical compositions can be administered for prophylactic and/or therapeutic treatments. Toxicity and therapeutic efficacy of the active ingredient can be determined according to

standard pharmaceutical procedures in cell cultures and/or experimental animals, including, for example, determining the LD50 (the dose lethal to 50% of the population) and the ED50 (the dose therapeutically effective in 50% of the population). The dose ratio between toxic and therapeutic effects is the therapeutic index and it can be expressed as the ratio LD50/ED50. Therapies that exhibit large therapeutic indices are preferred.

**[00307]** The data obtained from cell culture and/or animal studies can be used in formulating a range of dosages for humans. The dosage of the active ingredient typically lies within a range of circulating concentrations that include the ED50 with low toxicity. The dosage can vary within this range depending upon the dosage form employed and the route of administration utilized. The components used to formulate the pharmaceutical compositions are preferably of high purity and are substantially free of potentially harmful contaminants (e.g., at least National Food (NF) grade, generally at least analytical grade, and more typically at least pharmaceutical grade). Moreover, compositions intended for in vivo use are usually sterile. To the extent that a given compound must be synthesized prior to use, the resulting product is typically substantially free of any potentially toxic agents, particularly any endotoxins, which may be present during the synthesis or purification process. Compositions for parental administration are also sterile, substantially isotonic and made under GMP conditions.

**[00308]** The effective amount of a therapeutic composition to be given to a particular patient will depend on a variety of factors, several of which will differ from patient to patient. A competent clinician will be able to determine an effective amount of a therapeutic agent to administer to a patient to halt or reverse the progression of the disease condition as required. Utilizing LD50 animal data, and other information available for the agent, a clinician can determine the maximum safe dose for an individual, depending on the route of administration. For instance, an intravenously administered dose may be more than an intrathecally administered dose, given the greater body of fluid into which the therapeutic composition is being administered. Similarly, compositions which are rapidly cleared from the body may be administered at higher doses, or in repeated doses, in order to maintain a therapeutic concentration. Utilizing ordinary skill, the competent clinician will be able to optimize the dosage of a particular therapeutic in the course of routine clinical trials.

**[00309]** Genetically Modified Host Cells

**[00310]** The present disclosure provides genetically modified host cells, including isolated genetically modified host cells, where a genetically modified host cell comprises (has been genetically modified with: 1) an exogenous guide RNA; 2) an exogenous nucleic acid comprising a nucleotide sequence encoding a guide RNA; 3) an exogenous site-directed modifying polypeptide (e.g., a naturally occurring Cas9; a modified, i.e., mutated or variant, Cas9; a chimeric Cas9; etc.); 4) an exogenous nucleic acid comprising a nucleotide sequence encoding a site-directed modifying polypeptide; or 5) any combination of the above. A genetically modified cell is generated by genetically modifying a host cell with, for example: 1) an exogenous guide RNA;

2) an exogenous nucleic acid comprising a nucleotide sequence encoding a guide RNA; 3) an exogenous site-directed modifying polypeptide; 4) an exogenous nucleic acid comprising a nucleotide sequence encoding a site-directed modifying polypeptide; or 5) any combination of the above.).

**[00311]** All cells suitable to be a target cell are also suitable to be a genetically modified host cell. For example, a genetically modified host cells of interest can be a cell from any organism (e.g. a bacterial cell, an archaeal cell, a cell of a single-cell eukaryotic organism, a plant cell, an algal cell, e.g., *Botryococcus braunii*, *Chlamydomonas reinhardtii*, *Nannochloropsis gaditana*, *Chlorella pyrenoidosa*, *Sargassum patens* C. Agardh, and the like, a fungal cell (e.g., a yeast cell), an animal cell, a cell from an invertebrate animal (e.g. fruit fly, cnidarian, echinoderm, nematode, etc.), a cell from a vertebrate animal (e.g., fish, amphibian, reptile, bird, mammal), a cell from a mammal (e.g., a pig, a cow, a goat, a sheep, a rodent, a rat, a mouse, a non-human primate, a human, etc.), etc.

**[00312]** In some embodiments, a genetically modified host cell has been genetically modified with an exogenous nucleic acid comprising a nucleotide sequence encoding a site-directed modifying polypeptide (e.g., a naturally occurring Cas9; a modified, i.e., mutated or variant, Cas9; a chimeric Cas9; etc.). The DNA of a genetically modified host cell can be targeted for modification by introducing into the cell a guide RNA (or a DNA encoding a guide RNA, which determines the genomic location/sequence to be modified) and optionally a donor nucleic acid. In some embodiments, the nucleotide sequence encoding a site-directed modifying polypeptide is operably linked to an inducible promoter (e.g., heat shock promoter, Tetracycline-regulated promoter, Steroid-regulated promoter, Metal-regulated promoter, estrogen receptor-regulated promoter, etc.). In some embodiments, the nucleotide sequence encoding a site-directed modifying polypeptide is operably linked to a spatially restricted and/or temporally restricted promoter (e.g., a tissue specific promoter, a cell type specific promoter, etc.). In some embodiments, the nucleotide sequence encoding a site-directed modifying polypeptide is operably linked to a constitutive promoter.

**[00313]** In some embodiments, a genetically modified host cell is *in vitro*. In some embodiments, a genetically modified host cell is *in vivo*. In some embodiments, a genetically modified host cell is a prokaryotic cell or is derived from a prokaryotic cell. In some embodiments, a genetically modified host cell is a bacterial cell or is derived from a bacterial cell. In some embodiments, a genetically modified host cell is an archaeal cell or is derived from an archaeal cell. In some embodiments, a genetically modified host cell is a eukaryotic cell or is derived from a eukaryotic cell. In some embodiments, a genetically modified host cell is a plant cell or is derived from a plant cell. In some embodiments, a genetically modified host cell is an animal cell or is derived from an animal cell. In some embodiments, a genetically modified host cell is an invertebrate cell or is derived from an invertebrate cell. In some embodiments, a genetically modified host cell is a vertebrate cell or is derived from a vertebrate cell. In some embodiments, a genetically modified host cell is a mammalian cell or is derived from a mammalian cell. In some embodiments, a

genetically modified host cell is a rodent cell or is derived from a rodent cell. In some embodiments, a genetically modified host cell is a human cell or is derived from a human cell.

**[00314]** The present disclosure further provides progeny of a genetically modified cell, where the progeny can comprise the same exogenous nucleic acid or polypeptide as the genetically modified cell from which it was derived. The present disclosure further provides a composition comprising a genetically modified host cell.

**[00315]** Genetically modified stem cells and genetically modified progenitor cells

**[00316]** In some embodiments, a genetically modified host cell is a genetically modified stem cell or progenitor cell. Suitable host cells include, e.g., stem cells (adult stem cells, embryonic stem cells, iPS cells, etc.) and progenitor cells (e.g., cardiac progenitor cells, neural progenitor cells, etc.). Suitable host cells include mammalian stem cells and progenitor cells, including, e.g., rodent stem cells, rodent progenitor cells, human stem cells, human progenitor cells, etc. Suitable host cells include *in vitro* host cells, e.g., isolated host cells.

**[00317]** In some embodiments, a genetically modified host cell comprises an exogenous guide RNA nucleic acid. In some embodiments, a genetically modified host cell comprises an exogenous nucleic acid comprising a nucleotide sequence encoding a guide RNA. In some embodiments, a genetically modified host cell comprises an exogenous site-directed modifying polypeptide (e.g., a naturally occurring Cas9; a modified, i.e., mutated or variant, Cas9; a chimeric Cas9; etc.). In some embodiments, a genetically modified host cell comprises an exogenous nucleic acid comprising a nucleotide sequence encoding a site-directed modifying polypeptide. In some embodiments, a genetically modified host cell comprises exogenous nucleic acid comprising a nucleotide sequence encoding 1) a guide RNA and 2) a site-directed modifying polypeptide.

**[00318]** In some cases, the site-directed modifying polypeptide comprises an amino acid sequence having at least about 75%, at least about 80%, at least about 85%, at least about 90%, at least about 95%, at least about 99%, or 100%, amino acid sequence identity to amino acids 7-166 and/or 731-1003 of SEQ ID NO: 8, or to the corresponding portions in any of the amino acid sequences set forth as SEQ ID NOS: 1-800.

**[00319]** Compositions

**[00320]** The present disclosure provides a composition comprising a guide RNA and/or a site-directed modifying polypeptide. In some cases, the site-directed modifying polypeptide is a chimeric polypeptide. A composition is useful for carrying out a method of the present disclosure, e.g., a method for site-specific modification of a target DNA; a method for site-specific modification of a polypeptide associated with a target DNA; etc.

**[00321]** Compositions comprising a guide RNA

**[00322]** The present disclosure provides a composition comprising a guide RNA. The composition can comprise, in addition to the guide RNA, one or more of: a salt, e.g., NaCl, MgCl<sub>2</sub>, KCl, MgSO<sub>4</sub>, etc.; a buffering agent, e.g., a Tris buffer, N-(2-Hydroxyethyl)piperazine-N'-(2-ethanesulfonic acid) (HEPES), 2-(N-Morpholino)ethanesulfonic acid (MES), MES sodium salt, 3-(N-Morpholino)propanesulfonic acid (MOPS), N-tris[Hydroxymethyl]methyl-3-aminopropanesulfonic acid (TAPS), etc.; a solubilizing agent; a detergent, e.g., a non-ionic detergent such as Tween-20, etc.; a nuclease inhibitor; and the like. For example, in some cases, a composition comprises a guide RNA and a buffer for stabilizing nucleic acids.

**[00323]** In some embodiments, a guide RNA present in a composition is pure, e.g., at least about 75%, at least about 80%, at least about 85%, at least about 90%, at least about 95%, at least about 98%, at least about 99%, or more than 99% pure, where "% purity" means that guide RNA is the recited percent free from other macromolecules, or contaminants that may be present during the production of the guide RNA.

**[00324]** Compositions comprising a chimeric polypeptide

**[00325]** The present disclosure provides a composition a chimeric polypeptide. The composition can comprise, in addition to the guide RNA, one or more of: a salt, e.g., NaCl, MgCl<sub>2</sub>, KCl, MgSO<sub>4</sub>, etc.; a buffering agent, e.g., a Tris buffer, HEPES, MES, MES sodium salt, MOPS, TAPS, etc.; a solubilizing agent; a detergent, e.g., a non-ionic detergent such as Tween-20, etc.; a protease inhibitor; a reducing agent (e.g., dithiothreitol); and the like.

**[00326]** In some embodiments, a chimeric polypeptide present in a composition is pure, e.g., at least about 75%, at least about 80%, at least about 85%, at least about 90%, at least about 95%, at least about 98%, at least about 99%, or more than 99% pure, where "% purity" means that the site-directed modifying polypeptide is the recited percent free from other proteins, other macromolecules, or contaminants that may be present during the production of the chimeric polypeptide.

**[00327]** Compositions comprising a guide RNA and a site-directed modifying polypeptide

**[00328]** The present disclosure provides a composition comprising: (i) a guide RNA or a DNA polynucleotide encoding the same; and ii) a site-directed modifying polypeptide, or a polynucleotide encoding the same. In some cases, the site-directed modifying polypeptide is a chimeric site-directed modifying polypeptide. In other cases, the site-directed modifying polypeptide is a naturally-occurring site-directed modifying polypeptide. In some instances, the site-directed modifying polypeptide exhibits enzymatic activity that modifies a target DNA. In other cases, the site-directed modifying polypeptide exhibits enzymatic activity that modifies a polypeptide that is associated with a target DNA. In still other cases, the site-directed modifying polypeptide modulates transcription of the target DNA.



**[00329]** The present disclosure provides a composition comprising: (i) a guide RNA, as described above, or a DNA polynucleotide encoding the same, the guide RNA comprising: (a) a first segment comprising a nucleotide sequence that is complementary to a sequence in a target DNA; and (b) a second segment that interacts with a site-directed modifying polypeptide; and (ii) the site-directed modifying polypeptide, or a polynucleotide encoding the same, the site-directed modifying polypeptide comprising: (a) an RNA-binding portion that interacts with the guide RNA; and (b) an activity portion that exhibits site-directed enzymatic activity, wherein the site of enzymatic activity is determined by the guide RNA.

**[00330]** In some instances, a composition comprises: a composition comprising: (i) a guide RNA, the guide RNA comprising: (a) a first segment comprising a nucleotide sequence that is complementary to a sequence in a target DNA; and (b) a second segment that interacts with a site-directed modifying polypeptide; and (ii) the site-directed modifying polypeptide, the site-directed modifying polypeptide comprising: (a) an RNA-binding portion that interacts with the guide RNA; and (b) an activity portion that exhibits site-directed enzymatic activity, wherein the site of enzymatic activity is determined by the guide RNA.

**[00331]** In other embodiments, a composition comprises: (i) a polynucleotide encoding a guide RNA, the guide RNA comprising: (a) a first segment comprising a nucleotide sequence that is complementary to a sequence in a target DNA; and (b) a second segment that interacts with a site-directed modifying polypeptide; and (ii) a polynucleotide encoding the site-directed modifying polypeptide, the site-directed modifying polypeptide comprising: (a) an RNA-binding portion that interacts with the guide RNA; and (b) an activity portion that exhibits site-directed enzymatic activity, wherein the site of enzymatic activity is determined by the guide RNA.

**[00332]** In some embodiments, a composition includes both RNA molecules of a double-molecule guide RNA. As such, in some embodiments, a composition includes an activator-RNA that comprises a duplex-forming segment that is complementary to the duplex-forming segment of a targeter-. The duplex-forming segments of the activator-RNA and the targeter-RNA hybridize to form the dsRNA duplex of the protein-binding segment of the guide RNA. The targeter-RNA further provides the DNA-targeting segment (single stranded) of the guide RNA and therefore targets the guide RNA to a specific sequence within the target DNA. As one non-limiting example, the duplex-forming segment of the activator-RNA comprises a nucleotide sequence that has at least about 70%, at least about 80%, at least about 90%, at least about 95%, at least about 98%, or 100% identity with a tracrRNA sequence set out in Supplementary Table S5. As another non-limiting example, the duplex-forming segment of the targeter-RNA comprises a nucleotide sequence that has at least about 70%, at least about 80%, at least about 90%, at least about 95%, at least about 98%, or 100% identity with a CRISPR repeat sequence set out in Supplementary Table S5.

**[00333]** The present disclosure provides a composition comprising: (i) a guide RNA, or a DNA polynucleotide encoding the same, the guide RNA comprising: (a) a first segment comprising a nucleotide sequence that is complementary to a sequence in a target DNA; and (b) a second segment that interacts with a site-directed modifying polypeptide; and (ii) the site-directed modifying polypeptide, or a polynucleotide encoding the same, the site-directed modifying polypeptide comprising: (a) an RNA-binding portion that interacts with the guide RNA; and (b) an activity portion that modulates transcription within the target DNA, wherein the site of modulated transcription within the target DNA is determined by the guide RNA.

**[00334]** For example, in some cases, a composition comprises: (i) a guide RNA, the guide RNA comprising: (a) a first segment comprising a nucleotide sequence that is complementary to a sequence in a target DNA; and (b) a second segment that interacts with a site-directed modifying polypeptide; and (ii) the site-directed modifying polypeptide, the site-directed modifying polypeptide comprising: (a) an RNA-binding portion that interacts with the guide RNA; and (b) an activity portion that modulates transcription within the target DNA, wherein the site of modulated transcription within the target DNA is determined by the guide RNA.

**[00335]** As another example, in some cases, a composition comprises: (i) a DNA polynucleotide encoding a guide RNA, the guide RNA comprising: (a) a first segment comprising a nucleotide sequence that is complementary to a sequence in a target DNA; and (b) a second segment that interacts with a site-directed modifying polypeptide; and (ii) a polynucleotide encoding the site-directed modifying polypeptide, the site-directed modifying polypeptide comprising: (a) an RNA-binding portion that interacts with the guide RNA; and (b) an activity portion that modulates transcription within the target DNA, wherein the site of modulated transcription within the target DNA is determined by the guide RNA. A composition can comprise, in addition to i) a guide RNA, or a DNA polynucleotide encoding the same; and ii) a site-directed modifying polypeptide, or a polynucleotide encoding the same, one or more of: a salt, e.g., NaCl, MgCl<sub>2</sub>, KCl, MgSO<sub>4</sub>, etc.; a buffering agent, e.g., a Tris buffer, HEPES, MES, MES sodium salt, MOPS, TAPS, etc.; a solubilizing agent; a detergent, e.g., a non-ionic detergent such as Tween-20, etc.; a protease inhibitor; a reducing agent (e.g., dithiothreitol); and the like.

**[00336]** In some cases, the components of the composition are individually pure, e.g., each of the components is at least about 75%, at least about 80%, at least about 90%, at least about 95%, at least about 98%, at least about 99%, or at least 99%, pure. In some cases, the individual components of a composition are pure before being added to the composition.

**[00337]** For example, in some embodiments, a site-directed modifying polypeptide present in a composition is pure, e.g., at least about 75%, at least about 80%, at least about 85%, at least about 90%, at least about 95%, at least about 98%, at least about 99%, or more than 99% pure, where "% purity" means that the site-directed modifying polypeptide is the recited percent free from other proteins (e.g.,

proteins other than the site-directed modifying polypeptide), other macromolecules, or contaminants that may be present during the production of the site-directed modifying polypeptide.

**[00338]** Kits

**[00339]** The present disclosure provides kits for carrying out a method. A kit can include one or more of: a site-directed modifying polypeptide; a nucleic acid comprising a nucleotide encoding a site-directed modifying polypeptide; a guide RNA; a nucleic acid comprising a nucleotide sequence encoding a guide RNA; an activator-RNA; a nucleic acid comprising a nucleotide sequence encoding an activator-RNA; a targeter-RNA; and a nucleic acid comprising a nucleotide sequence encoding a targeter-RNA. A site-directed modifying polypeptide; a nucleic acid comprising a nucleotide encoding a site-directed modifying polypeptide; a guide RNA; a nucleic acid comprising a nucleotide sequence encoding a guide RNA; an activator-RNA; a nucleic acid comprising a nucleotide sequence encoding an activator-RNA; a targeter-RNA; and a nucleic acid comprising a nucleotide sequence encoding a targeter-RNA, are described in detail above. A kit may comprise a complex that comprises two or more of: a site-directed modifying polypeptide; a nucleic acid comprising a nucleotide encoding a site-directed modifying polypeptide; a guide RNA; a nucleic acid comprising a nucleotide sequence encoding a guide RNA; an activator-RNA; a nucleic acid comprising a nucleotide sequence encoding an activator-RNA; a targeter-RNA; and a nucleic acid comprising a nucleotide sequence encoding a targeter-RNA. In some embodiments, a kit comprises a site-directed modifying polypeptide, or a polynucleotide encoding the same. In some embodiments, the site-directed modifying polypeptide comprises: (a) an RNA-binding portion that interacts with the guide RNA; and (b) an activity portion that modulates transcription within the target DNA, wherein the site of modulated transcription within the target DNA is determined by the guide RNA. In some cases, the activity portion of the site-directed modifying polypeptide exhibits reduced or inactivated nuclease activity. In some cases, the site-directed modifying polypeptide is a chimeric site-directed modifying polypeptide.

**[00340]** In some embodiments, a kit comprises: a site-directed modifying polypeptide, or a polynucleotide encoding the same, and a reagent for reconstituting and/or diluting the site-directed modifying polypeptide. In other embodiments, a kit comprises a nucleic acid (e.g., DNA, RNA) comprising a nucleotide encoding a site-directed modifying polypeptide. In some embodiments, a kit comprises: a nucleic acid (e.g., DNA, RNA) comprising a nucleotide encoding a site-directed modifying polypeptide; and a reagent for reconstituting and/or diluting the site-directed modifying polypeptide.

**[00341]** A kit comprising a site-directed modifying polypeptide, or a polynucleotide encoding the same, can further include one or more additional reagents, where such additional reagents can be selected from: a buffer for introducing the site-directed modifying polypeptide into a cell; a wash buffer; a control reagent; a control expression vector or RNA polynucleotide; a reagent for *in vitro* production of the site-directed modifying polypeptide from DNA, and the like. In some cases, the site-directed modifying polypeptide included in a kit is a chimeric site-directed modifying polypeptide, as described above.

**[00342]** In some embodiments, a kit comprises a guide RNA, or a DNA polynucleotide encoding the same, the guide RNA comprising: (a) a first segment comprising a nucleotide sequence that is complementary to a sequence in a target DNA; and (b) a second segment that interacts with a site-directed modifying polypeptide. In some embodiments, the guide RNA further comprises a third segment (as described above). In some embodiments, a kit comprises: (i) a guide RNA, or a DNA polynucleotide encoding the same, the guide RNA comprising: (a) a first segment comprising a nucleotide sequence that is complementary to a sequence in a target DNA; and (b) a second segment that interacts with a site-directed modifying polypeptide; and (ii) a site-directed modifying polypeptide, or a polynucleotide encoding the same, the site-directed modifying polypeptide comprising: (a) an RNA-binding portion that interacts with the guide RNA; and (b) an activity portion that exhibits site-directed enzymatic activity, wherein the site of enzymatic activity is determined by the guide RNA. In some embodiments, the activity portion of the site-directed modifying polypeptide does not exhibit enzymatic activity (comprises an inactivated nuclease, e.g., via mutation). In some cases, the kit comprises a guide RNA and a site-directed modifying polypeptide. In other cases, the kit comprises: (i) a nucleic acid comprising a nucleotide sequence encoding a guide RNA; and (ii) a nucleic acid comprising a nucleotide sequence encoding site-directed modifying polypeptide. As another example, a kit can include: (i) a guide RNA, or a DNA polynucleotide encoding the same, comprising: (a) a first segment comprising a nucleotide sequence that is complementary to a sequence in a target DNA; and (b) a second segment that interacts with a site-directed modifying polypeptide; and (ii) the site-directed modifying polypeptide, or a polynucleotide encoding the same, comprising: (a) an RNA-binding portion that interacts with the guide RNA; and (b) an activity portion that modulates transcription within the target DNA, wherein the site of modulated transcription within the target DNA is determined by the guide RNA. In some cases, the kit comprises: (i) a guide RNA; and a site-directed modifying polypeptide. In other cases, the kit comprises: (i) a nucleic acid comprising a nucleotide sequence encoding a guide RNA; and (ii) a nucleic acid comprising a nucleotide sequence encoding site-directed modifying polypeptide. The present disclosure provides a kit comprising: (1) a recombinant expression vector comprising (i) a nucleotide sequence encoding a guide RNA, wherein the guide RNA comprises: (a) a first segment comprising a nucleotide sequence that is complementary to a sequence in a target DNA; and (b) a second segment that interacts with a site-directed modifying polypeptide; and (ii) a nucleotide sequence encoding the site-directed modifying polypeptide, wherein the site-directed modifying polypeptide comprises: (a) an RNA-binding portion that interacts with the guide RNA; and (b) an activity portion that exhibits site-directed enzymatic activity, wherein the site of enzymatic activity is determined by the guide RNA.; and (2) a reagent for reconstitution and/or dilution of the expression vector.

**[00343]** The present disclosure provides a kit comprising: (1) a recombinant expression vector comprising: (i) a nucleotide sequence encoding a guide RNA, wherein the guide RNA comprises: (a) a first segment comprising a nucleotide sequence that is complementary to a sequence in a target DNA; and (b) a second segment that interacts with a site-directed modifying polypeptide; and (ii) a nucleotide sequence

encoding the site-directed modifying polypeptide, wherein the site-directed modifying polypeptide comprises: (a) an RNA-binding portion that interacts with the guide RNA; and (b) an activity portion that modulates transcription within the target DNA, wherein the site of modulated transcription within the target DNA is determined by the guide RNA; and (2) a reagent for reconstitution and/or dilution of the recombinant expression vector.

**[00344]** The present disclosure provides a kit comprising: (1) a recombinant expression vector comprising a nucleic acid comprising a nucleotide sequence that encodes a DNA targeting RNA comprising: (i) a first segment comprising a nucleotide sequence that is complementary to a sequence in a target DNA; and (ii) a second segment that interacts with a site-directed modifying polypeptide; and (2) a reagent for reconstitution and/or dilution of the recombinant expression vector. In some embodiments of this kit, the kit comprises: a recombinant expression vector comprising a nucleotide sequence that encodes a site-directed modifying polypeptide, wherein the site-directed modifying polypeptide comprises: (a) an RNA-binding portion that interacts with the guide RNA; and (b) an activity portion that exhibits site-directed enzymatic activity, wherein the site of enzymatic activity is determined by the guide RNA. In other embodiments of this kit, the kit comprises: a recombinant expression vector comprising a nucleotide sequence that encodes a site-directed modifying polypeptide, wherein the site-directed modifying polypeptide comprises: (a) an RNA-binding portion that interacts with the guide RNA; and (b) an activity portion that modulates transcription within the target DNA, wherein the site of modulated transcription within the target DNA is determined by the guide RNA.

**[00345]** In some embodiments of any of the above kits, the kit comprises an activator-RNA or a targeter-RNA. In some embodiments of any of the above kits, the kit comprises a single-molecule guide RNA. In some embodiments of any of the above kits, the kit comprises two or more double-molecule or single-molecule guide RNAs. In some embodiments of any of the above kits, a guide RNA (e.g., including two or more guide RNAs) can be provided as an array (e.g., an array of RNA molecules, an array of DNA molecules encoding the guide RNA(s), etc.). Such kits can be useful, for example, for use in conjunction with the above described genetically modified host cells that comprise a site-directed modifying polypeptide. In some embodiments of any of the above kits, the kit further comprises a donor polynucleotide to effect the desired genetic modification. Components of a kit can be in separate containers; or can be combined in a single container.

**[00346]** In some cases, a kit further comprises one or more variant Cas9 site-directed polypeptides that exhibits reduced endodeoxyribonuclease activity relative to wild-type Cas9.

**[00347]** In some cases, a kit further comprises one or more nucleic acids comprising a nucleotide sequence encoding a variant Cas9 site-directed polypeptide that exhibits reduced endodeoxyribonuclease activity relative to wild-type Cas9.

**[00348]** Any of the above-described kits can further include one or more additional reagents, where such additional reagents can be selected from: a dilution buffer; a reconstitution solution; a wash buffer; a control reagent; a control expression vector or RNA polynucleotide; a reagent for *in vitro* production of the site-directed modifying polypeptide from DNA, and the like.

**[00349]** In addition to above-mentioned components, a kit can further include instructions for using the components of the kit to practice the methods. The instructions for practicing the methods are generally recorded on a suitable recording medium. For example, the instructions may be printed on a substrate, such as paper or plastic, etc. As such, the instructions may be present in the kits as a package insert, in the labeling of the container of the kit or components thereof (i.e., associated with the packaging or subpackaging) etc. In other embodiments, the instructions are present as an electronic storage data file present on a suitable computer readable storage medium, e.g. CD-ROM, diskette, flash drive, etc. In yet other embodiments, the actual instructions are not present in the kit, but means for obtaining the instructions from a remote source, e.g. via the internet, are provided. An example of this embodiment is a kit that includes a web address where the instructions can be viewed and/or from which the instructions can be downloaded. As with the instructions, this means for obtaining the instructions is recorded on a suitable substrate.

**[00350]** Non-Human Genetically Modified Organisms

**[00351]** In some embodiments, a genetically modified host cell has been genetically modified with an exogenous nucleic acid comprising a nucleotide sequence encoding a site-directed modifying polypeptide (e.g., a naturally occurring Cas9; a modified, i.e., mutated or variant, Cas9; a chimeric Cas9; etc.). If such a cell is a eukaryotic single-cell organism, then the modified cell can be considered a genetically modified organism. In some embodiments, the non-human genetically modified organism is a Cas9 transgenic multicellular organism.

**[00352]** In some embodiments, a genetically modified non-human host cell (e.g., a cell that has been genetically modified with an exogenous nucleic acid comprising a nucleotide sequence encoding a site-directed modifying polypeptide, e.g., a naturally occurring Cas9; a modified, i.e., mutated or variant, Cas9; a chimeric Cas9; etc.) can generate a genetically modified nonhuman organism (e.g., a mouse, a fish, a frog, a fly, a worm, etc.). For example, if the genetically modified host cell is a pluripotent stem cell (i.e., PSC) or a germ cell (e.g., sperm, oocyte, etc.), an entire genetically modified organism can be derived from the genetically modified host cell. In some embodiments, the genetically modified host cell is a pluripotent stem cell (e.g., ESC, iPSC, pluripotent plant stem cell, etc.) or a germ cell (e.g., sperm cell, oocyte, etc.), either *in vivo* or *in vitro*, that can give rise to a genetically modified organism. In some embodiments the genetically modified host cell is a vertebrate PSC (e.g., ESC, iPSC, etc.) and is used to generate a genetically modified organism (e.g. by injecting a PSC into a blastocyst to produce a chimeric/mosaic animal, which could then be mated to generate non-chimeric/non-mosaic genetically modified organisms; grafting in the case of

plants; etc.). Any convenient method/protocol for producing a genetically modified organism, including the methods described herein, is suitable for producing a genetically modified host cell comprising an exogenous nucleic acid comprising a nucleotide sequence encoding a site-directed modifying polypeptide (e.g., a naturally occurring Cas9; a modified, i.e., mutated or variant, Cas9; a chimeric Cas9; etc.). Methods of producing genetically modified organisms are known in the art. For example, see Cho et al., *Curr Protoc Cell Biol.* 2009 Mar;Chapter 19:Unit 19.11: Generation of transgenic mice; Gama et al., *Brain Struct Funct.* 2010 Mar;214(2-3):91-109. Epub 2009 Nov 25: Animal transgenesis: an overview; Husaini et al., *GM Crops.* 2011 Jun-Dec;2(3):150-62. Epub 2011 Jun 1: Approaches for gene targeting and targeted gene expression in plants.

**[00353]** In some embodiments, a genetically modified organism comprises a target cell for methods of the invention, and thus can be considered a source for target cells. For example, if a genetically modified cell comprising an exogenous nucleic acid comprising a nucleotide sequence encoding a site-directed modifying polypeptide (e.g., a naturally occurring Cas9; a modified, i.e., mutated or variant, Cas9; a chimeric Cas9; etc.) is used to generate a genetically modified organism, then the cells of the genetically modified organism comprise the exogenous nucleic acid comprising a nucleotide sequence encoding a site-directed modifying polypeptide (e.g., a naturally occurring Cas9; a modified, i.e., mutated or variant, Cas9; a chimeric Cas9; etc.). In some such embodiments, the DNA of a cell or cells of the genetically modified organism can be targeted for modification by introducing into the cell or cells a guide RNA (or a DNA encoding a guide RNA) and optionally a donor nucleic acid. For example, the introduction of a guide RNA (or a DNA encoding a guide RNA) into a subset of cells (e.g., brain cells, intestinal cells, kidney cells, lung cells, blood cells, etc.) of the genetically modified organism can target the DNA of such cells for modification, the genomic location of which will depend on the DNA-targeting sequence of the introduced guide RNA.

**[00354]** In some embodiments, a genetically modified organism is a source of target cells for methods of the invention. For example, a genetically modified organism comprising cells that are genetically modified with an exogenous nucleic acid comprising a nucleotide sequence encoding a site-directed modifying polypeptide (e.g., a naturally occurring Cas9; a modified, i.e., mutated or variant, Cas9; a chimeric Cas9; etc.) can provide a source of genetically modified cells, for example PSCs (e.g., ESCs, iPSCs, sperm, oocytes, etc.), neurons, progenitor cells, cardiomyocytes, etc.

**[00355]** In some embodiments, a genetically modified cell is a PSC comprising an exogenous nucleic acid comprising a nucleotide sequence encoding a site-directed modifying polypeptide (e.g., a naturally occurring Cas9; a modified, i.e., mutated or variant, Cas9; a chimeric Cas9; etc.). As such, the PSC can be a target cell such that the DNA of the PSC can be targeted for modification by introducing into the PSC a guide RNA (or a DNA encoding a guide RNA) and optionally a donor nucleic acid, and the genomic location of the modification will depend on the DNA-targeting sequence of the introduced guide RNA. Thus, in some embodiments, the methods described herein can be used to modify the DNA (e.g., delete and/or

replace any desired genomic location) of PSCs derived from a genetically modified organism. Such modified PSCs can then be used to generate organisms having both (i) an exogenous nucleic acid comprising a nucleotide sequence encoding a site-directed modifying polypeptide (e.g., a naturally occurring Cas9; a modified, i.e., mutated or variant, Cas9; a chimeric Cas9; etc.) and (ii) a DNA modification that was introduced into the PSC.

**[00356]** An exogenous nucleic acid comprising a nucleotide sequence encoding a site-directed modifying polypeptide (e.g., a naturally occurring Cas9; a modified, i.e., mutated or variant, Cas9; a chimeric Cas9; etc.) can be under the control of (i.e., operably linked to) an unknown promoter (e.g., when the nucleic acid randomly integrates into a host cell genome) or can be under the control of (i.e., operably linked to) a known promoter. Suitable known promoters can be any known promoter and include constitutively active promoters (e.g., CMV promoter), inducible promoters (e.g., heat shock promoter, Tetracycline-regulated promoter, Steroid-regulated promoter, Metal-regulated promoter, estrogen receptor-regulated promoter, etc.), spatially restricted and/or temporally restricted promoters (e.g., a tissue specific promoter, a cell type specific promoter, etc.), etc.

**[00357]** A genetically modified organism (e.g. an organism whose cells comprise a nucleotide sequence encoding a site-directed modifying polypeptide, e.g., a naturally occurring Cas9; a modified, i.e., mutated or variant, Cas9; a chimeric Cas9; etc.) can be any organism including for example, a plant; algae; an invertebrate (e.g., a cnidarian, an echinoderm, a worm, a fly, etc.); a vertebrate (e.g., a fish (e.g., zebrafish, puffer fish, gold fish, etc.), an amphibian (e.g., salamander, frog, etc.), a reptile, a bird, a mammal, etc.); an ungulate (e.g., a goat, a pig, a sheep, a cow, etc.); a rodent (e.g., a mouse, a rat, a hamster, a guinea pig); a lagomorpha (e.g., a rabbit); etc.

**[00358]** In some cases, the site-directed modifying polypeptide comprises an amino acid sequence having at least about 75%, at least about 80%, at least about 85%, at least about 90%, at least about 95%, at least about 99%, or 100%, amino acid sequence identity to amino acids 7-166 and/or 731-1003 of SEQ ID NO: 8, or to the corresponding portions in any of the amino acid sequences set forth as SEQ ID NOs: 1-800.

**[00359]** Transgenic non-human animals

**[00360]** As described above, in some embodiments, a nucleic acid (e.g., a nucleotide sequence encoding a site-directed modifying polypeptide, e.g., a naturally occurring Cas9; a modified, i.e., mutated or variant, Cas9; a chimeric Cas9; etc.) or a recombinant expression vector is used as a transgene to generate a transgenic animal that produces a site-directed modifying polypeptide. Thus, the present disclosure further provides a transgenic non-human animal, which animal comprises a transgene comprising a nucleic acid comprising a nucleotide sequence encoding a site-directed modifying polypeptide, e.g., a naturally occurring Cas9; a modified, i.e., mutated or variant, Cas9; a chimeric Cas9; etc., as described above. In some embodiments, the genome of the transgenic non-human animal



comprises a nucleotide sequence encoding a site-directed modifying polypeptide. In some embodiments, the transgenic non-human animal is homozygous for the genetic modification. In some embodiments, the transgenic non-human animal is heterozygous for the genetic modification. In some embodiments, the transgenic non-human animal is a vertebrate, for example, a fish (e.g., zebra fish, gold fish, puffer fish, cave fish, etc.), an amphibian (frog, salamander, etc.), a bird (e.g., chicken, turkey, etc.), a reptile (e.g., snake, lizard, etc.), a mammal (e.g., an ungulate, e.g., a pig, a cow, a goat, a sheep, etc.; a lagomorph (e.g., a rabbit); a rodent (e.g., a rat, a mouse); a nonhuman primate; etc.), etc.

**[00361]** An exogenous nucleic acid comprising a nucleotide sequence encoding a site-directed modifying polypeptide (e.g., a naturally occurring Cas9; a modified, i.e., mutated or variant, Cas9; a chimeric Cas9; etc.) can be under the control of (i.e., operably linked to) an unknown promoter (e.g., when the nucleic acid randomly integrates into a host cell genome) or can be under the control of (i.e., operably linked to) a known promoter. Suitable known promoters can be any known promoter and include constitutively active promoters (e.g., CMV promoter), inducible promoters (e.g., heat shock promoter, Tetracycline-regulated promoter, Steroid-regulated promoter, Metal-regulated promoter, estrogen receptor-regulated promoter, etc.), spatially restricted and/or temporally restricted promoters (e.g., a tissue specific promoter, a cell type specific promoter, etc.), etc.

**[00362]** In some cases, the site-directed modifying polypeptide comprises an amino acid sequence having at least about 75%, at least about 80%, at least about 85%, at least about 90%, at least about 95%, at least about 99%, or 100%, amino acid sequence identity to amino acids 7-166 and/or 731-1003 of SEQ ID NO: 8, or to the corresponding portions in any of the amino acid sequences set forth as SEQ ID NOs: 1-800.

**[00363]** Transgenic plants

**[00364]** As described above, in some embodiments, a nucleic acid (e.g., a nucleotide sequence encoding a site-directed modifying polypeptide, e.g., a naturally occurring Cas9; a modified, i.e., mutated or variant, Cas9; a chimeric Cas9; etc.) or a recombinant expression vector is used as a transgene to generate a transgenic plant that produces a site-directed modifying polypeptide. Thus, the present disclosure further provides a transgenic plant, which plant comprises a transgene comprising a nucleic acid comprising a nucleotide sequence encoding site-directed modifying polypeptide, e.g., a naturally occurring Cas9; a modified, i.e., mutated or variant, Cas9; a chimeric Cas9; etc., as described above. In some embodiments, the genome of the transgenic plant comprises a nucleic acid. In some embodiments, the transgenic plant is homozygous for the genetic modification. In some embodiments, the transgenic plant is heterozygous for the genetic modification.

**[00365]** Methods of introducing exogenous nucleic acids into plant cells are well known in the art. Such plant cells are considered "transformed," as defined above. Suitable methods include viral infection (such as double stranded DNA viruses), transfection, conjugation, protoplast fusion, electroporation,

particle gun technology, calcium phosphate precipitation, direct microinjection, silicon carbide whiskers technology, *Agrobacterium-mediated* transformation and the like. The choice of method is generally dependent on the type of cell being transformed and the circumstances under which the transformation is taking place (i.e. *in vitro*, *ex vivo*, or *in vivo*). Transformation methods based upon the soil bacterium *Agrobacterium tumefaciens* are particularly useful for introducing an exogenous nucleic acid molecule into a vascular plant. The wild type form of *Agrobacterium* contains a Ti (tumor-inducing) plasmid that directs production of tumorigenic crown gall growth on host plants. Transfer of the tumor-inducing T-DNA region of the Ti plasmid to a plant genome requires the Ti plasmid-encoded virulence genes as well as T-DNA borders, which are a set of direct DNA repeats that delineate the region to be transferred. An *Agrobacterium-based* vector is a modified form of a Ti plasmid, in which the tumor inducing functions are replaced by the nucleic acid sequence of interest to be introduced into the plant host.

**[00366]** *Agrobacterium-mediated* transformation generally employs cointegrate vectors or binary vector systems, in which the components of the Ti plasmid are divided between a helper vector, which resides permanently in the *Agrobacterium* host and carries the virulence genes, and a shuttle vector, which contains the gene of interest bounded by T-DNA sequences. A variety of binary vectors are well known in the art and are commercially available, for example, from Clontech (Palo Alto, Calif.). Methods of coculturing *Agrobacterium* with cultured plant cells or wounded tissue such as leaf tissue, root explants, hypocotyledons, stem pieces or tubers, for example, also are well known in the art. See., e.g., Glick and Thompson, (eds.), *Methods in Plant Molecular Biology and Biotechnology*, Boca Raton, Fla.: CRC Press (1993).

**[00367]** Microprojectile-mediated transformation also can be used to produce a transgenic plant. This method, first described by Klein et al. (*Nature* 327:70-73 (1987)), relies on microprojectiles such as gold or tungsten that are coated with the desired nucleic acid molecule by precipitation with calcium chloride, spermidine or polyethylene glycol. The microprojectile particles are accelerated at high speed into an angiosperm tissue using a device such as the BIOLISTIC PD-1000 (Biorad; Hercules Calif.).

**[00368]** A nucleic acid may be introduced into a plant in a manner such that the nucleic acid is able to enter a plant cell(s), e.g., via an *in vivo* or *ex vivo* protocol. By "*in vivo*," it is meant in the nucleic acid is administered to a living body of a plant e.g. infiltration. By "*ex vivo*" it is meant that cells or explants are modified outside of the plant, and then such cells or organs are regenerated to a plant. A number of vectors suitable for stable transformation of plant cells or for the establishment of transgenic plants have been described, including those described in Weissbach and Weissbach, (1989) *Methods for Plant Molecular Biology* Academic Press, and Gelvin et al., (1990) *Plant Molecular Biology Manual*, Kluwer Academic Publishers. Specific examples include those derived from a Ti plasmid of *Agrobacterium tumefaciens*, as well as those disclosed by Herrera-Estrella et al. (1983) *Nature* 303: 209, Bevan (1984) *Nucl Acid Res.* 12: 8711-8721, Klee (1985) *Bio/Technolo* 3: 637-642. Alternatively, non-Ti vectors can be used to transfer the DNA into plants and cells by using free DNA delivery techniques. By using these

methods transgenic plants such as wheat, rice (Christou (1991) *Bio/Technology* 9:957-9 and 4462) and corn (Gordon-Kamm (1990) *Plant Cell* 2: 603-618) can be produced. An immature embryo can also be a good target tissue for monocots for direct DNA delivery techniques by using the particle gun (Weeks et al. (1993) *Plant Physiol* 102: 1077-1084; Vasil (1993) *Bio/Technolo* 10: 667-674; Wan and Lemeaux (1994) *Plant Physiol* 104: 37-48 and for *Agrobacterium*-mediated DNA transfer (Ishida et al. (1996) *Nature Biotech* 14: 745-750). Exemplary methods for introduction of DNA into chloroplasts are biolistic bombardment, polyethylene glycol transformation of protoplasts, and microinjection (Daniell et al *Nat. Biotechnol* 16:345-348, 1998; Staub et al *Nat. Biotechnol* 18: 333-338, 2000; O'Neill et al *Plant J.* 3:729-738, 1993; Knoblauch et al *Nat. Biotechnol* 17: 906-909; U.S. Pat. Nos. 5,451,513, 5,545,817, 5,545,818, and 5,576,198; in Intl. Application No. WO 95/16783; and in Boynton et al., *Methods in Enzymology* 217: 510-536 (1993), Svab et al., *Proc. Natl. Acad. Sci. USA* 90: 913-917 (1993), and McBride et al., *Proc. Nati. Acad. Sci. USA* 91: 7301-7305 (1994)). Any vector suitable for the methods of biolistic bombardment, polyethylene glycol transformation of protoplasts and microinjection will be suitable as a targeting vector for chloroplast transformation. Any double stranded DNA vector may be used as a transformation vector, especially when the method of introduction does not utilize *Agrobacterium*.

**[00369]** Plants which can be genetically modified include grains, forage crops, fruits, vegetables, oil seed crops, palms, forestry, and vines. Specific examples of plants which can be modified follow: maize, banana, peanut, field peas, sunflower, tomato, canola, tobacco, wheat, barley, oats, potato, soybeans, cotton, carnations, sorghum, lupin and rice.

**[00370]** Also provided by the disclosure are transformed plant cells, tissues, plants and products that contain the transformed plant cells. A feature of the transformed cells, and tissues and products that include the same is the presence of a nucleic acid integrated into the genome, and production by plant cells of a site-directed modifying polypeptide, e.g., a naturally occurring Cas9; a modified, i.e., mutated or variant, Cas9; a chimeric Cas9; etc. Recombinant plant cells of the present invention are useful as populations of recombinant cells, or as a tissue, seed, whole plant, stem, fruit, leaf, root, flower, stem, tuber, grain, animal feed, a field of plants, and the like.

**[00371]** A nucleic acid comprising a nucleotide sequence encoding a site-directed modifying polypeptide (e.g., a naturally occurring Cas9; a modified, i.e., mutated or variant, Cas9; a chimeric Cas9; etc.) can be under the control of (i.e., operably linked to) an unknown promoter (e.g., when the nucleic acid randomly integrates into a host cell genome) or can be under the control of (i.e., operably linked to) a known promoter. Suitable known promoters can be any known promoter and include constitutively active promoters, inducible promoters, spatially restricted and/or temporally restricted promoters, etc.

**[00372]** In some cases, the site-directed modifying polypeptide comprises an amino acid sequence having at least about 75%, at least about 80%, at least about 85%, at least about 90%, at least about 95%, at least about 99%, or 100%, amino acid sequence identity to amino acids 7-166 and/or 731-1003 of SEQ ID NO: 8, or to the

corresponding portions in any of the amino acid sequences set forth as SEQ ID NOs: 1-800. Also provided by the disclosure is reproductive material of a transgenic plant, where reproductive material includes seeds, progeny plants and clonal material.

**[00373]** Detailed Description — Part II

**[00374]** The present disclosure provides methods of modulating transcription of a target nucleic acid in a host cell. The methods generally involve contacting the target nucleic acid with an enzymatically inactive Cas9 polypeptide and a single-guide RNA. The methods are useful in a variety of applications, which are also provided.

**[00375]** A transcriptional modulation method of the present disclosure overcomes some of the drawbacks of methods involving RNAi. A transcriptional modulation method of the present disclosure finds use in a wide variety of applications, including research applications, drug discovery (e.g., high throughput screening), target validation, industrial applications (e.g., crop engineering; microbial engineering, etc.), diagnostic applications, therapeutic applications, and imaging techniques.

**[00376]** Methods of Modulating Transcription

**[00377]** The present disclosure provides a method of selectively modulating transcription of a target DNA in a host cell. The method generally involves: a) introducing into the host cell: i) a guide RNA, or a nucleic acid comprising a nucleotide sequence encoding the guide RNA; and ii) a variant Cas9 site-directed polypeptide ("variant Cas9 polypeptide"), or a nucleic acid comprising a nucleotide sequence encoding the variant Cas9 polypeptide, where the variant Cas9 polypeptide exhibits reduced endodeoxyribonuclease activity.

**[00378]** The guide RNA (also referred to herein as "guide RNA"; or "gRNA") comprises: i) a first segment comprising a nucleotide sequence that is complementary to a target sequence in a target DNA; ii) a second segment that interacts with a site-directed polypeptide; and iii) a transcriptional terminator. The first segment, comprising a nucleotide sequence that is complementary to a target sequence in a target DNA, is referred to herein as a "targeting segment". The second segment, which interacts with a site-directed polypeptide, is also referred to herein as a "protein-binding sequence" or "dCas9-binding hairpin," or "dCas9 handle." By "segment" it is meant a segment/section/region of a molecule, e.g., a contiguous stretch of nucleotides in an RNA. The definition of "segment," unless otherwise specifically defined in a particular context, is not limited to a specific number of total base pairs, and may include regions of RNA molecules that are of any total length and may or may not include regions with complementarity to other molecules. As described above, guide RNA according to the present disclosure can be a single-molecule guide RNA or a two-molecule guide RNA. The term "guide RNA" or "gRNA" is inclusive, referring both to two-molecule guide RNAs and to single-molecule guide RNAs (i.e., sgRNAs).

**[00379]** The variant Cas9 site-directed polypeptide comprises: i) an RNA-binding portion that interacts with the guide RNA; and an activity portion that exhibits reduced endodeoxyribonuclease activity.

**[00380]** The guide RNA and the variant Cas9 polypeptide form a complex in the host cell; the complex selectively modulates transcription of a target DNA in the host cell.

**[00381]** In some cases, a transcription modulation method of the present disclosure provides for selective modulation (e.g., reduction or increase) of a target nucleic acid in a host cell. For example, "selective" reduction of transcription of a target nucleic acid reduces transcription of the target nucleic acid by at least about 10%, at least about 20%, at least about 30%, at least about 40%, at least about 50%, at least about 60%, at least about 70%, at least about 80%, at least about 90%, or greater than 90%, compared to the level of transcription of the target nucleic acid in the absence of a guide RNA/variant Cas9 polypeptide complex. Selective reduction of transcription of a target nucleic acid reduces transcription of the target nucleic acid, but does not substantially reduce transcription of a non-target nucleic acid, e.g., transcription of a non-target nucleic acid is reduced, if at all, by less than 10% compared to the level of transcription of the non-target nucleic acid in the absence of the guide RNA/variant Cas9 polypeptide complex.

**[00382]** Increased transcription

**[00383]** "Selective" increased transcription of a target DNA can increase transcription of the target DNA by at least about 1.1 fold (e.g., at least about 1.2 fold, at least about 1.3 fold, at least about 1.4 fold, at least about 1.5 fold, at least about 1.6 fold, at least about 1.7 fold, at least about 1.8 fold, at least about 1.9 fold, at least about 2 fold, at least about 2.5 fold, at least about 3 fold, at least about 3.5 fold, at least about 4 fold, at least about 4.5 fold, at least about 5 fold, at least about 6 fold, at least about 7 fold, at least about 8 fold, at least about 9 fold, at least about 10 fold, at least about 12 fold, at least about 15 fold, or at least about 20-fold) compared to the level of transcription of the target DNA in the absence of a guide RNA/variant Cas9 polypeptide complex. Selective increase of transcription of a target DNA increases transcription of the target DNA, but does not substantially increase transcription of a non-target DNA, e.g., transcription of a non-target DNA is increased, if at all, by less than about 5-fold (e.g., less than about 4-fold, less than about 3-fold, less than about 2-fold, less than about 1.8-fold, less than about 1.6-fold, less than about 1.4-fold, less than about 1.2-fold, or less than about 1.1-fold) compared to the level of transcription of the non-targeted DNA in the absence of the guide RNA/variant Cas9 polypeptide complex.

**[00384]** As a non-limiting example, increased transcription can be achieved by fusing dCas9 to a heterologous sequence. Suitable fusion partners include, but are not limited to, a polypeptide that provides an activity that indirectly increases transcription by acting directly on the target DNA or on a polypeptide (e.g., a histone or other DNA-binding protein) associated with the target DNA. Suitable fusion partners include, but are not limited to, a polypeptide that provides for methyltransferase activity, demethylase

activity, acetyltransferase activity, deacetylase activity, kinase activity, phosphatase activity, ubiquitin ligase activity, deubiquitinating activity, adenylation activity, deadenylation activity, SUMOylating activity, deSUMOylating activity, ribosylation activity, deribosylation activity, myristoylation activity, or demyristoylation activity.

**[00385]** Additional suitable fusion partners include, but are not limited to, a polypeptide that directly provides for increased transcription of the target nucleic acid (e.g., a transcription activator or a fragment thereof, a protein or fragment thereof that recruits a transcription activator, a small molecule/drug-responsive transcription regulator, etc.).

**[00386]** A non-limiting example of a method using a dCas9 fusion protein to increase transcription in a prokaryote includes a modification of the bacterial one-hybrid (B1H) or two-hybrid (B2H) system. In the B1H system, a DNA binding domain (BD) is fused to a bacterial transcription activation domain (AD, e.g., the alpha subunit of the *Escherichia coli* RNA polymerase (RNAP $\alpha$ )). Thus, a dCas9 can be fused to a heterologous sequence comprising an AD. When the dCas9 fusion protein arrives at the upstream region of a promoter (targeted there by the guide RNA) the AD (e.g., RNAP $\alpha$ ) of the dCas9 fusion protein recruits the RNAP holoenzyme, leading to transcription activation. In the B2H system, the BD is not directly fused to the AD; instead, their interaction is mediated by a protein-protein interaction (e.g., GAL11P - GAL4 interaction). To modify such a system for use in the methods, dCas9 can be fused to a first protein sequence that provides for protein-protein interaction (e.g., the yeast GAL11P and/or GAL4 protein) and RNA $\alpha$  can be fused to a second protein sequence that completes the protein-protein interaction (e.g., GAL4 if GAL11P is fused to dCas9, GAL11P if GAL4 is fused to dCas9, etc.). The binding affinity between GAL11P and GAL4 increases the efficiency of binding and transcription firing rate.

**[00387]** A non-limiting example of a method using a dCas9 fusion protein to increase transcription in a eukaryotes includes fusion of dCas9 to an activation domain (AD) (e.g., GAL4, herpesvirus activation protein VP16 or VP64, human nuclear factor NF- $\kappa$ B p65 subunit, etc.). To render the system inducible, expression of the dCas9 fusion protein can be controlled by an inducible promoter (e.g., Tet-ON, Tet-OFF, etc.). The guide RNA can be design to target known transcription response elements (e.g., promoters, enhancers, etc.), known upstream activating sequences (UAS), sequences of unknown or known function that are suspected of being able to control expression of the target DNA, etc.

**[00388]** Additional fusion partners

**[00389]** Non-limiting examples of fusion partners to accomplish increased or decreased transcription include, but are not limited to, transcription activator and transcription repressor domains (e.g., the Kriippel associated box (KRAB or SKD); the Mad mSIN3 interaction domain (SID); the ERF repressor domain (ERD), etc). In some such cases, the dCas9 fusion protein is targeted by the guide RNA to a specific location (i.e., sequence) in the target DNA and exerts locus-specific regulation such as blocking RNA polymerase binding to a promoter (which selectively inhibits transcription activator function), and/or

modifying the local chromatin status (e.g., when a fusion sequence is used that modifies the target DNA or modifies a polypeptide associated with the target DNA). In some cases, the changes are transient (e.g., transcription repression or activation). In some cases, the changes are inheritable (e.g., when epigenetic modifications are made to the target DNA or to proteins associated with the target DNA, e.g., nucleosomal histones).

**[00390]** In some embodiments, the heterologous sequence can be fused to the C-terminus of the dCas9 polypeptide. In some embodiments, the heterologous sequence can be fused to the N-terminus of the dCas9 polypeptide. In some embodiments, the heterologous sequence can be fused to an internal portion (i.e., a portion other than the N- or C- terminus) of the dCas9 polypeptide.

**[00391]** The biological effects of a method using a dCas9 fusion protein can be detected by any convenient method (e.g., gene expression assays; chromatin-based assays, e.g., Chromatin immunoPrecipitation (ChIP), Chromatin in vivo Assay (CiA), etc.; and the like).

**[00392]** In some cases, a method involves use of two or more different guide RNAs. For example, two different guide RNAs can be used in a single host cell, where the two different guide RNAs target two different target sequences in the same target nucleic acid.

**[00393]** Thus, for example, a transcriptional modulation method can further comprise introducing into the host cell a second guide RNA, or a nucleic acid comprising a nucleotide sequence encoding the second guide RNA, where the second guide RNA comprises: i) a first segment comprising a nucleotide sequence that is complementary to a second target sequence in the target DNA; ii) a second segment that interacts with the site-directed polypeptide; and iii) a transcriptional terminator. In some cases, use of two different guide RNAs targeting two different targeting sequences in the same target nucleic acid provides for increased modulation (e.g., reduction or increase) in transcription of the target nucleic acid.

**[00394]** As another example, two different guide RNAs can be used in a single host cell, where the two different guide RNAs target two different target nucleic acids. Thus, for example, a transcriptional modulation method can further comprise introducing into the host cell a second guide RNA, or a nucleic acid comprising a nucleotide sequence encoding the second guide RNA, where the second guide RNA comprises: i) a first segment comprising a nucleotide sequence that is complementary to a target sequence in at least a second target DNA; ii) a second segment that interacts with the site-directed polypeptide; and iii) a transcriptional terminator.

**[00395]** In some embodiments, a nucleic acid (e.g., a guide RNA, e.g., a single-molecule guide RNA, an activator-RNA, a targeter-RNA, etc.; a donor polynucleotide; a nucleic acid encoding a site-directed modifying polypeptide; etc.) comprises a modification or sequence that provides for an additional desirable feature (e.g., modified or regulated stability; subcellular targeting; tracking, e.g., a fluorescent label; a binding site for a protein or protein complex; etc.). Non-limiting examples include: a 5' cap (e.g., a 7-

methylguanylate cap (m<sup>7</sup>G)); a 3' polyadenylated tail (i.e., a 3' poly(A) tail); a riboswitch sequence or an aptamer sequence (e.g., to allow for regulated stability and/or regulated accessibility by proteins and/or protein complexes); a terminator sequence; a sequence that forms a dsRNA duplex (i.e., a hairpin)); a modification or sequence that targets the RNA to a subcellular location (e.g., nucleus, mitochondria, chloroplasts, and the like); a modification or sequence that provides for tracking (e.g., direct conjugation to a fluorescent molecule, conjugation to a moiety that facilitates fluorescent detection, a sequence that allows for fluorescent detection, etc.); a modification or sequence that provides a binding site for proteins (e.g., proteins that act on DNA, including transcriptional activators, transcriptional repressors, DNA methyltransferases, DNA demethylases, histone acetyltransferases, histone deacetylases, and the like); and combinations thereof.

**[00396]** DNA-targeting segment

**[00397]** The DNA-targeting segment (or "DNA-targeting sequence") of a guide RNA comprises a nucleotide sequence that is complementary to a specific sequence within a target DNA (the complementary strand of the target DNA).

**[00398]** In other words, the DNA-targeting segment of a guide RNA interacts with a target DNA in a sequence-specific manner via hybridization (i.e., base pairing). As such, the nucleotide sequence of the DNA-targeting segment may vary and determines the location within the target DNA that the guide RNA and the target DNA will interact. The DNA-targeting segment of a guide RNA can be modified (e.g., by genetic engineering) to hybridize to any desired sequence within a target DNA.

**[00399]** Stability control sequence (e.g., transcriptional terminator segment)

**[00400]** A stability control sequence influences the stability of an RNA (e.g., a guide RNA, a targeter-RNA, an activator-RNA, etc.). One example of a suitable stability control sequence is a transcriptional terminator segment (i.e., a transcription termination sequence). A transcriptional terminator segment of a guide RNA can have a total length of from about 10 nucleotides to about 100 nucleotides, e.g., from about 10 nucleotides (nt) to about 20 nt, from about 20 nt to about 30 nt, from about 30 nt to about 40 nt, from about 40 nt to about 50 nt, from about 50 nt to about 60 nt, from about 60 nt to about 70 nt, from about 70 nt to about 80 nt, from about 80 nt to about 90 nt, or from about 90 nt to about 100 nt. For example, the transcriptional terminator segment can have a length of from about 15 nucleotides (nt) to about 80 nt, from about 15 nt to about 50 nt, from about 15 nt to about 40 nt, from about 15 nt to about 30 nt or from about 15 nt to about 25 nt.

**[00401]** In some cases, the transcription termination sequence is one that is functional in a eukaryotic cell. In some cases, the transcription termination sequence is one that is functional in a prokaryotic cell.

**[00402]** Nucleotide sequences that can be included in a stability control sequence (e.g., transcriptional termination segment, or in any segment of the guide RNA to provide for increased stability)



include, for example, 5'-UAAUCCCACAGCCGCCAGUUCGCGUGGCGGCAUUUU-5' (a Rho-independent *trp* termination site).

**[00403]** Additional sequences

**[00404]** In some embodiments, a guide RNA comprises at least one additional segment at either the 5' or 3' end. For example, a suitable additional segment can comprise a 5' cap (e.g., a 7-methylguanylate cap (m<sup>7</sup>G)); a 3' polyadenylated tail (i.e., a 3' poly(A) tail); a riboswitch sequence (e.g., to allow for regulated stability and/or regulated accessibility by proteins and protein complexes); a sequence that forms a dsRNA duplex (i.e., a hairpin); a sequence that targets the RNA to a subcellular location (e.g., nucleus, mitochondria, chloroplasts, and the like); a modification or sequence that provides for tracking (e.g., direct conjugation to a fluorescent molecule, conjugation to a moiety that facilitates fluorescent detection, a sequence that allows for fluorescent detection, etc.); a modification or sequence that provides a binding site for proteins (e.g., proteins that act on DNA, including transcriptional activators, transcriptional repressors, DNA methyltransferases, DNA demethylases, histone acetyltransferases, histone deacetylases, and the like) a modification or sequence that provides for increased, decreased, and/or controllable stability; and combinations thereof.

**[00405]** Multiple simultaneous guide RNAs

**[00406]** In some embodiments, multiple guide RNAs are used simultaneously in the same cell to simultaneously modulate transcription at different locations on the same target DNA or on different target DNAs. In some embodiments, two or more guide RNAs target the same gene or transcript or locus. In some embodiments, two or more guide RNAs target different unrelated loci. In some embodiments, two or more guide RNAs target different, but related loci.

**[00407]** Because the guide RNAs are small and robust they can be simultaneously present on the same expression vector and can even be under the same transcriptional control if so desired. In some embodiments, two or more (e.g., 3 or more, 4 or more, 5 or more, 10 or more, 15 or more, 20 or more, 25 or more, 30 or more, 35 or more, 40 or more, 45 or more, or 50 or more) guide RNAs are simultaneously expressed in a target cell (from the same or different vectors). The expressed guide RNAs can be differently recognized by Cas9 proteins from different bacteria, such as *S. pyogenes*, *S. thermophilus*, *L. innocua*, and *N. meningitidis*.

**[00408]** In some cases, multiple guide RNAs can be encoded in an array mimicking naturally occurring CRISPR arrays of targeter RNAs and corresponding tracrRNAs (activator RNAs). The targeting segments are encoded as approximately 30 nucleotide long sequences (can be about 16 to about 100 nt) and are separated by CRISPR repeat sequences. In some cases, the array and tracrRNAs are introduced to a cell by DNAs encoding the RNAs. In some cases, they are introduced to the cell as RNAs.

**[00409]** To express multiple guide RNAs, an artificial RNA processing system mediated by the Csy4 endoribonuclease can be used. Multiple guide RNAs can be concatenated into a tandem array on a precursor transcript (e.g., expressed from a U6 promoter), and separated by Csy4-specific RNA sequence. Co-expressed Csy4 protein cleaves the precursor transcript into multiple guide RNAs. Advantages for using an RNA processing system include: first, there is no need to use multiple promoters; second, since all guide RNAs are processed from a precursor transcript, their concentrations are normalized for similar dCas9-binding.

**[00410]** Csy4 is a small endoribonuclease (RNase) protein derived from bacteria *Pseudomonas aeruginosa*. Csy4 specifically recognizes a minimal 17-bp RNA hairpin, and exhibits rapid (<1 min) and highly efficient (>99.9%) RNA cleavage. Unlike most RNases, the cleaved RNA fragment remains stable and functionally active. The Csy4-based RNA cleavage can be repurposed into an artificial RNA processing system. In this system, the 17-bp RNA hairpins are inserted between multiple RNA fragments that are transcribed as a precursor transcript from a single promoter. Co-expression of Csy4 is effective in generating individual RNA fragments.

**[00411]** Site-directed polypeptide

**[00412]** As noted above, a guide RNA and a variant Cas9 site-directed polypeptide form a complex. The guide RNA provides target specificity to the complex by comprising a nucleotide sequence that is complementary to a sequence of a target DNA. The variant Cas9 site-directed polypeptide has reduced endodeoxyribonuclease activity. For example, a variant Cas9 site-directed polypeptide suitable for use in a transcription modulation method of the present disclosure exhibits less than about 20%, less than about 15%, less than about 10%, less than about 5%, less than about 1%, or less than about 0.1%, of the endodeoxyribonuclease activity of a wild-type Cas9 polypeptide, e.g., a wild-type Cas9 polypeptide comprising an amino acid sequence set out in SEQ ID NO:8. In some embodiments, the variant Cas9 site-directed polypeptide has substantially no detectable endodeoxyribonuclease activity. In some embodiments when a site-directed polypeptide has reduced catalytic activity (e.g., when a SEQ ID NO: 8 *S. pyogenes* Cas9 protein has a D10, G12, G17, E762, H840, N863, H982, H983, A984, D986, and/or a A987 mutation, e.g., D10A, G12A, G17A, E762A, H840A, N863A, H982A, H983A, A984A, and/or D986A), the polypeptide can still bind to target DNA in a site-specific manner (because it is still guided to a target DNA sequence by a guide RNA) as long as it retains the ability to interact with the guide RNA.

**[00413]** In some cases, a suitable variant Cas9 site-directed polypeptide comprises an amino acid sequence having at least about 75%, at least about 80%, at least about 85%, at least about 90%, at least about 95%, at least about 99% or 100% amino acid sequence identity to amino acids 7-166 and/or 731-1003 of SEQ ID NO: 8, or to the corresponding portions in any one of the amino acid sequences SEQ ID NOs: 1-800.

**[00414]** In some cases, the variant Cas9 site-directed polypeptide is a nickase that can cleave the complementary strand of the target DNA but has reduced ability to cleave the non-complementary strand of the target DNA. For example, the variant Cas9 site-directed polypeptide can have a mutation (amino acid substitution) that reduces the function of the RuvC domain. As a non-limiting example, in some cases, the variant Cas9 site-directed polypeptide is a D10A (aspartate to alanine) mutation of SEQ ID NO: 8 (or the corresponding mutation of any of the amino acid sequences set forth in SEQ ID NOs: 1-800).

**[00415]** In some cases, the variant Cas9 site-directed polypeptide is a nickase that can cleave the non-complementary strand of the target DNA but has reduced ability to cleave the complementary strand of the target DNA. For example, the variant Cas9 site-directed polypeptide can have a mutation (amino acid substitution) that reduces the function of the HNH domain (RuvC/HNH/RuvC domain motifs, "domain 2"). As a non-limiting example, in some cases, the variant Cas9 site-directed polypeptide is a H840A (histidine to alanine at amino acid position 840 of SEQ ID NO:8) or the corresponding mutation of any of the amino acid sequences set forth in SEQ ID NOs: 1-800).

**[00416]** In some cases, the variant Cas9 site-directed polypeptide has a reduced ability to cleave both the complementary and the non-complementary strands of the target DNA. As a non-limiting example, in some cases, the variant Cas9 site-directed polypeptide harbors both D10A and H840A mutations of SEQ ID NO: 8 (or the corresponding mutations of any of the amino acid sequences set forth in SEQ ID NOs: 1-800). Other residues can be mutated to achieve the same effect (i.e. inactivate one or the other nuclease portions). As non-limiting examples, *S. pyogenes* Cas9 residues D10, G12, G17, E762, H840, N863, H982, H983, A984, D986, and/or A987 of SEQ ID NO: 8 (or the corresponding mutations of any of the proteins set forth as SEQ ID NOs: 1-800) can be altered (i.e., substituted) (see Table 1 for examples of the conservation of Cas9 amino acid residues). Also, mutations other than alanine substitutions are contemplated.

**[00417]** In some embodiments, a variant Cas9 endonuclease comprises one or more mutations corresponding to a *S. pyogenes* Cas9 mutation E762A, HH983AA or D986A in SEQ ID NO: 8. In some embodiments, the modified Cas 9 endonuclease further comprises one or more mutations corresponding to a *S. pyogenes* Cas9 mutation D10A, H840A, G12A, G17A, N854A, N863A, N982A or A984A in SEQ ID NO: 8.

**[00418]** In some cases, the variant Cas9 site-directed polypeptide is a fusion polypeptide (a "variant Cas9 fusion polypeptide"), i.e., a fusion polypeptide comprising: i) a variant Cas9 site-directed polypeptide; and ii) a covalently linked heterologous polypeptide (also referred to as a "fusion partner").

**[00419]** The heterologous polypeptide may exhibit an activity (e.g., enzymatic activity) that will also be exhibited by the variant Cas9 fusion polypeptide (e.g., methyltransferase activity, acetyltransferase

activity, kinase activity, ubiquitinating activity, etc.). A heterologous nucleic acid sequence may be linked to another nucleic acid sequence (e.g., by genetic engineering) to generate a chimeric nucleotide sequence encoding a chimeric polypeptide. In some embodiments, a variant Cas9 fusion polypeptide is generated by fusing a variant Cas9 polypeptide with a heterologous sequence that provides for subcellular localization (i.e., the heterologous sequence is a subcellular localization sequence, e.g., a nuclear localization signal (NLS) for targeting to the nucleus; a mitochondrial localization signal for targeting to the mitochondria; a chloroplast localization signal for targeting to a chloroplast; an ER retention signal; and the like). In some embodiments, the heterologous sequence can provide a tag (i.e., the heterologous sequence is a detectable label) for ease of tracking and/or purification (e.g., a fluorescent protein, e.g., green fluorescent protein (GFP), YFP, RFP, CFP, mCherry, tdTomato, and the like; a histidine tag, e.g., a 6XHis tag; a hemagglutinin (HA) tag; a FLAG tag; a Myc tag; and the like). In some embodiments, the heterologous sequence can provide for increased or decreased stability (i.e., the heterologous sequence is a stability control peptide, e.g., a degron, which in some cases is controllable (e.g., a temperature sensitive or drug controllable degron sequence, see below). In some embodiments, the heterologous sequence can provide for increased or decreased transcription from the target DNA (i.e., the heterologous sequence is a transcription modulation sequence, e.g., a transcription factor/activator or a fragment thereof, a protein or fragment thereof that recruits a transcription factor/activator, a transcription repressor or a fragment thereof, a protein or fragment thereof that recruits a transcription repressor, a small molecule/drug-responsive transcription regulator, etc.). In some embodiments, the heterologous sequence can provide a binding domain (i.e., the heterologous sequence is a protein binding sequence, e.g., to provide the ability of a chimeric dCas9 polypeptide to bind to another protein of interest, e.g., a DNA or histone modifying protein, a transcription factor or transcription repressor, a recruiting protein, etc.).

**[00420]** Suitable fusion partners that provide for increased or decreased stability include, but are not limited to degron sequences. Degrons are readily understood by one of ordinary skill in the art to be amino acid sequences that control the stability of the protein of which they are part. For example, the stability of a protein comprising a degron sequence is controlled at least in part by the degron sequence. In some cases, a suitable degron is constitutive such that the degron exerts its influence on protein stability independent of experimental control (i.e., the degron is not drug inducible, temperature inducible, etc.) In some cases, the degron provides the variant Cas9 polypeptide with controllable stability such that the variant Cas9 polypeptide can be turned "on" (i.e., stable) or "off" (i.e., unstable, degraded) depending on the desired conditions. For example, if the degron is a temperature sensitive degron, the variant Cas9 polypeptide may be functional (i.e., "on", stable) below a threshold temperature (e.g., 42°C, 41°C, 40°C, 39°C, 38°C, 37°C, 36°C, 35°C, 34°C, 33°C, 32°C, 31°C, 30°C, etc.) but non-functional (i.e., "off", degraded) above the threshold temperature. As another example, if the degron is a drug inducible degron, the presence or absence of drug can switch the protein from an "off" (i.e., unstable) state to an "on" (i.e., stable) state or vice versa. An exemplary drug inducible degron is derived from the FKBP12

protein. The stability of the degron is controlled by the presence or absence of a small molecule that binds to the degron.

**[00421]** Examples of suitable degrons include, but are not limited to those degrons controlled by Shield-1, DHFR, auxins, and/or temperature. Non-limiting examples of suitable degrons are known in the art (e.g., Dohmen et al., *Science*, 1994. 263(5151): p. 1273-1276: Heat-inducible degron: a method for constructing temperature-sensitive mutants; Schoeber et al., *Am J Physiol Renal Physiol*. 2009 Jan;296(1):F204-11 : Conditional fast expression and function of multimeric TRPV5 channels using Shield-1 ; Chu et al., *Bioorg Med Chem Lett*. 2008 Nov 15;18(22):5941-4: Recent progress with FKBP-derived destabilizing domains ; Kanemaki, *Pflugers Arch*. 2012 Dec 28: Frontiers of protein expression control with conditional degrons; Yang et al., *Mol Cell*. 2012 Nov 30;48(4):487-8: Titivated for destruction: the methyl degron; Barbour et al., *Biosci Rep*. 2013 Jan 18;33(1):. Characterization of the bipartite degron that regulates ubiquitin-independent degradation of thymidylate synthase; and Greussing et al., *J Vis Exp*. 2012 Nov 10;(69): Monitoring of ubiquitin-proteasome activity in living cells using a Degron (dgn)-destabilized green fluorescent protein (GFP)-based reporter protein; all of which are hereby incorporated in their entirety by reference).

**[00422]** Exemplary degron sequences have been well-characterized and tested in both cells and animals. Thus, fusing Cas9 to a degron sequence produces a "tunable" and "inducible" Cas9 polypeptide. Any of the fusion partners described herein can be used in any desirable combination. As one non-limiting example to illustrate this point, a Cas9 fusion protein can comprise a YFP sequence for detection, a degron sequence for stability, and transcription activator sequence to increase transcription of the target DNA. Furthermore, the number of fusion partners that can be used in a Cas9 fusion protein is unlimited. In some cases, a Cas9 fusion protein comprises one or more (e.g. two or more, three or more, four or more, or five or more) heterologous sequences.

**[00423]** Suitable fusion partners include, but are not limited to, a polypeptide that provides for methyltransferase activity, demethylase activity, acetyltransferase activity, deacetylase activity, kinase activity, phosphatase activity, ubiquitin ligase activity, deubiquitinating activity, adenylation activity, deadenylation activity, SUMOylating activity, deSUMOylating activity, ribosylation activity, deribosylation activity, myristoylation activity, or demyristoylation activity, any of which can be directed at modifying the DNA directly (e.g., methylation of DNA) or at modifying a DNA-associated polypeptide (e.g., a histone or DNA binding protein). Further suitable fusion partners include, but are not limited to boundary elements (e.g., CTCF), proteins and fragments thereof that provide periphery recruitment (e.g., Lamin A, Lamin B, etc.), and protein docking elements (e.g., FKBP/FRB, Pil 1/Aby 1, etc.).

**[00424]** In some embodiments, a site-directed modifying polypeptide can be codon-optimized. This type of optimization is known in the art and entails the mutation of foreign-derived DNA to mimic the codon preferences of the intended host organism or cell while encoding the same protein. Thus, the

codons are changed, but the encoded protein remains unchanged. For example, if the intended target cell was a human cell, a human codon-optimized dCas9 (or dCas9 variant) would be a suitable site-directed modifying polypeptide. As another non-limiting example, if the intended host cell were a mouse cell, than a mouse codon-optimized Cas9 (or variant, e.g., enzymatically inactive variant) would be a suitable Cas9 site-directed polypeptide. While codon optimization is not required, it is acceptable and may be preferable in certain cases.

**[00425]** Polyadenylation signals can also be chosen to optimize expression in the intended host.

**[00426]** Host cells

**[00427]** A method of the present disclosure to modulate transcription may be employed to induce transcriptional modulation in mitotic or post-mitotic cells *in vivo* and/or *ex vivo* and/or *in vitro*. Because the guide RNA provides specificity by hybridizing to target DNA, a mitotic and/or post-mitotic cell can be any of a variety of host cell, where suitable host cells include, but are not limited to, a bacterial cell; an archaeal cell; a single-celled eukaryotic organism; a plant cell; an algal cell, e.g., *Botryococcus braunii*, *Chlamydomonas reinhardtii*, *Nannochloropsis gaditana*, *Chlorella pyrenoidosa*, *Sargassum patens*, *C. agardh*, and the like; a fungal cell; an animal cell; a cell from an invertebrate animal (e.g., an insect, a cnidarian, an echinoderm, a nematode, etc.); a eukaryotic parasite (e.g., a malarial parasite, e.g., *Plasmodium falkiparum*; a helminth; etc.); a cell from a vertebrate animal (e.g., fish, amphibian, reptile, bird, mammal); a mammalian cell, e.g., a rodent cell, a human cell, a non-human primate cell, etc. Suitable host cells include naturally-occurring cells; genetically modified cells (e.g., cells genetically modified in a laboratory, e.g., by the "hand of man"); and cells manipulated *in vitro* in any way. In some cases, a host cell is isolated.

**[00428]** Any type of cell may be of interest (e.g. a stem cell, e.g. an embryonic stem (ES) cell, an induced pluripotent stem (iPS) cell, a germ cell; a somatic cell, e.g. a fibroblast, a hematopoietic cell, a neuron, a muscle cell, a bone cell, a hepatocyte, a pancreatic cell; an *in vitro* or *in vivo* embryonic cell of an embryo at any stage, e.g., a 1-cell, 2-cell, 4-cell, 8-cell, etc. stage zebrafish embryo; etc.). Cells may be from established cell lines or they may be primary cells, where "primary cells", "primary cell lines", and "primary cultures" are used interchangeably herein to refer to cells and cells cultures that have been derived from a subject and allowed to grow *in vitro* for a limited number of passages, i.e. splittings, of the culture. For example, primary cultures include cultures that may have been passaged 0 times, 1 time, 2 times, 4 times, 5 times, 10 times, or 15 times, but not enough times go through the crisis stage. Primary cell lines can be are maintained for fewer than 10 passages *in vitro*. Target cells are in many embodiments unicellular organisms, or are grown in culture.

**[00429]** If the cells are primary cells, such cells may be harvest from an individual by any convenient method. For example, leukocytes may be conveniently harvested by apheresis, leukocytapheresis, density gradient separation, etc., while cells from tissues such as skin, muscle, bone marrow, spleen,

liver, pancreas, lung, intestine, stomach, etc. are most conveniently harvested by biopsy. An appropriate solution may be used for dispersion or suspension of the harvested cells. Such solution will generally be a balanced salt solution, e.g. normal saline, phosphate-buffered saline (PBS), Hank's balanced salt solution, etc., conveniently supplemented with fetal calf serum or other naturally occurring factors, in conjunction with an acceptable buffer at low concentration, e.g., from 5-25 mM. Convenient buffers include HEPES, phosphate buffers, lactate buffers, etc. The cells may be used immediately, or they may be stored, frozen, for long periods of time, being thawed and capable of being reused. In such cases, the cells will usually be frozen in 10% dimethyl sulfoxide (DMSO), 50% serum, 40% buffered medium, or some other such solution as is commonly used in the art to preserve cells at such freezing temperatures, and thawed in a manner as commonly known in the art for thawing frozen cultured cells.

**[00430]** Introducing nucleic acid into a host cell

**[00431]** A guide RNA, or a nucleic acid comprising a nucleotide sequence encoding same, can be introduced into a host cell by any of a variety of well-known methods. Similarly, where a method involves introducing into a host cell a nucleic acid comprising a nucleotide sequence encoding a variant Cas9 site-directed polypeptide, such a nucleic acid can be introduced into a host cell by any of a variety of well-known methods.

**[00432]** Methods of introducing a nucleic acid into a host cell are known in the art, and any known method can be used to introduce a nucleic acid (e.g., an expression construct) into a stem cell or progenitor cell. Suitable methods include, include e.g., viral or bacteriophage infection, transfection, conjugation, protoplast fusion, lipofection, electroporation, calcium phosphate precipitation, polyethyleneimine (PEI)-mediated transfection, DEAE-dextran mediated transfection, liposome-mediated transfection, particle gun technology, calcium phosphate precipitation, direct micro injection, nanoparticle-mediated nucleic acid delivery (see, e.g., Panyam et., al *Adv Drug Deliv Rev.* 2012 Sep 13. pii: 50169-409X(12)00283-9. doi: 10.1016/j.addr.2012.09.023 ), and the like.

**[00433]** Nucleic Acids

**[00434]** The present disclosure provides an isolated nucleic acid comprising a nucleotide sequence encoding a guide RNA. In some cases, a nucleic acid also comprises a nucleotide sequence encoding a variant Cas9 site-directed polypeptide.

**[00435]** In some embodiments, a method involves introducing into a host cell (or a population of host cells) one or more nucleic acids comprising nucleotide sequences encoding a guide RNA and/or a variant Cas9 site-directed polypeptide. In some embodiments a cell comprising a target DNA is *in vitro*. In some embodiments a cell comprising a target DNA is *in vivo*. Suitable nucleic acids comprising nucleotide sequences encoding a guide RNA and/or a site-directed polypeptide include expression vectors, where

an expression vector comprising a nucleotide sequence encoding a guide RNA and/or a site-directed polypeptide is a "recombinant expression vector."

**[00436]** In some embodiments, the recombinant expression vector is a viral construct, e.g., a recombinant adeno-associated virus construct (see, e.g., U.S. Patent No. 7,078,387), a recombinant adenoviral construct, a recombinant lentiviral construct, a recombinant retroviral construct, etc.

Suitable expression vectors include, but are not limited to, viral vectors (e.g. viral vectors based on vaccinia virus; poliovirus; adenovirus (see, e.g., Li et al., *Invest Ophthalmol Vis Sci* 35:2543-2549, 1994; Borrás et al., *Gene Ther* 6:515-524, 1999; Li and Davidson, *PNAS* 92:7700-7704, 1995; Sakamoto et al., *H Gene Ther* 5:1088-1097, 1999; WO 94/12649, WO 93/03769; WO 93/19191; WO 94/28938; WO 95/11984 and WO 95/00655); adeno-associated virus (see, e.g., Ali et al., *Hum Gene Ther* 9:81-86, 1998; Flannery et al., *PNAS* 94:6916-6921, 1997; Bennett et al., *Invest Ophthalmol Vis Sci* 38:2857-2863, 1997; Jomary et al., *Gene Ther* 4:683-690, 1997; Rolling et al., *Hum Gene Ther* 10:641-648, 1999; Ali et al., *Hum Mol Genet* 5:591-594, 1996; Srivastava in WO 93/09239, Samulski et al., *J. Vir.* (1989) 63:3822-3828; Mendelson et al., *Viol.* (1988) 166:154-165; and Flotte et al., *PNAS* (1993) 90:10613-10617); SV40; herpes simplex virus; human immunodeficiency virus (see, e.g., Miyoshi et al., *PNAS* 94:10319-23, 1997; Takahashi et al., *J Virol* 73:7812-7816, 1999); a retroviral vector (e.g., Murine Leukemia Virus, spleen necrosis virus, and vectors derived from retroviruses such as Rous Sarcoma Virus, Harvey Sarcoma Virus, avian leukosis virus, a lentivirus, human immunodeficiency virus, myeloproliferative sarcoma virus, and mammary tumor virus); and the like.

**[00437]** Numerous suitable expression vectors are known to those of skill in the art, and many are commercially available. The following vectors are provided by way of example; for eukaryotic host cells: pXT1, pSG5 (Stratagene), pSVK3, pBPV, pMSG, and pSVLSV40 (Pharmacia). However, any other vector may be used so long as it is compatible with the host cell.

**[00438]** Depending on the host/vector system utilized, any of a number of suitable transcription and translation control elements, including constitutive and inducible promoters, transcription enhancer elements, transcription terminators, etc. may be used in the expression vector (see e.g., Bitter et al. (1987) *Methods in Enzymology*, 153:516-544).

**[00439]** In some embodiments, a nucleotide sequence encoding a guide RNA and/or a variant Cas9 site-directed polypeptide is operably linked to a control element, e.g., a transcriptional control element, such as a promoter. The transcriptional control element may be functional in either a eukaryotic cell, e.g., a mammalian cell; or a prokaryotic cell (e.g., bacterial or archaeal cell). In some embodiments, a nucleotide sequence encoding a guide RNA and/or a variant Cas9 site-directed polypeptide is operably linked to multiple control elements that allow expression of the nucleotide sequence encoding a guide RNA and/or a variant Cas9 site-directed polypeptide in both prokaryotic and eukaryotic cells.



**[00440]** A promoter can be a constitutively active promoter (i.e., a promoter that is constitutively in an active/"ON" state), it may be an inducible promoter (i.e., a promoter whose state, active/"ON" or inactive/"OFF", is controlled by an external stimulus, e.g., the presence of a particular temperature, compound, or protein.), it may be a spatially restricted promoter (i.e., transcriptional control element, enhancer, etc.)(e.g., tissue specific promoter, cell type specific promoter, etc.), and it may be a temporally restricted promoter (i.e., the promoter is in the "ON" state or "OFF" state during specific stages of embryonic development or during specific stages of a biological process, e.g., hair follicle cycle in mice).

**[00441]** Suitable promoters can be derived from viruses and can therefore be referred to as viral promoters, or they can be derived from any organism, including prokaryotic or eukaryotic organisms. Suitable promoters can be used to drive expression by any RNA polymerase (e.g., pol I, pol II, pol III). Exemplary promoters include, but are not limited to the SV40 early promoter, mouse mammary tumor virus long terminal repeat (LTR) promoter; adenovirus major late promoter (Ad MLP); a herpes simplex virus (HSV) promoter, a cytomegalovirus (CMV) promoter such as the CMV immediate early promoter region (CMVIE), a rous sarcoma virus (RSV) promoter, a human U6 small nuclear promoter (U6) (Miyagishi et al. , Nature Biotechnology 20, 497 - 500 (2002)), an enhanced U6 promoter (e.g., Xia et al., Nucleic Acids Res. 2003 Sep 1;31(17)), a human H1 promoter (H1), and the like.

**[00442]** Examples of inducible promoters include, but are not limited to T7 RNA polymerase promoter, T3 RNA polymerase promoter, Isopropyl-beta-D-thiogalactopyranoside (IPTG)-regulated promoter, lactose induced promoter, heat shock promoter, Tetracycline-regulated promoter (e.g., Tet-ON, Tet-OFF, etc.), Steroid-regulated promoter, Metal-regulated promoter, estrogen receptor-regulated promoter, etc. Inducible promoters can therefore be regulated by molecules including, but not limited to, doxycycline; RNA polymerase, e.g., T7 RNA polymerase; an estrogen receptor; an estrogen receptor fusion; etc.

**[00443]** In some embodiments, the promoter is a spatially restricted promoter (i.e., cell type specific promoter, tissue specific promoter, etc.) such that in a multi-cellular organism, the promoter is active (i.e., "ON") in a subset of specific cells. Spatially restricted promoters may also be referred to as enhancers, transcriptional control elements, control sequences, etc. Any convenient spatially restricted promoter may be used and the choice of suitable promoter (e.g., a brain specific promoter, a promoter that drives expression in a subset of neurons, a promoter that drives expression in the germline, a promoter that drives expression in the lungs, a promoter that drives expression in muscles, a promoter that drives expression in islet cells of the pancreas, etc.) will depend on the organism. For example, various spatially restricted promoters are known for plants, flies, worms, mammals, mice, etc. Thus, a spatially restricted promoter can be used to regulate the expression of a nucleic acid encoding a site-directed polypeptide in a wide variety of different tissues and cell types, depending on the organism. Some spatially restricted promoters are also temporally restricted such that the promoter is in the "ON" state or "OFF" state during specific stages of embryonic development or during specific stages of a biological process (e.g., hair follicle cycle in mice).

**[00444]** For illustration purposes, examples of spatially restricted promoters include, but are not limited to, neuron-specific promoters, adipocyte-specific promoters, cardiomyocyte-specific promoters, smooth muscle-specific promoters, photoreceptor-specific promoters, etc. Neuron-specific spatially restricted promoters include, but are not limited to, a neuron-specific enolase (NSE) promoter (see, e.g., EMBL HSENO2, X51956); an aromatic amino acid decarboxylase (AADC) promoter; a neurofilament promoter (see, e.g., GenBank HUMNFL, L04147); a synapsin promoter (see, e.g., GenBank HUMSYNIB, M55301); a thy-1 promoter (see, e.g., Chen et al. (1987) *Cell* 51:7-19; and Llewellyn, et al. (2010) *Nat. Med.* 16(10):1161-1166); a serotonin receptor promoter (see, e.g., GenBank S62283); a tyrosine hydroxylase promoter (TH) (see, e.g., Oh et al. (2009) *Gene Ther* 16:437; Sasaoka et al. (1992) *Mol. Brain Res.* 16:274; Boundy et al. (1998) *J. Neurosci.* 18:9989; and Kaneda et al. (1991) *Neuron* 6:583-594); a GnRH promoter (see, e.g., Radovick et al. (1991) *Proc. Natl. Acad. Sci. USA* 88:3402-3406); an L7 promoter (see, e.g., Oberdick et al. (1990) *Science* 248:223-226); a DNMT promoter (see, e.g., Bartge et al. (1988) *Proc. Natl. Acad. Sci. USA* 85:3648-3652); an enkephalin promoter (see, e.g., Comb et al. (1988) *EMBO J.* 17:3793-3805); a myelin basic protein (MBP) promoter; a Ca<sup>2+</sup>-calmodulin-dependent protein kinase II-alpha (CamKIIa) promoter (see, e.g., Mayford et al. (1996) *Proc. Natl. Acad. Sci. USA* 93:13250; and Casanova et al. (2001) *Genesis* 31:37); a CMV enhancer/platelet-derived growth factor-0 promoter (see, e.g., Liu et al. (2004) *Gene Therapy* 11:52-60); and the like.

**[00445]** Adipocyte-specific spatially restricted promoters include, but are not limited to aP2 gene promoter/enhancer, e.g., a region from -5.4 kb to +21 bp of a human aP2 gene (see, e.g., Tozzo et al. (1997) *Endocrinol.* 138:1604; Ross et al. (1990) *Proc. Natl. Acad. Sci. USA* 87:9590; and Pavjani et al. (2005) *Nat. Med.* 11:797); a glucose transporter-4 (GLUT4) promoter (see, e.g., Knight et al. (2003) *Proc. Natl. Acad. Sci. USA* 100:14725); a fatty acid translocase (FAT/CD36) promoter (see, e.g., Kuriki et al. (2002) *Biol. Pharm. Bull.* 25:1476; and Sato et al. (2002) *J. Biol. Chem.* 277:15703); a stearyl-CoA desaturase-1 (SCD1) promoter (Tabor et al. (1999) *J. Biol. Chem.* 274:20603); a leptin promoter (see, e.g., Mason et al. (1998) *Endocrinol.* 139:1013; and Chen et al. (1999) *Biochem. Biophys. Res. Comm.* 262:187); an adiponectin promoter (see, e.g., Kita et al. (2005) *Biochem. Biophys. Res. Comm.* 331:484; and Chakrabarti (2010) *Endocrinol.* 151:2408); an adipsin promoter (see, e.g., Platt et al. (1989) *Proc. Natl. Acad. Sci. USA* 86:7490); a resistin promoter (see, e.g., Seo et al. (2003) *Molec. Endocrinol.* 17:1522); and the like.

**[00446]** Cardiomyocyte-specific spatially restricted promoters include, but are not limited to control sequences derived from the following genes: myosin light chain-2, a-myosin heavy chain, AE3, cardiac troponin C, cardiac actin, and the like. Franz et al. (1997) *Cardiovasc. Res.* 35:560-566; Robbins et al. (1995) *Ann. N.Y. Acad. Sci.* 752:492-505; Linn et al. (1995) *Circ. Res.* 76:584591; Parmacek et al. (1994) *Mol. Cell. Biol.* 14:1870-1885; Hunter et al. (1993) *Hypertension* 22:608-617; and Sartorelli et al. (1992) *Proc. Natl. Acad. Sci. USA* 89:4047-4051.

**[00447]** Smooth muscle-specific spatially restricted promoters include, but are not limited to an SM22a promoter (see, e.g., Akyilrek et al. (2000) *Mol. Med.* 6:983; and U.S. Patent No. 7,169,874); a smoothelin promoter (see, e.g., WO 2001/018048); an  $\alpha$ -smooth muscle actin promoter; and the like. For example, a 0.4 kb region of the SM22a promoter, within which lie two CArG elements, has been shown to mediate vascular smooth muscle cell-specific expression (see, e.g., Kim, et al. (1997) *Mol. Cell. Biol.* 17, 2266-2278; Li, et al., (1996) *J. Cell Biol.* 132, 849-859; and Moessler, et al. (1996) *Development* 122, 2415-2425).

**[00448]** Photoreceptor-specific spatially restricted promoters include, but are not limited to, a rhodopsin promoter; a rhodopsin kinase promoter (Young et al. (2003) *Ophthalmol. Vis. Sci.* 44:4076); a beta phosphodiesterase gene promoter (Nicoud et al. (2007) *J. Gene Med.* 9:1015); a retinitis pigmentosa gene promoter (Nicoud et al. (2007) *supra*); an interphotoreceptor retinoid-binding protein (IRBP) gene enhancer (Nicoud et al. (2007) *supra*); an IRBP gene promoter (Yokoyama et al. (1992) *Exp Eye Res.* 55:225); and the like.

**[00449]** Libraries

**[00450]** The present disclosure provides a library of guide RNAs. The present disclosure provides a library of nucleic acids comprising nucleotides encoding guide RNAs. A library of nucleic acids comprising nucleotides encoding guide RNAs can comprises a library of recombinant expression vectors comprising nucleotides encoding the guide RNAs.

**[00451]** A library can comprise from about 10 individual members to about  $10^{12}$  individual members; e.g., a library can comprise from about 10 individual members to about  $10^2$  individual members, from about  $10^2$  individual members to about  $10^3$  individual members, from about  $10^3$  individual members to about  $10^5$  individual members, from about  $10^5$  individual members to about  $10^7$  individual members, from about  $10^7$  individual members to about  $10^9$  individual members, or from about  $10^9$  individual members to about  $10^{12}$  individual members.

**[00452]** An "individual member" of a library differs from other members of the library in the nucleotide sequence of the DNA targeting segment of the guide RNA. Thus, e.g., each individual member of a library can comprise the same or substantially the same nucleotide sequence of the protein-binding segment as all other members of the library; and can comprise the same or substantially the same nucleotide sequence of the transcriptional termination segment as all other members of the library; but differs from other members of the library in the nucleotide sequence of the DNA targeting segment of the guide RNA. In this way, the library can comprise members that bind to different target nucleic acids.

**[00453]** Uses

**[00454]** A method for modulating transcription according to the present disclosure finds use in a variety of applications, which are also provided. Applications include research applications; diagnostic applications; industrial applications; and treatment applications.

**[00455]** Research applications include, e.g., determining the effect of reducing or increasing transcription of a target nucleic acid on, e.g., development, metabolism, expression of a downstream gene, and the like.

**[00456]** High through-put genomic analysis can be carried out using a transcription modulation method, in which only the DNA-targeting segment of the guide RNA needs to be varied, while the protein-binding segment and the transcription termination segment can (in some cases) be held constant. A library (e.g., a library) comprising a plurality of nucleic acids used in the genomic analysis would include: a promoter operably linked to a guide RNA-encoding nucleotide sequence, where each nucleic acid would include a different DNA-targeting segment, a common protein-binding segment, and a common transcription termination segment. A chip could contain over  $5 \times 10^4$  unique guide RNAs. Applications would include large-scale phenotyping, gene-to-function mapping, and meta-genomic analysis.

**[00457]** The methods disclosed herein find use in the field of metabolic engineering. Because transcription levels can be efficiently and predictably controlled by designing an appropriate guide RNA, as disclosed herein, the activity of metabolic pathways (e.g., biosynthetic pathways) can be precisely controlled and tuned by controlling the level of specific enzymes (e.g., via increased or decreased transcription) within a metabolic pathway of interest. Metabolic pathways of interest include those used for chemical (fine chemicals, fuel, antibiotics, toxins, agonists, antagonists, etc.) and/or drug production.

**[00458]** Biosynthetic pathways of interest include but are not limited to (1) the mevalonate pathway (e.g., HMG-CoA reductase pathway) (converts acetyl-CoA to dimethylallyl pyrophosphate (DMAPP) and isopentenyl pyrophosphate (IPP), which are used for the biosynthesis of a wide variety of biomolecules including terpenoids/isoprenoids), (2) the non-mevalonate pathway (i.e., the "2-C-methyl-D-erythritol 4-phosphate/1-deoxy-D-xylulose 5-phosphate pathway" or "MEP/DOXP pathway" or "DXP pathway")(also produces DMAPP and IPP, instead by converting pyruvate and glyceraldehyde 3-phosphate into DMAPP and IPP via an alternative pathway to the mevalonate pathway), (3) the polyketide synthesis pathway (produces a variety of polyketides via a variety of polyketide synthase enzymes. Polyketides include naturally occurring small molecules used for chemotherapy (e. g., tetracyclin, and macrolides) and industrially important polyketides include rapamycin (immunosuppressant), erythromycin (antibiotic), lovastatin (anticholesterol drug), and epothilone B (anticancer drug)), (4) fatty acid synthesis pathways, (5) the DAHP (3-deoxy-D-arabino-heptulosonate 7-phosphate) synthesis pathway, (6) pathways that produce potential biofuels (such as short-chain alcohols and alkane, fatty acid methyl esters and fatty alcohols, isoprenoids, etc.), etc.

**[00459]** Networks and Cascades

**[00460]** The methods disclosed herein can be used to design integrated networks (i.e., a cascade or cascades) of control. For example, a guide RNA / variant Cas9 site-directed polypeptide may be used to control (i.e., modulate, e.g., increase, decrease) the expression of another DNA-targeting RNA or another variant Cas9 site-directed polypeptide. For example, a first guide RNA may be designed to target the modulation of transcription of a second chimeric dCas9 polypeptide with a function that is different than the first variant Cas9 site-directed polypeptide (e.g., methyltransferase activity, demethylase activity, acetyltransferase activity, deacetylase activity, etc.). In addition, because different dCas9 proteins (e.g., derived from different species) may require a different Cas9 handle (i.e., protein binding segment), the second chimeric dCas9 polypeptide can be derived from a different species than the first dCas9 polypeptide above. Thus, in some cases, the second chimeric dCas9 polypeptide can be selected such that it may not interact with the first guide RNA. In other cases, the second chimeric dCas9 polypeptide can be selected such that it does interact with the first guide RNA. In some such cases, the activities of the two (or more) dCas9 proteins may compete (e.g., if the polypeptides have opposing activities) or may synergize (e.g., if the polypeptides have similar or synergistic activities). Likewise, as noted above, any of the complexes (i.e., guide RNA / dCas9 polypeptide) in the network can be designed to control other guide RNAs or dCas9 polypeptides. Because a guide RNA and variant Cas9 site-directed polypeptide can be targeted to any desired DNA sequence, the methods described herein can be used to control and regulate the expression of any desired target. The integrated networks (i.e., cascades of interactions) that can be designed range from very simple to very complex, and are without limit.

**[00461]** In a network wherein two or more components (e.g., guide RNAs, activator-RNAs, targeter-RNAs, or dCas9 polypeptides) are each under regulatory control of another guide RNA/dCas9 polypeptide complex, the level of expression of one component of the network may affect the level of expression (e.g., may increase or decrease the expression) of another component of the network. Through this mechanism, the expression of one component may affect the expression of a different component in the same network, and the network may include a mix of components that increase the expression of other components, as well as components that decrease the expression of other components. As would be readily understood by one of skill in the art, the above examples whereby the level of expression of one component may affect the level of expression of one or more different component(s) are for illustrative purposes, and are not limiting. An additional layer of complexity may be optionally introduced into a network when one or more components are modified (as described above) to be manipulable (i.e., under experimental control, e.g., temperature control; drug control, i.e., drug inducible control; light control; etc.).

**[00462]** As one non-limiting example, a first guide RNA can bind to the promoter of a second guide RNA, which controls the expression of a target therapeutic/metabolic gene. In such a case, conditional expression of the first guide RNA indirectly activates the therapeutic/metabolic gene. RNA cascades of

this type are useful, for example, for easily converting a repressor into an activator, and can be used to control the logics or dynamics of expression of a target gene.

[00463] A transcription modulation method can also be used for drug discovery and target validation.

### Examples

[00464] Various aspects of the invention make use of the following materials and methods and are illustrated by the following non-limiting examples, wherein Example 1 relates to Cas9 orthologs, Example 2 relates to exchangeability of bacterial RNase III enzymes, Example 3 relates to the Cas9 HNH and RuvC domains, Example 4 relates to exchangeability of Cas9 endonucleases in tracrRNA-directed pre-crRNA maturation by RNase III, Example 5 relates to PAMs of Cas9 orthologs and Example 6 relates to exchangeability of guide RNA and Cas9 endonucleases.

### Materials and Methods

[00465] Bacterial strains and culture conditions

[00466] Supplementary Table S1 lists bacterial strains used in this study. *S. pyogenes*, *Streptococcus mutans*, *Campylobacter jejuni*, *N. meningitidis*, *Escherichia coli* and *Francisella novicida* were grown as previously described (15,16). BHI (Brain Heart Infusion, Becton Dickinson) agar and BHI broth medium supplemented with 1% glucose and 1% lactose were used to culture *S. thermophilus* at 42°C in a 5% CO<sub>2</sub> environment (16). *Pasteurella multocida* and *Staphylococcus aureus* were grown at 37°C on BHI agar plates and in BHI broth with shaking. Cell growth was monitored by measuring the optical density of cultures at 620 nm (OD<sub>620</sub>) using a microplate reader (BioTek PowerWave).

[00467] Bacterial transformation

[00468] *E. coli* was transformed with plasmid DNA according to standard protocols (35).

Transformation of *S. pyogenes* was performed as previously described (36) with some modifications. *S. pyogenes* pre-cultures were diluted 1:100 in fresh THY medium and grown at 37°C, 5% CO<sub>2</sub> until OD<sub>620</sub> reached 0.3. Glycine was added to the medium to 10% final concentration and growth was maintained for an additional hour. Cells were spun down at 4°C at 2500 x g and washed three times with electroporation buffer (5 mM KH<sub>2</sub>PO<sub>4</sub>, 0.4 M D-sorbitol, 10% glycerol, pH 4.5), finally suspended in the same buffer and equalized to the same OD<sub>620</sub>. For electroporation, 1 µg of plasmid was incubated with the competent cells on ice for 10 min. The conditions were 25 µF, 600 Ω and 1.5 V using 1 mm electroporation cuvettes (Biorad). After a regeneration time of 3 h, bacteria were spread on agar medium supplemented with kanamycin (300 µg/ml). Transformation assays were performed at least three times independently with technical triplicates. The efficiencies were calculated as CFU (colony-forming units) per µg of plasmid

DNA. Positive and negative control transformations were done with backbone plasmid pEC85 and sterile H<sub>2</sub>O, respectively.

**[00469]** DNA manipulations

**[00470]** DNA manipulations including DNA preparation (QIAprep Spin MiniPrep Kit, Qiagen), PCR (*Phusion*® High-Fidelity DNA *Polymerase*, Finnzyme), DNA digestion (restriction enzymes, Fermentas), DNA ligation (T4 DNA ligase, Fermentas), DNA purification (QIAquick PCR Purification Kit, Qiagen) and agarose gel electrophoresis were performed according to standard techniques or manufacturers' protocols with some modifications (35). Site-directed mutagenesis was done using QuikChange II XL kit (Stratagene) or PCR-based mutagenesis (37). Synthetic oligonucleotides (Sigma-Aldrich & Biomers) and plasmids used and generated in this study are listed in Supplementary Table S1. The integrity of all constructed plasmids was verified by enzymatic digestion and sequencing at LGC Genomics.

**[00471]** Construction of plasmids for complementation studies in *S. pyogenes*

**[00472]** The backbone shuttle vector pEC85 was used for complementation study (38,39). The RNase-III encoding genes (*rnc* genes) of *S. pyogenes*, *S. mutans*, *S. thermophilus*, *C. jejuni*, *N. meningitidis*, *P. multocida*, *F. novicida*, *E. coli* and *S. aureus*, and the genes encoding truncated and inactive RNase III variants (truncated and inactive (D51A) *rnc* mutants) of *S. pyogenes* were cloned in pEC483 (pEC85 containing the native promoter of *S. pyogenes rnc*) using NcoI and EcoRI restriction sites (Supplementary Table S1, Supplementary Figure S6). The ortholog and mutant *cas9* genes were cloned in pEC342 (pEC85 containing a sequence encoding *tracrRNA*-171 nt (16) and the native promoter of the *S. pyogenes cas* operon) using Sall and SmaI restriction sites (Supplementary Table S1). Note that in a previous study, we observed low abundance of *tracrRNA* in the *cas9* deletion mutant. For this reason, plasmids used in *cas9* complementation studies were designed to encode *tracrRNA* in addition to *cas9* (16). The generated *rnc* and *cas9* recombinant plasmids were introduced in *S. pyogenes*  $\Delta rnc$  and  $\Delta cas9$  deletion strains, respectively (Supplementary Table S1). Plasmid integrity in all complemented strains was checked by plasmid DNA extraction and digestion.

**[00473]** Construction of plasmids for transformation studies in *S. pyogenes*

**[00474]** Plasmid pEC85 was used as backbone vector for transformation studies. A DNA fragment containing WT *speM* protospacer sequence was cloned in the PstI site of plasmids containing coding sequences of WT or mutated *cas9* from *S. pyogenes* (Supplementary Table S1).

**[00475]** Construction of plasmids for protein purification

**[00476]** The overexpression vector pET16b (Novagen) was modified by inserting three additional restriction sites (Sall, SacI, NotI) into the NdeI restriction site, generating pEC621. The genes coding for the orthologous Cas9 proteins were PCR amplified from genomic DNA of the corresponding strains using

primers containing a Sall and a NotI restriction site (Supplementary Table S1). The *S. pyogenes cas9* mutant genes were PCR amplified from the complementation plasmids mentioned above. All orthologous and mutant *cas9* genes were cloned into the Sall and NotI sites of pEC621.

**[00477]** Construction of substrate plasmids for *in vitro* cleavage assays

**[00478]** Plasmid pEC287 that contains the *speM* protospacer sequence was used as a vector to construct all substrate plasmids. The PAM sequence located in 3' just next to the crRNA-targeted sequence of the *speM* protospacer (GGG on this plasmid) was modified by PCR-mediated site-directed mutagenesis (37) using one standard oligonucleotide (OLEC 3140 or OLEC3194) that either introduced or removed a XbaI restriction site for screening purposes, and a second mutagenic oligonucleotide to exchange the protospacer adjacent sequence (Supplementary Table S1).

**[00479]** RNA preparation

**[00480]** Total RNA from *S. pyogenes* SF370 WT, deletion mutants and complemented strains was prepared from culture samples collected at the mid-logarithmic phase of growth using TRIzol (Sigma-Aldrich). The total RNA samples were treated with DNase I (Fermentas) according to the manufacturer's instructions. The concentration of RNA in each sample was measured using NanoDrop.

**[00481]** Northern blot analysis

**[00482]** Northern blot analysis was carried out essentially as described previously (40-42). Total RNA was separated on 10% polyacrylamide 8 M urea gels and further processed for blotting on nylon membranes (Hybond™ N+, GE healthcare; Trans-Blot® SD semi-dry transfer apparatus, Biorad; 1X TBE, 2 h at 10 V/cm), chemical crosslinking with EDC (1-Ethyl-3-(3-dimethylaminopropyl) carbodiimide hydrochloride) (41) and prehybridization (Rapid-hyb buffer, GE healthcare; 1 h at 42°C). Oligonucleotide probes (40 pmol) were labeled with <sup>32</sup>P (20 µCi) using the T4-polynucleotide kinase (10 U, Fermentas) and purified using G-25 columns (GE Healthcare) prior use. Visualization of the radioactive signal was done using a phosphorimager. 5S rRNA served as loading control.

**[00483]** Protein purification

**[00484]** *E. coli* Rosetta2(DE3) and *E. coli* NiCo21(DE3) (New England Biolabs) were transformed with overexpression plasmids coding for *S. pyogenes* WT and mutant or orthologous Cas9, respectively. Cells were grown at 37°C to reach an OD<sub>600</sub> of 0.7 - 0.8, protein expression was induced by adding IPTG to a final concentration of 0.5 mM and cultures were further grown at 13°C overnight. The cells were harvested by centrifugation and the pellet was resuspended in lysis-buffer (20 mM HEPES pH 7.5, 500 mM KCl [1 M for *S. thermophilus*\* Cas9], 0.1% Triton X-100, 25 mM imidazole) and lysed by sonication. The lysate was cleared by centrifugation (> 20 000 x g) and incubated with Ni-NTA (Qiagen) for 1 h at 4°C. After washing the Ni-NTA with lysis-buffer and wash-buffer (20 mM HEPES pH 7.5, 300 mM KCl,



0.1% Triton X-100, 25 mM imidazole), the recombinant protein was eluted with elution-buffer (20 mM HEPES pH 7.5, 150 mM KCl, 0.1 mM DTT, 250 mM imidazole, 1 mM EDTA) and the fractions were analyzed by SDS-PAGE. In the case of *S. pyogenes* Cas9 WT and mutants, the protein containing eluates were pooled and further purified via HiTrap SP FF (GE Healthcare) cation-exchange chromatography. Briefly, the protein was loaded on the column equilibrated with buffer A (20 mM HEPES pH 7.5, 100 mM KCl) using an FPLC system (Äkta, GE Healthcare). Cas9 was eluted with a gradient of buffer B (20 mM HEPES pH 7.5, 1 M KCl) over 12 ml. 1 ml fractions were collected and analyzed by SDS-PAGE. The protein containing fractions were pooled and dialyzed overnight (20 mM HEPES pH 7.5, 150 mM KCl, 50% glycerol). For Cas9 orthologs, the eluates from Ni-NTA purification were checked for purity by SDS-PAGE. In case of contaminants, a second purification over chitin beads was performed as described in the manual for NiCo21(DE3) cells from New England Biolabs. Briefly, 1 ml chitin beads (New England Biolabs) equilibrated with buffer A was incubated with the Ni<sup>2+</sup>-IMAC eluates for 1 h at 4°C. Afterwards the beads were added onto a column and the Cas9 containing flowthroughs were collected and again checked for purity by SDS-PAGE (Supplementary Figure S1). The purified proteins were dialyzed overnight. The protein concentration was calculated by measuring the OD<sub>280</sub> using the extinction coefficient. The detailed characteristics of purified proteins are summarized in Supplementary Figure S1A.

**[00485]** *In vitro* transcription

**[00486]** RNA for *in vitro* DNA cleavage assays was generated by *in vitro* transcription using the AmpliScribe™ T7-Flash™ Transcription Kit (Epicentre) according to the manufacturer's instructions. PCR products or synthetic oligonucleotides used as templates are listed in Supplementary Table S1. The synthesized tracrRNA and repeat region of crRNA from each bacterial species correspond to the mature forms of RNAs as determined by deep RNA sequencing (15) or bioinformatics predictions. The spacer region of all crRNAs used in this study targets the *speM* protospacer (encoding superantigen; targeted by spacer 2 of *S. pyogenes* SF370 CRISPR array, Spyo1h\_002 (16)). Transcribed RNAs were precipitated and further purified from 10% polyacrylamide 8 M urea denaturing gel. The RNA concentration was determined by measuring the OD<sub>260</sub> and the molarity was calculated. Equimolar amounts of crRNA and tracrRNA were mixed in 5X RNA annealing buffer (1 M NaCl, 100 mM HEPES pH 7.5), heated up to 95°C for 5 min and slowly cooled to room temperature before use.

**[00487]** *In vitro* DNA cleavage assays

**[00488]** For the cleavage assays using Cas9 mutant proteins, 25 nM of Cas9 were incubated with equimolar amounts of prehybridized *S. pyogenes* dual-RNA in cleavage buffer (20 mM HEPES pH 7.5, 150 mM KCl, 10 mM MgCl<sub>2</sub>, 0.5 mM DTT, 0.1 mM EDTA) for 15 min at 37°C. Plasmid DNA (5 nM) containing *speM* (NGG PAM) was added and further incubated for 1 h at 37°C. The reaction was stopped by addition of 5X loading buffer (250 mM EDTA, 30% glycerol, 1.2% SDS, 0.1% (w/v) bromophenol blue)

and analyzed by 1% agarose gel electrophoresis in 1X TAE. Cleavage products were visualized by ethidium bromide staining. All other cleavage assays were carried out using the same conditions with the following modifications: KGB (43) (100 mM potassium glutamate, 25 mM Tris/acetate pH 7.5, 10 mM Mg-acetate, 0.5 mM 2-mercaptoethanol, 10 µg/ml BSA) was used as cleavage buffer and different concentrations of dual-RNA:Cas9 complex were analyzed. The concentration of plasmid DNA was kept constant in all experiments, i.e. 5 nM.

**[00489]** Search for PAM motifs

**[00490]** Spacer sequences of the selected bacterial species were extracted from the CRISPRdatabase (<http://crispr.u-psud.fr/crispr/>) and used to find cognate protospacer candidates using megaBLAST (<http://blast.ncbi.nih.gov/Blast>). Protospacer candidates were defined as containing a sequence with ≥90% similarity to the crRNA spacer sequence and originating from phage, plasmid or genomic DNA related to the bacterial species of the targeting CRISPR-Cas. For the investigated CRISPR-Cas loci, the orientation of transcription was determined previously by RNA sequencing or Northern blot analysis (15,16). It was also shown before that in type II CRISPR-Cas, the PAM sequence is located in 3' of the protospacer, juxtaposed to the sequence targeted by cognate crRNA on the non-target strand (14,18,23,44). To identify possible PAMs in each bacterial species, 10 nt sequences on the non-target strand directly downstream of each protospacer sequence were aligned. A logo plot (<http://weblogo.berkeley.edu/>) showing the most abundant nucleotides was created and PAM sequences were predicted. In the cases of CRISPR-Cas loci for which no suitable protospacer sequences could be identified (*S. mutans* UA159, *C. jejuni* NCTC 11168, *P. multocida* Pm70, *F. novicida* U112), closely related strains of the same species were selected (Supplementary Table S2). The spacer contents of the type II CRISPR arrays in selected strains were analyzed (<http://crispr.u-psud.fr/Server/>). The spacer sequences were then used to select cognate protospacer sequences as described above.

**[00491]** Protein sequence analysis

**[00492]** Position-Specific Iterated (PSI)-BLAST program (45) was used to retrieve orthologs of the Cas9 family in the NCBI nr database. Sequences shorter than 800 amino acids were discarded. The BLASTClust program (46) set up with a length coverage cutoff of 0.8 and a score coverage threshold (bit score divided by alignment length) of 0.8 was used to cluster the remaining sequences (Supplementary Table S2). This procedure produced 82 clusters. In the case of sequences reported in this study, one or several representatives from each cluster were selected and aligned using the MUSCLE program (47) with default parameters, followed by a manual correction on the basis of local alignments obtained using PSI-BLAST (45) and HHpred programs (48). The confidently aligned blocks (Supplementary Figure S2) with 285 informative positions were used for maximum likelihood tree reconstruction using the FastTree program (49) with the default parameters: JTT evolutionary model, discrete gamma model with 20 rate categories. The same program was used to calculate the bootstrap values. Cas1 sequences were

selected from the corresponding *cas* operons (Supplementary Table S2). A few incomplete sequences were substituted by other Cas1 sequences from the same Cas9 cluster (Supplementary Table S2). Several Cas1 proteins from subtypes I-A, B, C and E were included as an outgroup. Cas1 sequences were aligned using the same approach described above and 252 informative positions (Supplementary Figure S3) were used for maximum likelihood tree reconstruction using the FastTree program. RNase III multiple sequence alignment was prepared using the MUSCLE program.

**[00493]** RNA sequence and structure analysis

**[00494]** RNA duplex secondary structures were predicted using RNAfold of the Vienna RNA package (50,51) and RNAhybrid (<http://bibiserv.techfak.uni-bielefeld.de/rnahybrid/>). The structure predictions were then visualized using VARNA (52).

**Supplementary Table S1. Strains, plasmids and primers used in the study.**

Strain	Relevant characteristics	Source
<b><u>Streptococcus pyogenes</u></b>		
<b><u>WT</u></b>		
EC904	SF370 (M1 serotype) (WT)	ATCC 700294
<b><u>Δ<i>cas9</i></u></b>		
EC1788	EC904Δ <i>cas9</i>	(16)
<b><u>Δ<i>rnc</i></u></b>		
EC1636	EC904Δ <i>rnc</i>	(16)
<b><u>Δ<i>cas9</i> in SF370 + <i>cas9</i> complementations <i>in trans</i></u></b>		
EC2121	EC1788 + pEC714 ( <i>Pcas9</i> (Spy)- <i>cas9</i> (Spy)-CtHis)	This study
EC2127	EC1788 + pEC710 (171 tracrRNA- <i>Pcas9</i> (Spy)-CtHis)	This study
EC2150	EC1788 + pEC553 ( <i>Pcas9</i> (Spy)- <i>cas9</i> -HH983AA(Spy)-CtHis)	This study
EC2151	EC1788 + pEC554 ( <i>Pcas9</i> (Spy)- <i>cas9</i> -D10A(Spy)-CtHis)	This study
EC2152	EC1788 + pEC555 ( <i>Pcas9</i> (Spy)- <i>cas9</i> -H840A(Spy)-CtHis)	This study
EC2153	EC1788 + pEC556 ( <i>Pcas9</i> (Spy)- <i>cas9</i> -N854A(Spy)-	This study

	CtHis	
EC2154	EC1788 + pEC557 ( <i>Pcas9(Spy)-cas9-N863A(Spy)-CtHis</i> )	This study
EC2155	EC1788 + pEC558 ( <i>Pcas9(Spy)-cas9-D986A(Spy)-CtHis</i> )	This study
EC2156	EC1788 + pEC559 ( <i>Pcas9(Spy)-cas9-E762A(Spy)-CtHis</i> )	This study
EC2118	EC1788 + pEC518 ( <i>Pcas9(Spy)-cas9(Cje)-CtHis</i> )	This study
EC2128	EC1788 + pEC538 ( <i>Pcas9(Spy)-cas9(Fno)-CtHis</i> )	This study
EC2199	EC1788 + pEC544 ( <i>Pcas9(Spy)-cas9(Nme)-CtHis</i> )	This study
EC2119	EC1788 + pEC520 ( <i>Pcas9(Spy)-cas9(Pmu)-CtHis</i> )	This study
EC2111	EC1788 + pEC519 ( <i>Pcas9(Spy)-cas9(Smu)-CtHis</i> )	This study
EC2112	EC1788 + pEC521 ( <i>Pcas9(Spy)-cas9(Sth*)-CtHis</i> )	This study
EC2120	EC1788 + pEC522 ( <i>Pcas9(Spy)-cas9(Sth**)-CtHis</i> )	This study

***Δrnc in SF370 + rnc complementations in trans***

EC2076	EC1636 + pEC484 ( <i>Pmrc(Spy)-mrc(Spy)</i> )	This study
EC2084	EC1636 + pEC505 ( <i>Pmrc(Spy)-mrc-catalytically inactive(Spy)</i> )	This study
EC2083	EC1636 + pEC504 ( <i>Pmrc(Spy)-mrc-RNA binding inactive(Spy)</i> )	This study
EC2078	EC1636 + pEC486 ( <i>Pmrc(Spy)-rnc(Cje)</i> )	This study
EC2080	EC1636 + pEC492 ( <i>Pmrc(Spy)-mrc(Eco)</i> )	This study
EC2126	EC1636 + pEC537 ( <i>Pmrc(Spy)-rnc(Fno)</i> )	This study
EC2085	EC1636 + pEC506 ( <i>Pmrc(Spy)-rnc(Nme)</i> )	This study
EC2077	EC1636 + pEC485 ( <i>Pmrc(Spy)-mrc(Pmu)</i> )	This study
EC2086	EC1636 + pEC507 ( <i>Pmrc(Spy)-mrc(Sau)</i> )	This study
EC2082	EC1636 + pEC494 ( <i>Pmrc(Spy)-rnc(Smu)</i> )	This study
EC2131	EC1636 + pEC534 ( <i>Pmrc(Spy)-mrc(Sth)</i> )	This study

***Campylobacter jejuni***

EC437	NCTC 11168; ATCC 700819 (WT), CIP 107370	Pasteur Institute
-------	--	-------------------

<b><u>Francisella novicida</u></b>		
EC1041	U112 (WT)	Anders Sjöstedt
<b><u>Neisseria meningitidis</u></b>		
EC438	CIP 107858	Pasteur Institute
<b><u>Pasteurella multocida</u></b>		
EC439	Pm70 (WT), ATCC BAA-1113	Pasteur Institute
<b><u>Staphylococcus aureus</u></b>		
EC36	COL (WT)	Lab strain collection
<b><u>Streptococcus mutans</u></b>		
EC1293	UA159 (WT)	(16)
<b><u>Streptococcus thermophilus</u></b>		
EC810	LMD-9 (WT)	(16)
<b><u>E. coli</u></b>		
RDN204	TOP10, host for cloning	Invitrogen
EC1265	Rosetta	Novagen

<sup>a</sup> Cje: *Campylobacter jejuni* NCTC 11168; Eco: *Escherichia coli* TOP10; Fno: *Francisella novicida* U112; Nme: *Neisseria meningitidis* A Z2491; Pmu: *Pasteurella multocida* Pm70; Sau: *Staphylococcus aureus* COL; Smu: *Streptococcus mutans* UA159; Spy: *Streptococcus pyogenes* SF370; Sth: *Streptococcus thermophilus* LMD-9.

Plasmid	Relevant characteristics	Source
<b><u>Vectors for <i>S. pyogenes</i></u></b>		
pEC85	repDEG-pAM $\beta$ 1, pJH1-aphIII, ColE1	Bernhard Roppenser
<b><u>Plasmids for cas9 domain functional and co-evolution analysis in <i>S. pyogenes</i> SF370</u></b>		
pEC268	pEC85 $\Omega$ 171 tracrRNA (171 nt form)	(16)
pEC309	pEC85 $\Omega$ Pcas9(Spy)-cas9(Spy)	(16)
pEC368	pEC85 $\Omega$ 171 tracrRNA-Pcas9(Spy)-cas9(Spy)	(16)
pEC710	pEC85 $\Omega$ 171 tracrRNA-Pcas9(Spy)-CtHis	This study
pEC714	pEC710 $\Omega$ cas9(Spy)	This study

pEC553	pEC710Ωcas9-HH983AA(Spy)-CtHis	This study
pEC615	pEC553ΩspeM	This study
pEC554	pEC710Ωcas9-D10A(Spy)-CtHis	This study
pEC659	pEC554ΩspeM	This study
pEC555	pEC710Ωcas9-H840A(Spy)-CtHis	This study
pEC660	pEC555ΩspeM	This study
pEC556	pEC710Ωcas9-N854A(Spy)-CtHis	This study
pEC661	pEC556ΩspeM	This study
pEC557	pEC710Ωcas9-N863A(Spy)-CtHis	This study
pEC618	pEC557ΩspeM	This study
pEC558	pEC710Ωcas9-D986A(Spy)-CtHis	This study
pEC662	pEC558ΩspeM	This study
pEC559	pEC710Ωcas9-E762A(Spy)-CtHis	This study
pEC619	pEC559ΩspeM	This study
pEC518	pEC710Ωcas9(Cje)-CtHis	This study
pEC538	pEC710Ωcas9(Fno)-CtHis	This study
pEC544	pEC710Ωcas9(Nme)-CtHis	This study
pEC520	pEC710Ωcas9(Pmu)-CtHis	This study
pEC519	pEC710Ωcas9(Smu)-CtHis	This study
pEC521	pEC710Ωcas9(Sth*)-CtHis	This study
pEC522	pEC710Ωcas9(Sth**)-CtHis	This study

---

**Plasmids for *rnc* co-evolution analysis in *S. pyogenes* SF370**

pEC483	pEC85ΩPrnc(Spy)	This study
pEC484	pEC85ΩPrnc(Spy)- <i>rnc</i> (Spy)	This study
pEC505	pEC85ΩPrnc(Spy)- <i>rnc</i> -catalytically inactive(Spy)	This study
pEC504	pEC85ΩPrnc(Spy)- <i>rnc</i> -RNA binding inactive(Spy)	This study
pEC486	pEC85ΩPrnc(Spy)- <i>rnc</i> (Cje)	This study

pEC492	pEC85ΩPmc(Spy)-rnc(Eco)	This study
pEC537	pEC85ΩPmc(Spy)-rnc(Fno)	This study
pEC506	pEC85ΩPmc(Spy)-rnc(Nme)	This study
pEC485	pEC85ΩPmc(Spy)-rnc(Pmu)	This study
pEC507	pEC85ΩPmc(Spy)-rnc(Sau)	This study
pEC494	pEC85ΩPmc(Spy)-rnc(Smu)	This study
pEC534	pEC85ΩPmc(Spy)-rnc(Sth)	This study

---

**Plasmids for protospacer study *in vitro***

pEC287	pEC85ΩPspeM-speM (10 bp downstream protospacer: GGGTATTGGG)	Lab plasmid collection
pEC691	pEC287 (10 bp downstream protospacer: TGGTATTGGG)	This study
pEC692	pEC287 (10 bp downstream protospacer: TGGTGTTGGG)	This study
pEC693	pEC287 (10 bp downstream protospacer: GGGTGATTGG)	This study
pEC694	pEC287 (10 bp downstream protospacer: GGAGAATGGG)	This study
pEC696	pEC287 (10 bp downstream protospacer: GGGTCATAGG)	This study
pEC697	pEC287 (10 bp downstream protospacer: AGAAACAGGG)	This study
pEC698	pEC287 (10 bp downstream protospacer: AGAACCAGGG)	This study
pEC701	pEC287 (10 bp downstream protospacer: GTTTGATTGG)	This study
pEC706	pEC287 (10 bp downstream protospacer: GGAAAATGGG)	This study

---

**Plasmids for Cas9 overexpression**

pEC225	pET16b	Novagen
pEC621	pEC225 inserted with cassette harboring NotI,	This study

	SacI, Sall site	
pEC626	pEC621Ωcas9(Spy)	This study
pEC627	pEC621Ωcas9-D10A(Spy)	This study
pEC628	pEC621Ωcas9-E762A(Spy)	This study
pEC629	pEC621Ωcas9-H840A(Spy)	This study
pEC630	pEC621Ωcas9-N854A(Spy)	This study
pEC631	pEC621Ωcas9-HH983AA(Spy)	This study
pEC632	pEC621Ωcas9(Cje)	This study
pEC633	pEC621Ωcas9(Pmu)	This study
pEC634	pEC621Ωcas9(Nme)	This study
pEC635	pEC621Ωcas9(Smu)	This study
pEC638	pEC621Ωcas9-N863A(Spy)	This study
pEC639	pEC621Ωcas9-D986A(Spy)	This study
pEC640	pEC621Ωcas9(Sth*)	This study
pEC641	pEC621Ωcas9(Sth**)	This study
pEC657	pEC621Ωcas9(Fno)	This study

Purpose	Primer	Sequence 5'-3' <sup>a</sup>	F/R <sup>b</sup>	Usage <sup>c</sup>
<b><u>tracrRNA expression in <i>S. pyogenes</i> SF370</u></b>				
tracrRNA	OLEC101 4	GGACTAGCCTTATTTAACTTG	R	NB (3' probe)
<b><u>crRNA (CRISPR01 (type II-A) expression in <i>S. pyogenes</i> SF370</u></b>				
crRNA	OLEC104 9	GGACCATTCAAAAACAGCATAGCTCTAAAAC	R	NB (repeat)
<b><u>Loading controls for Northern blots</u></b>				
5S rRNA	OLEC288	CTAAGCGACTACCTTATCTCA	R	NB
<b><u>His-tagged cas9 constructs (pEC85-based)</u></b>				



pEC710	OLEC215 1	GCAGGAATTCATCAGTGATGGTGATGGTGATGC CCGGGTTTGTGCGACCTCCTAAAAATAAAAGTTTA AATTAAATC	F	Cloning
	OLEC206 6	GGTGGTCTGCAGGTTTGCAGTCAGAGTAGAATA GAAG	R	
pEC714	OLEC209 6	ATGCAGGTCGACATGGATAAGAAATACTCAATAG GC	F	Expression cas9(Spy)
	OLEC209 7	ATGCAGCCCGGGTTCACCTCCTAGCTGACTCAA ATC	R	
speM	OLEC286 7	ATGCAGCCTGCAGGGTGACAGAGAGAAACTTGA TTCAAC	F	Cloning of <i>speM</i> in other plasmids
	OLEC286 8	ATGCAGCCTGCAGGCTTCGTTTAAGTAAACATCA AAGTG	R	
pEC518	OLEC210 4	ATGCAGGTCGACGTGGCAAGAATTTTGGCATTG	F	Cloning cas9(Cje)
	OLEC210 5	ATGCAGCCCGGGTTTTTAAATCTTCTCTTTGTC	R	
pEC538	OLEC284 0	ATTAGTCGACATGAATTTCAAATATTGCCAATAG	F	Cloning cas9(Fno)
	OLEC284 1	ATTACCCGGGATTATTAGATGTTTCATTATAAATAC	R	
pEC544	OLEC209 2	ATGCAGGTCGACATGGCTGCCTTCAAACCTAATC C	F	Cloning cas9(Nme)
	OLEC209 3	ATGCAGCCCGGGACGGACAGGCGGGCGTTTTT CAG	R	
pEC520	OLEC210 0	ATGCAGGTCGACATGCAAAACAACAATTTAAGTT A	F	Cloning cas9(Pmu)
	OLEC210 1	ATGCAGCCCGGGACGCACAGGTTGTCTTTGCTG AG	R	
pEC519	OLEC209 0	ATGCAGGTCGACATGAAAAACCTTACTCTATTG GAC	F	Cloning cas9(Smu)
	OLEC209 1	ATGCAGCCCGGGTCTCCTCCTAACTTATTGAGA TC	R	
pEC521	OLEC209 8	ATGCAGGTCGACATGACTAAGCCATACTCAATTG G	F	Cloning cas9(Sth*)

	OLEC209 9	ATGCAGCCCGGGACCCCTCTCCTAGTTTGGCAAG GTC	R	
pEC522	OLEC210 2	ATGCAGGTCGACATGAGTGACTTAGTTTTAGGAC TTG	F	Cloning cas9(Sth**)
	OLEC210 3	ATGCAGCCCGGGAAAATCTAGCTTAGGCTTATCA CC	R	
pEC553	OLEC222 9	GTACGTGAGATTAACAATTACGCTGCTGCCCATG ATGCGTATCTA	F	Mutagenesis cas9- HH983AA(Spy)
	OLEC223 0	TAGATACGCATCATGGGCAGCAGCGTAATTGTTA ATCTCACGTAC	R	
pEC554	OLEC212 8	GAAATACTCAATAGGCTTAGCTATCGGCACAAAT AGCGTCG	F	Mutagenesis cas9-D10A(Spy)
	OLEC212 9	CGACGCTATTTGTGCCGATAGCTAAGCCTATTGA GTATTC	R	
pEC555	OLEC222 3	TTTAAGTGATTATGATGTCGATGCCATTGTTCCAC AAAGTTTCCT	F	Mutagenesis cas9-H840A(Spy)
	OLEC222 4	AGGAAACTTTGTGGAACAATGGCATCGACATCAT AATCACTTAAA	R	
pEC556	OLEC222 5	CCTTAAAGACGATTCAATAGACGCTAAGGTCTTA ACGCGTTCTGA	F	Mutagenesis cas9-N854A(Spy)
	OLEC222 6	TCAGAACGCGTTAAGACCTTAGCGTCTATTGAAT CGTCTTTAAGG	R	
pEC557	OLEC222 7	GGTCTTAACGCGTTCTGATAAAGCTCGTGGTAAA TCGGATAACGT	F	Mutagenesis cas9-N863A(Spy)
	OLEC222 8	ACGTTATCCGATTTACCACGAGCTTATCAGAAC GCGTTAAGACC	R	
pEC558	OLEC223 1	GTAACAATTACCATCATGCCCATGCTGCGTATCT AAATGCCGTCG	F	Mutagenesis cas9-D986A(Spy)
	OLEC223 2	CGACGGCATTTAGATACGCAGCATGGGCATGAT GGTAATTGTTAC	R	
pEC559	OLEC222 1	CAGAAAATATCGTTATTGCAATGGCACGTGAAAA TCAGACA	F	Mutagenesis cas9-E762A(Spy)
	OLEC222 2	TGTCTGATTTTCACGTGCCATTGCAATAACGATAT TTTCTG	R	

<b><i>rnc</i> constructs (pEC85-based)</b>				
pEC483	OLEC214 9	ATGCAGGCATGCCCTGTAGTTTTGGCTTGTCTGATC	F	Cloning in pEC85
	OLEC327 4	ATGCAGAGCTCCATGGAAAAATCCCTTTCATATTTGTCAGTAGACC	R	
pEC484	OLEC210 9	ATGCAGCCATGGAACAGCTTGAAGAGTTACTCTCAAC	F	Cloning <i>mc</i> (Spy), SEQ
	OLEC166 8	CTTTTAAAAACATCTAACCTCAC	R	
pEC504	OLEC210 9	ATGCAGCCATGGAACAGCTTGAAGAGTTACTCTCAAC	F	Cloning of <i>rnc</i> RNA binding inactive(Spy)
	OLEC265 6	ATGCAGGAATTCCTACCCTTTTTCCACCTGAGGATC	R	
pEC505	OLEC214 2	GAACGCTTGGAATTTTTAGGAGCCGCTGTTCTACAATTGATTATT	F	Mutagenesis of <i>rnc</i> catalytically inactive(Spy)
	OLEC214 3	AATAATCAATTGTAGAACAGCGGCTCCTAAAAATCCAAGCGTTC	R	
pEC486	OLEC211 6	ATGCAGCCATGGAAAACATTGAAAAGCTAGAGCAGAG	F	Cloning <i>mc</i> (Cje), SEQ
	OLEC211 7	ATGCAGGAATTCCTATAAAGCTCCTAATTTCTCAAG	R	
pEC492	OLEC212 4	ATGCAGCCATGGACCCCATCGTAATTAATCGGCTTC	F	Cloning <i>mc</i> (Eco), SEQ
	OLEC212 5	ATGCAGGAATTCCTCATTCCAGCTCCAGTTTTTTCAACG	R	
pEC537	OLEC284 2	ATTACCATGGTTCCCTGAATATTCACGATTTTATAAC	F	Cloning <i>mc</i> (Fno), SEQ
	OLEC284 3	ATTGAATTCCTATTTTTTTTCATGTAAGCCTTGTTGTG	R	
pEC506	OLEC211 8	ATGCAGCCATGGAAGACGATGTTTTGAAACAGCAGG	F	Cloning <i>mc</i> (Nme), SEQ
	OLEC211 9	ATGCAGGAATTCCTCATTCTTTTTCTTCTTCAGCGGC	R	
pEC485	OLEC211	ATGCAGCCATGGCTCAAAATTTAGAACGTTTACA	F	Cloning <i>mc</i> (Pmu),

	4	ACG		SEQ
	OLEC211 5	ATGCAGGAATTCTCATTTCATTTC <del>CAATA</del> ATTGT	R	
pEC507	OLEC212 6	ATGCAGCCATGGCTAAACAAAAGAAAAGTGAGAT AG	F	Cloning <i>mc</i> (Sau), SEQ
	OLEC212 7	ATGCAGGAATTCTCTATTTAATTTGTTTTAATTGCT TATAGG	R	
pEC494	OLEC211 0	ATGCAGCCATGGAAACATTAGAAAAAAAACTGGC AG	F	Cloning <i>mc</i> (Smu), SEQ
	OLEC211 1	ATGCAGGAATTCTTAAGAACCTCGTTGAAGTTTT C	R	
pEC534	OLEC284 9	ATTACCATGGATCAACTGAACAAAACTTGAACA GGACTTTGG	F	Cloning <i>mc</i> (Sth), SEQ
	OLEC285 0	ATTAGAATTCTTAATTACCTAGTTGTTCAAGGGCA GACTTCGC	R	

**Cas9 overexpression (pEC621 based)**

pEC621	OLEC297 8	<u>TAGCGGCCGCGAGCTCGTCGACGC</u>	F	Cassette inserting NotI, SacI, Sall site in pEC225
	OLEC297 9	<u>TAGCGTCGACGAGCTCGCGGCCGC</u>	R	
pEC626, 627,  628, 629, 630, 631, 638, 639	OLEC209 7	ATGCAGGTCGACATGGATAAGAAATACTCAATAG GC	F	Cloning <i>cas9</i> (Spy and all mutants)
	OLEC298 3	AGCTAGCGGCCGCTCAGTCACCTCCTAGCTGAC TCAAATC	R	
pEC632	OLEC210 4	ATGCAGGTCGACGTGGCAAGAATTTGGCATTG	F	Cloning <i>cas9</i> (Cje)
	OLEC298 6	ATGCAGCGGCCGCTCATTTTTTAAAATCTTCTCTT TGTC	R	
pEC633	OLEC210 0	ATGCAGGTCGACATGCAAACAACAAATTTAAGTT A	F	Cloning <i>cas9</i> (Pmu)
	OLEC217 3	ATGACGCGGCCGCTTAACGCACAGGTTGTCTTT GCTG	R	
pEC634	OLEC209	ATGCAGGTCGACATGGCTGCCTTCAAACCTAATC	F	Cloning <i>cas9</i> (Nme)

	2	C		
	OLEC298 2	ATGACGCGGCCGCTTAACGGACAGGCGGGCGTT TTTTCAG	R	
pEC635	OLEC209 0	ATGCAGGT <u>CGAC</u> ATGAAAAACCTTACTCTATTG GAC	F	Cloning cas9(Smu)
	OLEC298 1	ATGACGCGGCCGCTTAGTCTCCTCCTAACTTATT GAG	R	
pEC640	OLEC209 8	ATGCAGGT <u>CGAC</u> ATGACTAAGCCATACTCAATTG G	F	Cloning cas9(Sth*)
	OLEC298 4	ATGACGCGGCCGCTTAACCCTCTCCTAGTTTGGC AAG	R	
pEC641	OLEC210 2	ATGCAGGT <u>CGAC</u> ATGAGTGACTTAGTTTTAGGAC TTG	F	Cloning cas9(Sth**)
	OLEC298 5	ATGACGCGGCCGCTTAAAAATCTAGCTTAGGCTT ATCAC	R	
pEC657	OLEC284 0	ATTAGT <u>CGAC</u> ATGAATTTCAAATATTGCCAATAG	F	Cloning cas9(Fno)
	OLEC298 7	ATGCAGCGGCCGCTAATTATTAGATGTTTCATT ATAAATAC	R	

**Mutagenesis 10 bp downstream of *speM* protospacer**

pEC691	OLEC314 0	CAACCACTAATTTCTAGAAAAATCTTCG	R	Mutagenesis on pEC287
	OLEC314 1	CAATTTGTAAAAAATGGTATTGGGGAATTC	F	
pEC692	OLEC314 0	CAACCACTAATTTCTAGAAAAATCTTCG	R	Mutagenesis on pEC287
	OLEC314 2	CAATTTGTAAAAAATGGTGTGGGGAATTC	F	
pEC693	OLEC314 0	CAACCACTAATTTCTAGAAAAATCTTCG	R	Mutagenesis on pEC287
	OLEC314 4	CAATTTGTAAAAAAGGGTGATTGGGGAATTC	F	
pEC694	OLEC314 0	CAACCACTAATTTCTAGAAAAATCTTCG	R	Mutagenesis on pEC287

	OLEC314 3	CAATTTGTAAAAAAGGAGAATGGGGAATTC	F	
pEC696	OLEC319 4	CAACCACTAATTTT TAGAAAAATCTTCG	R	Mutagenesis on pEC693
	OLEC319 7	CAATTTGTAAAAAAGGGTCATAGGGAATTC	F	
pEC697	OLEC319 4	CAACCACTAATTTT TAGAAAAATCTTCG	R	Mutagenesis on pEC694
	OLEC319 8	CAATTTGTAAAAAAGAAACAGGGGAATTC	F	
pEC698	OLEC319 4	CAACCACTAATTTT TAGAAAAATCTTCG	R	Mutagenesis on pEC694
	OLEC319 9	CAATTTGTAAAAAAGAACCAGGGGAATTC	F	
pEC701	OLEC319 4	CAACCACTAATTTT TAGAAAAATCTTCG	R	Mutagenesis on pEC693
	OLEC320 4	CAATTTGTAAAAAAGTTTGATTGGGAATTC	F	
pEC706	OLEC319 4	CAACCACTAATTTT TAGAAAAATCTTCG	R	Mutagenesis on pEC694
	OLEC320 8	CAATTTGTAAAAAAGGAAAATGGGGAATTC	F	

***In vitro* tracrRNA and crRNA of *Streptococcus pyogenes* SF370 (*speM* spacer underlined)**

T7- tracrRNA	OLEC152 1	GAAATTAATACGACTCACTATAGAAAACAGCATA GCAAGTTAAAATAA	F	T7-tracrRNA 5'
	OLEC152 2	AAAAAAAGCACCGACTCGGTGCCAC	R	T7-tracrRNA 3'
T7-crRNA (template)	OLEC217 7	GAAATTAATACGACTCACTATAGGATAACTCAAT <u>TTGTAAAAAAGTTTTAGAGCTATGCTGTTTTG</u>	F	crRNA <i>speM</i> 5'
	OLEC217 9	CAAACAGCATAGCTCTAAAACTTTTTACAAATT <u>GAGTTATCCTATAGTGAGTCGTATTAATTC</u>	R	crRNA <i>speM</i> 3'

***In vitro* tracrRNA and crRNA of *Neisseria meningitidis* A Z2491 (*speM* spacer underlined)**

T7- tracrRNA	OLEC308 3	GAAATTAATACGACTCACTATAGGGAGAGCGAA ATGAGAACCGTTGCTACAATAAGCGCTCTGAAAA GATGTGCCGCAACGCTCTGCCCTTAAAGCTTCT	F	T7-tracrRNA 5'
-----------------	--------------	--	---	----------------

(template)		GCTTTAAGGGGCATCGTTTATT		
	OLEC308 4	AATAACGATGCCCTTAAAGCAGAAGCTTTAAG GGGCAGAGCGTTGCGGCACATCTTTTCAGACGC CTTATTGTAGCAACGGTTCTCATTTGCTCTCCCT ATAGTGAGTCGTATTAATTC	R	T7-tracrRNA 3'
T7-crRNA (template)	OLEC220 9	<u>GAAATTAATACGACTCACTATAGATGATAACTCA</u> <u>ATTTGTAAAAA</u> AGTTGTAGCTCCCTTTCTCATT	F	crRNA speM 5'
	OLEC221 4	AAATGAGAAAGGGAGCTACAAC <u>TTTTTACAAATT</u> <u>GAGTTATCATCTATAGTGAGTCGTATTAATTC</u>	R	crRNA speM 3'

***In vitro* tracrRNA and crRNA of *Streptococcus mutans* UA159 (*speM* spacer underlined)**

T7- tracrRNA	OLEC309 8	<u>GAAATTAATACGACTCACTATAGGAAACAACACA</u> GCAAGTTAAAAAAG	F	T7-tracrRNA 5'
	OLEC309 9	AAATAAAAAAGCACCGAATCGG	R	T7-tracrRNA 3'
T7-crRNA (template)	OLEC308 5	<u>GAAATTAATACGACTCACTATAGGATAACTCAAT</u> <u>TTGTAAAAA</u> AGTTTAGAGCTGTGTTGT	F	crRNA speM 5'
	OLEC308 6	ACAACACAGCTCTAAAAC <u>TTTTTACAAATTGAGT</u> <u>TATCCTATAGTGAGTCGTATTAATTC</u>	R	crRNA speM 3'

***In vitro* tracrRNA and crRNA of *Campylobacter jejuni* NCTC 11168 (*speM* spacer underlined)**

T7- tracrRNA  (template)	OLEC312 8	<u>GAAATTAATACGACTCACTATAGGAAGGGACTA</u> AAATAAAGAGTTTGCGGGACTCTGCGGGGTTACA ATCCCCTAAAACCGC	F	T7-tracrRNA 5'
	OLEC312 9	GCGGTTTTAGGGGATTGTAACCCCGCAGAGTCC CGCAAACCTTTATTTTAGTCCCTTCTATAGTGA GTCGTATTAATTC	R	T7-tracrRNA 3'
T7-crRNA (template)	OLEC308 7	<u>GAAATTAATACGACTCACTATAGGATAACTCAAT</u> <u>TTGTAAAAA</u> AGTTTAGTCCCT	F	crRNA speM 5'
	OLEC308 8	AGGGACTAAAAC <u>TTTTTACAAATTGAGTTATCCT</u> <u>ATAGTGAGTCGTATTAATTC</u>	R	crRNA speM 3'

***In vitro* tracrRNA and crRNAs of *Francisella novicida* U112 (*speM* spacer underlined)**

T7- tracrRNA	OLEC310 2	<u>GAAATTAATACGACTCACTATAGGGTACCAAATA</u> ATTAATGCTCTG	F	T7-tracrRNA 5'
	OLEC310	GTTATTCAGACGTGTCAAACAG	R	T7-tracrRNA 3'

	3			
T7-crRNA (template)	OLEC308 9	<u>GAAATTAATACGACTCACTATAGGATAACTCAAT</u> <u>TTGTAAAAAAGTTTCAGTTGCTGAATTATTTGGTA</u> AAC	F	crRNA speM 5'
	OLEC309 0	GTTTACCAAATAATTCAGCAACTGAAACTTTTTTA <u>CAAATTGAGTTATCCTATAGTGAGTCGTATTAAT</u> TTC	R	crRNA speM 3'

***In vitro* tracrRNA and crRNAs of *Streptococcus thermophilus*\* LMD-9 (speM spacer underlined)**

T7- tracrRNA	OLEC310 4	<u>GAAATTAATACGACTCACTATAGGAACAACACA</u> GCGAGTTAAAAAAGG	F	T7-tracrRNA 5'
	OLEC310 5	AAAAAAAACACCGAATCGGTG	R	T7-tracrRNA 3'
T7-crRNA (template)	OLEC308 5	<u>GAAATTAATACGACTCACTATAGGATAACTCAAT</u> <u>TTGTAAAAAAGTTTTAGAGCTGTGTTGT</u>	F	crRNA speM 5'
	OLEC308 6	ACAACACAGCTCTAAAACTTTTTACAAATTGAGT <u>TATCCTATAGTGAGTCGTATTAATTC</u>	R	crRNA speM 3'

***In vitro* tracrRNA and crRNAs of *Streptococcus thermophilus*\*\* LMD-9 (speM spacer underlined)**

T7- tracrRNA	OLEC310 6	<u>GAAATTAATACGACTCACTATAGGCTTAAATCTT</u> GCAGAAGCTACAAAG	F	T7-tracrRNA 5'
	OLEC310 7	AAATAACGAAAACACCCTGCC	R	T7-tracrRNA 3'
T7-crRNA (template)	OLEC309 1	<u>GAAATTAATACGACTCACTATAGGATAACTCAAT</u> <u>TTGTAAAAAAGTTTTGTACTCTCAAGATTTA</u>	F	crRNA speM 5'
	OLEC309 2	TAAATCTTGAGAGTACAAAACTTTTTTACAAATT <u>GAGTTATCCTATAGTGAGTCGTATTAATTC</u>	R	crRNA speM 3'

***In vitro* tracrRNA and crRNAs of *Pasteurella multocida* Pm70 (speM spacer underlined)**

T7- tracrRNA	OLEC310 8	<u>GAAATTAATACGACTCACTATAGGCTGCGAAAT</u> GAGAGACGTTGCTAC	F	T7-tracrRNA 5'
	OLEC310 9	AAAAACGATGCCCTTGCAATTAAG	R	T7-tracrRNA 3'
T7-crRNA (template)	OLEC309 3	<u>GAAATTAATACGACTCACTATAGGATAACTCAAT</u> <u>TTGTAAAAAAGTTGTAGTTCCCTCTCTCATTTCCG</u>	F	crRNA speM 5'
	OLEC309 4	GCGAAATGAGAGAGGGAAC TACAAC TTTTTTACA <u>AATTGAGTTATCCTATAGTGAGTCGTATTAATTC</u>	R	crRNA speM 3'



**Primers for sequencing analysis****cas9 *Streptococcus mutans* UA159**

cas9(Smu)	OLEC279 2	ATGAAAAAACCTTACTCTATTGGA	F	SEQ
	OLEC279 3	GATTTTAAAAAGCATTTTGAATTA	F	SEQ
	OLEC279 4	TACTTGCCAAATCAAAAAGTTCTT	F	SEQ
	OLEC279 5	ATTATGGGACATCAACCTGAAAAT	F	SEQ
	OLEC279 6	TACCCACAATTGGAACCTGAATTT	F	SEQ

**cas9 *Neisseria meningitidis* A Z2491**

cas9(Nme)	OLEC279 7	ATGGCTGCCTTCAAACCTAATCCA	F	SEQ
	OLEC279 8	GTTCAAAAAATGTTGGGGCATTGC	F	SEQ
	OLEC279 9	ATCCATATTGAAACTGCAAGGGAA	F	SEQ
	OLEC280 0	AACGCGTTTGACGGTAAAACCATA	F	SEQ

**cas9 *Streptococcus thermophilus*\* LMD-9**

cas9(Sth*)	OLEC280 7	ATGACTAAGCCATACTCAATTGGA	F	SEQ
	OLEC280 8	GATTTTAGGAAATGTTTAAATTA	F	SEQ
	OLEC280 9	TATTTGCCAGAAGAGAAGGTACTT	F	SEQ
	OLEC281 0	GTAATGGGAGGAAGAAAACCCGAG	F	SEQ
	OLEC281	GCAAGTGCTTTACTTAAGAAATAC	F	SEQ

	1			
	OLEC281 2	TTACTTTATCATGCTAAGAGAATA	F	SEQ
<b><u>cas9 Streptococcus thermophilus** LMD-9</u></b>				
	OLEC281 7	ATGAGTGACTTAGTTTTAGGACTT	F	SEQ
	OLEC281 8	ATTTTTGGAATTCTAATTGGGAAA	F	SEQ
cas9(Sth**)	OLEC281 9	GGAGACTTTGACAATATTGTCATC	F	SEQ
	OLEC282 0	TTGAATTTGTGGAAAAACAAAAG	F	SEQ
	OLEC282 1	CAGGAAAAATACAATGACATTAAG	F	SEQ
<b><u>cas9 Pasteurella multocida Pm70</u></b>				
	OLEC281 3	ATGCAAACAACAAATTTAAGTTAT	F	SEQ
	OLEC281 4	ACGCATGAAAAAATGAGTTTAAA	F	SEQ
cas9(Pmu)	OLEC281 5	CTTGGGAAATCTTTTAAAGAACGT	F	SEQ
	OLEC281 6	TATGAAATGGTGGATCAAGAAAGC	F	SEQ
<b><u>cas9 Campylobacter jejuni NCTC 11168</u></b>				
	OLEC282 2	GTGGCAAGAATTTTGGCATTGAT	F	SEQ
	OLEC282 3	GATGAAAAAAGAGCGCCAAAAAAT	F	SEQ
cas9(Cje)	OLEC282 4	AACTACAAGGCCAAAAAAGACGCC	F	SEQ
	OLEC282 5	AACAAAAGGAAGTTTTTTGAGCCT	F	SEQ
<b><u>cas9 Francisella novicida U112</u></b>				

cas9(Fno)	OLEC286 9	<i>ATGAATTTCAA</i> <u>AAATATTGCCAATA</u>	F	SEQ
	OLEC287 0	<i>TTAGATACTCTTTTAACTGATGAT</i>	F	SEQ
	OLEC287 1	<i>TTAAAAGTCTTAAAGTCAAGTAAA</i>	F	SEQ
	OLEC287 2	<i>GGTTCAGAAGATAAAAAAGGTAAT</i>	F	SEQ
	OLEC287 3	<i>AGAATTTTCTGCCTACGTGATCTT</i>	F	SEQ
	OLEC287 4	<i>CCAATACTAATCCATAAAGAACTA</i>	F	SEQ
	OLEC287 5	<i>ACATCAAAAAATATTTTTTGGCTG</i>	F	SEQ

<sup>a</sup> *italic*, sequence annealing to the template; underlined, restriction site; **bold**, T7 promoter.

<sup>b</sup> F, forward primer; R, reverse primer.

<sup>c</sup> NB, probe for Northern blot; SEQ, sequencing

Example 1

[00495] Diversity of Cas9 orthologs.

[00496] To investigate the evolution and diversity of dual-RNA:Cas9 systems, publicly available genomes were subjected to multiple rounds of BLAST search using previously retrieved Cas9 sequences as queries (15). Cas9 orthologs were identified in 653 bacterial strains representing 347 species (Supplementary Table S2). After removing incomplete or highly similar sequences, we selected 83 diverse, representative Cas9 orthologs for multiple sequence alignment and phylogenetic tree reconstruction (Figure 1A, Supplementary Table S2, Supplementary Figures S2 and S4, see Materials and Methods). The Cas9 tree topology largely agrees with the phylogeny of the corresponding Cas1 proteins (Supplementary Table S2, Supplementary Figures S3 and S4) and fully supports the previously described classification of type II CRISPR-Cas into three subtypes, II-A (specified by *csn2*), II-B (characterized by long and most diverged *cas9* variants (formerly *csx12*) and *cas4*), and II-C (three-*cas* gene operon) (15).

Supplementary Table S2. List of bacterial strains with identified Cas9 orthologs.

Cluster <sup>a</sup>	Strain <sup>b</sup>	Cas9 length (aa)	Cas9 GI	Cas1GI <sup>c</sup>	Subtype <sup>d</sup>
1	<i>Dolosigranulum pigrum</i> ATCC 51524	1332	375088882		Type II-A
	<i>Enterococcus faecalis</i> ATCC 29200	1337	229548613		
	<i>Enterococcus faecalis</i> ATCC 4200	1337	256617555		
	<i>Enterococcus faecalis</i> D6	1337	257086028		
	<i>Enterococcus faecalis</i> E1Sol	1337	257080914		
	<i>Enterococcus faecalis</i> OG1RF	1337	384512368		
	<i>Enterococcus faecalis</i> TX0470	1337	312900261		
	<i>Enterococcus faecalis</i> TX4244	1337	422695652		
	<i>Enterococcus faecium</i> 1,141,733	1339	257888853		
	<i>Enterococcus faecium</i> 1,231,408	1340	257893735		
	<i>Enterococcus faecium</i> E1133	1339	430847551		
	<i>Enterococcus faecium</i> E3083	1340	431757680		
	<i>Enterococcus faecium</i> PC4.1	1340	293379700		
	<i>Enterococcus faecium</i> TX1330	1340	227550972		
	<i>Enterococcus faecium</i> TX1337RF	1340	424765774		
	<i>Enterococcus hirae</i> ATCC 9790	1336	392988474		
	<i>Enterococcus italicus</i> DSM 15952	1330	315641599		
	<i>Lactobacillus animalis</i> KCTC 3501	1314	335357451		
	<i>Listeria innocua</i> ATCC 33091	1337	423101383		
	<i>Listeria innocua</i> Clip11262	1334	16801805		
	<i>Listeria innocua</i> FSL S4-378	1103	422414122		
	<i>Listeria ivanovii</i> FSL F6-596	953	315305353		
	<i>Listeria monocytogenes</i> 10403S	1334	386044902		
<i>Listeria monocytogenes</i> FSL J1-175	1099	255520581			

Cluster <sup>a</sup>	Strain <sup>b</sup>	Cas9 length (aa)	Cas9 GI	Cas1GI <sup>c</sup>	Subtype <sup>d</sup>
	<i>Listeria monocytogenes</i> FSL J1-194	1334	254825045		
	<i>Listeria monocytogenes</i> FSL J1-208	1334	422810631		
	<i>Listeria monocytogenes</i> FSL N3-165	1334	254829042		
	<i>Listeria monocytogenes</i> FSL R2-503	1334	254854201		
	<i>Listeria monocytogenes</i> str. 1/2a F6854	1334	47097148		
	<i>Streptococcus agalactiae</i> 2603V/R	1370	22537057		
	<i>Streptococcus agalactiae</i> 515	1377	77413160		
	<i>Streptococcus agalactiae</i> A909	1370	76788458		
	<i>Streptococcus agalactiae</i> ATCC 13813	1378	339301617		
	<i>Streptococcus agalactiae</i> CJB111	1370	77411010		
	<i>Streptococcus agalactiae</i> COH1	1370	77407964		
	<i>Streptococcus agalactiae</i> FSL S3-026	1370	417005168		
	<i>Streptococcus agalactiae</i> GB00112	1370	421147428		
	<i>Streptococcus agalactiae</i> H36B	1370	77405721		
	<i>Streptococcus agalactiae</i> NEM316	1377	25010965		
	<i>Streptococcus agalactiae</i> SA20-06	1370	410594450		
	<i>Streptococcus agalactiae</i> STIR-CD-17	1370	421532069		
	<i>Streptococcus anginosus</i> F0211	1345	315223162		
	<i>Streptococcus anginosus</i> SK1138	1386	421490579		
	<i>Streptococcus anginosus</i> SK52 = DSM 20563	1396	335031483		
	<i>Streptococcus bovis</i> ATCC 700338	1373	306833855		
	<i>Streptococcus canis</i> FSL Z3-227	1375	392329410		
	<i>Streptococcus constellatus</i> subsp. <i>constellatus</i> SK53	1345	418965022		
	<i>Streptococcus dysgalactiae</i> subsp. <i>equisimilis</i> AC-2713	1371	410494913		
	<i>Streptococcus dysgalactiae</i> subsp. <i>equisimilis</i> ATCC 12394	1371	386317166		
	<i>Streptococcus dysgalactiae</i> subsp. <i>equisimilis</i> GGS_124	1371	251782637		
	<i>Streptococcus dysgalactiae</i> subsp. <i>equisimilis</i> RE378	1371	408401787		
	<i>Streptococcus equi</i> subsp. <i>zooepidemicus</i> MGCS10565	1348	195978435		
	<i>Streptococcus equinus</i> ATCC 9812	1377	320547102		
	<i>Streptococcus gallolyticus</i> subsp. <i>gallolyticus</i> ATCC BAA-2069	1370	325978669		
	<i>Streptococcus gallolyticus</i> subsp. <i>gallolyticus</i> TX20005	1370	306831733		
	<i>Streptococcus gallolyticus</i> UCN34	1371	288905639		
	<i>Streptococcus infantarius</i> subsp. <i>infantarius</i> CJ18	1375	379705580		
	<i>Streptococcus iniae</i> 9117	1368	406658208		
	<i>Streptococcus macacae</i> NCTC 11558	1338	357636406		
	<i>Streptococcus mitis</i> SK321	1392	307710946		
	<i>Streptococcus mutans</i> 11SSST2	1345	449165720		
	<i>Streptococcus mutans</i> 11SSST2	1345	449951835		
1 (continued)	<i>Streptococcus mutans</i> 11VS1	1345	449976542		Type II-A
	<i>Streptococcus mutans</i> 14D	1345	450149988		
	<i>Streptococcus mutans</i> 15VF2	1355	449170557		
	<i>Streptococcus mutans</i> 15VF2	1355	449965974		
	<i>Streptococcus mutans</i> 1SM1	1345	449158457		

Cluster <sup>a</sup>	Strain <sup>b</sup>	Cas9 length (aa)	Cas9 GI	Cas1GI <sup>c</sup>	Subtype <sup>d</sup>
	<i>Streptococcus mutans</i> 1SM1	1345	449920643		
	<i>Streptococcus mutans</i> 24	1350	449247589		
	<i>Streptococcus mutans</i> 24	1350	450180942		
	<i>Streptococcus mutans</i> 2VS1	1345	449174812		
	<i>Streptococcus mutans</i> 2VS1	1345	449968746		
	<i>Streptococcus mutans</i> 3SN1	1345	449162653		
	<i>Streptococcus mutans</i> 3SN1	1345	449931425		
	<i>Streptococcus mutans</i> 4SM1	1345	449159838		
	<i>Streptococcus mutans</i> 4SM1	1345	449927152		
	<i>Streptococcus mutans</i> 4VF1	1345	449167132		
	<i>Streptococcus mutans</i> 4VF1	1345	449961027		
	<i>Streptococcus mutans</i> 5SM3	1345	449176693		
	<i>Streptococcus mutans</i> 5SM3	1345	449980571		
	<i>Streptococcus mutans</i> 66-2A	1359	449240165		
	<i>Streptococcus mutans</i> 66-2A	1359	450160342		
	<i>Streptococcus mutans</i> 8ID3	1345	449154769		
	<i>Streptococcus mutans</i> 8ID3	1345	449872064		
	<i>Streptococcus mutans</i> A19	1345	449187668		
	<i>Streptococcus mutans</i> A19	1345	450013175		
	<i>Streptococcus mutans</i> B	1345	450166294		
	<i>Streptococcus mutans</i> G123	1345	450029806		
	<i>Streptococcus mutans</i> GS-5	1345	397650022		
	<i>Streptococcus mutans</i> LJ23	1345	387785882		
	<i>Streptococcus mutans</i> M21	1345	449194333		
	<i>Streptococcus mutans</i> M21	1345	450036249		
	<i>Streptococcus mutans</i> M230	1345	449260994		
	<i>Streptococcus mutans</i> M230	1345	449903532		
	<i>Streptococcus mutans</i> M2A	1345	449209586		
	<i>Streptococcus mutans</i> M2A	1345	450074072		
	<i>Streptococcus mutans</i> N29	1345	449182997		
	<i>Streptococcus mutans</i> N29	1345	450003067		
	<i>Streptococcus mutans</i> N3209	1345	449210660		
	<i>Streptococcus mutans</i> N3209	1345	450077860		
	<i>Streptococcus mutans</i> N66	1345	449212466		
	<i>Streptococcus mutans</i> N66	1345	450083993		
	<i>Streptococcus mutans</i> NFSM1	1350	449202104		
	<i>Streptococcus mutans</i> NFSM1	1350	450051112		
	<i>Streptococcus mutans</i> NLML1	1345	450140393		
	<i>Streptococcus mutans</i> NLML4	1338	449202681		
	<i>Streptococcus mutans</i> NLML4	1338	450059882		
	<i>Streptococcus mutans</i> NLML9	1345	449209148		
	<i>Streptococcus mutans</i> NLML9	1345	450066176		
	<i>Streptococcus mutans</i> NMT4863	1355	449186850		
	<i>Streptococcus mutans</i> NMT4863	1355	450007078		
	<i>Streptococcus mutans</i> NN2025	1345	290580220		
	<i>Streptococcus mutans</i> NV1996	1345	450086338		
	<i>Streptococcus mutans</i> NVAB	1345	449181424		
	<i>Streptococcus mutans</i> NVAB	1345	449990810		
	<i>Streptococcus mutans</i> R221	1345	449258042		
	<i>Streptococcus mutans</i> R221	1345	449899675		
	<i>Streptococcus mutans</i> S1B	1345	449251227		
	<i>Streptococcus mutans</i> S1B	1345	449877120		

Cluster <sup>a</sup>	Strain <sup>b</sup>	Cas9 length (aa)	Cas9 GI	Cas1GI <sup>c</sup>	Subtype <sup>d</sup>
1 (continued)	<i>Streptococcus mutans</i> SF1	1345	450098705		
	<i>Streptococcus mutans</i> SF14	1345	449221374		
	<i>Streptococcus mutans</i> SF14	1345	450107816		
	<i>Streptococcus mutans</i> SM1	1345	449245264		
	<i>Streptococcus mutans</i> SM1	1345	450176410		
	<i>Streptococcus mutans</i> SM4	1345	449246010		
	<i>Streptococcus mutans</i> SM4	1345	450170248		
	<i>Streptococcus mutans</i> SM6	1345	449223000		
	<i>Streptococcus mutans</i> SM6	1345	450112022		
	<i>Streptococcus mutans</i> ST6	1350	449227252		
	<i>Streptococcus mutans</i> ST6	1350	450123011		
	<b><i>Streptococcus mutans</i> UA159</b>	<b>1345</b>	<b>24379809</b>	<b>24379808</b>	
	<i>Streptococcus mutans</i> W6	1345	450094364		
	<i>Streptococcus oralis</i> SK304	1373	421488030		
	<i>Streptococcus oralis</i> SK610	1371	419782534		
	<i>Streptococcus pseudoporcinus</i> LQ 940-04	1374	416852857		
	<b><i>Streptococcus pyogenes</i> SF370 (M1 GAS)</b>	<b>1368</b>	<b>13622193</b>	<b>13622194</b>	
	<i>Streptococcus pyogenes</i> MGAS10270	1368	94543903		
	<i>Streptococcus pyogenes</i> MGAS10750	1371	94994317		
	<i>Streptococcus pyogenes</i> MGAS15252	1367	383479946		
	<i>Streptococcus pyogenes</i> MGAS2096	1368	94992340		
	<i>Streptococcus pyogenes</i> MGAS315	1368	21910213		Type II-A
	<i>Streptococcus pyogenes</i> MGAS5005	1368	71910582		
	<i>Streptococcus pyogenes</i> MGAS6180	1368	71903413		
	<i>Streptococcus pyogenes</i> MGAS9429	1368	94988516		
	<i>Streptococcus pyogenes</i> NZ131	1368	209559356		
	<i>Streptococcus pyogenes</i> SSI-1	1368	28896088		
	<i>Streptococcus rattii</i> FA-1 = DSM 20564	1370	400290495		
	<i>Streptococcus salivarius</i> K12	1385	421452908		
	<i>Streptococcus sanguinis</i> SK115	1377	422848603		
	<i>Streptococcus sanguinis</i> SK330	1392	422860049		
	<i>Streptococcus sanguinis</i> SK353	1370	422821159		
	<i>Streptococcus</i> sp. C300	1377	322375978		
	<i>Streptococcus</i> sp. F0441	1371	414157437		
<i>Streptococcus</i> sp. M334	1375	322378004			
<i>Streptococcus</i> sp. oral taxon 56 str. F0418	1371	339640839			
<i>Streptococcus suis</i> ST1	1381	389856936			
<i>Streptococcus thermophilus</i>	1388	343794781			
<b><i>Streptococcus thermophilus</i> LMD-9</b>	<b>1388</b>	<b>116628213</b>	<b>116628212</b>		
<i>Streptococcus thermophilus</i> MN-ZLW-002	1388	387910220			
<i>Streptococcus thermophilus</i> ND03	1388	386087120			
2	<i>Campylobacter coli</i> 1098	984	419564797		
	<i>Campylobacter coli</i> 111-3	984	419536531		
	<i>Campylobacter coli</i> 132-6	987	419572019		
	<i>Campylobacter coli</i> 151-9	984	419603415		
	<i>Campylobacter coli</i> 1909	984	419576091		
	<i>Campylobacter coli</i> 1957	965	419581876		Type II-C
	<i>Campylobacter coli</i> 2692	984	419553162		
	<i>Campylobacter coli</i> 59-2	984	419578074		
	<i>Campylobacter coli</i> 67-8	965	419587721		
	<i>Campylobacter coli</i> 80352	965	419558307		
	<i>Campylobacter coli</i> 80352	987	419559505		

Cluster <sup>a</sup>	Strain <sup>b</sup>	Cas9 length (aa)	Cas9 GI	Cas1GI <sup>c</sup>	Subtype <sup>d</sup>
	<i>Campylobacter jejuni</i> subsp. <i>doylei</i> 269.97	984	153952471		
	<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 110-21	987	419676124		
	<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 129-258	987	419619138		
	<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 1336	987	283956897		
	<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 140-16	984	419681578		
	<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 1577	984	419685099		
	<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 1854	987	419689467		
	<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 1997-10	984	419666522		
	<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 2008-1025	987	419650041		
	<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 2008-872	984	419654778		
	<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 2008-979	987	419660762		
	<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 2008-988	965	419656328		
	<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 2008-988	984	419655317		
	<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 260.94	961	86152042		
	<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 414	985	283953849		
	<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 51037	984	419674189		
	<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 51494	984	419619463		
	<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 53161	987	419647275		
	<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 60004	984	419629136		
	<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 81116	984	157415744		
	<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 84-25	984	88596565		
	<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 87459	984	419680124		
	<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> ATCC 33560	984	419643715		
	<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> CF93-6	987	86149266		
	<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> CG8486	984	148925683		
	<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> HB93-13	984	86152450		
2 (continued)	<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> LMG 23210	987	419696801		Type II-C
	<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> LMG 23211	984	419697443		
	<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> LMG 23263	984	419628620		
	<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> LMG 23264	984	419632476		
	<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> LMG 23269	987	419634246		
	<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> LMG 23357	987	419641132		
	<b><i>Campylobacter jejuni</i> subsp. <i>jejuni</i> NCTC 11168</b>	<b>984</b>	<b>218563121</b>	<b>218563120</b>	
	<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> NW	983	424845990		
	<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> PT14	987	407942868		
	<i>Campylobacter lari</i>	1003	345468028		
	<i>Helicobacter canadensis</i> MIT 98-5491	1007	253828136		
	<i>Helicobacter cinaedi</i> ATCC BAA-847	1023	396079277		
	<i>Helicobacter cinaedi</i> CCUG 18818	1023	313144862		
	<i>Helicobacter cinaedi</i> PAGU611	1023	386762035		
	<i>Catellibacillus marimammalium</i> M35/04/3	1140	424780480		
	<i>Lactobacillus farciminis</i> KCTC 3681	1126	336394701		
	<i>Listeriaceae bacterium</i> TTU M1-001	1087	381184145		
	<i>Streptococcus anginosus</i> 1_2_62CV	1125	319939170		
	<i>Streptococcus gallolyticus</i> UCN34	1130	288905632		
3	<i>Streptococcus gordonii</i> str. Challis substr. CH1	1136	157150687		Type II-A
	<i>Streptococcus infantarius</i> ATCC BAA-102	1129	171779984		
	<i>Streptococcus macedonicus</i> ACA-DC 198	1130	374338350		
	<i>Streptococcus mitis</i> ATCC 6249	1134	306829274		
	<i>Streptococcus mutans</i> NLML5	1128	449203378		
	<i>Streptococcus mutans</i> NLML5	1128	450064617		



Cluster <sup>a</sup>	Strain <sup>b</sup>	Cas9 length (aa)	Cas9 GI	Cas1GI <sup>c</sup>	Subtype <sup>d</sup>
3 (continued)	<i>Streptococcus mutans</i> NLML8	1125	449151037		
	<i>Streptococcus mutans</i> NLML8	1125	450133520		
	<i>Streptococcus mutans</i> ST1	1134	449228751		
	<i>Streptococcus mutans</i> ST1	1134	450114718		
	<i>Streptococcus mutans</i> U2A	1125	449232458		
	<i>Streptococcus mutans</i> U2A	1125	450125471		
	<i>Streptococcus oralis</i> SK1074	1121	418974877		
	<i>Streptococcus oralis</i> SK313	1134	417940002		
	<i>Streptococcus parasanguinis</i> F0449	1140	419799964		
	<i>Streptococcus pasteurianus</i> ATCC 43144	1130	336064611		
	<i>Streptococcus salivarius</i> JIM8777	1127	387783792		
	<i>Streptococcus salivarius</i> PS4	1135	419707401		
	<i>Streptococcus</i> sp. BS35b	1026	401684660		
	<i>Streptococcus</i> sp. C150	1139	322372617		
	<i>Streptococcus</i> sp. GMD6S	1121	406576934		
	<i>Streptococcus suis</i> 89/1591	1122	223932525		
	<i>Streptococcus suis</i> D9	1122	386584496		
	<i>Streptococcus suis</i> ST3	1122	330833104		
	<i>Streptococcus thermophilus</i> CNRZ1066	1128	55822627		
	<i>Streptococcus thermophilus</i> JIM 8232	1121	386344353		
	<b><i>Streptococcus thermophilus</i> LMD-9</b>	<b>1121</b>	<b>116627542</b>	<b>116627543</b>	
<i>Streptococcus thermophilus</i> LMG 18311	1122	55820735			
<i>Streptococcus thermophilus</i> MN-ZLW-002	1121	387909441			
<i>Streptococcus thermophilus</i> MTCC 5460	1122	445374534			
<i>Streptococcus thermophilus</i> ND03	1121	386086348		Type II-A	
<i>Streptococcus vestibularis</i> ATCC 49124	1128	322517104			
4	<i>Actinobacillus minor</i> NM305	1056	240949037		
	<i>Actinobacillus pleuropneumoniae</i> serovar 10 str. D13039	1054	307256472		
	<i>Actinobacillus succinogenes</i> 130Z	1062	152978060		
	<i>Actinobacillus suis</i> H91-0380	1054	407692091		
	<i>Haemophilus parainfluenzae</i> ATCC 33392	1054	325578067		
	<i>Haemophilus parainfluenzae</i> CCUG 13788	1052	359298684		
	<i>Haemophilus parainfluenzae</i> T3T1	1052	345430422		
	<i>Haemophilus sputorum</i> HK 2154	1052	402304649		
	<i>Kingella kingae</i> PYKK081	1060	381401699		
	<i>Neisseria bacilliformis</i> ATCC BAA-1200	1077	329117879		
	<i>Neisseria cinerea</i> ATCC 14685	1082	261378287		
	<i>Neisseria flavescens</i> SK114	1081	241759613		Type II-C
	<i>Neisseria lactamica</i> 020-06	1082	313669044		
	<i>Neisseria meningitidis</i> 053442	1082	161869390		
	<i>Neisseria meningitidis</i> 2007056	1082	433531983		
	<i>Neisseria meningitidis</i> 63049	1082	433514137		
	<i>Neisseria meningitidis</i> 8013	1082	385324780		
	<i>Neisseria meningitidis</i> 92045	1082	421559784		
	<i>Neisseria meningitidis</i> 93003	1081	421538794		
	<i>Neisseria meningitidis</i> 93004	1081	421541126		
	<i>Neisseria meningitidis</i> 96023	1082	433518260		
<i>Neisseria meningitidis</i> 98008	1081	421555531			
<i>Neisseria meningitidis</i> alpha14	1082	254804356			
<i>Neisseria meningitidis</i> alpha275	1082	254672046			
<i>Neisseria meningitidis</i> ATCC 13091	1082	304388355			

Cluster <sup>a</sup>	Strain <sup>b</sup>	Cas9 length (aa)	Cas9 GI	Cas1GI <sup>c</sup>	Subtype <sup>d</sup>
	<i>Neisseria meningitidis</i> N1568	1081	416164244		
	<i>Neisseria meningitidis</i> NM140	1081	421545139		
	<i>Neisseria meningitidis</i> NM220	1082	418291220		
	<i>Neisseria meningitidis</i> NM233	1082	418288950		
	<i>Neisseria meningitidis</i> WUE 2594	1082	385337435		
	<b><i>Neisseria meningitidis</i> Z2491</b>	<b>1082</b>	<b>218767588</b>	<b>218767587</b>	
	<i>Neisseria</i> sp. oral taxon 14 str. F0314	1089	298369677		
	<i>Neisseria wadsworthii</i> 9715	1097	350570326		
	<i>Pasteurella multocida</i> subsp. <i>gallicida</i> X73	1058	425063822		
	<i>Pasteurella multocida</i> subsp. <i>multocida</i> str. P52VAC	1056	421263876		
	<b><i>Pasteurella multocida</i> subsp. <i>multocida</i> str. Pm70</b>	<b>1056</b>	<b>15602992</b>	<b>15602991</b>	
	<i>Simonsiella muelleri</i> ATCC 29453	1063	404379108		
	<i>Lactobacillus brevis</i> subsp. <i>gravesensis</i> ATCC 27305	1377	227509761		
	<i>Lactobacillus buchneri</i> CD034	1371	406027703		
	<i>Lactobacillus buchneri</i> NRRL B-30929	1371	331702228		
	<i>Lactobacillus casei</i> BL23	1361	191639137		
	<i>Lactobacillus casei</i> Lc-10	1361	418010298		
	<i>Lactobacillus casei</i> M36	1363	417996992		
	<i>Lactobacillus casei</i> str. Zhang	1361	301067199		
	<i>Lactobacillus casei</i> T71499	1360	417999832		
	<i>Lactobacillus casei</i> UCD174	1366	418002962		
	<i>Lactobacillus casei</i> W56	1389	409997999		
5	<i>Lactobacillus coryniformis</i> subsp. <i>coryniformis</i> KCTC 3167	1354	333394446		Type II-A
	<i>Lactobacillus curvatus</i> CRL 705	1368	354808135		
	<i>Lactobacillus fermentum</i> 28-3-CHN	1313	260662220		
	<i>Lactobacillus fermentum</i> ATCC 14931	1381	227514633		
	<i>Lactobacillus florum</i> 2F	1327	408790128		
	<i>Lactobacillus gasseri</i> JV-V03	1391	300361537		
	<i>Lactobacillus hominis</i> CRBIP 24.179	1386	395244248		
	<i>Lactobacillus jensenii</i> 269-3	1391	238854567		
	<i>Lactobacillus jensenii</i> 27-2-CHN	1395	256852176		
	<i>Lactobacillus johnsonii</i> DPC 6026	1375	385826041		
	<i>Lactobacillus mucosae</i> LM1	1382	377831443		
	<i>Lactobacillus paracasei</i> subsp. <i>paracasei</i> 8700:2	1362	239630053		
	<i>Lactobacillus pentosus</i> IG1	1382	339637353		
	<i>Lactobacillus pentosus</i> KCA1	1361	392947436		
	<i>Lactobacillus pentosus</i> MP-10	1358	334881121		
	<i>Lactobacillus plantarum</i> ZJ316	1358	448819853		
	<b><i>Lactobacillus rhamnosus</i> GG</b>	<b>1363</b>	<b>258509199</b>	<b>258509198</b>	
	<i>Lactobacillus rhamnosus</i> HN001	1361	199597394		
5	<i>Lactobacillus rhamnosus</i> R0011	1361	418072660		Type II-A
(continued)	<i>Lactobacillus ruminis</i> ATCC 25644	1375	323340068		
	<i>Lactobacillus salivarius</i> SMXD51	1339	418960525		
	<i>Lactobacillus sanfranciscensis</i> TMW 1.1304	1331	347534532		
	<i>Lactobacillus</i> sp. 66c	1419	408410332		
	<i>Pediococcus acidilactici</i> DSM 20284	1364	304386254		
	<i>Pediococcus acidilactici</i> MA18/5M	1366	418068659		
	<i>Psychroflexus torquus</i> ATCC 700755	1509	408489713		

Cluster <sup>a</sup>	Strain <sup>b</sup>	Cas9 length (aa)	Cas9 GI	Cas1GI <sup>c</sup>	Subtype <sup>d</sup>
6	<i>Anaerophaga</i> sp. HS1	1552	371776944		
	<i>Anaerophaga thermohalophila</i> DSM 12881	1515	346224232		
	<i>Bacteroides coprophilus</i> DSM 18228	1509	224026357		
	<i>Bacteroides coprosuis</i> DSM 18011	1504	333031006		
	<i>Bacteroides dorei</i> DSM 17855	1504	212694363		
	<i>Bacteroides eggerthii</i> 1_2_48FAA	1509	317474201		
	<i>Bacteroides faecis</i> 27-5	1526	380696107		
	<i>Bacteroides fluxus</i> YIT 12057	1509	329965125		
	<i>Bacteroides nordii</i> CL02T12C05	1512	393788929		
	<b><i>Bacteroides</i> sp. 20_3</b>	<b>1517</b>	<b>301311869</b>	<b>301311870</b>	Type II-C
	<i>Bacteroides</i> sp. D2	1510	383115507		
	<i>Bacteroides uniformis</i> CL03T00C23	1508	423303159		
	<i>Bacteroides vulgatus</i> CL09T03C04	1504	423312075		
	<i>Capnocytophaga gingivalis</i> ATCC 33624	1436	228473057		
	<i>Capnocytophaga</i> sp. CM59	1437	402830627		
	<i>Capnocytophaga</i> sp. oral taxon 324 str. F0483	1471	429756885		
	<i>Capnocytophaga</i> sp. oral taxon 326 str. F0382	1450	429752492		
	<i>Capnocytophaga</i> sp. oral taxon 412 str. F0487	1450	393778597		
	<i>Chryseobacterium</i> sp. CF314	1419	399023756		
	<i>Fibrobacter succinogenes</i> subsp. <i>succinogenes</i> S85	1512	261414553		
	<i>Flavobacteriaceae</i> bacterium S85	1516	372210605		
	<i>Flavobacterium columnare</i> ATCC 49512	1459	365960762		
	<i>Fluviicola taffensis</i> DSM 16823	1458	327405121		
	<i>Mucilaginibacter paludis</i> DSM 18603	1473	373954054		
	<i>Myroides odoratus</i> DSM 2801	1466	374597806		
	<i>Ornithobacterium rhinotracheale</i> DSM 15997	1535	392391493		Type II-C
	<i>Prevotella bivia</i> JCVIHMP010	1485	282858617		
	<i>Prevotella buccae</i> ATCC 33574	1457	315607525		
	<i>Prevotella nigrescens</i> ATCC 33563	1506	340351024		
	<i>Prevotella</i> sp. MSX73	1483	402307189		
	<i>Prevotella timonensis</i> CRIS 5C-B1	1487	282881485		
	<i>Prevotella veroralis</i> F0319	1496	260592128		
<i>Sphingobacterium spiritivorum</i> ATCC 33861	1426	300771242			
<i>Weeksella virosa</i> DSM 16922	1440	325955459			
7	<i>Bacteroides fragilis</i> 638R	1436	375360193		
	<b><i>Bacteroides fragilis</i> NCTC 9343</b>	<b>1436</b>	<b>60683389</b>	<b>60683388</b>	
	<i>Bacteroides</i> sp. 2_1_16	1436	265767599		
	<i>Bacteroides</i> sp. 3_1_19	1424	298377533		
	<i>Bacteroides</i> sp. D2	1436	383110723		
	<i>Bacteroidetes</i> oral taxon 274 str. F0058	1434	298373376		
	<i>Belliella baltica</i> DSM 15883	1352	390944707		
	<i>Bergeyella zoohelcum</i> CCUG 30536	1430	406673990		Type II-C
	<i>Capnocytophaga canimorsus</i> Cc5	1430	340622236		
	<i>Capnocytophaga ochracea</i> DSM 7271	1426	256819408		
	<i>Capnocytophaga</i> sp. oral taxon 329 str. F0087	1435	332882466		
	<i>Capnocytophaga</i> sp. oral taxon 335 str. F0486	1426	420149252		
	<i>Capnocytophaga</i> sp. oral taxon 380 str. F0488	1432	429748017		
	<i>Capnocytophaga sputigena</i> Capno	1426	213962376		
	<i>Flavobacterium psychrophilum</i> JIP02/86	1354	150025575		
	7 (continued)	<i>Galbibacter</i> sp. ck-12-15	1391	408370397	
<i>Indibacter alkaliphilus</i> LW1		1354	404451234		

Cluster <sup>a</sup>	Strain <sup>b</sup>	Cas9 length (aa)	Cas9 GI	Cas1GI <sup>c</sup>	Subtype <sup>d</sup>
	<i>Joostella marina</i> DSM 19592	1397	386818981		
	<i>Kordia algicida</i> OT-1	1391	163754820		
	<i>Marinilabilia</i> sp. AK2	1345	410030899		
	<i>Myroides injenensis</i> M09-0166	1401	399927444		
	<i>Niabella soli</i> DSM 19437	1426	374372722		
	<i>Parabacteroides johnsonii</i> DSM 18315	1443	218258638		
	<i>Parabacteroides</i> sp. D13	1424	256840409		
	<i>Prevotella histicola</i> F0411	1375	357042839		
	<i>Prevotella intermedia</i> 17	1380	387132277		
	<i>Prevotella nigrescens</i> F0103	1380	445119230		
	<i>Prevotella oralis</i> ATCC 33269	1391	323344874		
	<i>Prevotella</i> sp. oral taxon 306 str. F0472	1375	383811446		
	<i>Riemerella anatipestifer</i> RA-CH-1	1405	407451859		
	<i>Riemerella anatipestifer</i> RA-GD	1400	386321727		
	<i>Zunongwangia profunda</i> SM-A87	1388	295136244		
	<i>Actinomyces coleocanis</i> DSM 15436	1105	227494853		
	<i>Actinomyces georgiae</i> F0490	1113	420151340		
	<i>Actinomyces naeslundii</i> str. Howell 279	1101	400293272		
	<i>Actinomyces</i> sp. ICM47	1144	396585058		
	<i>Actinomyces</i> sp. oral taxon 175 str. F0384	1095	343523232		
	<i>Actinomyces</i> sp. oral taxon 181 str. F0379	1103	429758968		
	<i>Actinomyces</i> sp. oral taxon 848 str. F0332	1120	269219760		
	<i>Actinomyces turicensis</i> ACS-279-V-Col4	1114	405979650		
	<i>Bifidobacterium dentium</i> Bd1	1138	283456135		
	<b><i>Bifidobacterium longum</i> DJO10A</b>	<b>1187</b>	<b>189440764</b>	189440765	
	<i>Bifidobacterium longum</i> subsp. <i>longum</i> 2-2B	1124	419852381		
	<i>Bifidobacterium longum</i> subsp. <i>longum</i> KACC 91563	1138	384200944		
	<i>Bifidobacterium</i> sp. 12_1_47BFAA	1151	317482066	<b>317482065</b>	
	<i>Corynebacterium accolens</i> ATCC 49725	1099	227502575		
	<i>Corynebacterium accolens</i> ATCC 49726	1099	306835141		
8	<i>Corynebacterium diphtheriae</i> 241	1084	375289763		Type II-C
	<i>Corynebacterium diphtheriae</i> 31A	1084	376283539		
	<i>Corynebacterium diphtheriae</i> BH8	1084	376286566		
	<i>Corynebacterium diphtheriae</i> bv. <i>intermedius</i> str. NCTC 5011	1084	419861895		
	<i>Corynebacterium diphtheriae</i> C7 (beta)	1084	376289243		
	<i>Corynebacterium diphtheriae</i> HC02	1084	376292154		
	<i>Corynebacterium diphtheriae</i> NCTC 13129	1084	38232678		
	<i>Corynebacterium diphtheriae</i> VA01	1084	376256051		
	<i>Corynebacterium matruchotii</i> ATCC 14266	1089	305681510		
	<i>Corynebacterium matruchotii</i> ATCC 33806	1069	225021644		
	<i>Gardnerella vaginalis</i> 1500E	1186	415717744		
	<i>Gardnerella vaginalis</i> 284V	1186	415703177		
	<i>Gardnerella vaginalis</i> 5-1	1186	298252606		
	<i>Mobiluncus curtisii</i> subsp. <i>holmesii</i> ATCC 35242	1123	315656340		
	<i>Mobiluncus mulieris</i> 28-1	1091	269977848		
	<i>Mobiluncus mulieris</i> FB024-16	1091	307700167		
	<i>Scardovia inopinata</i> F0304	1178	294790575		
9	<i>Bacillus cereus</i> BAG4X12-1	1068	423439645		Type II-C
	<i>Bacillus cereus</i> BAG4X2-1	1078	423445130		
	<i>Bacillus cereus</i> Rock1-15	1069	229113166		

Cluster <sup>a</sup>	Strain <sup>b</sup>	Cas9 length (aa)	Cas9 GI	Cas1GI <sup>c</sup>	Subtype <sup>d</sup>
	<b><i>Bacillus smithii</i> 7_3_47FAA</b>	<b>1088</b>	<b>365156657</b>	365156658	
	<i>Bacillus thuringiensis</i> serovar <i>finitimus</i> YBT-020	1069	384183447		
	<i>Brevibacillus laterosporus</i> GI-9	1092	421874297	<b>421874296</b>	
	<i>Clostridium perfringens</i> C str. JGS1495	1065	169343975		
	<i>Clostridium perfringens</i> D str. JGS1721	1065	182624245		
	<i>Sporolactobacillus vineae</i> DSM 21990 = SL153	1084	404330915		
10	<i>Gemella haemolysans</i> ATCC 10379	1392	241889924		Type II-A
	<i>Gemella morbillorum</i> M424	1385	317495358		
	<i>Megasphaera</i> sp. UPII 135-E	1352	342218215		
10	<b><i>Veillonella atypica</i> ACS-134-V-Col7a</b>	<b>1398</b>	<b>303229466</b>	<b>303229394</b>	Type II-A
(continued)	<i>Veillonella parvula</i> ATCC 17745	1398	282849530		
	<i>Veillonella</i> sp. 6_1_27	1395	294792465		
	<i>Veillonella</i> sp. oral taxon 780 str. F0422	1120	342213964		
	<i>Treponema denticola</i> AL-2	1395	449103686		
	<i>Treponema denticola</i> ASLM	1395	449106292		
11	<b><i>Treponema denticola</i> ATCC 35405</b>	<b>1395</b>	<b>42525843</b>	<b>42525844</b>	Type II-A
	<i>Treponema denticola</i> H1-T	1395	449118593		
	<i>Treponema denticola</i> H-22	1395	449117322		
	<i>Treponema denticola</i> OTK	1395	449125136		
	<i>Treponema denticola</i> SP37	1395	449130155		
	<b><i>Mycoplasma canis</i> PG 14</b>	<b>1233</b>	<b>384393286</b>	<b>384393287</b>	
	<i>Mycoplasma canis</i> PG 14	1233	419703974		
12	<i>Mycoplasma canis</i> UF31	1233	384937953		Type II-A
	<i>Mycoplasma canis</i> UF33	1233	419704625		
	<i>Mycoplasma canis</i> UFG1	1233	419705269		
	<i>Mycoplasma canis</i> UFG4	1233	419705920		
	<i>Mycoplasma cynos</i> C142	1239	433625054		
	<i>Enterococcus faecalis</i> Fly1	1150	257084992		
	<i>Enterococcus faecalis</i> R508	1150	424761124		
	<i>Enterococcus faecalis</i> T11	1150	257419486		
13	<b><i>Enterococcus faecalis</i> TX0012</b>	<b>1150</b>	<b>315149830</b>	<b>315149831</b>	Type II-A
	<i>Enterococcus faecalis</i> TX0012	1150	422729710		
	<i>Enterococcus faecalis</i> TX1342	1150	422701955		
	<i>Facklamia hominis</i> CCUG 36813	1142	406671118		
	<i>Gluconacetobacter diazotrophicus</i> PAI 5	1003	209542524		
	<i>Gluconacetobacter diazotrophicus</i> PAI 5	1050	162147907		
	<i>Methylocystis</i> sp. ATCC 49242	1080	323139312		
14	<b><i>Methylosinus trichosporium</i> OB3b</b>	<b>1082</b>	<b>296446027</b>	<b>296446028</b>	Type II-C
	<i>Rhodopseudomonas palustris</i> BisB18	1066	90425961		
	<i>Rhodopseudomonas palustris</i> BisB5	1064	91975509		
	<i>Tistrella mobilis</i> KA081020-065	1049	389874754		
	<i>Francisella</i> cf. <i>novicida</i> 3523	1646	387824704		
	<i>Francisella</i> cf. <i>novicida</i> Fx1	1629	385792694		
	<i>Francisella novicida</i> FTG	1629	208779141		
15	<i>Francisella novicida</i> GA99-3548	1629	254374175		Type II-B
	<b><i>Francisella novicida</i> U112</b>	<b>1629</b>	<b>118497352</b>	<b>118497353</b>	
	<i>Francisella tularensis</i> subsp. <i>novicida</i> GA99-3549	1629	254372717		
	<i>Acidovorax avenae</i> subsp. <i>avenae</i> ATCC 19860	1045	326315085		
	<i>Alicyclophilus denitrificans</i> BC	1029	319760940		
16	<b><i>Alicyclophilus denitrificans</i> K601</b>	<b>1029</b>	<b>330822845</b>	<b>330822846</b>	Type II-C
	gamma proteobacterium HdN1	1025	304313029		

Cluster <sup>a</sup>	Strain <sup>b</sup>	Cas9 length (aa)	Cas9 GI	Cas1GI <sup>c</sup>	Subtype <sup>d</sup>
	<i>Nitrosomonas</i> sp. AL212	1044	325983496		
	<i>Verminephrobacter eiseniae</i> EF01-2	1068	121608211		
17	<i>Mycoplasma gallisepticum</i> NC95_13295-2-2P	1269	401767318		
	<i>Mycoplasma gallisepticum</i> NY01_2001.047-5-1P	1224	401768851		
	<b><i>Mycoplasma gallisepticum</i> str. F</b>	<b>1269</b>	<b>284931710</b>	<b>284931711</b>	Type II-A
	<i>Mycoplasma gallisepticum</i> str. F	1269	385326554		
	<i>Mycoplasma gallisepticum</i> str. R(low)	1270	294660600		
18	<i>Prevotella buccalis</i> ATCC 35310	1218	282878504		
	<i>Prevotella ruminicola</i> 23	1204	294674019		
	<i>Prevotella stercorea</i> DSM 18206	1216	359406728		Type II-C
	<i>Prevotella tanneriae</i> ATCC 51259	1234	258648111		
	<b><i>Prevotella timonensis</i> CRIS 5C-B1</b>	<b>1218</b>	<b>282880052</b>	<b>282880053</b>	
19	<i>Phascolarctobacterium succinatutens</i> YIT 12067	1087	323142435		
	<i>Roseburia intestinalis</i> L1-82	1140	257413184		
	<i>Roseburia intestinalis</i> M50/1	1128	291537230		Type II-C
	<b><i>Roseburia inulinivorans</i> DSM 16841</b>	<b>1152</b>	<b>225377804</b>	<b>225377803</b>	
	<i>Subdoligranulum</i> sp. 4_3_54A2FAA	1084	365132400		
20	<b><i>Coriobacterium glomerans</i> PW2</b>	<b>1384</b>	<b>328956315</b>	<b>328956316</b>	Type II-A
	<i>Eggerthella</i> sp. YY7918	1380	339445983		
20	<i>Gordonibacter pamelaee</i> 7-10-1-b	1371	295106015		Type II-A
(continued)	<i>Olsenella uli</i> DSM 7084	1399	302336020		Type II-A
21	<b><i>Fusobacterium nucleatum</i> subsp. <i>vincentii</i> ATCC 49256</b>	<b>1374</b>	<b>34762592</b>	34762593	Type II-A
	<i>Fusobacterium</i> sp. 1_1_41FAA	1367	294782278		
	<i>Fusobacterium</i> sp. 3_1_27	1367	294785695		
	<i>Fusobacterium</i> sp. 3_1_36A2	1367	256845019	<b>256845020</b>	
22	<i>Fingoldia magna</i> ACS-171-V-Col3	1347	302380288		
	<b><i>Fingoldia magna</i> ATCC 29328</b>	<b>1348</b>	<b>169823755</b>	<b>169823756</b>	Type II-A
	<i>Fingoldia magna</i> SY403409CC001050417	1348	417926052		
	<i>Helcococcus kunzii</i> ATCC 51366	1338	375092427		
23	<i>Prevotella denticola</i> CRIS 18C-A	1422	325859619		
	<i>Prevotella micans</i> F0438	1425	373501184		Type II-C
	<b><i>Prevotella</i> sp. C561</b>	<b>1424</b>	<b>345885718</b>	<b>345885719</b>	
24	<i>Leuconostoc gelidum</i> KCTC 3527	1355	333398273		
	<b><i>Oenococcus kitaharae</i> DSM 17330</b>	<b>1389</b>	<b>366983953</b>	<b>366983954</b>	Type II-A
	<i>Oenococcus kitaharae</i> DSM 17330	1389	372325145		
25	<i>Anaerococcus tetradius</i> ATCC 35098	1361	227501312		
	<i>Lactobacillus iners</i> LactinV 11V1-d	1369	309803917		Type II-A
	<b><i>Peptoniphilus duerdenii</i> ATCC BAA-1640</b>	<b>1364</b>	<b>304438954</b>	<b>304438953</b>	
26	<b><i>Coprococcus catus</i> GD17</b>	<b>1338</b>	<b>291520705</b>	<b>291520706</b>	Type II-A
	<i>Dorea longicatena</i> DSM 13814	1340	153855454		
	<i>Ruminococcus lactaris</i> ATCC 29176	1341	197301447		
27	<b><i>Staphylococcus pseudintermedius</i> ED99</b>	<b>1334</b>	<b>323463801</b>	<b>323463802</b>	Type II-A
	<i>Staphylococcus pseudintermedius</i> ED99	1334	386318630		
	<i>Staphylococcus simulans</i> ACS-120-V-Sch1	1112	414160476		
28	<b><i>Dinoroseobacter shibae</i> DFL 12</b>	<b>1079</b>	<b>159042956</b>	<b>159042957</b>	Type II-C
	<i>Sphingobium</i> sp. AP49	1110	398385143		
	<i>Sphingomonas</i> sp. S17	1090	332188827		
29	<b><i>Flavobacterium branchiophilum</i> FL-15</b>	<b>1473</b>	<b>347536497</b>	no cas1	Type II-C
	<i>Flavobacterium columnare</i> ATCC 49512	1535	365959402		

Cluster <sup>a</sup>	Strain <sup>b</sup>	Cas9 length (aa)	Cas9 GI	Cas1GI <sup>c</sup>	Subtype <sup>d</sup>
30	<i>Bifidobacterium bifidum</i> S17	1420	310286728	310286727	Type II-A
	<i>Scardovia wiggisiae</i> F0424	1471	423349694		
31	<i>Burkholderiales</i> bacterium 1_1_47	1428	303257695		Type II-B
	<i>Parasutterella excrementihominis</i> YIT 11859	1428	331001027	331001028	
32	<i>Streptococcus sanguinis</i> SK49	1421	422884106	422884107	Type II-A
	<i>Streptococcus</i> sp. oral taxon 71 str. 73H25AP	1420	306826314		
33	<i>Eubacterium</i> sp. AS15	1391	402309258		Type II-A
	<i>Eubacterium yurii</i> subsp. <i>margaretiae</i> ATCC 43715	1391	306821691	306821690	
34	<i>Legionella pneumophila</i> 130b	1372	307608922		Type II-B
	<i>Legionella pneumophila</i> str. Paris	1372	54296138	54296139	
35	<i>Acidaminococcus intestini</i> RyC-MR95	1358	352684361		Type II-A
	<i>Acidaminococcus</i> sp. D21	1358	227824983	227824982	
36	<i>Lactobacillus farciminis</i> KCTC 3681	1356	336394882	336394883	Type II-A
	<i>Lactobacillus versmoldensis</i> KCTC 3814	1289	365906066		
37	<i>Mycoplasma synoviae</i> 53	1304	144575181		Type II-A
	<i>Mycoplasma synoviae</i> 53	1314	71894592	71894593	
38	<i>Elusimicrobium minutum</i> Pei191	1195	187250660	187250661	Type II-C
	uncultured Termite group 1 bacterium phylotype Rs-D17	1032	189485059		
39	<i>Clostridium spiroforme</i> DSM 1552	1116	169349750		Type II-A
	<i>Eubacterium dolichum</i> DSM 3991	1096	160915782	160915783	
40	<i>Eubacterium rectale</i> ATCC 33656	1114	238924075	238924076	Type II-A
	<i>Eubacterium ventriosum</i> ATCC 27560	1107	154482474		
41	<i>Staphylococcus aureus</i> subsp. <i>aureus</i>	1053	403411236		Type II-A
	<i>Staphylococcus lugdunensis</i> M23590	1054	315659848	315659847	
42	<i>Ignavibacterium album</i> JCM 16511	1688	385811609	385811610	Type II-C
43	<i>Odoribacter laneus</i> YIT 12061	1498	374384763	374384762	Type II-C
44	<i>Caenispirillum salinarum</i> AK4	1442	427429481	427429479	Type II-C
45	<i>Sutterella wadsworthensis</i> 3_1_45B	1422	319941583	319941582	Type II-B
46	<i>Bergeyella zoohelcum</i> ATCC 43767	1415	423317190	423317188	Type II-C
47	<i>Wolinella succinogenes</i> DSM 1740	1409	34557932	34557933	Type II-B
48	<i>gamma proteobacterium</i> HTCC5015	1397	254447899	no cas1	Type II-B
49	<i>Filifactor alocis</i> ATCC 35896	1365	374307738	374307737	Type II-A
50	<i>Planococcus antarcticus</i> DSM 14505	1333	389815359	389815358	Type II-A
51	<i>Catenibacterium mitsuokai</i> DSM 15897	1329	224543312	224543313	Type II-A
52	<i>Solobacterium moorei</i> F0204	1327	320528778	320528779	Type II-A
53	<i>Fructobacillus fructosus</i> KCTC 3544	1323	339625081	339625080	Type II-A
54	<i>Mycoplasma ovipneumoniae</i> SC01	1265	363542550	363542551	Type II-A
54	<i>Streptobacillus moniliformis</i> DSM 12112	1259	269123826		
55	<i>Mycoplasma mobile</i> 163K	1236	47458868	47458867	Type II-A
56	<i>Porphyromonas</i> sp. oral taxon 279 str. F0450	1197	402847315	402847305	Type II-C
57	<i>Actinomyces</i> sp. oral taxon 180 str. F0310	1181	315605738	315605739	Type II-C
58	<i>Sphaerochaeta globus</i> str. Buddy	1179	325972003	325972002	Type II-C
59	<i>Rhodospirillum rubrum</i> ATCC 11170	1173	83591793	83591790	Type II-C
60	<i>Azospirillum</i> sp. B510	1168	288957741	288957738	Type II-C
61	<i>Nitrobacter hamburgensis</i> X14	1166	92109262	no cas1	Type II-C
62	<i>Ruminococcus albus</i> 8	1156	325677756	325677757	Type II-C
63	<i>Barnesiella intestinhominis</i> YIT 11860	1153	404487228	404487227	Type II-C
64	<i>Alicyclobacillus hesperidum</i> URH17-3-68	1146	403744858	403744859	Type II-C

Cluster <sup>a</sup>	Strain <sup>b</sup>	Cas9 length (aa)	Cas9 GI	Cas1GI <sup>c</sup>	Subtype <sup>d</sup>
65	<i>Acidothermus cellulolyticus</i> 11B	1138	117929158	117929157	Type II-C
66	<i>Nitratifractor salsuginis</i> DSM 16511	1132	319957206	319957207	Type II-C
67	<i>Acidovorax ebreus</i> TPSY	1131	222109285	222109284	Type II-C
67	<i>Francisella tularensis</i> subsp. <i>tularensis</i> WY96-3418	1125	134302318		
68	<i>Lactobacillus coryniformis</i> subsp. <i>torquens</i> KCTC 3535	1119	336393381	336393380	Type II-C
69	<i>Alcanivorax</i> sp. W11-5	1113	407803669	407803668	Type II-C
70	<i>Akkermansia muciniphila</i> ATCC BAA-835	1101	187736489	187736488	Type II-C
71	<i>Ilyobacter polytropus</i> DSM 2926	1092	310780384	310780383	Type II-C
72	<i>Bradyrhizobium</i> sp. BTAi1	1064	148255343	no cas1	Type II-C
73	<i>Ralstonia syzygii</i> R24	1062	344171927	344171926	Type II-C
74	<i>Treponema</i> sp. JC4	1062	384109266	384109265	Type II-C
75	<i>Wolinella succinogenes</i> DSM 1740	1059	34557790	34557789	Type II-C
76	<i>Rhodovulum</i> sp. PH10	1059	402849997	402849996	Type II-C
77	<i>Aminomonas paucivorans</i> DSM 12260	1052	312879015	312879014	Type II-C
77	<i>Bacteroides</i> sp. 3_1_33FAA	1055	265750948		
78	<i>Parvibaculum lavamentivorans</i> DS-1	1037	154250555	154250554	Type II-C
79	<i>Candidatus Puniceispirillum marinum</i> IMCC1322	1035	294086111	294086112	Type II-C
80	<i>Blastopirellula marina</i> DSM 3645	1027	87307579		
80	<i>Helicobacter mustelae</i> 12198	1024	291276265	291276264	Type II-C
81	<i>Clostridium cellulolyticum</i> H10	1021	220930482	220930481	Type II-C
82	<i>Lactobacillus crispatus</i> FB077-07	857	423321767		
82	uncultured delta proteobacterium HF0070_07E19	1011	297182908	no cas1	Type II-C
	<i>Acetobacter aceti</i> NBRC 14818	240	340779894		
	<i>Acetobacter aceti</i> NBRC 14818	376	340779669		
	<i>Acetobacter aceti</i> NBRC 14818	400	340779439		
	<i>Actinobacillus ureae</i> ATCC 25976	239	322514756		
	<i>Actinobacillus ureae</i> ATCC 25976	400	322514772		
	<i>Bacillus cereus</i> BAG2X1-3	333	423408783		
	<i>Bacteroides cellulosilyticus</i> DSM 14838	206	224535831		
	<i>Bacteroides cellulosilyticus</i> DSM 14838	1219	224535832		
	<i>Bacteroides coprosuis</i> DSM 18011	349	333031028		
	<i>Bacteroides oleiciplenus</i> YIT 12058	653	427387687		
	<i>Bacteroides oleiciplenus</i> YIT 12058	779	427387686		
	<i>Bacteroides</i> sp. 9_1_42FAA	1055	237710146		
	<i>Bacteroides uniformis</i> CL03T12C37	286	423308124		
	<i>Bacteroides uniformis</i> CL03T12C37	1210	423308121		
	<i>Bifidobacterium bifidum</i> IPLA 20015	1281	421736922		
	<i>Bifidobacterium dentium</i> ATCC 27678	1121	171742822		
	<i>Bifidobacterium longum</i> subsp. <i>longum</i> 1-6B	182	419848319		
	<i>Bifidobacterium longum</i> subsp. <i>longum</i> 1-6B	354	419847807		
	<i>Bifidobacterium longum</i> subsp. <i>longum</i> 1-6B	441	419848320		
	<i>Bifidobacterium longum</i> subsp. <i>longum</i> 44B	166	419856168		
	<i>Bifidobacterium longum</i> subsp. <i>longum</i> 44B	967	419856216		
	<i>Butyrivibrio fibrisolvens</i> 16/4	103	291518094		
	<i>Butyrivibrio fibrisolvens</i> 16/4	177	291518096		
	<i>Butyrivibrio fibrisolvens</i> 16/4	765	291518097		
	<i>Campylobacter coli</i> 2685	933	419548338		



Cluster <sup>a</sup>	Strain <sup>b</sup>	Cas9 length (aa)	Cas9 GI	Cas1GI <sup>c</sup>	Subtype <sup>d</sup>
	<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 2008-894	666	419652996		
	<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 305	190	317510779		
	<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 305	759	317510780		
	<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 327	462	415747744		
	<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 327	512	415747743		
	<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> CG8421	721	205356639		
	<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> M1	861	384442103		
	candidate division TM7 single-cell isolate TM7c	372	167957190		
	<i>Capnocytophaga ochracea</i> F0287	303	315224863		
	<i>Capnocytophaga ochracea</i> F0287	1117	315224862		
	<i>Coprococcus comes</i> ATCC 27758	686	226325213		
	<i>Diplosphaera colitermitum</i> TAV2	210	225184109		
	<i>Enterococcus faecalis</i> TX1467	921	422867931		
	<i>Enterococcus faecalis</i> TX4248	936	307270261		
	<i>Enterococcus faecium</i> E2620	892	431752788		
	<i>Enterococcus</i> sp. 7L76	116	295113136		
	<i>Francisella tularensis</i> subsp. <i>holarctica</i> 257	878	254367943		
	<i>Francisella tularensis</i> subsp. <i>holarctica</i> FSC022	158	254369498		
	<i>Francisella tularensis</i> subsp. <i>holarctica</i> FSC022	244	254369502		
	<i>Francisella tularensis</i> subsp. <i>holarctica</i> FSC022	292	254369497		
	<i>Francisella tularensis</i> subsp. <i>holarctica</i> FSC022	393	254369499		
	<i>Francisella tularensis</i> subsp. <i>holarctica</i> FSC022	501	254369496		
	<i>Francisella tularensis</i> subsp. <i>holarctica</i> LVS	158	89256630		
	<i>Francisella tularensis</i> subsp. <i>holarctica</i> LVS	393	89256631		
	<i>Francisella tularensis</i> subsp. <i>holarctica</i> URFT1	53	290953529		
	<i>Francisella tularensis</i> subsp. <i>holarctica</i> URFT1	285	290953528		
	<i>Francisella tularensis</i> subsp. <i>tularensis</i> SCHU S4	1123	56707712		
	<i>Gemella haemolysans</i> M341	1258	329766883		
	<i>Haemophilus pittmaniae</i> HK 85	121	343519651		
	<i>Haemophilus pittmaniae</i> HK 85	203	343519677		
	<i>Haemophilus pittmaniae</i> HK 85	650	343519679		
	<i>Helicobacter hepaticus</i> ATCC 51449	131	32266975		
	<i>Helicobacter pullorum</i> MIT 98-5489	344	242308998		
	<i>Helicobacter pullorum</i> MIT 98-5489	702	242309214		
	<i>Kingella kingae</i> ATCC 23330	1000	333374624		
	<i>Lactobacillus buchneri</i> ATCC 11577	1239	227512703		
	<i>Lactobacillus casei</i> 21/1	234	417984225		
	<i>Lactobacillus casei</i> 21/1	1128	417984226		
	<i>Lactobacillus casei</i> CRF28	566	417994652		
	<i>Lactobacillus casei</i> CRF28	700	417993346		
	<i>Lactobacillus casei</i> UW1	315	418005912		
	<i>Lactobacillus casei</i> UW1	330	418005913		
	<i>Lactobacillus casei</i> UW1	412	418005908		
	<i>Lactobacillus casei</i> UW4	236	418008739		
	<i>Lactobacillus casei</i> UW4	330	418008740		
	<i>Lactobacillus crispatus</i> 214-1	534	293381764		
	<i>Lactobacillus crispatus</i> CTV-05	298	312978192		
	<i>Lactobacillus crispatus</i> FB049-03	206	423318602		
	<i>Lactobacillus crispatus</i> FB049-03	347	423318603		
	<i>Lactobacillus crispatus</i> FB049-03	857	423318600		
	<i>Lactobacillus crispatus</i> JV-V01	278	227878395		

Cluster <sup>a</sup>	Strain <sup>b</sup>	Cas9 length (aa)	Cas9 GI	Cas1 GI <sup>c</sup>	Subtype <sup>d</sup>
	<i>Lactobacillus crispatus</i> JV-V01	544	227878705		
	<i>Lactobacillus crispatus</i> MV-1A-US	277	256850790		
	<i>Lactobacillus crispatus</i> MV-1A-US	538	256850346		
	<i>Lactobacillus crispatus</i> MV-3A-US	279	262048056		
	<i>Lactobacillus delbrueckii</i> subsp. <i>bulgaricus</i> 2038	544	385815564		
	<i>Lactobacillus delbrueckii</i> subsp. <i>bulgaricus</i> 2038	669	385815562		
	<i>Lactobacillus iners</i> LactinV 09V1-c	255	309804524		
	<i>Lactobacillus iners</i> LactinV 09V1-c	343	309804534		
	<i>Lactobacillus iners</i> LactinV 09V1-c	447	309804536		
	<i>Lactobacillus iners</i> SPIN 2503V10-D	270	309809475		
	<i>Lactobacillus iners</i> SPIN 2503V10-D	667	309809480		
	<i>Lactobacillus ruminis</i> ATCC 25644	1352	417973941		
	<i>Lactobacillus salivarius</i> ACS-116-V-Col5a	629	301299400		
	<i>Lactobacillus salivarius</i> CECT 5713	897	385839899		
	<i>Lactobacillus salivarius</i> UCC118	1149	90961083		
	<i>Leptospira inadai</i> serovar Lyme str. 10	125	398345609		
	<i>Leptospira inadai</i> serovar Lyme str. 10	418	398341884		
	<i>Leptospira inadai</i> serovar Lyme str. 10	907	398345610		
	<i>Leuconostoc pseudomesenteroides</i> 4882	468	399517481		
	<i>Leuconostoc pseudomesenteroides</i> 4882	883	399517482		
	<i>Listeria ivanovii</i> FSL F6-596	232	315301622		
	<i>Listeria ivanovii</i> FSL F6-596	849	315301624		
	<i>Listeria monocytogenes</i> FSL F2-208	782	422410878		
	<i>Listeria monocytogenes</i> FSL J1-208	300	255024093		
	<i>Listeria seeligeri</i> FSL N1-067	874	313631816		
	<i>Listeria seeligeri</i> FSL N1-067	874	422420175		
	<i>Maritimibacter alkaliphilus</i> HTCC2654	997	84685065		
	<i>Mycoplasma iowae</i> 695	226	350547050		
	<i>Mycoplasma iowae</i> 695	933	350546886		
	<i>Neisseria lactamica</i> ATCC 23970	408	269215119		
	<i>Neisseria lactamica</i> ATCC 23970	666	269215120		
	<i>Neisseria lactamica</i> Y92-1009	241	422110930		
	<i>Neisseria lactamica</i> Y92-1009	828	422110931		
	<i>Neisseria meningitidis</i> NM3001	67	421568320		
	<i>Neisseria meningitidis</i> NM3001	976	421568319		
	<i>Neisseria mucosa</i> C102	220	319639577		
	<i>Neisseria</i> sp. oral taxon 20 str. F0370	392	429743981		
	<i>Neisseria</i> sp. oral taxon 20 str. F0370	701	429743980		
	<i>Neisseria subflava</i> NJ9703	587	284799897		
	<i>Nitritalea hafalkaliphila</i> LW7	79	390445315		
	<i>Nitrobacter hamburgensis</i> X14	641	92118334		
	<i>Oribacterium sinus</i> F0268	653	227873236		
	<i>Parabacteroides merdae</i> ATCC 43184	103	154493351		
	<i>Parabacteroides merdae</i> CL03T12C32	84	423346601		
	<i>Parabacteroides merdae</i> CL09T00C40	82	423723156		
	<i>Pasteurella bettyae</i> CCUG 2042	398	387770127		
	<i>Pasteurella bettyae</i> CCUG 2042	610	387770112		
	<i>Pasteurella multocida</i> subsp. <i>multocida</i> str. Anand1_buffalo	199	421253447		
	<i>Pasteurella multocida</i> subsp. <i>multocida</i> str. Anand1_cattle	53	421259752		
	<i>Pasteurella multocida</i> subsp. <i>multocida</i> str. Anand1_cattle	63	421259756		

Cluster <sup>a</sup>	Strain <sup>b</sup>	Cas9 length (aa)	Cas9 GI	Cas1GI <sup>c</sup>	Subtype <sup>d</sup>
Anand1_cattle	<i>Pasteurella multocida</i> subsp. <i>multocida</i> str.				
Anand1_cattle		134	421259749		
	<i>Pediococcus acidilactici</i> 7_4	1229	270290729		
	<i>Pediococcus lolii</i> NGRI 0510Q	270	427443367		
	<i>Pediococcus lolii</i> NGRI 0510Q	1016	427441502		
	<i>Peptoniphilus</i> sp. oral taxon 386 str. F0131	1341	299144352		
	<i>Porphyromonas catoniae</i> F0037	211	429741290		
	<i>Porphyromonas catoniae</i> F0037	1009	429741242		
	<i>Prevotella denticola</i> F0289	1218	327314511		
	<i>Prevotella disiens</i> FB035-09AN	443	303235616		
	<i>Prevotella disiens</i> FB035-09AN	795	303237415		
	<i>Prevotella melaninogenica</i> D18	1354	288802595		
	<i>Prevotella multiformis</i> DSM 16608	129	325268382		
	<i>Prevotella multiformis</i> DSM 16608	535	325268383		
	<i>Prevotella oulorum</i> F0390	691	345881543		
	<i>Prevotella oulorum</i> F0390	774	345881542		
	<i>Prevotella saccharolytica</i> F0055	242	429739781		
	<i>Prevotella</i> sp. oral taxon 317 str. F0108	593	288929745		
	<i>Prevotella</i> sp. oral taxon 317 str. F0108	1174	288930149		
	<i>Prevotella</i> sp. oral taxon 472 str. F0295	241	260910968		
	<i>Prevotella</i> sp. oral taxon 472 str. F0295	992	260910970		
	<i>Pseudoramibacter alactolyticus</i> ATCC 23263	586	315926102		
	<i>Pseudoramibacter alactolyticus</i> ATCC 23263	770	315926103		
	<i>Rhizobium etli</i> GR56	103	218671711		
	<i>Riemerella anatipestifer</i> ATCC 11845 = DSM 15868	1145	383485594		
	<i>Sphingobacterium spiritivorum</i> ATCC 33300	116	227540450		
	<i>Sphingobacterium spiritivorum</i> ATCC 33300	1306	227540451		
	<i>Staphylococcus massiliensis</i> S46	475	425737243		
	<i>Staphylococcus massiliensis</i> S46	581	425737242		
	<i>Staphylococcus simulans</i> ACS-120-V-Sch1	1112	410873248		
	<i>Streptococcus agalactiae</i> 18RS21	773	76799343		
	<i>Streptococcus downei</i> F0415	994	312866154		
	<i>Streptococcus dysgalactiae</i> subsp. <i>equisimilis</i> SK1249	538	417753185		
	<i>Streptococcus dysgalactiae</i> subsp. <i>equisimilis</i> SK1250	1155	417926916		
	<i>Streptococcus mutans</i> SA38	1229	449253007		
	<i>Streptococcus mutans</i> SA38	1229	449880497		
	<i>Streptococcus oralis</i> SK255	550	417794716		
	<i>Streptococcus oralis</i> SK255	670	417793840		
	<i>Streptococcus pseudoparcinus</i> SPIN 20026	1326	313890160		
	<i>Streptococcus pyogenes</i> M49 591	1052	56808315		
	<i>Streptococcus sanguinis</i> VMC66	1167	323351495		
	<i>Streptococcus</i> sp. BS35b	93	401683465		
	<i>Streptococcus</i> sp. GMD4S	206	419816637		
	<i>Streptococcus</i> sp. GMD4S	317	419819606		
	<i>Streptococcus thermophilus</i> CNCM I-1630	302	418027683		
	<i>Streptococcus thermophilus</i> CNCM I-1630	595	418027684		
	<i>Streptococcus thermophilus</i> MTCC 5461	39	445389093		
	<i>Streptococcus vestibularis</i> F0396	97	312863468		

Cluster <sup>a</sup>	Strain <sup>b</sup>	Cas9 length (aa)	Cas9 GI	Cas1GI <sup>c</sup>	Subtype <sup>d</sup>
	<i>Streptococcus vestibularis</i> F0396	1038	312863582		
	<i>Sutterella parvirubra</i> YIT 11816	406	378822098		
	<i>Sutterella parvirubra</i> YIT 11816	951	378821855		
	<i>Sutterella wadsworthensis</i> 2_1_59BFAA	389	422348538		
	<i>Tannerella</i> sp. 6_1_58FAA_CT1	976	365118488		
	<i>Treponema denticola</i> ATCC 33520	631	449107910		
	<i>Treponema denticola</i> ATCC 33520	769	449107911		
	<i>Treponema denticola</i> F0402	357	422340642		
	<i>Treponema denticola</i> F0402	370	422340641		
	<i>Treponema denticola</i> F0402	631	422340640		
	<i>Treponema phagedenis</i> F0421	591	320536383		
	<i>Treponema phagedenis</i> F0421	738	320536384		
	<i>Treponema vincentii</i> ATCC 35580	281	257456747		
	<i>Treponema vincentii</i> ATCC 35580	992	257456748		
	uncultured bacterium	600	406975829		
	uncultured bacterium	1017	406999582		
	uncultured bacterium T3_7_42578	675	411001094		
	uncultured Termite group 1 bacterium phylotype Rs-D17	166	189485058		
	uncultured Termite group 1 bacterium phylotype Rs-D17	1032	189485225		
	<i>Verminephrobacter aporrectodeae</i> subsp. <i>tuberculatae</i> At4	983	347820874		

<sup>a</sup>Cas9 sequences are grouped according to the BLASTclust clustering program. Truncated sequences were not selected for the analysis and are listed at bottom of the table without any cluster number (see Materials and Methods).

<sup>b</sup>Bacterial strains harboring *cas9* gene orthologue are listed; GI, GenInfo Identifier. Bold, cluster representatives chosen for the alignment and tree reconstruction. Grey, discarded, incomplete Cas9 sequences (see Materials and Methods). Note, that the incomplete sequences were all confirmed to be truncated Cas9 orthologues due to the presence of conserved motifs and similarity to the other Cas9 orthologues.

<sup>c</sup>Cas1 GenInfo Identifier of the representative sequences chosen for the alignment and tree reconstruction are given. Grey, discarded, incomplete sequences. When possible, alternative Cas1 sequence from the same cluster as the discarded Cas1 sequence was selected (clusters 8, 9 and 21, in bold).

<sup>d</sup>Type II CRISPR subtype of the CRISPR loci of the Cas9 cluster as inferred from the representative Cas1 and Cas9 trees topology.

**[00497]** Analysis of the composition of *cas* genes, transcription direction of the CRISPR arrays with respect to that of the *cas* operon, and location and orientation of *tracr*RNAs resulted in the division of subtypes into groups with distinct locus characteristics, especially within the subtype II-A (Figure 1, clusters marked with different colors) (15). We selected Cas9 enzymes representative of the major type II groups. Cas9 orthologs of *S. pyogenes*, *S. thermophilus*\* (CRISPR3) and *S. mutans* were chosen for type II-A systems associated with shorter, ~220 amino acid Csn2 variants (Csn2a). Cas9 of *S. thermophilus*\*\* (CRISPR1) represents a distinct group of type II-A sequences associated with longer, ~350 amino acid

version of Csn2 orthologs (Csn2b). Cas9 of *F. novicida* was selected for type II-B. The closely related Cas9 orthologs of *P. multocida* and *N. meningitidis* and the distinct, short Cas9 of *C. jejuni* were chosen for type II-C (Figure 1B). Expression of associated tracrRNAs and crRNAs in *S. pyogenes*, *S. mutans*, *F. novicida*, *N. meningitidis* and *C. jejuni* was already validated by deep RNA sequencing (15,16). The RNAs in *S. thermophilus* and *P. multocida* were predicted bioinformatically based on the sequences from related species within the same type II group. Figure 1B shows the organization of the eight selected type II CRISPR-Cas loci and highlights our previous findings demonstrating that the type II loci architectures are highly variable among subtypes, yet conserved within each group (15). These variations are in good agreement with the clustering derived from the Cas9 and Cas1 phylogenetic trees (Figure 1A, Supplementary Figure S4).

**[00498]** Thus, to evaluate dual-RNA:Cas9 diversity, the bioinformatics analysis of type II CRISPR-Cas systems from available genomes identified Cas9 orthologs in a plethora of bacterial species that belong to 12 phyla and were isolated from diverse environments (Supplementary Tables S2 and S4). Most of the strains that harbor type II CRISPR-Cas systems (and accordingly Cas9) are pathogens and commensals of vertebrates. A majority of these strains were isolated from gastrointestinal tracts and feces of mammals, fish and birds, but also from wounds, abscesses and spinocerebral fluid of septicemia patients. Strains were also isolated from invertebrates and environmental samples, including fresh and sea water, plant material, soil and food, the latter comprising species used in fermentation processes. Cas9 is also present in species from extreme environments such as deep sea sediments, hot springs and Antarctic ice, further demonstrating the wide spread of type II CRISPR-Cas systems in bacteria. A comparison of the taxonomy and habitats of representative strains with the phylogenetic clustering of Cas9 sequences shows little correlation (Supplementary Figure S11). In particular, clusters of Cas9 genes were identified from taxonomically distant bacteria that were isolated from similar habitats. Examples include diverse Firmicutes, Mollicutes, Spirochaete and Fusobacteria, that were all isolated from gastrointestinal tracts of mammals, and members of different Proteobacteria, Firmicutes and Fusobacteria families mostly found in environmental samples (Supplementary Figure S11, clusters 1 and 3). A few exceptions involve grouping of Cas9 genes from closely related species isolated from diverse habitats such as Actinobacteria isolated from human and dog specimens but also from hot springs (Supplementary Figure S11, clusters 2, 4 and 5). This complex distribution of Cas9 across bacterial genomes indicates that evolution of dual-RNA:Cas9 systems in bacteria occurs both vertically and horizontally (55).

#### Example 2

**[00499]** Bacterial RNases III are interchangeable in dual-RNA maturation.

**[00500]** As described in *S. pyogenes* and *S. thermophilus*, RNase III plays an essential role in the biogenesis of dual-RNA:Cas9 systems by co-processing tracrRNA and pre-crRNA at the level of antirepeat:repeat duplexes (16,17). The interchangeability of *S. pyogenes* RNase III with RNases III from selected bacterial species was analyzed in the co-processing of *S. pyogenes* tracrRNA:pre-crRNA, including strains that lack type II CRISPR-Cas (*S. aureus* COL, *E. coli* TOP10). Northern blot analysis showed that all RNases III studied can co-process the RNA duplex (Figure 2, Supplementary Figure S5), indicating that there is no species-specificity for tracrRNA:pre-crRNA cleavage by RNase III. Multiple sequence alignment of RNase III orthologs demonstrates conservation of the catalytic aspartate residue and the dsRNA binding domain (Figure 2, Supplementary Figure S6) that are both required for RNA co-processing (Figure 2, Supplementary Figure S5). These data imply that the conservation of tracrRNA:pre-crRNA co-processing by bacterial RNase III provides a degree of flexibility allowing the functionality of dual-RNA:Cas9 systems in multiple species upon horizontal transfer.

**[00501]** Thus, to investigate the basis for the horizontal dissemination of CRISPR-Cas modules among bacteria, the specificity of RNase III utilized by type II CRISPR-Cas for dual-RNA maturation was analyzed. Complementation analysis shows that RNase III from a variety of species, including bacteria that lack type II CRISPR-Cas, can process *S. pyogenes* tracrRNA:pre-crRNA, suggesting that type II CRISPR-Cas systems can exploit any double-stranded RNA cleavage activity. This finding is consistent with the observation of *S. pyogenes* dual-RNA maturation in human cells which is apparently mediated by host RNases (2).

### Example 3

**[00502]** Cas9 HNH and split RuvC domains are the catalytic moieties for DNA interference.

**[00503]** Comparison of Cas9 sequences revealed high diversity in amino acid composition and length (984 amino acid for *C. jejuni* to 1648 amino acids for *F. novicida*), especially in the linker sequence between the highly conserved N-terminal RuvC and central RuvC-HNH-RuvC regions and in the C-terminal extension (Supplementary Figure S2). Several studies demonstrated the importance of the nuclease motifs for dsDNA cleavage activity by mutating one aspartate in the N-terminal motif of the RuvC domain and one or several residues in the predicted catalytic motif of the HNH domain of the Cas9 enzyme (14,22,23). To investigate the relevance of all catalytic motifs for tracrRNA:pre-crRNA processing and/or DNA interference, alanine substitutions of selected residues were created (Figure 3A). In addition to the already published catalytic amino acids, we created Cas9 point mutants of conserved amino acid residues in the central RuvC motifs (14) (Figure 3A, Supplementary Figure S2). Northern blot analysis of *S. pyogenes* cas9 deletion mutant complemented with each of the cas9 point mutants revealed the presence of mature tracrRNA and crRNA forms, demonstrating that none of the catalytic motifs is involved in dual-RNA maturation by RNase III. This is in agreement with previous data showing that

RNase III is the enzyme that specifically cleaves tracrRNA:pre-crRNA duplex (16). Cas9 seems to have a stabilizing function on dual-RNA. We show that the catalytic motifs are not involved in RNA duplex stabilization (Figure 3B, Supplementary Figure S7).

**[00504]** To investigate the involvement of the conserved motifs of Cas9 in DNA interference *in vivo*, a previously described plasmid-based read-out system was used that mimics infection with invading protospacer-containing DNA elements (16). Transformation assays were done in *S. pyogenes* WT or a *cas9* deletion mutant using plasmids containing the *speM* protospacer gene (complementary to the second spacer of *S. pyogenes* SF370 type II CRISPR array (16)) and WT or mutant *cas9* (Figure 3C). In this assay, Cas9 expressed following plasmid delivery in bacterial cells catalyzes its own vector cleavage, when active. Control experiments showed that the *speM* protospacer-containing plasmid was not tolerated in WT *S. pyogenes*, demonstrating activity of WT CRISPR-Cas. Similarly, a plasmid containing the *speM* protospacer and encoding WT Cas9 could not be maintained in the *cas9* deletion mutant, demonstrating that Cas9 is able to cleave the plasmid from which it is expressed. Except for Cas9 N854A, all plasmids encoding Cas9 mutants were tolerated in the *cas9* deletion strain, indicating abrogation of Cas9 interference activity for these variants.

**[00505]** The *in vivo* DNA targeting data were confirmed with *in vitro* DNA cleavage assays. Purified WT and mutant Cas9 proteins were incubated with tracrRNA:crRNA targeting *speM* and subjected to cleavage of plasmid DNA containing the *speM* protospacer. WT and N854A Cas9 show dsDNA cleavage activity, whereas the other Cas9 mutants cleave only one strand of the dsDNA substrate, yielding nicked open circular plasmid DNA (Figure 3D). This corroborates the results obtained *in vivo* showing the importance of the conserved nuclease motifs for DNA interference by Cas9. In addition to the previously published data demonstrating the importance of the N-terminal RuvC motif and the catalytic motif of HNH, we thus defined new catalytic residues in the central RuvC motifs.

**[00506]** Dual-RNA and Cas9 sequences have widely evolved in bacteria (15). However, despite the high sequence variability among Cas9 sequences, certain motifs are conserved. In addition to the previously identified central HNH and N-terminal RuvC catalytic motifs (20,21,44,56), we show that the two middle RuvC motifs are required for interference activity *in vivo* and *in vitro*. In agreement with previous findings, deactivation of either one of the catalytic motifs (RuvC or HNH) results in nicking activity of Cas9 originating from the other motif (2,8,24,25). None of the mutations introduced in these conserved motifs affected the role of Cas9 in tracrRNA:pre-crRNA maturation by RNase III *in vivo*.

#### Example 4

**[00507]** Only Cas9 from closely related CRISPR-Cas systems can substitute for *S. pyogenes* Cas9 in tracrRNA-directed pre-crRNA maturation by RNase III.

**[00508]** Beside the conservation of the HNH and split RuvC domains involved in DNA cleavage (14,15), the length of Cas9 orthologs and the amino acid sequences of Cas9 are highly variable among the different groups of type II CRISPR-Cas systems (Figure 4A, Supplementary Figure S2). Hence, whether this variability plays a role in the specificity of Cas9 with regard to tracrRNA:pre-crRNA duplex and mature crRNA stabilization was investigated. A *S. pyogenes* *cas9* deletion mutant was complemented with Cas9 from selected bacterial species representative of the various type II groups and analyzed tracrRNA:pre-crRNA processing by Northern blot. Cas9 proteins from *S. mutans* and *S. thermophilus*\* can substitute for the stabilizing role of *S. pyogenes* Cas9 in RNA processing by RNase III (Figure 4B, Supplementary Figure S8). By contrast, Cas9 from *S. thermophilus*\*\**, C. jejuni, N. meningitidis, P. multocida* and *F. novicida* could not complement the lack of RNA processing in the *cas9* mutant of *S. pyogenes*. In these strains, the 75-nt processed form of tracrRNA is observed as a very weak signal of background level of dual-RNA processed by RNase III in the absence of Cas9. Overall, only Cas9 from closely related systems of *S. pyogenes* in the type II-A cluster can substitute endogenous Cas9 role in dual-RNA stabilization and subsequent maturation by RNase III.

**[00509]** Thus, substitution of orthologs from the selected species for the endogenous *S. pyogenes* Cas9 shows that only Cas9 proteins from the *S. pyogenes* subcluster are capable of assisting tracrRNA:pre-crRNA processing by RNase III. This result indicates that the less-conserved inter-motif regions, which are the basis for the Cas9 subgrouping, could be responsible for Cas9 specificity for certain dual-RNAs.

#### Example 5

**[00510]** Cas9 orthologs require their specific PAM sequence for DNA cleavage activity.

**[00511]** In *S. pyogenes* and *S. thermophilus*\* types II-A, PAMs were identified as NGG and NGGNG, respectively. In these two species, mutating the PAM abrogates DNA interference by dual-RNA:Cas9 (14,22,23). To identify the functional PAMs for Cas9 from bacterial species other than *S. pyogenes* and *S. thermophilus*, potential protospacers matching spacer sequences in the selected CRISPR arrays were searched using BLAST. For *S. mutans* UA159, *C. jejuni* NCTC 11168, *P. multocida* Pm70 and *F. novicida* U112, potential protospacers were identified. Therefore, strains that harbor a closely related variant of Cas9 (Supplementary Table S2) were searched and their spacer sequences analyzed following the same approach (Supplementary Table S3). The identified 10 nt sequences located directly downstream of the protospacer sequence were aligned and the most common nucleotides that could represent PAM sequences were delineated. Based on the data visualized as a logo plot (Figure 5A), plasmid DNA substrates were designed containing the *speM* protospacer followed by different adjacent sequences either comprising the predicted PAM or not (Figure 5B). The Cas9 orthologous proteins were purified (Supplementary Figure S1) and dual-RNA orthologs were designed based on deep RNA sequencing data



(15), with the spacer sequence of crRNA targeting *speM*. To determine the protospacer-adjacent sequences critical for efficient DNA targeting, the purified Cas9 orthologs and their cognate dual-RNAs were used in DNA cleavage assays with different plasmid substrates (Figure 5C, Supplementary Figure S9). The previously published PAMs for Cas9 from *S. pyogenes* (NGG), *S. mutans* (NGG), *S. thermophilus*\* (NGGNG) and *N. meningitidis* (NNNNGATT) (27,28,53,54) were confirmed by multiple sequence alignments and *in vitro* cleavage assay, validating our approach. However, dual-RNA guided Cas9 from *S. thermophilus*\* could efficiently cleave target DNA in the presence of only NGG instead of NGGNG (Supplementary Figure S9). This is in contrast to data obtained *in vivo*, where mutation of the third G abrogates interference by Cas9 of *S. thermophilus*\* (23). For *S. thermophilus*\*\*<sup>†</sup>, the PAM was published as NNAGAAW (27), which differs by one base from the sequence that we derived (NNAAA<sup>†</sup>AW). *In vitro* cleavage assays with these two sequences demonstrate that the DNA substrate with the “NNAAA<sup>†</sup>AW” PAM is cleaved more efficiently by Cas9 of *S. thermophilus*\*\* compared to the “NNAGAAW” PAM (Supplementary Figure S9).

**[00512]** Using the same approach, the PAM activity of the most common protospacer-downstream sequences for *C. jejuni*, *F. novicida* and *P. multocida* were validated by *in vitro* cleavage assays, resulting in the most probable PAM sequences being NNNNACA (*C. jejuni*), GNNNCNNA (*P. multocida*) and NG (*F. novicida*) (Figure 5C, Supplementary Figure S9). Analysis of the protospacer-adjacent sequence from *C. jejuni* shows the same frequency of C and A (“NNNNCCA” or “NNNNACA”) at position 5 downstream of the protospacer (Supplementary Table S3). Hence, both substrates were tested for cleavage activity by *C. jejuni* dual-RNA:Cas9. Only the DNA target containing A at this position was cleaved efficiently (Supplementary Figure S9). This result could be explained by the origin of the protospacer, with the “NNNNCCA” PAM being mostly found in genomic DNA or prophages of *Campylobacter* strains. In this case, the mutated PAM sequence on the chromosomally located protospacer prevents self-targeting. The *P. multocida* PAM requires further verification given that the multiple sequence alignment was derived from only two protospacer sequences. Thus, a series of specific PAMs that enable dsDNA cleavage by dual-RNA:Cas9 complexes from different bacterial species *in vitro* were identified. For gene editing purposes, it is contemplated that a range of potential motifs be analyzed to select those PAMs that would allow efficient targeting with limited off-site effect.

**Supplementary Table S3. Overview of type II CRISPR-Cas spacer sequences from selected bacterial strains with BLAST candidate protospacers and their downstream sequence.**

Strain <sup>a</sup>	Number of spacers	CRISPR Spacer <sup>b</sup>	Spacer sequence	Blast candidate <sup>c</sup>	% Identity <sup>d</sup>	10 bp downstream protospacer <sup>e</sup>
<i>Streptococcus pyogenes</i> SF370 (Accession: NC_002737)	1		TGCGCTGGTTGATTTCCTTTCGCGCTTTT	<i>S. pyogenes</i> MGAS1882 (MGAS1882_1116), MGAS8232 (spyM18_0769), MGAS10394 (M6_Spy0995, M6_Spy1349), SSI-1 (SPs0926), ΦP9 <b>endopeptidase gene</b>	100	
				<i>S. pyogenes</i> MGAS2096 (MGAS2096_Spy1450), A20 (A20_1472c), M1 476 (M1GAS476_1503), MGAS9429 (MGAS9429_Spy1426), MGAS5005 (M5005_Spy1424) <b>endopeptidase gene</b>	97	
				<i>S. pyogenes</i> M1 GAS (SPy_0700), MGAS2096 (MGAS2096_Spy0592) <b>endopeptidase gene</b>	97	
				<i>S. pyogenes</i> MGAS6180 (M28_Spy1234); NIH1 (NIH1.1_43), SSI-1 (SPs0647), MGAS315 (SpyM3_0930, SpyM3_1215) <b>phage related gene</b>	100	
	2		TTATATGAACATAACTCAATTTG TAAAAAA	<b>gene for pyrogenic exotoxin M (speM) of several Streptococci strains</b>	100	
	6	3	AGGAATATCCGCAATAATTAATT GCGCTCT	<i>S. pyogenes</i> MGAS8232 (spyM18_0742), MGAS10750 (MGAS10750_Spy0588), MGAS10270 (MGAS10270_Spy0563) <b>adenine specific methylase gene</b>	100	
				<i>S. pyogenes</i> Manfredo (SpyM50653) <b>adenine specific methylase gene</b>	97	
	4		AGTGCCGAGGAAAAATTAGGTGC GCTTGCC	<i>S. pyogenes</i> Alab49 (SPYALAB49_001176), MGAS10750 (MGAS10750_Spy1285), MGAS9429 (MGAS9429_Spy0843), MGAS10394 (M6_Spy1203), SSI-1 (SPs0763), MGAS315 (SpyM3_1101), ΦH4489A (hylP) <b>hyaluronoglucosaminidase gene</b>	100	
				<i>S. pyogenes</i> MGAS8232 (spyM18_1254), NZ131 (Spy49_0785) <b>hyaluronoglucosaminidase gene</b>	97	
	5		TAAATTTGTTTAGCAGGTAAACC GTGCTTT	<i>S. pyogenes</i> MGAS10750 (MGAS10750_Spy0839), MGAS10270 (MGAS10270_Spy0546, MGAS10270_Spy0804), SSI-1 (SPs0517, SPs0888), MGAS1882 (MGAS1882_1156), MGAS8232, NZ131(Spy49_1511c), MGAS315 (SpyM3_0965, SpyM3_1347) <b>phage protein gene or intergenic region</b>	100	

Strain <sup>a</sup>	Number of spacers	CRIS PR Spacer <sup>b</sup>	Spacer sequence	Blast candidate <sup>c</sup>	% Identity <sup>d</sup>	10 bp downstream protospacer <sup>e</sup>
<i>Streptococcus mutans</i> UA159 (Accession: NC_004350)	5	3	CTAACTATGATGACACAACAGCT TTTAGCG	ΦM102 (orf13) putative tail protein gene	100	TTGAAAT TTC
<i>Streptococcus mutans</i> LJ23 (Accession: NC_017768)	8	2	TGAAGTGCAAGCTTACGTGACTG ACTCGCG	ΦM102 (orf15) putative minor structural protein	90	AGGATATG CAG
<i>Streptococcus mutans</i> GS-5 (Accession: NC_018089)	21	3	TAATAGCAATCGTGACGGACGTA TTGATTT	ΦM102 (orf15) putative minor structural protein	97	AGGTTGAA ATT
		5	GTTGAGTGCAACAGCTAGCTAAT AGCTTTT	ΦM102	100	AGGCTGG CAC
		16	AGGCATTTTCTGATTGAGATTTT CGATATT	ΦM102 (orf3) putative large terminase gene	93	TGGAAAG ATG
		18	TATAGCTAATATGTTTATACTGA CAGCGCA	ΦM102 (orf7) putative DNA packaging protein gene	100	AGAAAGA TTG
<i>Streptococcus mutans</i> NN2025 (Accession: NC_013928)	69	2	GATTGTGCCCGCTAGTAAACCGC CTCGCGC	ΦM102 (orf20) putative endolysin gene	93	TGGAGAT TTG
		6	GATTGTATCAGTAATCGAACTTC TGCTTAT	ΦM102 (orf38, orf39) hypothetical protein gene	93	GGGATTT GAC
		8	TGGTCCAAAGTGCAAGCCAAAG AAAAACA	ΦM102 (orf11) putative major tail protein gene	97	AAACGGT CTT
		9	ATTGTCAATCGCCGTTCTGCGCT TGCGACG	ΦM102 (orf17) hypothetical protein gene	90	CGTTTTT GAA
		17	GCTTGAATATAATTGTGTATCCG CCAATGA	ΦM102 (orf21) putative replisome organizer gene	93	CGAATTA CGA
		23	AAAAGAAACGCCTTTTGATTTG ACCAATC	ΦM102 (orf14) putative receptor-binding protein gene	90	AAAGCA AGA
		29	AGTTATTAATATCTATGACAGTC TCAAAGA	ΦM102 (orf14) putative receptor-binding protein gene	93	GGCTAC AGA
		37	TTCTGGCTGTCTTTCAGAGTGAT AAGCGCA	ΦM102 (orf2) putative small terminase gene	100	GGATTTT TCA
		40	TGCAAGTTATCTTGCTATGTGGA CGAATTG	ΦM102 (orf9) hypothetical protein gene	93	GGAAACA ATC
		43	GCAATTTAGTTTTATCCGTGGG AGCAGCA	ΦM102 (orf3) putative large terminase gene	93	AGCTCC ATT

Strain <sup>a</sup>	Number of spacers	CRISPR Spacer <sup>b</sup>	Spacer sequence	Blast candidate <sup>c</sup>	% Identity <sup>d</sup>	10 bp downstream protospacer <sup>e</sup>
	48		AGAGTATAGCCAGTGTTTTCAAG GCCTTTA	ΦM102 (orf12) <b>putative tape measure protein gene</b>	93	GTGGTGA CAA
	49		CGCAACAATGACTATTAATATCA ACGGTGG	ΦM102 (orf15) <b>putative minor structural protein gene</b>	93	CGGAGC AAT
	56		AATCGCTTCTTTGCTAACCACAA TTTGTGC	ΦM102 (orf26) <b>putative RecT family single-strand annealing protein gene</b>	93	AGCGCA GAG
	60		AAATGCTCTTGAAGAACCTGATA GATGACA	ΦM102 (orf3) <b>putative large terminase gene</b>	93	GAGACGA AAA
	66		TGCAAAAGATGGCCTCGAGCAAT TATCGCA	ΦM102 (orf33) <b>hypothetical protein gene</b>	100	TGGATTA AGC
	<i>Streptococcus thermophilus</i> LMD-9 CASS4 locus (Accession: NC_008532)	2		TCAATGAGTGGTATCCAAGACGA AAACTTA	<i>Streptococcus thermophilus</i> plasmid pSt106 <b>putative resolvase gene</b>	100
3			CCTTGTCGTGGCTCTCCATACGC CCATATA	<i>Streptococcus thermophilus</i> plasmid pND103	100	AGGGGG GGT
4			TGTTTGGGAAACCGCAGTAGCCA TGATFAA	Φ7201 (orf33)	100	AGCTTC GCT
5			ACAGAGTACAATATTGTCTCAT TGGAGACAC	Φ TP-J34 (orf11) <b>hypothetical protein gene</b>	94	TGGGTA GGA
				ΦSfi19 (orf1626) <b>minor tail protein gene</b>	100	TGCTGCT AAT
6			CTCATATTCGTTAGTTGCTTTTG TCATAAA	ΦYMC 2011 (Ssal_phage00063) <b>putative minor tail protein gene</b>	90	GGTCT AAC
				Φ7201 (orf33)	90	TGCTCT AGA
<i>Streptococcus thermophilus</i> LMD-9 CASS4a locus (Accession: NC_008532)		2		CTTCACCTCAAATCTTAGAGCTG GACTAAA	Φ7201 (orf39)	100
	3		ATGTCTGAAAAATAACCGACCAT CATTACT	Φ TP-J34 (orf49), ΦSfi11 (orf669) <b>putative minor structural protein gene</b>	93	CCGAT GTC
	4		GAAGCTCATCATGTTAAGGCTAA AACCTAT	ΦALQ13.2 (orf35) <b>helicase gene</b>	90	CTAAAA TTA
				ΦSfi11 (orf443), ΦSfi18 (orf443), ΦSfi21 (orf443), ΦSfi19 (orf443), ΦO1205 (orf10) <b>putative helicase gene</b>	90	CTCAAA TTA
	5		TAGTCTAAATAGATTTCTTGAC CATTGTA	Φ1033, Φ 1042 <b>nonfunctional host specificity protein gene</b>	97	ATGAT TCA

Strain <sup>a</sup>	Number of spacers	CRISPR Spacer <sup>b</sup>	Spacer sequence	Blast candidate <sup>c</sup>	% Identity <sup>d</sup>	10 bp downstream protospacer <sup>e</sup>
				ΦDT1.1 (orf18), ΦDT1.2 (orf18), ΦDT1.3 (orf18), ΦDT1.4 (orf18), ΦDT1.5 (orf18), ΦMD4 (orf18) <b>host specificity protein gene</b>	93	ATGAAAT TCA
	6		ATTCGTGAAAAATATCGTGAAATAGGCAA	pSt08 plasmid	97	CCGAAATA
	7		TCTAGGCTCATCTAAAGATAAATCAGTAGC	ΦALQ13.2 (orf25), Φ858 (orf30), ΦST3 (orf253) <b>endonuclease gene</b>	90	TGAAAA TTA
				ΦJ1 (orf253), ΦS3b (orf253) <b>endonuclease gene</b>	90	TGAAAG TTA
				ΦSfi11	100	TCAGAT TAT
	13		AACTACCAAGCAAATCAGCAATCAATAAGT	ΦYMC-2011 (Ssal_phage00051) <b>predicted clp-protease gene</b>	93	TTAGAA CAT
				ΦSfi21 (orf221) <b>clp-protease gene</b>	90	TCAGAA TAT
				Φ858 (orf22)	93	AAAAAA ACT
	16		AACAGTTACTATTAATCACGATTCCAACGG	Φ2972 (orf21) <b>structural protein gene</b>	93	AAAAAA ACT
				ΦAbc2 (orf17) <b>tail protein gene</b>	93	TAAAAAG ACT
<hr/>						
<i>Campylobacter jejuni</i>						
subsp. jejuni NCTC 11168 (Accession: NC_002163)	5	1-5		no significant BLAST hits		
<hr/>						
<i>Campylobacter jejuni</i>						
subsp. jejuni CF93-6 (Accession: AANJ00000000)	5	3	TCATCATCACTTAAACCTTAAATTTACC	<i>C. jejuni</i> RM1221 (CJE1445) <b>hypothetical protein gene</b>	93	ATAACGC AAG

Strain <sup>a</sup>	Number of spacers	CRISPR Spacer <sup>b</sup>	Spacer sequence	Blast candidate <sup>c</sup>	% Identity <sup>d</sup>	10 bp downstream protospacer <sup>e</sup>
<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> HB93-13c_jejunisubsp_jejunihb_13_42 (Accession: AANQ000000)	9	1	GCATTGCTTTACTACATAGCCAGTCGTGTA	<i>C. jejuni</i> subsp. <i>doylei</i> 269.97 (JJD26997_1148) <b>conserved hypothetical protein gene</b>	100	TCACA CGC
			2	TTATTTTGTCGCTAATTGCACCTAAAGAC	<i>C. jejuni</i> subsp. <i>doylei</i> 269.97 (JJD26997_0867) <b>putative primase gene</b>	97
<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> NW genomic scaffold Mich_State_University:Contig3 (Accession: JH376989 REGION: 13521..15062)	5	5	GGGACACGAGGAATCCTGTCTGATCCGGG	<i>C. jejuni</i> subsp. <i>jejuni</i> PT14 (A911_r08426, A911_r08428, A911_r08430), NCTC 11168-BN148 (BN148_r02, BN148_r05, BN148_r08), S3 (CJS3_1811, CJS3_1817, CJS3_1830), ICDCCJ07001 (ICDCCJ07001_29, ICDCCJ07001_396, ICDCCJ07001_718), M1 (CJM1_0031, CJM1_0413, CJM1_0727), IA3902 (CJSA_Cj23SA, CJSA_Cj23SB, CJSA_Cj23SAC), BABS091400, 81116 (C8J_Cj23SA, C8J_Cj23SB, C8J_Cj23SC), 81-176 (CJJ81176_1714, CJJ81176_1727, CJJ81176_1707), NCTC 11168; <i>C. jejuni</i> DSM 4688, UNSW091300, strain 100, RP0001, 102-27 (rrlC, rrlB, rrlA), 69-30 (rrlC, rrlB, rrlA), 140-16 (rrlC, rrlB, rrlA), 110-21 (rrlC, rrlB, rrlA), RM1221 (CJE_Cj23SA, CJE_Cj23SB, CJE_Cj23SC), TGH9011_ATCC43431 (rrl); <i>C. coli</i> 59-2 (rrlC, rrlB, rrlA); <i>C. jejuni</i> subsp. <i>doylei</i> 269.97 (JJD26997_0040, JJD26997_1264, JJD26997_1520) <b>23S rRNA gene</b>	100	TCGAC CGA
<i>Campylobacter jejuni</i> subsp. <i>doylei</i> 269.97 (Accession: NC_009707)	5	2	CTAAGCAATCTTATTTTACCATCTTTTTTA	<i>C. jejuni</i> strain TGH 9011 (Tgh093)	97	TAAAA CTT
				<i>C. jejuni</i> RM1221 (CJE1099) <b>hypothetical protein gene</b>	93	TAAAA CTT
<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 1336 (Accession: NZ_CM000854 NZ_ADGL0100000)	2	1	TTACTGATATTTAAAATTAACCTCATAATTT	<i>C. jejuni</i> 00-3477 (cje0227), <i>C. jejuni</i> subsp. <i>jejuni</i> S3 (CJS3_0723), ΦCGC-2007 <b>prophage related genes</b>	100	GCTGC TTA
				<i>C. jejuni</i> NCTC 13255 (putative CJIE1-2-like prophage), 99-7046 (putative CJIE1-3-like prophage), 00-2425 (putative CJIE1 prophage), RM1221 (CJE0227) <i>C. jejuni</i> subsp. <i>jejuni</i> ICDCCJ07001 (ICDCCJ07001_691) <b>major tail sheath protein</b>	93	GCTGC TTA

Strain <sup>a</sup>	Number of spacers	CRISPR Spacer <sup>b</sup>	Spacer sequence	Blast candidate <sup>c</sup>	% Identity <sup>d</sup>	10 bp downstream protospacer <sup>e</sup>
		2	ATAAAGCTAATGCAAAAGTTGAA AACAAA	<i>C. jejuni</i> NCTC 13255 (putative CJIE1-2-like prophage), 99-7046 (putative CJIE1-3-like prophage), 00-3477 (putative CJIE1-4 Mu-like prophage), 00-2425 (putative CJIE1 prophage), RM1221 (CJE0238), <i>C. jejuni</i> subsp. <i>jejuni</i> S3 (CJS3_0704), ICDCJ07001, <i>C. huyoilei</i> <b>hypothetical protein gene</b>	100	AGAGCT TAA
<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 414 (Accession: NZ_CM000855 NZ_ADGM01000000)	33	2	TTTATCTGCATCCATAATGGCAA TGAGTGA	<i>C. jejuni</i> subsp. <i>jejuni</i> PT14 (A911_03310), NCTC 11168-BN148 (BN148_0680c), S3 (CJS3_0675), ICDCJ07001 (ICDCJ07001_619), M1 (CJM1_0650), IA3902 (CJSA_0644), 81116 (C8J_0632), 81-176 (CJJ81176_0703), NCTC 11168 (Cj0680c), P694a (Cj0680c), P569a (Cj0680c), P179a (Cj0680c), H73020 (Cj0680c), P569a (Cj0680c), P179a (Cj0680c), H73020 (Cj0680c), H704a (Cj0680c), <i>C. jejuni</i> RM1221 (CJE0778), <i>C. jejuni</i> subsp. <i>doylei</i> 269.97 (JJD26997_1327) <b>excinuclease ABC subunit B gene</b>	97	AACTT GGC
		2	CTTCTGCCTTTTACAAGCTCGC TTTCTTT	<i>N. gonorrhoeae</i> (NGU65994, PivNG), FA 1090 (NGO1137, NGO1164, NGO1262) <b>invertase related genes, phage associated protein genes</b>	97	CGCCGAC CGG
				<i>N. meningitidis</i> NZ-05/33 (NMBNZ0533_1722), M04-240196 (NMBNZ0533_1722), M01-240149 (NMBH4476_1701), H44/76 (NMBH4476_1701) <b>hypothetical proteins upstream of transposase gene</b>	100	GTCTGAT TTT
				<i>N. lactamica</i> isolate 3207487 (plasmid pNL3.2), <i>N. lactamica</i> (plasmid pNL9)	97	GTCTGAT TTT
		3	TTTGGTAAAGGTTTCTGTTGCGA CCCCAAT	<i>N. gonorrhoeae</i> TCDC-NG08107, NCCP11945 <b>intergenic region (putative phage proteins)</b>	93	GGCTTT TTT
	16			<i>N. gonorrhoeae</i> NCCP11945 (NGK_1948, NGK_1990, NGK_2023) <b>hypothetical protein genes</b>	93	GTCTGAT TTT
				<i>N. gonorrhoeae</i> <b>intergenic region PivNG</b>	93	GTCTGAT TTT
<i>Neisseria meningitidis</i> serogroup A strain Z2491 (Accession: NC_003116)				<i>N. gonorrhoeae</i> FA 1090 <b>numerous intergenic regions in prophages</b>	93	GTCTGAT TTT
		7	AAATTCGTTTCAGATAGCAAACG CAGTAGT	<i>N. gonorrhoeae</i> TCDC-NG08107, <i>N. gonorrhoeae</i> NCCP11945 <b>intergenic region (putative phage proteins)</b>	97	GGACGAT TTT
		12	GGGTAGCCAGTGCTAAAACCGCA CCCCGCTT	<i>N. lactamica</i> plasmid pNL9	93	TGCGGC ATA
				<i>N. meningitidis</i> plasmid pJS-B	100	TACGAAC ATT
		13	CCAAATAGAAATACATACGCCGA GTAATTA	<i>N. lactamica</i> plasmid pNL9	93	AGCTC TTT

Strain <sup>a</sup>	Number of spacers	CRIS PR Spacer <sup>b</sup>	Spacer sequence	Blast candidate <sup>c</sup>	% Identity <sup>d</sup>	10 bp downstream protospacer <sup>e</sup>
				<i>N. meningitidis</i> plasmid pJS-B	97	AGCCCTG
	14		TTTCTTTTGTGTAATTGTTCTGCC TTTTTTA	<i>N. lactamica</i> plasmid pNL9	100	ATTCGAT
				<i>N. meningitidis</i> plasmid pJS-B	100	ATTCGAT
	15		TACCCACGGCGGAAACCATTGCC ACAAAAC	<i>N. meningitidis</i> strain alpha522 draft genome (NMALPHA522_0671), H44/76 (NMBH4476_0684), 053442 ( NMCC_0153), <i>N. meningitidis</i> serogroup C FAM18 (NMC1864) <b>hypothetical protein gene</b>	100	CCATGAC
				<i>N.meningitidis</i> M04-240196 (NMBM04240196_0048, NMBM04240196_0749) <b>putative membrane protein gene</b>	100	CCATGAC
<i>Pasteurella multocida</i> str. Pm70 (Accession: NC_002663)	5	1-5		no significant BLAST hits		
<i>Pasteurella multocida</i> subsp. <i>gallicida</i> X73 (Accession: CM001580 AMBP01000000)	20	15	AAAGAATACACCCTTATTCAAA AAGTTTG	<i>P. multocida</i> 1.8 kb plasmid	100	CCGACAG ATG
			GTCTGAACAGTATTAACACTTC TGTTTCT	<i>P. multocida</i> subsp. <i>multocida</i> str. HN06(PMCN06_2098) <b>hypothetical protein gene</b>	97	CGATCGT ACT
<i>Francisella tularensis</i> subsp. <i>novicida</i> U112 (Accession: NC_008601)	13	1-13		no significant BLAST hits		
<i>Francisella novicida</i> FTG	22	15	ATCTCAAAAGCAGCTCTTTTCGCG TGTAATATCGTT	<i>F. cf. novicida</i> 3523 (FN3523_1002) <b>phage protein gene</b>	91	TCGATTA CAT



Strain <sup>a</sup>	Number of spacers	CRISPR Spacer <sup>b</sup>	Spacer sequence	Blast candidate <sup>c</sup>	% Identity <sup>d</sup>	10 bp downstream protospacer <sup>e</sup>
FTG scaffold 1 genomic scaffold (Accession: NZ_DS99536_3 NZ_ABXZ010_00000)	19		CTATCTAAGAGAACTTACAAGAC AAGAGAAAATACT	<i>F. cf. novicida</i> 3523 (FN3523_0993) hypothetical protein gene	94	
<i>Francisella tularensis</i> subsp. <i>novicida</i> GA99-3548 supercont1.3 (Accession: DS264589 ABAH010000_00)	2		AGCCCTATCAGAAATATATGCAA GTTTGAATATAG	<i>F. cf. novicida</i> 3523 (FN3523_1009) phage-related baseplate assembly protein gene	89	
	3		AGATAACTCTTATATTGATTTGT ATATTGAAGATA	<i>F. cf. novicida</i> 3523 (FN3523_1006) hypothetical protein gene	94	
	4		CGCAAAAAAGGCCGAATTTGAGCA GAAAAATTTGGGC	<i>F. cf. novicida</i> 3523 (FN3523_0999) hypothetical protein gene	91	

<sup>a</sup>Selected strains used in this study. No potential protospacers were found for *Streptococcus mutans* UA159, *Campylobacter jejuni* subsp. *jejuni* NCTC 11168, *Pasteurella multocida* str. Pm70 and *Francisella tularensis* subsp. *novicida* U112. Therefore, closely related strains were analyzed for the presence of type II CRISPR-Cas arrays. Spacer sequences from selected arrays were then used to search for protospacer candidates.

<sup>b</sup>Numbering of spacers starts from the leader proximal end based on RNAseq data (15). Spacers with no significant protospacer BLAST hit are not listed in the table.

<sup>c</sup>A BLAST candidate was considered a potential protospacer when the identity to the spacer was ≥ 90% and when the protospacer originated either from phage, plasmid or genomic DNA related to the analyzed species. For each identified protospacer, the strain name, the protospacer-containing gene locus and the potential function of the gene are given.

<sup>d</sup>Percentage identity between spacer and protospacer sequence. e10 nt sequence located directly 3' of the protospacer sequence. The identified sequences for each bacterial species were aligned using GeneDoc (<http://www.nrbsc.org/gfx/genedoc/>). The degree of conservation is indicated with a color code (black: 100%, dark grey: ≥ 80%, light grey: ≥ 60%). These sequences were used to create the logo plot represented in Figure 5.

**Supplementary Table S4 – Cas9 is present in bacteria from 12 different phyla and diverse habitats**

Strain <sup>a</sup>	Class	Isolation/habitat <sup>b</sup>
<b>Actinobacteria</b>		
Actinobacteridae		
<i>Acidothermus cellulolyticus</i> 11B	<i>Acidothermaceae</i>	extremophile (hot water spring)
<i>Actinomyces coleocanis</i>	<i>Actinomycetaceae</i>	dog genital tract

Strain <sup>a</sup>	Class	Isolation/habitat <sup>b</sup>
<i>Actinomyces georgiae</i> F0490	Actinomycetaceae	oral cavity
<i>Actinomyces naeslundii</i> str. Howell 279	Actinomycetaceae	oral cavity
<i>Actinomyces</i> sp. ICM47	Actinomycetaceae	ND
<i>Actinomyces</i> sp. oral taxon 175 str. F0384	Actinomycetaceae	oral cavity
<i>Actinomyces</i> sp. oral taxon 180 str. F0310	Actinomycetaceae	oral cavity
<i>Actinomyces</i> sp. oral taxon 181 str. F0379	Actinomycetaceae	oral cavity
<i>Actinomyces</i> sp. oral taxon 848 str. F0332	Actinomycetaceae	oral cavity
<i>Actinomyces turicensis</i> ACS-279-V-Col4	Actinomycetaceae	genital tract
<i>Bifidobacterium bifidum</i> S17	Bifidobacteriaceae	gastrointestinal tract/feces
<i>Bifidobacterium dentium</i> Bd1	Bifidobacteriaceae	oral cavity
<i>Bifidobacterium longum</i> DJO10A	Bifidobacteriaceae	gastrointestinal tract/feces
<i>Bifidobacterium</i> sp. 12_1_47BFAA	Bifidobacteriaceae	gastrointestinal tract/feces
<i>Corynebacterium accolens</i> ATCC 49726	Corynebacterineae	wound
<i>Corynebacterium diphtheriae</i> NCTC 13129	Corynebacterineae	oral cavity
<i>Corynebacterium matruchotii</i> ATCC 14266	Corynebacterineae	oral cavity
<i>Gardnerella vaginalis</i> 5-1	Bifidobacteriaceae	genital tract
<i>Mobiluncus curtisii</i> ATCC 35242	Actinomycetaceae	genital tract
<i>Mobiluncus mulieris</i> 28-1	Actinomycetaceae	genital tract
<i>Scardovia inopinata</i> F0304	Bifidobacteriaceae	oral cavity
<i>Scardovia wiggsiae</i> F0424	Bifidobacteriaceae	oral cavity
<b>Coriobacteridae</b>		
<i>Coriobacterium glomerans</i> PW2	Coriobacteriaceae	invertebrate (red soldier bug)
<i>Eggerthella</i> sp. YY7918	Coriobacteriaceae	gastrointestinal tract/feces
<i>Gordonibacter pamelaee</i> 7-10-1-b	Coriobacteriaceae	gastrointestinal tract/feces
<i>Olsenella uli</i> DSM 7084	Coriobacteriaceae	oral cavity
<b>Bacteroidetes</b>		
<b>Bacteroidia</b>		
<i>Anaerophaga</i> sp. HS1	Marinilabiliaceae	extremophile (hot water spring)
<i>Anaerophaga thermohalophila</i> DSM 12881	Marinilabiliaceae	environmental sample (oil residue)
<i>Bacteroides cellulosilyticus</i> DSM 14838	Bacteroidaceae	gastrointestinal tract/feces
<i>Bacteroides coprophilus</i> DSM 18228	Bacteroidaceae	gastrointestinal tract/feces
<i>Bacteroides coprosuis</i> DSM 18011	Bacteroidaceae	pig feces
<i>Bacteroides dorei</i> DSM 17855	Bacteroidaceae	gastrointestinal tract/feces
<i>Bacteroides eggerthii</i> 1_2_48FAA	Bacteroidaceae	gastrointestinal tract/feces
<i>Bacteroides faecis</i> MAJ27	Bacteroidaceae	gastrointestinal tract/feces
<i>Bacteroides fluxus</i> YIT 12057	Bacteroidaceae	gastrointestinal tract/feces
<i>Bacteroides fragilis</i> NCTC9343	Bacteroidaceae	gastrointestinal tract/feces
<i>Bacteroides nordii</i> CL02T12C05	Bacteroidaceae	gastrointestinal tract/feces
<i>Bacteroides oleiciplenus</i> YIT 12058	Bacteroidaceae	gastrointestinal tract/feces
<i>Bacteroides</i> sp. 2_1_16	Bacteroidaceae	gastrointestinal tract/feces
<i>Bacteroides</i> sp. 203	Bacteroidaceae	gastrointestinal tract/feces
<i>Bacteroides</i> sp. 3_1_19	Bacteroidaceae	gastrointestinal tract/feces
<i>Bacteroides</i> sp. 3_1_33FAA	Bacteroidaceae	gastrointestinal tract/feces
<i>Bacteroides</i> sp. 9_1_42FAA	Bacteroidaceae	gastrointestinal tract/feces
<i>Bacteroides</i> sp. D2	Bacteroidaceae	gastrointestinal tract/feces
<i>Bacteroides uniformis</i> CL03T00C23	Bacteroidaceae	gastrointestinal tract/feces
<i>Bacteroides vulgatus</i> CL09T03C04	Bacteroidaceae	gastrointestinal tract/feces
<i>Bacteroidetes</i> oral taxon 274 str. F0058	Bacteroidaceae	oral cavity
<i>Barnesiella intestinhominis</i> YIT 11860	Bacteroidaceae	gastrointestinal tract/feces
Bacteroidia (continued)		

Strain <sup>a</sup>	Class	Isolation/habitat <sup>b</sup>
<i>Marinilabilia</i> sp. AK2	<i>Marinilabiliaceae</i>	extremophile (solar saltern)
<i>Odoribacter laneus</i> YIT 12061	<i>Porphyromonadaceae</i>	gastrointestinal tract/feces
<i>Parabacteroides johnsonii</i> DSM 18315	<i>Bacteroidaceae</i>	gastrointestinal tract/feces
<i>Parabacteroides</i> sp. D13	<i>Bacteroidaceae</i>	gastrointestinal tract/feces
<i>Porphyromonas catoniae</i> F0037	<i>Porphyromonadaceae</i>	oral cavity
<i>Porphyromonas</i> sp. oral taxon 279 str. F0450	<i>Porphyromonadaceae</i>	oral cavity
<i>Prevotella bivia</i> JCVIHMP010	<i>Prevotellaceae</i>	genital tract
<i>Prevotella buccae</i> ATCC 33574	<i>Prevotellaceae</i>	oral cavity
<i>Prevotella buccalis</i> ATCC 35310	<i>Prevotellaceae</i>	oral cavity
<i>Prevotella denticola</i> F0289	<i>Prevotellaceae</i>	oral cavity
<i>Prevotella disiens</i> FB035-09AN	<i>Prevotellaceae</i>	oral cavity
<i>Prevotella histicola</i> F0411	<i>Prevotellaceae</i>	oral cavity
<i>Prevotella intermedia</i> 17	<i>Prevotellaceae</i>	oral cavity
<i>Prevotella melaninogenica</i> D18	<i>Prevotellaceae</i>	oral cavity/rumen
<i>Prevotella micans</i> F0438	<i>Prevotellaceae</i>	oral cavity
<i>Prevotella multiformis</i> DSM 16608	<i>Prevotellaceae</i>	oral cavity
<i>Prevotella nigrescens</i> ATCC 33563	<i>Prevotellaceae</i>	oral cavity
<i>Prevotella oralis</i> ATCC 33269	<i>Prevotellaceae</i>	oral cavity
<i>Prevotella oulorum</i> F0390	<i>Prevotellaceae</i>	oral cavity
<i>Prevotella ruminicola</i> 23	<i>Prevotellaceae</i>	rumen
<i>Prevotella saccharolytica</i> F0055	<i>Prevotellaceae</i>	oral cavity
<i>Prevotella</i> sp. C561	<i>Prevotellaceae</i>	oral cavity
<i>Prevotella</i> sp. MSX73	<i>Prevotellaceae</i>	oral cavity
<i>Prevotella</i> sp. oral taxon 306 str. F0472	<i>Prevotellaceae</i>	oral cavity
<i>Prevotella</i> sp. oral taxon 317 str. F0108	<i>Prevotellaceae</i>	oral cavity
<i>Prevotella</i> sp. oral taxon 472 str. F0295	<i>Prevotellaceae</i>	oral cavity
<i>Prevotella stercorea</i> DSM 18206	<i>Prevotellaceae</i>	gastrointestinal tract/feces
<i>Prevotella tanneriae</i> ATCC 51259	<i>Prevotellaceae</i>	oral cavity
<i>Prevotella timonensis</i> CRIS 5C-B1	<i>Prevotellaceae</i>	wound (breast abscess)
<i>Prevotella veroralis</i> F0319	<i>Prevotellaceae</i>	oral cavity
<i>Tannerella</i> sp. 6_1_58FAA_CT1	<i>Porphyromonadaceae</i>	gastrointestinal tract/feces
Cytophagia		
<i>Belliella baltica</i> DSM 15883	<i>Cyclobacteriaceae</i>	environmental sample (groundwater)
<i>Indibacter alkaliphilus</i> LW1	<i>Cyclobacteriaceae</i>	extremophile (soda lake)
<i>Nitritalea halalkaliphila</i> LW7	<i>Cyclobacteriaceae</i>	extremophile (saline soda lake)
Flavobacteria		
<i>Bergeyella zoohelcum</i> ATCC 43767	<i>Flavobacteriaceae</i>	oral cavity dog and cat oral cavity/zoonotic infections
<i>Capnocytophaga canimorsus</i> Cc5	<i>Flavobacteriaceae</i>	infections
<i>Capnocytophaga gingivalis</i> ATCC 33624	<i>Flavobacteriaceae</i>	oral cavity
<i>Capnocytophaga ochracea</i> DSM 7271	<i>Flavobacteriaceae</i>	oral cavity
<i>Capnocytophaga</i> sp. CM59	<i>Flavobacteriaceae</i>	oral cavity
<i>Capnocytophaga</i> sp. oral taxon 324 str. F0483	<i>Flavobacteriaceae</i>	oral cavity
<i>Capnocytophaga</i> sp. oral taxon 326 str. F0382	<i>Flavobacteriaceae</i>	oral cavity
<i>Capnocytophaga</i> sp. oral taxon 329 str. F0087	<i>Flavobacteriaceae</i>	oral cavity
<i>Capnocytophaga</i> sp. oral taxon 335 str. F0486	<i>Flavobacteriaceae</i>	oral cavity
<i>Capnocytophaga</i> sp. oral taxon 380 str. F0488	<i>Flavobacteriaceae</i>	oral cavity
<i>Capnocytophaga</i> sp. oral taxon 412 str. F0487	<i>Flavobacteriaceae</i>	oral cavity
<i>Capnocytophaga sputigena</i> ATCC 33612	<i>Flavobacteriaceae</i>	oral cavity
<i>Chryseobacterium</i> sp. CF314	<i>Flavobacteriaceae</i>	vegetation
<i>Flavobacteriaceae</i> bacterium S85	<i>Flavobacteriaceae</i>	environmental sample (seawater)
<i>Flavobacterium branchiophilum</i> FL-15	<i>Flavobacteriaceae</i>	fish pathogen

Strain <sup>a</sup>	Class	Isolation/habitat <sup>b</sup>
<i>Flavobacterium columnare</i> ATCC 49512	Flavobacteriaceae	fish pathogen
<i>Flavobacterium psychrophilum</i> JIP02/86	Flavobacteriaceae	fish pathogen
<i>Fluviicola taffensis</i> DSM 16823	Cryomorpaceae	environmental sample (fresh water)
<i>Galbibacter</i> sp. ck-12-15	Flavobacteriaceae	extremophile (deep sea sediment)
<i>Joostella marina</i> DSM 19592	Flavobacteriaceae	environmental sample (seawater)
<i>Kordia algicida</i> OT-1	Flavobacteriaceae	environmental sample (seawater)
<i>Myroides injenensis</i> M09-0166	Flavobacteriaceae	human clinical specimens
<i>Myroides odoratus</i> DSM 2801	Flavobacteriaceae	fish
Flavobacteria (continued)		
<i>Ornithobacterium rhinotracheale</i> DSM 15997	Flavobacteriaceae	bird respiratory tract
<i>Psychroflexus torquis</i> ATCC 700755	Flavobacteriaceae	extremophile (antarctic ice)
<i>Riemerella anapestifer</i> ATCC 11845 = DSM 15868	Flavobacteriaceae	bird
<i>Weeksella virosa</i> DSM 16922	Flavobacteriaceae	genital tract/urine
<i>Zunongwangia profunda</i> SM-A87	Flavobacteriaceae	extremophile (deep sea sediment)
Sphingobacteria		
<i>Mucilaginibacter paludis</i> DSM 18603	Sphingobacteriaceae	food (fermented)
<i>Niabella soli</i> DSM 19437	Chitinophagaceae	environmental sample (soil)
<i>Sphingobacterium spiritivorum</i> ATCC 33861	Sphingobacteriaceae	human clinical specimens
Firmicutes		
Bacilli		
<i>Alicyclophilus denitrificans</i>	Alicyclobacillaceae	environmental sample (sewage)
<i>Alicyclobacillus hesperidum</i> URH17-3-68	Alicyclobacillaceae	extremophile (hot water spring)
<i>Bacillus cereus</i> Rock1-15	Bacillaceae	environmental sample (soil)
<i>Bacillus smithii</i> 7 3 47FAA	Bacillaceae	human clinical specimens
<i>Bacillus thuringiensis</i> serovar <i>finitimus</i> YBT-020	Bacillaceae	environmental sample (soil)
<i>Brevibacillus laterosporus</i> GI-9	Paenibacillaceae	environmental sample (soil)
<i>Catelicoccus marimammalium</i> M35/04/3	Enterococcaceae	grey seal gastrointestinal tract
<i>Dolosigranulum pigrum</i> ATCC 51524	Carnobacteriaceae	human clinical specimens
<i>Enterococcus faecalis</i> TX0012	Enterococcaceae	gastrointestinal tract/feces
<i>Enterococcus faecium</i> 1231408	Enterococcaceae	gastrointestinal tract/feces
<i>Enterococcus hirae</i> ATCC 9790	Enterococcaceae	gastrointestinal tract/feces
<i>Enterococcus italicus</i> DSM 15952	Enterococcaceae	food (fermented)
<i>Enterococcus</i> sp. 7L76	Enterococcaceae	gastrointestinal tract/feces
<i>Facklamia hominis</i> CCUG 36813	Aerococcaceae	burbuncle (human)
<i>Fructobacillus fructosus</i> KCTC 3544	Leuconostocaceae	vegetation
<i>Gemella haemolysans</i> ATCC 10379	Streptococcaceae	oral cavity
<i>Gemella moribillum</i> M424	Streptococcaceae	gastrointestinal tract/feces
<i>Lactobacillus animalis</i> KCTC 3501	Lactobacillaceae	food (fermented)
<i>Lactobacillus brevis</i> subsp. <i>gravesensis</i> ATCC 27305	Lactobacillaceae	food (fermented)
<i>Lactobacillus buchneri</i> ATCC 11577	Lactobacillaceae	food (fermented)
<i>Lactobacillus casei</i> str. Zhang	Lactobacillaceae	gastrointestinal tract/feces
<i>Lactobacillus coryniformis</i> subsp. <i>coryniformis</i> KCTC 3167	Lactobacillaceae	food (fermented)
<i>Lactobacillus coryniformis</i> subsp. <i>torquens</i> KCTC 3535	Lactobacillaceae	food (fermented)
<i>Lactobacillus crispatus</i> FB049-03	Lactobacillaceae	genital tract
<i>Lactobacillus curvatus</i> CRL 705	Lactobacillaceae	food (fermented)
<i>Lactobacillus delbrueckii</i> subsp. <i>bulgaricus</i> 2038	Lactobacillaceae	food (fermented)

Strain <sup>a</sup>	Class	Isolation/habitat <sup>b</sup>
<i>Lactobacillus farciminis</i> KCTC 3681	Lactobacillaceae	food (fermented)
<i>Lactobacillus fermentum</i> ATCC 14931	Lactobacillaceae	food (fermented)
<i>Lactobacillus florum</i> 2F	Lactobacillaceae	vegetation
<i>Lactobacillus gasseri</i> JV-V03	Lactobacillaceae	oral cavity
<i>Lactobacillus hominis</i> CRBIP 24.179	Lactobacillaceae	gastrointestinal tract/feces
<i>Lactobacillus iners</i> LactinV 11V1-d	Lactobacillaceae	genital tract/urine
<i>Lactobacillus jensenii</i> 269-3	Lactobacillaceae	genital tract/blood
<i>Lactobacillus johnsonii</i> DPC 6026	Lactobacillaceae	pig gastrointestinal tract
<i>Lactobacillus mucosae</i> LM1	Lactobacillaceae	wild pig gastrointestinal tract
<i>Lactobacillus paracasei</i> subsp. <i>paracasei</i> 8700:2	Lactobacillaceae	food (fermented)
<i>Lactobacillus pentosus</i> IG1	Lactobacillaceae	food (fermented)
<i>Lactobacillus plantarum</i> ZJ316	Lactobacillaceae	gastrointestinal tract/feces
<i>Lactobacillus rhamnosus</i> GG	Lactobacillaceae	gastrointestinal tract/feces
<i>Lactobacillus ruminis</i> ATCC 25644	Lactobacillaceae	rumen
<i>Lactobacillus salivarius</i> UCC118	Lactobacillaceae	oral cavity
<i>Lactobacillus sanfranciscensis</i> TMW 1-1304	Lactobacillaceae	food (fermented)
<i>Lactobacillus</i> sp. 66c	Lactobacillaceae	ND
<i>Lactobacillus versmoldensis</i> KCTC 3814	Lactobacillaceae	food (fermented)
<i>Leuconostoc gelidum</i> KCTC 3527	Leuconostocaceae	food (fermented)
<i>Leuconostoc pseudomesenteroides</i> 4882	Leuconostocaceae	food (fermented)

## Bacilli (continued)

<i>Listeria innocua</i> Clip11262	Listeriaceae	environmental sample (soil)
<i>Listeria ivanovii</i> FSL F6-596	Listeriaceae	animal and human/environmental samples
<i>Listeria monocytogenes</i> str. 1/2a F6854	Listeriaceae	animal and human/environmental samples
<i>Listeria seeligeri</i> FSL N1-067	Listeriaceae	animal and human/environmental samples
Listeriaceae bacterium TTU M1-001	Listeriaceae	environmental sample (soil)
<i>Oenococcus kitaharae</i> DSM 17330	Leuconostocaceae	food (fermented)
<i>Pediococcus acidilactici</i> DSM 20284	Lactobacillaceae	vegetation
<i>Pediococcus lolii</i> NGRI 0510Q	Lactobacillaceae	vegetation (fermented)
<i>Planococcus antarcticus</i> DSM 14505	Planococcaceae	extremophile (antarctic)
<i>Sporolactobacillus vineae</i> DSM 21990 = SL153	Sporolactobacillaceae	environmental sample (soil)
<i>Staphylococcus aureus</i> subsp. <i>aureus</i>	Staphylococcaceae	human clinical specimens
<i>Staphylococcus lugdunensis</i> M23590	Staphylococcaceae	human clinical specimens
<i>Staphylococcus massiliensis</i> S46	Staphylococcaceae	skin
<i>Staphylococcus pseudintermedius</i> ED99	Staphylococcaceae	dog skin
<i>Staphylococcus simulans</i> ACS-120-V-Sch1	Staphylococcaceae	genital tract
<i>Streptococcus agalactiae</i> 2603V/R	Streptococcaceae	gastrointestinal tract/feces
<i>Streptococcus anginosus</i> F0211	Streptococcaceae	oral cavity
<i>Streptococcus bovis</i> ATCC 700338	Streptococcaceae	rumen/zoonotic infections
<i>Streptococcus canis</i> FSL Z3-227	Streptococcaceae	food (fermented)
<i>Streptococcus constellatus</i> subsp. <i>constellatus</i> SK53	Streptococcaceae	human clinical specimens
<i>Streptococcus downei</i> F0415	Streptococcaceae	monkey oral cavity
<i>Streptococcus dysgalactiae</i> DSM 12112	Streptococcaceae	various animals/zoonotic infections
<i>Streptococcus equi</i> subsp. <i>zooepidemicus</i> MGCS10565	Streptococcaceae	horse respiratory tract
<i>Streptococcus equinus</i> ATCC 9812	Streptococcaceae	ruminants alimentary tract
<i>Streptococcus gallolyticus</i> UCN34	Streptococcaceae	ruminants alimentary tract
<i>Streptococcus gordonii</i> str. Challis substr. CH1	Streptococcaceae	oral cavity
<i>Streptococcus infantarius</i> ATCC BAA-102	Streptococcaceae	gastrointestinal tract/feces

Strain <sup>a</sup>	Class	Isolation/habitat <sup>b</sup>
<i>Streptococcus iniae</i> 9117	<i>Streptococcaceae</i>	fish/human pathogen
<i>Streptococcus macacae</i> NCTC 11558	<i>Streptococcaceae</i>	monkey oral cavity
<i>Streptococcus macedonicus</i> ACA-DC 198	<i>Streptococcaceae</i>	food (fermented)
<i>Streptococcus mitis</i> ATCC 6249	<i>Streptococcaceae</i>	oral cavity
<i>Streptococcus mutans</i> UA159	<i>Streptococcaceae</i>	oral cavity
<i>Streptococcus oralis</i> SK1074	<i>Streptococcaceae</i>	oral cavity
<i>Streptococcus parasanguinis</i> F0449	<i>Streptococcaceae</i>	oral cavity
<i>Streptococcus pasteurianus</i> ATCC 43144	<i>Streptococcaceae</i>	blood
<i>Streptococcus pseudoporcinus</i> SPIN 20026	<i>Streptococcaceae</i>	genital tract
<i>Streptococcus pyogenes</i> SF370	<i>Streptococcaceae</i>	oral cavity/wounds
<i>Streptococcus rattus</i> FA-1 = DSM 20564	<i>Streptococcaceae</i>	rat oral cavity
<i>Streptococcus salivarius</i> JIM8777	<i>Streptococcaceae</i>	oral cavity
<i>Streptococcus sanguinis</i> VMC66	<i>Streptococcaceae</i>	oral cavity
<i>Streptococcus</i> sp. BS35b	<i>Streptococcaceae</i>	oral cavity
<i>Streptococcus</i> sp. C150	<i>Streptococcaceae</i>	oral cavity (expectorated sputum)
<i>Streptococcus</i> sp. C300	<i>Streptococcaceae</i>	oral cavity (expectorated sputum)
<i>Streptococcus</i> sp. F0441	<i>Streptococcaceae</i>	oral cavity
<i>Streptococcus</i> sp. GMD4S	<i>Streptococcaceae</i>	oral cavity
<i>Streptococcus</i> sp. GMD6S	<i>Streptococcaceae</i>	oral cavity
<i>Streptococcus</i> sp. M334	<i>Streptococcaceae</i>	oral cavity (expectorated sputum)
<i>Streptococcus</i> sp. oral taxon 056 str. F0418	<i>Streptococcaceae</i>	oral cavity
<i>Streptococcus</i> sp. oral taxon 071 str. 73H25AP	<i>Streptococcaceae</i>	oral cavity
<i>Streptococcus suis</i> 89/1591	<i>Streptococcaceae</i>	pig
<i>Streptococcus thermophilus</i> LMD-9	<i>Streptococcaceae</i>	food (fermented)
<i>Streptococcus vestibularis</i> ATCC 49124	<i>Streptococcaceae</i>	oral cavity
Clostridia		
<i>Acidaminococcus intestini</i> RyC-MR95	<i>Acidaminococcaceae</i>	wound/abscess
<i>Acidaminococcus</i> sp. D21	<i>Acidaminococcaceae</i>	gastrointestinal tract/feces
<i>Aminomonas paucivorans</i> DSM 12260	<i>Syntrophomonadaceae</i>	environmental sample (sewage)
<i>Anaerococcus tetradius</i> ATCC 35098	<i>Peptostreptococcaceae</i>	human clinical specimens
<i>Butyrivibrio fibrisolvens</i> 16/4	<i>Lachnospiraceae</i>	rumen
<i>Catenibacterium mitsuokai</i> DSM 15897	<i>Lachnospiraceae</i>	gastrointestinal tract/feces
<i>Clostridium cellulolyticum</i> H10	<i>Clostridiaceae</i>	vegetation (composted)
Clostridia (continued)		
<i>Clostridium perfringens</i> D str. JGS1721	<i>Clostridiaceae</i>	environmental sample (vegetation/marine sediment)
<i>Clostridium spiroforme</i> DSM 1552	<i>Clostridiaceae</i>	gastrointestinal tract/feces
<i>Coprococcus catus</i> GD/7	<i>Lachnospiraceae</i>	gastrointestinal tract/feces
<i>Coprococcus comes</i> ATCC 27758	<i>Lachnospiraceae</i>	gastrointestinal tract/feces
<i>Dorea longicatena</i> DSM 13814	<i>Clostridiaceae</i>	gastrointestinal tract/feces
<i>Eubacterium dolichum</i> DSM 3991	<i>Eubacteriaceae</i>	gastrointestinal tract/feces
<i>Eubacterium rectale</i> ATCC 33656	<i>Eubacteriaceae</i>	gastrointestinal tract/feces
<i>Eubacterium</i> sp. AS15	<i>Eubacteriaceae</i>	oral cavity
<i>Eubacterium ventriosum</i> ATCC 27560	<i>Eubacteriaceae</i>	gastrointestinal tract/feces
<i>Eubacterium yurii</i> subsp. <i>margaretiae</i> ATCC 43715	<i>Peptostreptococcaceae</i>	oral cavity
<i>Filifactor alocis</i> ATCC 35896	<i>Peptostreptococcaceae</i>	cat and human oral cavity
<i>Fingoldia magna</i> ATCC 29328	<i>Peptostreptococcaceae</i>	oral cavity
<i>Helcococcus kunzii</i> ATCC 51366	<i>Clostridiales Family XI</i>	wound
<i>Oribacterium sinus</i> F0268	<i>Lachnospiraceae</i>	human clinical specimens
<i>Peptoniphilus duerdenii</i> ATCC BAA-1640	<i>Peptostreptococcaceae</i>	wound
<i>Peptoniphilus</i> sp. oral taxon 386 str. F0131	<i>Peptostreptococcaceae</i>	oral cavity

Strain <sup>a</sup>	Class	Isolation/habitat <sup>b</sup>
<i>Phascolarctobacterium</i> sp. YIT 12067	Acidaminococcaceae	gastrointestinal tract/feces
<i>Phascolarctobacterium succinatutens</i> YIT 12067	Acidaminococcaceae	gastrointestinal tract/feces
<i>Pseudoramibacter alactolyticus</i> ATCC 23263	Clostridiaceae	oral cavity
<i>Roseburia intestinalis</i> L1-82	Lachnospiraceae	gastrointestinal tract/feces
<i>Roseburia inulinivorans</i> DSM 16841	Lachnospiraceae	gastrointestinal tract/feces
<i>Ruminococcus albus</i> 8	Ruminococcaceae	gastrointestinal tract/feces
<i>Ruminococcus lactaris</i> ATCC 29176	Ruminococcaceae	gastrointestinal tract/feces
<i>Subdoligranulum</i> sp. 4_3_54A2FAA	Ruminococcaceae	gastrointestinal tract/feces
Negativicutes		
<i>Megasphaera</i> sp. UPII 135-E	Veillonellaceae	rumen
<i>Veillonella atypica</i> ACS-134-V-Col7a	Veillonellaceae	oral cavity
<i>Veillonella parvula</i> ATCC17745	Veillonellaceae	gastrointestinal/genital tract
<i>Veillonella</i> sp. 6_1_27	Veillonellaceae	gastrointestinal tract/feces
<i>Veillonella</i> sp. oral taxon 780 str. F0422	Veillonellaceae	oral cavity
Proteobacteria		
Alphaproteobacteria		
<i>Acetobacter acetii</i> NBRC 14818	Acetobacteraceae	environmental sample
<i>Azospirillum</i> sp. B510	Rhodospirillaceae	vegetation
<i>Bradyrhizobium</i> sp. BTAi1	Bradyrhizobiaceae	vegetation
<i>Caenispirillum salinarum</i> AK4	Rhodospirillaceae	extremophile (solar saltern)
<i>Dinoroseobacter shibae</i> DFL 12	Rhodobacteraceae	environmental sample (seawater)
<i>Gluconacetobacter diazotrophicus</i> PAI5	Acetobacteriaceae	vegetation
<i>Maritimibacter alkaliphilus</i> ATCC2654	Rhodobacteraceae	environmental sample (seawater)
<i>Methylocystis</i> sp. ATCC 49242	Methylocystaceae	environmental sample (sewage, fresh water)
<i>Methylosinus trichosporium</i> OB3b	Methylocystaceae	environmental sample (soil, fresh water)
<i>Nitrobacter hamburgensis</i> X14	Bradyrhizobiaceae	environmental sample (soil)
<i>Parvibaculum lavamentivorans</i> DS-1	Phyllobacteriaceae	environmental sample (sewage)
<i>Puniceispirillum marinum</i> IMCC1322	SAR16 clade	environmental sample (seawater)
<i>Rhodopseudomonas palustris</i> BisB18	Bradyrhizobiaceae	environmental sample (soil)
<i>Rhodospirillum rubrum</i> ATCC 11170	Rhodospirillaceae	environmental sample (sea mud)
<i>Rhodovulum</i> sp. PH10	Rhodobacteraceae	environmental sample (soil)
<i>Sphingobium</i> sp. AP49	Sphingomonadaceae	vegetation
<i>Sphingomonas</i> sp. S17	Sphingomonadaceae	environmental sample (stromatolite)
<i>Tistrella mobilis</i> KA081020-065	Rhodospirillaceae	environmental sample (seawater)
Betaproteobacteria		
<i>Acidovorax avenae</i> subsp. <i>avenae</i> ATCC 19860	Comamonadaceae	environmental sample (soil)
<i>Acidovorax ebreus</i> TPSY	Comamonadaceae	environmental sample (water)
<i>Burkholderiales</i> bacterium 1 1 47	Burkholderiales	gastrointestinal tract/feces
<i>Kingella kingae</i> ATCC 23330	Neisseriaceae	oral cavity
<i>Neisseria bacilliformis</i> ATCC BAA-1200	Neisseriaceae	oral cavity
Betaproteobacteria (continued)		
<i>Neisseria cinerea</i> ATCC 14685	Neisseriaceae	oral cavity
<i>Neisseria flavescens</i> SK114	Neisseriaceae	human clinical specimens
<i>Neisseria lactamica</i> 020-06	Neisseriaceae	oral cavity
<i>Neisseria meningitidis</i> A Z2491	Neisseriaceae	oral cavity
<i>Neisseria mucosa</i> C102	Neisseriaceae	oral cavity (expectorated sputum)
<i>Neisseria</i> sp. oral taxon 014 str. F0314	Neisseriaceae	oral cavity

Strain <sup>a</sup>	Class	Isolation/habitat <sup>b</sup>
<i>Neisseria subflava</i> NJ9703	<i>Neisseriaceae</i>	oral cavity
<i>Neisseria wadsworthii</i> 9715	<i>Neisseriaceae</i>	skin
<i>Nitrosomonas</i> sp. AL212	<i>Nitrosomonadaceae</i>	environmental sample (fresh water)
<i>Parasutterella excrementihominis</i> YIT 11859	<i>Alcaligenaceae</i>	gastrointestinal tract/feces
<i>Ralstonia syzygii</i> R24	<i>Burkholderiaceae</i>	environmental sample (soil)
<i>Simonsiella muelleri</i> ATCC 29453	<i>Neisseriaceae</i>	oral cavity
<i>Sutterella parvirubra</i> YIT 11816	<i>Alcaligenaceae</i>	gastrointestinal tract/feces
<i>Sutterella wadsworthensis</i> 3 1 45B	<i>Alcaligenaceae</i>	gastrointestinal tract/feces
<i>Verminephrobacter aporrectodeae</i> subsp. <i>tuberculatae</i> At4	<i>Comamonadaceae</i>	invertebrate (earthworm)
<i>Verminephrobacter eiseniae</i> EF01-2	<i>Comamonadaceae</i>	invertebrate (earthworm)
<b>Gamma</b> proteobacteria		
<i>Actinobacillus minor</i> NM305	<i>Pasteurellaceae</i>	pig respiratory tract
<i>Actinobacillus pleuropneumoniae</i> serovar 10 D13039	<i>Pasteurellaceae</i>	pig respiratory tract
<i>Actinobacillus succinogenes</i> 130Z	<i>Pasteurellaceae</i>	rumen
<i>Actinobacillus suis</i> H91-0380	<i>Pasteurellaceae</i>	pig pathogen
<i>Actinobacillus ureae</i> ATCC 25976	<i>Pasteurellaceae</i>	respiratory tract
<i>Alcanivorax</i> sp. W11-5	<i>Alcanivoracaceae</i>	extremophile (deep sea sediment)
<i>Francisella tularensis</i> subsp. <i>holarctica</i> LVS	<i>Francisellaceae</i>	engineered live vaccine strain human/environmental sample (water)
<i>Francisella tularensis</i> subsp. <i>novicida</i> U112	<i>Francisellaceae</i>	wound
<i>Francisella tularensis</i> subsp. <i>tularensis</i> WY96-3418	<i>Francisellaceae</i>	environmental sample (seawater)
gamma proteobacterium HTCC5015	<i>Unclassified</i>	environmental sample (sewage)
gammaproteobacterium HdN1	<i>Unclassified</i>	oral cavity/genital tract
<i>Haemophilus parainfluenzae</i> T3T1	<i>Pasteurellaceae</i>	oral cavity
<i>Haemophilus pittmaniae</i> HK 85	<i>Pasteurellaceae</i>	oral cavity
<i>Haemophilus sputorum</i> HK 2154	<i>Pasteurellaceae</i>	oral cavity
<i>Legionella pneumophila</i> str. Paris	<i>Legionellaceae</i>	human clinical specimens
<i>Pasteurella bettyae</i> CCUG 2042	<i>Pasteurellaceae</i>	genital tract
<i>Pasteurella multocida</i> subsp. <i>gallicida</i> X73	<i>Pasteurellaceae</i>	bird pathogen
<i>Pasteurella multocida</i> Pm70	<i>Pasteurellaceae</i>	bird respiratory tract/zoonotic infections
<b>Delta</b> proteobacteria		
uncultured delta proteobacterium HF0070_07E19	<i>Unclassified</i>	environmental sample (seawater)
<b>Epsilon</b> proteobacteria		
<i>Campylobacter coli</i> 2962	<i>Campylobacteraceae</i>	animals/human pathogen
<i>Campylobacter jejuni</i> NCTC11168	<i>Campylobacteraceae</i>	bird
<i>Campylobacter jejuni</i> subsp. <i>doylei</i> 269-97	<i>Campylobacteraceae</i>	blood
<i>Campylobacter lari</i>	<i>Campylobacteraceae</i>	gastrointestinal tract/feces
<i>Helicobacter canadensis</i> MIT 98-5491	<i>Helicobacteriaceae</i>	gastrointestinal tract/feces
<i>Helicobacter cinaedi</i> CCUG 18818	<i>Helicobacteriaceae</i>	gastrointestinal tract/feces
<i>Helicobacter hepaticus</i> ATCC 51449	<i>Helicobacteriaceae</i>	mouse liver
<i>Helicobacter mustelae</i> 12198	<i>Helicobacteriaceae</i>	ferret
<i>Helicobacter pullorum</i> MIT 98-5489	<i>Helicobacteriaceae</i>	bird/zoonotic infections
<i>Nitratifractor salsuginis</i> DSM 16511	<i>Unclassified</i>	extremophile (deep sea sediment)
<i>Wolinella succinogenes</i> DSM 1740	<i>Helicobacteraceae</i>	rumen
<b>Fusobacteria</b>		
<i>Fusobacterium nucleatum</i> subsp. <i>vincentii</i> ATCC	<i>Fusobacteriaceae</i>	oral cavity



Strain <sup>a</sup>	Class	Isolation/habitat <sup>b</sup>
49256		
<i>Fusobacterium</i> sp. 1_1_41FAA	<i>Fusobacteriaceae</i>	gastrointestinal tract/feces
<i>Fusobacterium</i> sp. 3_1_27	<i>Fusobacteriaceae</i>	gastrointestinal tract/feces
<i>Fusobacterium</i> sp. 3_1_36A2	<i>Fusobacteriaceae</i>	gastrointestinal tract/feces
<i>Ilyobacter polytropus</i> DSM 2926	<i>Fusobacteriaceae</i>	environmental sample (sea mud)
<i>Streptobacillus moniliformis</i> DSM 12112	<i>Leptotrichiaceae</i>	rodent/human pathogen
<b>Spirochaetes</b>		
<i>Leptospira inadai</i> serovar Lyme str. 10	<i>Leptospiraceae</i>	human clinical specimens
<i>Sphaerochaeta globus</i> str. Buddy	<i>Spirochaetaceae</i>	extremophile (marine hot spring)
<i>Treponema denticola</i> ATCC 35405	<i>Spirochaetaceae</i>	oral cavity
<i>Treponema phagedenis</i> F0421	<i>Spirochaetaceae</i>	monkey genital tracts
<i>Treponema</i> sp. JC4	<i>Spirochaetaceae</i>	rumen
<i>Treponema vincentii</i> ATCC 35580	<i>Spirochaetaceae</i>	oral cavity
<b>Tenericutes</b>		
<b>Mollicutes</b>		
<i>Mycoplasma canis</i> PG 14	<i>Mycoplasmataceae</i>	dog oral cavity
<i>Mycoplasma cynos</i> C142	<i>Mycoplasmataceae</i>	dog respiratory tract
<i>Mycoplasma gallisepticum</i> str. F	<i>Mycoplasmataceae</i>	bird pathogen
<i>Mycoplasma iowae</i> 695	<i>Mycoplasmataceae</i>	bird
<i>Mycoplasma mobile</i> 163K	<i>Mycoplasmataceae</i>	fish pathogen
<i>Mycoplasma ovipneumoniae</i> SC01	<i>Mycoplasmataceae</i>	goat respiratory tract
<i>Mycoplasma synoviae</i> 53	<i>Mycoplasmataceae</i>	bird pathogen
<i>Solobacterium moorei</i> F0204	<i>Erysipelotrichaceae</i>	gastrointestinal tract/feces
<b>Elusimicrobia</b>		
<i>Elusimicrobium minutum</i> Pei191	<i>Elusimicrobiaceae</i>	invertebrate (scarab beetle)
Uncultured Termite group 1 bacterium phylotype Rs-D17	<i>Elusimicrobiaceae</i>	invertebrate
<b>Fibrobacteres</b>		
<i>Fibrobacter succinogenes</i> S85	<i>Fibrobacteraceae</i>	rumen
<b>Ignavibacteria</b>		
<i>Ignavibacterium album</i> JCM 16511	<i>Ignavibacteriaceae</i>	extremophile (hot water spring)
<b>Planctomycetes</b>		
<i>Blastopirellula marina</i> DSM 3645	<i>Planctomycetaceae</i>	environmental sample (seawater)
<b>Verrucomicrobia</b>		
<i>Diplosphaera colitermitum</i> TAV2	<i>Opitutaceae</i>	invertebrate (termite)
<i>Akkermansia muciniphila</i> ATCC BAA-835	<i>Verrucomicrobiaceae</i>	gastrointestinal tract/feces
<b>Unclassified</b>		
candidate division TM7 single-cell isolate TM7c	<i>Unclassified</i>	oral cavity
uncultured bacterium	<i>Unclassified</i>	environmental sample (groundwater)
uncultured bacterium	<i>Unclassified</i>	environmental sample (groundwater)
uncultured bacterium T3_7_42578	<i>Unclassified</i>	invertebrate (honeybee)

<sup>a</sup>Single strains representing every species found to harbor the cas9 gene are listed.

<sup>b</sup>The origin of the specific strain and/or typical habitat of the species are given for every strain. ND, no data available. Note that if not specified otherwise, isolates from body sites and feces are human commensals and pathogens.

Supplementary Table S5 – tracrRNA and CRISPR repeats associated with the examined type II CRISPR-Cas systems.

strain	Cas9 tree position <sup>a</sup>	tracrRNA sequence <sup>b</sup>	Repeat <sup>c</sup>	Number of repeats <sup>d</sup>	tracrRNA length	Repeat length <sup>e</sup>
Francisella novicida U112	1	AUCUAAAAUUUAAAUGUACCAAUAAUUAAUAAUGCUCUG UAAUCAUUUAAAAGUAAUUUGAACGGACCUCUGUUUGA CACGUCUGAAUAAC (SEQ ID NO:2702)	GUUUCAGUUGCUGAAUUUUU GGUAAACUACUGUUAG (SEQ ID NO:2765)	14	90	37
Gamma proteobacterium HTCC5015	2	UCAGAAUGCAUCCCAUCAUUCUAUACACUGAAAUCAUA GAAAAUCACGUUUGGCCCCGACCAACUGCUUCGGCAU GUCGGUUUUUU (SEQ ID NO:2703)	GUUUCAGCUGUUGGUUUGUUG GGAUAAGCUCUGAAAC (SEQ ID NO:2766)	27	88	37
Parasutterella excrementihominis YIT11859	3	UUAAUUACAUCUUUUUAAACAACGAAGUCGCCUUCGGGC GAGCUGAAAUCAAUUUGAUUUAAAUUUAGAUCCGGCUA CUGAGGCUUUUGACCUUAUCCGGAUUAAACGAAGAGCCU CCGAGGAGGCUUUUU (SEQ ID NO:2704)	GUUUCAGUAGUUGUUAGAAGA AUGUAGUAUUGAAGCC (SEQ ID NO:2767)	>2	129	37
Sutterella wadsworthensis 3_1_45B	4	UUAGAGAUCAUAACCGCUAUGAGCUAUAGGAAAUCACCU UCGGGUGAGCUGAAAUCGCCUAAAGCUAAGAUAUGAAUC CGGCCACUAUCUAUUAGUAGAUUCCGGAUUUCU (SEQ ID NO:2705)	GUUUCAGUGCUAUAGCUCGUA GCGUUUAGAUUCUCCG (SEQ ID NO:2768)	13	111	37
Legionella pneumophila	5	UAAAAUAGAAAUCAUCUAAAAUUUCGAUACCCUGAAAUC AACAAAAUUAAAGAUUGAAUCGUUUUUUCUAUGCUCGUC	GUUUCAGUGGUUGGAUUUUUA GAUGAGGGAUUUAUUGG (SEQ ID NO:2769)	34	100	37

str. Paris	UUAAUAGCGAGCAUUAACGAUUU (SEQ ID NO: 2706)	ID NO: 2769)			
<i>Wolinella succinogenes</i> DSM 1740	UUGUUAGAAUUGUUC CGCAACACUUUUUAGCAAUCCG UUCGAUCCUUGAAAUCAUCAAAAAGAUUAAUAGACC CGCCACUGUUAUUGUACAUUGCGGGGACUUUUU (SEQ ID NO: 2707)	GUUUCACAGGCUAAGCGGAUU UGCUAUAAAAGUGUUGC (SEQ ID NO: 2770)	23	108	37
<i>Staphylococcus pseudintermedius</i> ED99	GUUUUACUUUUUUCUAAAUAACAUAAGUUUAAA CAAGCUAAAAGCGUCAUUAUUUUUUAACACCC UACUGUGACAGUGGGUUUUUUU (SEQ ID NO: 2708)	GUUUUAGCACUAUUGUUUUUU AGAAAAGAGUAAAAC (SEQ ID NO: 2771)	24	99	36
<i>Planococcus antartcticus</i> DSM 14505	AUUUCAAAAUAUUCCCCUUUAUAUUUUUCAAAGAAA AUGUACGUAAAGAGUGUUACUACUCUGUAACAUUACAU UGGUACGUAAAUAAGCUUAAAAGCGUAAAAGUUGGCC CUAUGAGGUCUCCGCCAUCGACUUCGUCGGUGGCCUUUU UU (SEQ ID NO: 2709)	GUUUUAGACCAAUGUAAUUUU AGAGAGUAGUAAAAC (SEQ ID NO: 2772)	>5	154	36
<i>Streptococcus sanguinis</i> SK49	AACUACGUUGGAACUUAUUCGAAAACAACACAGCCAAAAG AUUUUUUUUUUUGAGUUAAAUAUUGGUUAUCCAUAAUCA GUUAUGCGCACCGAUUCGGUGUUUUUU (SEQ ID NO: 2710)	GUUUUAGAGCUGUGUUUUUC GAAUGGUUCCAAAAC (SEQ ID NO: 2773)	11	104	36
<i>Listeria innocua</i> Clip11262	AUUGUUAGUUAUUCAAAAUAACAUAAGCAAAGUUAAAUA GGUUUUUGCCGUUAUCAACUUUUUAAUUAAGUAGCGCUG UUUCGGCGUUUUU (SEQ ID NO: 2711)	GUUUUAGAGCUAUGUUUUUU GAAUGCUAACAACAAC (SEQ ID NO: 2774)	11	90	36
<i>Streptococcus pyrogenes</i> SF370 (M1 GAS)	GUUGGAACCAUUCAAAAACAGCAUAAGCAAGUUAAAUA GGCUAGUCCGUUAUCAACUUUAAAAGUGGCACCGAGU CGGUGCUUUUUUU (SEQ ID NO: 2712)	GUUUUAGAGCUAUGUGUUUU GAAUGGUCCAAAAC (SEQ ID NO: 2775)	7	89	36

Streptococcus thermophilus LMD-9	11	UUUGUUUUGAAACCAUUCGAAACAACACAGCGAGUUA AAUAAAGGUUAGUCGUAACUCACUAAUUGAAAAGGUGGC ACCGAUUCGGUUUUUUUUU (SEQ ID NO:2713)	GUUUUAGAGCUGUGUUUC GAAUGGUUCCAAAAC (SEQ ID NO:2776)	9	96	36
Streptococcus mutans UA159	12	GUUGAAUCAUUCGAAACAACACAGCAAGUAAAAUA GGCAGUUAUUUAAUCCAGUCCGUACACAACUUGAAA AAGUGCGCACCGAUUCGGUUCUUUU (SEQ ID NO:2714)	GUUUUAGAGCUGUGUUUC GAAUGGUUCCAAAAC (SEQ ID NO:2777)	6	102	36
Coriobacterium glomerans PW2	13	CGUUUAUACAGUCAGGACAGCACUGCGAGUCAAA AUACGGCUUUGCCAAACUUGCCUCCUUCGGAGCGUC UCGUAGGACAAUUUGAAGCCCUUAGGGGUUCAU UUUUCU (SEQ ID NO:2715)	GUUUUGGAGCAGUGUUCU GACUGGUAUCCAAC (SEQ ID NO:2778)	10	120	36
Lactobacillus farciminis KCTC 3681	14	GUUUACUUAUUUAGAUUUUUAAGAUCAAAAAAU AAGGAUUUCCGAAUUUACCACCUUUUUUUUUUU AGGUGUUUUUUU (SEQ ID NO:2716)	GUUUUUGUACCUUAAAAGAAUC UAGAAUAGUAAAAC (SEQ ID NO:2779)	9	89	36
Catenibacterium mitsuokai DSM 15897	15	Too short contig	GUUUUAGGGUUUUGUUUUUU GAACUGAAUUAAAAC (SEQ ID NO:2780)	>4	-	36
Lactobacillus rhamnasus GG	16	UGUUGAGACGACAUCCUACAACAUCUUGAAUUGAU CUGACAUUACGAGUUGAGAUCAACAAGCUUCAGCU GAGUUCAUUUUCUGAGCCCAUGUUGGCCCAUACAU GCCACCCGAGUGCAAAUCGGUGGUUUUUUU (SEQ ID NO:2718)	GUCUCAGGUAGAUGCAGAU AAUCAGUUAAGAGC (SEQ ID NO:2781)	25	146	36
Bifidobacterium	17	GGAUUGUUUGGUCGAAUCCAUCAAGGUCAUUGAC CUGACAGGCAUAAAUUGAAAUAAAGCAAGGUUUCGACC	GUUUCAGAUCCUGUAGAU AAUGACUUUGACCAC (SEQ	45	144	36

bifidum S17	AAGCUUCAGAAGUUUUUAUACCCUGGGCCUUUAGGCUGUG AGGCUCGCCGAUAAUGUCGGGAGCCUCUUUU (SEQ ID NO:2719)		ID NO:2782)		
Oenococcus kitaharae DSM 17330	18 UUGGGAUUGAUCAUCCCAAAACAUCAUUGGGUUCUACCU CAUUGAUCUGACACACAGCAUUGAAGUAAGCAAGAUU AAUUCAAAGCUAAUUUCUUCACAUUUUAUGUGCAGA AGGCCUUUAGCCCAAAUACAUAFAAAAGUCGCAUUCA CUUGCGGACUUUUU (SEQ ID NO:2720)	18	GCUUCAGAUGUGUCAGAUC AAUGAGGUAGAACC (SEQ ID NO:2783)	58	167 36
Fructibacil lus fructosus KCTC 3544	19 CAUGUUUAGCUACCAUACAAGCAAGAAUUGUUUAGCUA ACUAAUUCUUGCUAGGAAGAACCCCAUUAAUCUGACAUAAC AGGUUAAAAGUAACGCAAGGGCUUCAGCCCCAAGCUUCA UGAACUUUUAAAAGUUGGCCUUUAGGCCUUUUUUU (SEQ ID NO:2721)	19	GCUUUAGAUGAUGCGGAUU AAUGGGUUUCUCC (SEQ ID NO:2784)	>2	149 36
Fingoldia magna ATCC 29328	20 UAAUCUCAGUUUUAAUACCUAAUUGAGAUAUACAUCAUAGA GUUCAAAUAAAAGUUUACUCAAAUCGCCCGAAAAGAGCC CACAUUGGGGACUAAAACAAAUCUUCGGAUUUUGUUUUU UU (SEQ ID NO:2722)	20	GUUUGAGAAUGAUGAAUUUC AAUAGGUAAUAAAAC (SEQ ID NO:2785)	15	116 36
Viellonella atypical ACS-134-V- Co17a	21 AUAAGUAAUCCAAUUAGUUUUUGGAGGUUUACAGAAUUA CACUACGAGUUCAAAUAACAUAUUUUUAUUAUCAAUGCCUU CGGGCCACCCGACGAGGGUAUCAUCUCAAUUUCUUCUG AAUUGGGAUUUUUU (SEQ ID NO:2723)	21	GUUUGCGGAGUAGUAAUUUCU GUAAAUCUCUAAAAC (SEQ ID NO:2786)	33	129 36
Solobacteri um moorei F0204	22 GAAUUUGUCUUUAUACCAGUAAGAAUUUUACAUAAGUAA GUUCAAAAACAGCUUUUAGCGAAAUUUACCGCUUUUGCGGA UUCACAUUGUGUGAAGUUUAACUCUCGAAAGAGAGUUUU UUCUUU (SEQ ID NO:2724)	22	CUUUGAGAACUAUGUAAAUA UGCUGGUAAGCAAAAAC (SEQ ID NO:2787)	>2	120 36
Acidaminoco ccus sp.	23 none	23	GUUUGAGAGAAUUGUAAAUAUC AAAGGAUAAUCAAAC (SEQ ID NO:2788)	40	- 36

D21				ID NO:2788)				
Eubacterium yurii subsp. Margaretiae ATCC 43715	24	AUAUCAUUUCAUUUGAUUUUACAAGGUGAGUUCAAAACAA GGAAUUUAUCCGGUAAUUGAUUGCUCGCAUUGUGCGACAU UUUUUUAUGUAAAUCGGAAGUCGGACUUUCGACUUUCU UUUUUUU (SEQ ID NO:2725)	GUUUGAGAAUUGAUUUUUUUU GUAUGGUACUCAAGC (SEQ ID NO:2790)	17	121	36		
Coprococcus catus GD/7	25	none	GUUUGAGAAUUGAUUUUUUUU GUAUGGUACUCAAGC (SEQ ID NO:2790)	16	-	36		
Fusobacterium nucleotum subsp. Vincentii ATCC 49256	26	none	GUUUGAGAAUUGAUUUUUUUU AAUAGAUUCAAAAC (SEQ ID NO:2791)	>3	-	36		
Filifactor alocis ATCC 35896	27	GUUGACUACCAUUGAGAUUACACUACACGGUUCAAAU AAAGAAUUUUUUAUUCGAAUUGCCCAUUGGGCCCAUUAUGAU AUGGAUAAAACUCGCUUAGCGAUUUUUUUU (SEQ ID NO:2726)	GUUUGAGAGUAGUAAUUUC AUUAGGUAGUCAAAAC (SEQ ID NO:2792)	26	106	36		
Peptoniphilus duerdenii ATCC BAA-1640	28	none	GUUUGAGAGUAAUUUGAUUUUC AUUAGGACUAAAAC (SEQ ID NO:2793)	30	-	36		
Treponema denticola ATCC 35405	29	AUUUAAGAUCUUAUUAAAUAACAACAACGAGUUCAAAU AAGAAUUAUCAAAAUCGUCCUUUUUGGGACCGCUCAU UGUGGAGCAUCAAGGCUUAAACAUGGUUUAAGCCUUUUUUU	GUUUGAGAGUAGUAAUUUA AGAUUGAUUCAAAAC (SEQ ID NO:2794)	58	115	36		

Staphylococcus lugenensis M23590	30	U (SEQ ID NO:2727)	GUACUUUAUACCUAAAAUUACAGAAUCUACUGAAAAAAGACAAUAUGUCGUGUUUAUCCCAUCAAAUUUAUUGGGGAAAAUUUUU (SEQ ID NO:)	GUUUUAGUACUCUGUAAAAUUU AGGUAAAGUGAUAC (SEQ ID NO:2795)	3	84	36
Eubacterium dolichum DSM 2991	31	UGAUCAUAAUCUAGCAAAAAGUUUAUAUGAUCUAACAAAACAAGGGUUUAUCCCGAAUCAAGUUCCAAGUAUAUGC UUGGAGCUUUUUUUU (SEQ ID NO:2728)	UGAUCAUAAUCUAGCAAAAAGUUUAUAUGAUCUAACAAAACAAGGGUUUAUCCCGAAUCAAGUUCCAAGUAUAUGC UUGGAGCUUUUUUUU (SEQ ID NO:2728)	GUUUUUGUUACCAUAUUGGAUUU UUGCUGAUAUAAGAC (SEQ ID NO:2796)	16	92	36
Streptococcus thermophilus LMD-9	32	UGUAAGGGACGCCUUACACAGUUACUUAAAUCUUGCAG AAGCUACAAGAUAGGCUUCAUGCCGAAAUAACACC CUGUCAUUUAUUGGCAGGGUUUUUCGUUUUUU (SEQ ID NO:2729)	UGUAAGGGACGCCUUACACAGUUACUUAAAUCUUGCAG AAGCUACAAGAUAGGCUUCAUGCCGAAAUAACACC CUGUCAUUUAUUGGCAGGGUUUUUCGUUUUUU (SEQ ID NO:2729)	GUUUUUGUACUCUCAAGAUUU AAGUAACUGUACAAC (SEQ ID NO:2797)	17	109	36
Enterococcus faecalis TX0012	33	UGUAGUCGACGGACUACCGUGUUUGACGAAACACGUCU UUAUUUUUUACUGAUAAAGAAAUUUAUUGAGAAUCUAC AAAAAUAGGCAUCUUGCCGAAUUUACCGCCCUACAUA UGUAGGGCGUUUUUUU (SEQ ID NO:2730)	UGUAGUCGACGGACUACCGUGUUUGACGAAACACGUCU UUAUUUUUUACUGAUAAAGAAAUUUAUUGAGAAUCUAC AAAAAUAGGCAUCUUGCCGAAUUUACCGCCCUACAUA UGUAGGGCGUUUUUUU (SEQ ID NO:2730)	GUUUUUGUACUCUCAUAAAAU UCUUAUCAGUAAAAAC (SEQ ID NO:2798)	3	130	36
Eubacterium rectale ATCC 33656	34	UGUAGAAAAUUUUUAUAGUCACGUAUUUUUUUCAGAU CUACUAAAAACAAGGCUUUUAUGCCGAAAUCAGGACACC GACGGUGCUUUUUUUU (SEQ ID NO:2731)	UGUAGAAAAUUUUUAUAGUCACGUAUUUUUUUCAGAU CUACUAAAAACAAGGCUUUUAUGCCGAAAUCAGGACACC GACGGUGCUUUUUUUU (SEQ ID NO:2731)	AUUUUAGUAAACUGAAAAUUU ACGUGACUGUAAAAAC (SEQ ID NO:2799)	45	95	36
Mycoplasma mobile 163K	35	UGUUUUUCGAAUAACAGAUACAGUUUAAGAAUACAUA AGAAUGAUACAUCACUAAAAAAAAGCCUUUAUGCCGUA CUACUACUUUUUUCAAAAUAAGUAGUUUUUUUUU (SEQ ID NO:2732)	UGUUUUUCGAAUAACAGAUACAGUUUAAGAAUACAUA AGAAUGAUACAUCACUAAAAAAAAGCCUUUAUGCCGUA CUACUACUUUUUUCAAAAUAAGUAGUUUUUUUUU (SEQ ID NO:2732)	GUUUUUGGUGUAGUAUCAUUU UAUGUAUUUUAAAAAC (SEQ ID NO:2800)	64	110	36
Mycoplasma ovipneumoniae SC01	36	CUAGUAAAGAAUUUGCGCACAAAAAUAAGACGCAUUUAU GCUGUCGAAUUUUUUUUUUUUUUUUUUUUUUUUUUUU (SEQ ID NO:2733)	CUAGUAAAGAAUUUGCGCACAAAAAUAAGACGCAUUUAU GCUGUCGAAUUUUUUUUUUUUUUUUUUUUUUUUUUUU (SEQ ID NO:2733)	GUUUUUGGUGUAGUACAAUUUC UUACUAGUAAAAAC (SEQ ID NO:2801)	>14	72	36



Mycoplasma galliseptic um str. F	37	AUUUUUUGCUUACACAAUUUUUUUGUGCGUAUUUUUAAGG CGCUGUUUUAUGCAGCUGCCCGCAUCCGCCAGCAUUUA UGCUCUGGCUUUUUUUU (SEQ ID NO:2734)	GUUUUAGCACUGUACAAUACU UGUGUUAAGCAAUAAAC (SEQ ID NO:2802)	40	92	36
Mycoplasma synoviae 53	38	UAUAUAUAUAUAUAUAUAUAUAUAUAUAUAUAUAUAUA AAAAUAAGGGCGCUUUAUGUAAGAUGCAAUAUAUGCACUUA UAUAAGCUGCCGUAACGCCGAGGUAACUCGGUUUUUUU U (SEQ ID NO:2735)	GUUUUUGGGUUUGUACAAUUAU UUUGUUAAGUAAAAC (SEQ ID NO:2803)	12	115	36
Mycoplasma canis PG 14	39	AGUACAAAUUAUAUAUAUAUAUAUAUAUAUAUAUAUA CCUAAAUAAGGGCGCUUUAUAUAUAUAUAUAUAUAUAUA AAGCGUAGCUUUUCAGUUUUUUUU (SEQ ID NO:2736)	GUUUUAGUGUUGUACAAUUAU UGGUAACCAAUAUAAAC (SEQ ID NO:2804)	11	98	36
Waliniella succioenes DSM 1740	40	CGUCGUCGUCGCGGAAAUUGGUCGAGUAGGCAGCGGCU AUAAAUAAGGGGUGUGGAGGCAUCCUGCGAAUUAUAUA CUACGGAGUAUUAUAUAUAUAUAUAUAUAUAUAUAUAUA	GUAUAGCCGCCUACUCAGGCC AUCCUCGUUAUAU (SEQ ID NO:2805)	10	92	36
Campylobact er jejuni subsp. jejuni NCTC 1168	41	AAGAAAAUUUUUUUUUUUUUUUUUUUUUUUUUUUUUU GACUCUGCGGGUUUACAAUCCCUUUUUUUUUUUUUUUUU (SEQ ID NO:2738)	GUUUUAGUCCUUUUUUUUUUUU UCUUUAUGGUAAAUAU (SEQ ID NO:2806)	5	73	36
Hellobacter mustelae 12198	42	none	GUUUUAGCCACUUCUAAUAUA UGUUUAUGCUAAAUAU (SEQ ID NO:2807)	11	-	36
Methylosinu s trichospori um OB3b	43	UAUGGAAAUUCGGAAGGGAAGCCACGCCAAGGUGGUUU CAUAGAAAUAUACUGAAGGAUUAUCCUUCUGUACAGAAA GUGGGGGGGGAUUUUUAUAUAUAUAUAUAUAUAUAUAUA	GCCGUGGCUUCCUGCCGGAU UCCUGUGGUAGGCU (SEQ ID NO:2808)	24	96	36

Ilyobacter polytopus DSM 2926	44	none	GUUGUACUCCCUAAUUUUU UAGCUAUGUUACAAU (SEQ ID NO:2809)	23	-	36
Bacillus smithii 7_3_47FAA	45	UAAGAUAUAUCACAGCAAUGAUUUAGGGUUACUAUG AUAAGGGCUUUCUACUUUAGGGUAGAGAUGUCCCGCG GCGUUGGGGAUCGCCUAUUUGCCUUUAAAAGGCACUCCC CAUUUUAAAAUU (SEQ ID NO:2740)	GUCAUAGUCCCUAAGAUUA UUGCUGUAUAGAU (SEQ ID NO:2810)	28	125	36
Clostridium perfringens D str. JGS1721	45	UUAUCAGGAACUAGGUUAGCAUUCGAGAGUUUAAC UAGUUACUUAACAAGCAUUAAGCCGUAAAGUAUCCC CUAUGUUCAUUUGAACCUAAGGGUAUCUUUU (SEQ ID NO:2741)	GUUAUAGUCCCUAAGAUUAUC UCGAUAGCUAUAUU (SEQ ID NO:2811)	27	107	36
Clostridium cellulolyticum H10	46	AUGGCAUUCGGAGCCUGAAUUUUGCUAUAUAAGGU GCUGGGUUUAGCCAGACCCCGUCCGUAACCCCGGCAU UUUUUUGCUGGGUAUCUUUUUUU (SEQ ID NO:2742)	GUUAUAGCUCCAAUUCAGGCU CCGAUAGCUAUAUU (SEQ ID NO:2812)	9	100	36
Acidovorax ebreus TPSY	47	CGAUUGGUUUUCCGGGGUGAGAGCCGUUGCUGCAAU AAGGAGGGGUCGAAAGACCCCGUCCGUAACCCAAAAGCC UGGCAGGGAACCCUGUCAGGCCUUUUUU (SEQ ID NO:2743)	GUUGUAGCUCCCUUCUCACC CCGGAUAGCUACACU (SEQ ID NO:2813)	15	103	36
Neisseria meningitidis Z2491	48	ACAUUUUGCGCACUGCGAAAUGAGAACCGUUGCUACA AUAAGCCGUCUGAAAAGAUUGCCGCAACGCUCUGCC CCUUAAAAGCUUCUGCUUUUAAAGGGGCAUCGUUUUUU (SEQ ID NO:2744)	GUUGUAGCUCCCUUCUCAUU UCCGAGUCUACAAU (SEQ ID NO:2814)	17	111	36
Pasteurella multocida str. Pm70	49	GCAUUVUUGCAGCUGCGAAAUGAGAGACGGUUGCUACA AUAAGCCUUCUGAAAAGAUACCCGUAACGCUCUGCCC CUUGUAUUCUUUAAUUGCAAGGGGCAUCGUUUUUU (SEQ ID NO:2745)	GUUGUAGUCCCUUCUCAUU UCCGAGUCUACAAU (SEQ ID NO:2815)	6	110	36

Aminomonas paucivorans DSM 12260	50	none		GUCAUAGCUCUCCGCGCACU CCGAAAUGCUAUGCU (SEQ ID NO:2816)	7	-	36
Roseburia intestinalis L1-82	52	CUAAGAGAAUUUAUCAUACCAAGUGAUAAUUAGGUUA UUACAAUAGGUAAAGAAACCUAAAAGCUCUAAUCCCAU UCUUCGGAAUGGGAAUUAUCUUUU (SEQ ID NO:2746)		GUUGUAAUUCUCCUGUUAUCAC UUGGUUAGGUAAU (SEQ ID NO:2817)	62	99	36
Lactobacillus corniformis subsp. Torquens KCTC 3535	51	No contig information			-	-	-
Alicyclobacillus hesperidum URH17-3-68	53	GCGAGGGAUUAUCAUACCAUACCAAGGCUUGCGAGGUUG CUAUGAUAGGCAACAGGCCGAAAGCACUGACCCGCA UUCCAAUGAAUGCGGGUCAUCUACUUUUU (SEQ ID NO:2747)		GUCAUAGUUCUCCUACAAGCC UCGAUGGUUAGAU (SEQ ID NO:2818)	54	105	36
Roseburia inulinivora DSM 16841	52	None			-	-	-
Uncult. delta proteobact. HF0070_07E19	54	none		GUCCUAGUUUCCCUCCAAUC AAAGCCUGCUACACU (SEQ ID NO:2819)	8	-	36
Caenispiril	58	AUCACAGGGUGCCAUUACCAAGAUUGGAGCAGGGUGG		GUCCUGUAGCCCGGUCCGUUC	11	-	36

lum salinarum AK4	AACGGACCGGCACCUACAGGACAAAGUGAUCAUACACGU GACAGCCGCCUCCCCCGCCAGUGGCCAAAGGGAGGC GGCUUUUCU (SEQ ID NO:2748)	CACCGUGCUAGCUUC (SEQ ID NO:2820)			
Ruminococcus albus 8	Too short contig		-	-	-
Trepanema sp. JC4	Too short cotig		-	-	-
Alcanivorax sp. W11-5	none		-	-	-
Rhodospirillum rubrum ATCC 11170	none	GUUCCAUGGCCCGUCCCACA CCGCCAUGGUAGAGU (SEQ ID NO:2821)	8	-	36
Ralstonia syzygii R24	GACUUUCCAGCAGACUGGGAAUUGCGCUUUUGCUACUAA CAAGCUGAAUCCGUUAGGAGUAAUAGCACCAAUAGAGA GGGCCGGCUUUUUGCGGCCCUUUGCUUUU (SEQ ID NO:2749)	GUUGUAGCCAGAGCGCAAUUC CCGAUCUGCUAACCUC (SEQ ID NO:2822)	35	105	36
Rhodovulum sp. PH10	CGUCUAGCAAGGAACGGCGGUGGCCUCUCCGUUAAACA AGGCACAUGCCACCAGAUUCGAGCGGGUCUCCGGUCCCGC CUCUUUGCUUU (SEQ ID NO:2750)	GUUGCGGUUGGCCCGGCCGC GUUCCCUUGCUVAGACC (SEQ ID NO:2823)	12	87	36
Alicyclophilus dentrificans K601	CACUCAGUUCACUGGGAUUGCGUCUGACCGCUAACA AGCUGAAAAGAUAGCACCAAUUGGAAAGCCCGCAUGCGG GGCUUUCGUCUUU (SEQ ID NO:2751)	GUUCCGGCCAGUGCGCAUAUC CCAGUGAUCUAGAUAU (SEQ ID NO:2824)	75	90	36
Cand. Puniceispirillum	GCCAUUAAUUAUUGAUAUUGCAAUAACACUCUGGUGAUUG AGGAGCCUAUGGUUAACAAGUGGGUUUCCUGCACAAAUA CUAAGAGCUGCCUCCGGGGCCUCUUUUGCUUU (SEQ	GUUGCUCUAGGCUCUCAAUCA CCAGAGUGCUAUAUCU (SEQ	27	109	36

	ID NO:2752)	ID NO:2825)				
marinum IMCC1322						
64 Azospirillum m sp. B510	UGGAAAUUCGAUGGGGAUCGGGGUCCAGCCGUUAAACA UGUCCCUUCGGGGAGCACGAAAUUGCGGGGGGGCCAC GGUCCGCCCUUUUUUU (SEQ ID NO:2753)	GUUGCGGCUGGACCCCGAUC CCCAUCGGCUACACU (SEQ ID NO:2826)	25	93	36	
67 Dinoroseobacter shibae DFL 12	GUUCAGAAUUCGGGUCGAGCCGUUAAACAAGCUCGAAG AAGCACCAUAUAAAACGCGUCGCGGGGGCGGUUUU CUUUUUU (SEQ ID NO:2754)	GUUGCGGCUGGACCCCGAUAU CUGAACAGCUAAAACU (SEQ ID NO:2827)	19		36	
65 Nitrobacter hamburgensis X14	None		-	-	-	
66 Bradyrhizobium sp. BTai1	none		-	-	-	
68 Parvibaculum lavamentivans DS-1	UAGCAAUUCGAGAGGGGUCGUUUUCGCAAGCAAUU GACCCUUGUGCGGGCUCGGCAUCCCAAGGUCAGCUGC CGUUUAUUUCGAAAGGCCACCGCAAGCAGCGCGGUG GGCCUUUUUUU (SEQ ID NO:2755)	GCUGCGGAUUGCGGGCUCUC UCGAUUUGCUACUCU (SEQ ID NO:2828)	49	125	36	
70 Bacteroides sp. 20_3	None	GUUGUAUUUUUUUUUUUUUU AGUAUUUUUGAUCCAUUGGAA ACAGC (SEQ ID NO:2829)	10	-	47	
69 Bergeyella zoohelcum ATCC 43767	Too short contig		-	-	-	

Ignavibacterium album JCM 16511	71	UUUCUGUCCCAUUUGUGAUUUUGCUUUUUGCACAGCAU CCUUUGGACAACUUGUUCUUUGAGGGAUUUAAAACCAA CCUAUCUGUUUAAAGAUAGUCAAUUCUUUUU (SEQ ID NO:2756)	GUUGUUUUAAUCCUAAAGA ACAAGUUGAAAGCAAUCACA AC (SEQ ID NO:2830)	9	107	45
Bacteroides fragilis NCTC 9343	72	none	GUUGUGAUUUUGCUUUUCAAUU AGUAUCUUUGAACCAUUGGAA ACAGCG (SEQ ID NO:2831)	28	4	48
Barnesiella interinhomina YIT 11860	74	UUAAUGUGUAAAUAUAAAUAUUUUCGAAAAUAACA AUAGUAUUAAAAUUUAUUAUUAUUUGUCAACACAA AUUUGAAAGCAAUCACAAUAGGAUUUCCGUUGUG AAAACAUUUGGAGGGGAGUAUUUAUACUCCUCGUUC UUUUUU (SEQ ID NO:2757)	GUUGUGAUUCGCUUUCAAUU UGUAUCUUUGACAUUUAAUU ACAGC (SEQ ID NO:2832)	>2	158	47
Porphyromonas sp. Oral taxon 279 str. F0450	73	none		-	-	-
Odoribacter laneus YIT 12061	75	CGUUGAUUAAACAAUCAAUUUUUACAUCUUUCACAG CAAGCCUAUUGCCGAGGAUGUAUCCUAUACUCCCG CUUCGGGGGAGUUUUU (SEQ ID NO:2758)	GCUGUGAUUUUGAUUAAUAC UUGAUUAGAUUACC (SEQ ID NO:2833)	>3	94	36
Flavobacterium branchiophilum FL-15	76	AUGUUUUUAUUUUUGCAGCAUGAUUAAUUUUUUAU CUUUAAUCUUUACACAAUAGCCUAUUGCCGUAGAUG AAAAUCUUUAGUCCUUCGGUGGGACUUUUUUUU (SEQ ID NO:2759)	GUUGUGUUUUUAAAGAUU AGAAAACACGAUUG (SEQ ID NO:2834)	29	112	36
Prevotella sp. C561	77	GUUUUUUUUACAGAAAUAAGUUAUUUUUGCACUCA GAUACACAGUGAAGACUUUUUCACAACAAGCCUAUAAAGC CGAAGAUUUUCUUUGUACCCUCCGGGUCACACACAGGGUC	GUUGUACGUGCUAAUUGCAAAG AUACACAUUUUGAAAGCAAUC ACAAC (SEQ ID	>3	121	47

	UUUUUUU (SEQ ID NO:2760)							
Prevotella timonensis CRIS 5C-B1	None							36
Elusimicrobium minutum Pei191	none							36
Sphaerochaeta globosa str. Buddy	GUUAUAUCUUAAACAAAAACCAGGAUUAUCUCUAAUAA GACUUAAGUCGCAAAAUGCCUUAUUUUGGGAGCUUU UUUU (SEQ ID NO:2761)						80	36
Acidothermus cellulolyticus 11B	GAGACAGGCUACCUAGCAAGACCCUUCGUGGGUCGC AUUCUUACCCUUCGAGCAGCGAGGGGUUCGUUU (SEQ ID NO:2762)						75	36
Actinomyces sp. Oral taxon 180 str. F0310	None							36
Bifidobacterium longum DJ010A	none							36
Akkermansia muciniphila ATCC BAA-835	ACAAAACAUCUGAAACAUCACUUUAACUCCCAACGGAAU CAAGACAAAUUUGAAAUGCAAAACCGAUUUUCCUGACU GCCAGCCAGUCACACCGGUAAACAAAAGCAUUUU (SEQ ID NO:2763)						109	36

<p>Nitratifrac tor salsuginis DSM 16511</p>	<p>86</p>	<p>GUUGUAAACAGGGUAGGGUUUUUUUAGGGGUCUUAAAAU CAAGAACUGUUACAACAGUUCCAUUCUAGGGCCCAUCU UCGGACGGGCCUCAGCCUUUUUUU (SEQ ID NO: 2764)</p>	<p>GUUUUAAGACCCCUCAAACC CCACCCUGUUACAAU (SEQ ID NO: 2843)</p>	<p>12</p>	<p>100</p>	<p>36</p>
---	-----------	--	---	-----------	------------	-----------

<sup>a</sup> The position of the strain on the Cas9 tree is given. Color shading corresponds to the color branch of the tree.

<sup>b</sup> Predicted or previously validated tracrRNA sequence is given. none, no tracrRNA was found; too short contig, the type II CRISPR-Cas locus is at the end of the genomic sequence contig and it was not possible to identify a tracrRNA ortholog; no contig information, genomic sequence contig encoding a type II CRISPR-Cas locus was not available.

<sup>c</sup> Predicted or previously validated CRISPR repeat sequence is given. none, no repeat-spacer array was found; too short contig, the type II CRISPR-Cas locus is at the end of the genomic sequence contig and it was not possible to identify a repeat-spacer array; no contig information, genomic sequence contig encoding a type II CRISPR-Cas locus was not available.

<sup>d</sup> Amount of the CRISPR repeats of the repeat-spacer array is given. Values preceded by ">" indicate a minimal amount of repeats in the array given that the array is at the end of the genomic sequence contig.

<sup>e</sup> The length of the CRISPR repeats is given. Values higher than the typical 36 nt are highlighted.



Supplemental Table S6

Strain	Cas9 GI	Cluster
<i>Acidaminococcus intestini</i> RYC-MR95	352684361	1
<i>Acidaminococcus</i> sp. D21	227824983	1
<i>Anaerococcus tetradius</i> ATCC 35098	227501312	1
<i>Bifidobacterium bifidum</i> S17	310286728	1
<i>Catenibacterium mitsuokai</i> DSM 15897	224543312	1
<i>Coprococcus catus</i> GD/7	291520705	1
<i>Coriobacterium glomerans</i> PW2	328956315	1
<i>Dolosigranulum pigrum</i> ATCC 51524	375088882	1
<i>Dorea longicatena</i> DSM 13814	153855454	1
<i>Eggerthella</i> sp. YY7918	339445983	1
<i>Enterococcus faecalis</i> ATCC 29200	229548613	1
<i>Enterococcus faecalis</i> ATCC 4200	256617555	1
<i>Enterococcus faecalis</i> D6	257086028	1
<i>Enterococcus faecalis</i> E1Sol	257080914	1
<i>Enterococcus faecalis</i> OG1RF	384512368	1
<i>Enterococcus faecalis</i> TX0470	312900261	1
<i>Enterococcus faecalis</i> TX4244	422695652	1
<i>Enterococcus faecium</i> 1,141,733	257888853	1
<i>Enterococcus faecium</i> 1,231,408	257893735	1
<i>Enterococcus faecium</i> E1133	430847551	1
<i>Enterococcus faecium</i> E3083	431757680	1
<i>Enterococcus faecium</i> PC4.1	293379700	1
<i>Enterococcus faecium</i> TX1330	227550972	1
<i>Enterococcus faecium</i> TX1337RF	424765774	1
<i>Enterococcus hirae</i> ATCC 9790	392988474	1
<i>Enterococcus italicus</i> DSM 15952	315641599	1
<i>Eubacterium</i> sp. ASI5	402309258	1
<i>Eubacterium yurii</i> subsp. <i>margaretiae</i> ATCC 43715	306821691	1
<i>Filifactor aloisii</i> ATCC 35896	374307738	1
<i>Finegoldia magna</i> ACS-171-V-Col3	302380288	1
<i>Finegoldia magna</i> ATCC 29328	169823755	1
<i>Finegoldia magna</i> SY403409CC001050417	417926052	1
<i>Fructobacillus fructosus</i> KCTC 3544	339625081	1
<i>Fusobacterium nucleatum</i> subsp. <i>vincentii</i> ATCC 49256	34762592	1
<i>Fusobacterium</i> sp. 1_1_41FAA	294782278	1

<i>Fusobacterium</i> sp. 3_1_27	294785695	1
<i>Fusobacterium</i> sp. 3_1_36A2	256845019	1
<i>Gemella haemolysons</i> ATCC 10379	241889924	1
<i>Gemella morbillorum</i> M424	317495358	1
<i>Gordonibacter pamelaiae</i> 7-10-1-b	295106015	1
<i>Helcococcus kunzii</i> ATCC 51366	375092427	1
<i>Lactobacillus animalis</i> KCTC 3501	335357451	1
<i>Lactobacillus brevis</i> subsp. <i>gravesensis</i> ATCC 27305	227509761	1
<i>Lactobacillus buchneri</i> CD034	406027703	1
<i>Lactobacillus buchneri</i> NRRL B-30929	331702228	1
<i>Lactobacillus casei</i> BL23	191639137	1
<i>Lactobacillus casei</i> Lc-10	418010298	1
<i>Lactobacillus casei</i> M36	417996992	1
<i>Lactobacillus casei</i> str. Zhang	301067199	1
<i>Lactobacillus casei</i> T71499	417999832	1
<i>Lactobacillus casei</i> UCD174	418002962	1
<i>Lactobacillus casei</i> W56	409997999	1
<i>Lactobacillus coryniformis</i> subsp. <i>coryniformis</i> KCTC 3167	333394446	1
<i>Lactobacillus curvatus</i> CRL 705	354808135	1
<i>Lactobacillus farciminis</i> KCTC 3681	336394882	1
<i>Lactobacillus fermentum</i> 28-3-CHN	260662220	1
<i>Lactobacillus fermentum</i> ATCC 14931	227514633	1
<i>Lactobacillus florum</i> 2F	408790128	1
<i>Lactobacillus gasseri</i> JV-V03	300361537	1
<i>Lactobacillus hominis</i> CRBIP 24.179	395244248	1
<i>Lactobacillus iners</i> LactinV 11V1-d	309803917	1
<i>Lactobacillus jensenii</i> 269-3	238854567	1
<i>Lactobacillus jensenii</i> 27-2-CHN	256852176	1
<i>Lactobacillus johnsonii</i> DPC 6026	385826041	1
<i>Lactobacillus mucosae</i> LM1	377831443	1
<i>Lactobacillus paracasei</i> subsp. <i>paracasei</i> 8700:2	239630053	1
<i>Lactobacillus pentosus</i> IG1	339637353	1
<i>Lactobacillus pentosus</i> KCA1	392947436	1
<i>Lactobacillus pentosus</i> MP-10	334881121	1
<i>Lactobacillus plantarum</i> Z1316	448819853	1
<i>Lactobacillus rhamnosus</i> GG	258509199	1
<i>Lactobacillus rhamnosus</i> HN001	199597394	1
<i>Lactobacillus rhamnosus</i> R0011	418072660	1
<i>Lactobacillus ruminis</i> ATCC 25644	323340068	1

<i>Lactobacillus salivarius</i> SMXD51	418960525	1
<i>Lactobacillus sanfranciscensis</i> TMW 1.1304	347534532	1
<i>Lactobacillus</i> sp. 66c	408410332	1
<i>Lactobacillus versmoldensis</i> KCTC 3814	365906066	1
<i>Leuconostoc gelidum</i> KCTC 3527	333398273	1
<i>Listeria innocua</i> ATCC 33091	423101383	1
<i>Listeria innocua</i> Clip11262	16801805	1
<i>Listeria innocua</i> FSL S4-378	422414122	1
<i>Listeria ivanovii</i> FSL F6-596	315305353	1
<i>Listeria monocytogenes</i> 10403S	386044902	1
<i>Listeria monocytogenes</i> FSL J1-175	255520581	1
<i>Listeria monocytogenes</i> FSL J1-194	254825045	1
<i>Listeria monocytogenes</i> FSL J1-208	422810631	1
<i>Listeria monocytogenes</i> FSL N3-165	254829042	1
<i>Listeria monocytogenes</i> FSL R2-503	254854201	1
<i>Listeria monocytogenes</i> str. 1/2a F6854	47097148	1
<i>Megaspheara</i> sp. UPII 135-E	342218215	1
<i>Oenococcus kitaharae</i> DSM 17330	366983953	1
<i>Oenococcus kitaharae</i> DSM 17330	372325145	1
<i>Olsenella uli</i> DSM 7084	302336020	1
<i>Pediococcus acidilactici</i> DSM 20284	304386254	1
<i>Pediococcus acidilactici</i> MA18/5M	418068659	1
<i>Peptoniphilus duerdenii</i> ATCC BAA-1640	304438954	1
<i>Planococcus antarcticus</i> DSM 14505	389815359	1
<i>Psychroflexus torquis</i> ATCC 700755	408489713	1
<i>Ruminococcus lactaris</i> ATCC 29176	197301447	1
<i>Scardovia wiggisiae</i> F0424	423349694	1
<i>Solobacterium moorei</i> F0204	320528778	1
<i>Staphylococcus pseudintermedius</i> ED99	323463801	1
<i>Staphylococcus pseudintermedius</i> ED99	386318630	1
<i>Staphylococcus simulans</i> ACS-120-V-Sch1	414160476	1
<i>Streptococcus agalactiae</i> 2603V/R	22537057	1
<i>Streptococcus agalactiae</i> 515	77413160	1
<i>Streptococcus agalactiae</i> A909	76788458	1
<i>Streptococcus agalactiae</i> ATCC 13813	339301617	1
<i>Streptococcus agalactiae</i> CJB111	77411010	1
<i>Streptococcus agalactiae</i> COH1	77407964	1
<i>Streptococcus agalactiae</i> FSL S3-026	417005168	1
<i>Streptococcus agalactiae</i> GB00112	421147428	1

<i>Streptococcus agalactiae</i> H36B	77405721	1
<i>Streptococcus agalactiae</i> NEM316	25010965	1
<i>Streptococcus agalactiae</i> SA20-06	410594450	1
<i>Streptococcus agalactiae</i> STIR-CD-17	421532069	1
<i>Streptococcus anginosus</i> F0211	315223162	1
<i>Streptococcus anginosus</i> SK1138	421490579	1
<i>Streptococcus anginosus</i> SK52 = DSM 20563	335031483	1
<i>Streptococcus bovis</i> ATCC 700338	306833855	1
<i>Streptococcus canis</i> FSL Z3-227	392329410	1
<i>Streptococcus constellatus</i> subsp. <i>constellatus</i> SK53	418965022	1
<i>Streptococcus dysgalactiae</i> subsp. <i>equisimilis</i> AC-2713	410494913	1
<i>Streptococcus dysgalactiae</i> subsp. <i>equisimilis</i> ATCC 12394	386317166	1
<i>Streptococcus dysgalactiae</i> subsp. <i>equisimilis</i> GGS_124	251782637	1
<i>Streptococcus dysgalactiae</i> subsp. <i>equisimilis</i> RE378	408401787	1
<i>Streptococcus equi</i> subsp. <i>zooepidermicus</i> MGCS10565	195978435	1
<i>Streptococcus equinus</i> ATCC 9812	320547102	1
<i>Streptococcus gallolyticus</i> subsp. <i>gallolyticus</i> ATCC BAA-2069	325978669	1
<i>Streptococcus gallolyticus</i> subsp. <i>gallolyticus</i> TX20005	306831733	1
<i>Streptococcus gallolyticus</i> UCN34	288905639	1
<i>Streptococcus infantarius</i> subsp. <i>infantarius</i> CJ18	379705580	1
<i>Streptococcus iniae</i> 9117	406658208	1
<i>Streptococcus macacae</i> NCTC 11558	357636406	1
<i>Streptococcus mitis</i> SK321	307710946	1
<i>Streptococcus mutans</i> 11SSST2	449165720	1
<i>Streptococcus mutans</i> 11SSST2	449951835	1
<i>Streptococcus mutans</i> 11VS1	449976542	1
<i>Streptococcus mutans</i> 14D	450149988	1
<i>Streptococcus mutans</i> 15VF2	449170557	1
<i>Streptococcus mutans</i> 15VF2	449965974	1
<i>Streptococcus mutans</i> 1SM1	449158457	1
<i>Streptococcus mutans</i> 1SM1	449920643	1
<i>Streptococcus mutans</i> 24	449247589	1
<i>Streptococcus mutans</i> 24	450180942	1
<i>Streptococcus mutans</i> 2VS1	449174812	1
<i>Streptococcus mutans</i> 2VS1	449968746	1
<i>Streptococcus mutans</i> 3SN1	449162653	1
<i>Streptococcus mutans</i> 3SN1	449931425	1
<i>Streptococcus mutans</i> 4SM1	449159838	1
<i>Streptococcus mutans</i> 4SM1	449927152	1

<i>Streptococcus mutans</i> 4VF1	449167132	1
<i>Streptococcus mutans</i> 4VF1	449961027	1
<i>Streptococcus mutans</i> 5SM3	449176693	1
<i>Streptococcus mutans</i> 5SM3	449980571	1
<i>Streptococcus mutans</i> 66-2A	449240165	1
<i>Streptococcus mutans</i> 66-2A	450160342	1
<i>Streptococcus mutans</i> 8ID3	449154769	1
<i>Streptococcus mutans</i> 8ID3	449872064	1
<i>Streptococcus mutans</i> A19	449187668	1
<i>Streptococcus mutans</i> A19	450013175	1
<i>Streptococcus mutans</i> B	450166294	1
<i>Streptococcus mutans</i> GI23	450029806	1
<i>Streptococcus mutans</i> GS-5	397650022	1
<i>Streptococcus mutans</i> LJ23	387785882	1
<i>Streptococcus mutans</i> M21	449194333	1
<i>Streptococcus mutans</i> M21	450036249	1
<i>Streptococcus mutans</i> M230	449260994	1
<i>Streptococcus mutans</i> M230	449903532	1
<i>Streptococcus mutans</i> M2A	449209586	1
<i>Streptococcus mutans</i> M2A	450074072	1
<i>Streptococcus mutans</i> N29	449182997	1
<i>Streptococcus mutans</i> N29	450003067	1
<i>Streptococcus mutans</i> N3209	449210660	1
<i>Streptococcus mutans</i> N3209	450077860	1
<i>Streptococcus mutans</i> N66	449212466	1
<i>Streptococcus mutans</i> N66	450083993	1
<i>Streptococcus mutans</i> NFSM1	449202104	1
<i>Streptococcus mutans</i> NFSM1	450051112	1
<i>Streptococcus mutans</i> NLML1	450140393	1
<i>Streptococcus mutans</i> NLML4	449202681	1
<i>Streptococcus mutans</i> NLML4	450059882	1
<i>Streptococcus mutans</i> NLML9	449209148	1
<i>Streptococcus mutans</i> NLML9	450066176	1
<i>Streptococcus mutans</i> NMT4863	449186850	1
<i>Streptococcus mutans</i> NMT4863	450007078	1
<i>Streptococcus mutans</i> NN2025	290580220	1
<i>Streptococcus mutans</i> NV1996	450086338	1
<i>Streptococcus mutans</i> NVAB	449181424	1

<i>Streptococcus mutans</i> NVAB	449990810	1
<i>Streptococcus mutans</i> R221	449258042	1
<i>Streptococcus mutans</i> R221	449899675	1
<i>Streptococcus mutans</i> S1B	449251227	1
<i>Streptococcus mutans</i> S1B	449877120	1
<i>Streptococcus mutans</i> SF1	450098705	1
<i>Streptococcus mutans</i> SF14	449221374	1
<i>Streptococcus mutans</i> SF14	450107816	1
<i>Streptococcus mutans</i> SM1	449245264	1
<i>Streptococcus mutans</i> SM1	450176410	1
<i>Streptococcus mutans</i> SM4	449246010	1
<i>Streptococcus mutans</i> SM4	450170248	1
<i>Streptococcus mutans</i> SM6	449223000	1
<i>Streptococcus mutans</i> SM6	450112022	1
<i>Streptococcus mutans</i> ST6	449227252	1
<i>Streptococcus mutans</i> ST6	450123011	1
<i>Streptococcus mutans</i> UA159	24379809	1
<i>Streptococcus mutans</i> W6	450094364	1
<i>Streptococcus oralis</i> SK304	421488030	1
<i>Streptococcus aralis</i> SK610	419782534	1
<i>Streptococcus pseudoporcinus</i> LQ 940-04	416852857	1
<i>Streptococcus pyogenes</i> M1	13622193	1
<i>Streptococcus pyogenes</i> MGAS10750	94543903	1
<i>Streptococcus pyogenes</i> MGAS15252	94994317	1
<i>Streptococcus pyogenes</i> MGAS2096	383479946	1
<i>Streptococcus pyogenes</i> MGAS315	94992340	1
<i>Streptococcus pyogenes</i> MGAS5005	21910213	1
<i>Streptococcus pyogenes</i> MGAS6180	71910582	1
<i>Streptococcus pyogenes</i> MGAS9429	71903413	1
<i>Streptococcus pyogenes</i> NZ131	94988516	1
<i>Streptococcus pyogenes</i> SSI-1	28896088	1
<i>Streptococcus rattii</i> FA-1 = DSM 20564	400290495	1
<i>Streptococcus salivarius</i> K12	421452908	1
<i>Streptococcus sanguinis</i> SK115	422848603	1
<i>Streptococcus sanguinis</i> SK330	422860049	1
<i>Streptococcus sanguinis</i> SK353	422821159	1
<i>Streptococcus sanguinis</i> SK49	422884106	1
<i>Streptococcus</i> sp. C300	322375978	1

<i>Streptococcus</i> sp. F0441	414157437	1
<i>Streptococcus</i> sp. M334	322378004	1
<i>Streptococcus</i> sp. oral taxon 56 str. F0418	339640839	1
<i>Streptococcus</i> sp. oral taxon 71 str. 73H25AP	306826314	1
<i>Streptococcus suis</i> ST1	389856936	1
<i>Streptococcus thermophilus</i>	343794781	1
<i>Streptococcus thermophilus</i> LMD-9	116628213	1
<i>Streptococcus thermophilus</i> MN-ZLW-002	387910220	1
<i>Streptococcus thermophilus</i> ND03	386087120	1
<i>Treponema denticola</i> AL-2	449103686	1
<i>Treponema denticola</i> ASLM	449106292	1
<i>Treponema denticola</i> ATCC 35405	42525843	1
<i>Treponema denticola</i> H1-T	449118593	1
<i>Treponema denticola</i> H-22	449117322	1
<i>Treponema denticola</i> OTK	449125136	1
<i>Treponema denticola</i> SP37	449130155	1
<i>Veillonella atypica</i> ACS-134-V-Col7a	303229466	1
<i>Veillonella parvula</i> ATCC 17745	282849530	1
<i>Veillonella</i> sp. 6_1_27	294792465	1
<i>Veillonella</i> sp. oral taxon 780 str. F0422	342213964	1
<i>Streptococcus pyogenes</i> SF370 (M1 GAS)	209559356	1
<i>Streptococcus pyogenes</i> MGAS10270	56808315	1
<i>Acidovorax ebreus</i> TPSY	222109285	2
<i>Actinobacillus minor</i> NM305	240949037	2
<i>Actinobacillus pleuropneumoniae</i> serovar 10 str. D13039	307256472	2
<i>Actinobacillus succinogenes</i> 1302	152978060	2
<i>Actinobacillus suis</i> H91-0380	407692091	2
<i>Alicyclobacillus hesperidum</i> URH17-3-68	403744858	2
<i>Aminomonas paucivorans</i> DSM 12260	312879015	2
<i>Bacillus cereus</i> BAG4X12-1	423439645	2
<i>Bacillus cereus</i> BAG4X2-1	423445130	2
<i>Bacillus cereus</i> Rock1-15	229113166	2
<i>Bacillus smithii</i> 7_3_47FAA	365156657	2
<i>Bacillus thuringiensis</i> serovar finitimus YBT-020	384183447	2
<i>Bacteroides</i> sp. 3_1_33FAA	265750948	2
<i>Brevibacillus laterosporus</i> GI-9	421874297	2
<i>Campylobacter coli</i> 1098	419564797	2
<i>Campylobacter coli</i> 111-3	419536531	2

<i>Campylobacter coli</i> 132-6	419572019	2
<i>Campylobacter coli</i> 151-9	419603415	2
<i>Campylobacter coli</i> 1909	419576091	2
<i>Campylobacter coli</i> 1957	419581876	2
<i>Campylobacter coli</i> 2692	419553162	2
<i>Campylobacter coli</i> 59-2	419578074	2
<i>Campylobacter coli</i> 67-8	419587721	2
<i>Campylobacter coli</i> 80352	419559505	2
<i>Campylobacter coli</i> 80352	419558307	2
<i>Campylobacter jejuni</i> subsp. <i>doylei</i> 269.97	153952471	2
<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 110-21	419676124	2
<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 129-258	419619138	2
<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 1336	283956897	2
<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 140-16	419681578	2
<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 1577	419685099	2
<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 1854	419689467	2
<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 1997-10	419666522	2
<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 2008-1025	419650041	2
<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 2008-872	419654778	2
<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 2008-979	419660762	2
<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 2008-988	419655317	2
<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 2008-988	419656328	2
<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 260.94	86152042	2
<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 414	283953849	2
<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 51037	419674189	2
<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 51494	419619463	2
<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 53161	419647275	2
<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 60004	419629136	2
<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 81116	157415744	2
<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 84-25	88596565	2
<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> 87459	419680124	2
<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> ATCC 33560	419643715	2
<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> CF93-6	86149266	2
<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> CG8486	148925683	2
<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> HB93-13	86152450	2
<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> LMG 23210	419696801	2
<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> LMG 23211	419697443	2
<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> LMG 23263	419628620	2



<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> LMG 23264	419632476	2
<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> LMG 23269	419634246	2
<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> LMG 23357	419641132	2
<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> NCTC 11168	218563121	2
<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> NW	424845990	2
<i>Campylobacter jejuni</i> subsp. <i>jejuni</i> PT14	407942868	2
<i>Campylobacter lari</i>	345468028	2
<i>Clostridium cellulolyticum</i> H10	220930482	2
<i>Clostridium perfringens</i> C str. JGS1495	169343975	2
<i>Clostridium perfringens</i> D str. JGS1721	182624245	2
<i>Haemophilus parainfluenzae</i> ATCC 33392	325578067	2
<i>Haemophilus parainfluenzae</i> CCUG 13788	359298684	2
<i>Haemophilus parainfluenzae</i> T3T1	345430422	2
<i>Haemophilus sputorum</i> HK 2154	402304649	2
<i>Helicobacter canadensis</i> MIT 98-5491	253828136	2
<i>Helicobacter cinaedi</i> ATCC BAA-847	396079277	2
<i>Helicobacter cinaedi</i> CCUG 18818	313144862	2
<i>Helicobacter cinaedi</i> PAGU611	386762035	2
<i>Helicobacter mustelae</i> 12198	291276265	2
<i>Ilyobacter polytropus</i> DSM 2926	310780384	2
<i>Kingella kingae</i> PYKK081	381401699	2
<i>Lactobacillus coryniformis</i> subsp. <i>torquens</i> KCTC 3535	336393381	2
<i>Neisseria bacilliformis</i> ATCC BAA-1200	329117879	2
<i>Neisseria cinerea</i> ATCC 14685	261378287	2
<i>Neisseria flavescens</i> SK114	241759613	2
<i>Neisseria lactamica</i> 020-06	313669044	2
<i>Neisseria meningitidis</i> 053442	161869390	2
<i>Neisseria meningitidis</i> 2007056	433531983	2
<i>Neisseria meningitidis</i> 63049	433514137	2
<i>Neisseria meningitidis</i> 8013	385324780	2
<i>Neisseria meningitidis</i> 92045	421559784	2
<i>Neisseria meningitidis</i> 93003	421538794	2
<i>Neisseria meningitidis</i> 93004	421541126	2
<i>Neisseria meningitidis</i> 96023	433518260	2
<i>Neisseria meningitidis</i> 98008	421555531	2
<i>Neisseria meningitidis</i> alpha14	254804356	2
<i>Neisseria meningitidis</i> alpha275	254672046	2
<i>Neisseria meningitidis</i> ATCC 13091	304388355	2

<i>Neisseria meningitidis</i> N1568	416164244	2
<i>Neisseria meningitidis</i> NM140	421545139	2
<i>Neisseria meningitidis</i> NM220	418291220	2
<i>Neisseria meningitidis</i> NM233	418288950	2
<i>Neisseria meningitidis</i> WUE 2594	385337435	2
<i>Neisseria meningitidis</i> Z2491	218767588	2
<i>Neisseria</i> sp. oral taxon 14 str. F0314	298369677	2
<i>Neisseria wadsworthii</i> 9715	350570326	2
<i>Pasteurella multocida</i> subsp. gallicida X73	425063822	2
<i>Pasteurella multocida</i> subsp. multocida str. P52VAC	421263876	2
<i>Pasteurella multocida</i> subsp. multocida str. Pm70	15602992	2
<i>Phascolarctobacterium succinatutens</i> YIT 12067	323142435	2
<i>Roseburia intestinalis</i> L1-82	257413184	2
<i>Roseburia intestinalis</i> M50/1	291537230	2
<i>Roseburia inulinivorans</i> DSM 16841	225377804	2
<i>Simonsiella muelleri</i> ATCC 29453	404379108	2
<i>Sporolactobacillus vineae</i> DSM 21990 = SL153	404330915	2
<i>Subdoligranulum</i> sp. 4_3_54A2FAA	365132400	2
<i>Wolinella succinogenes</i> DSM 1740	34557790	2
<i>Catellacoccus marimammalium</i> M35/04/3	424780480	3
<i>Clostridium spiroforme</i> DSM 1552	169349750	3
<i>Enterococcus faecalis</i> Fly1	257084992	3
<i>Enterococcus faecalis</i> R508	424761124	3
<i>Enterococcus faecalis</i> T11	257419486	3
<i>Enterococcus faecalis</i> TX0012	315149830	3
<i>Enterococcus faecalis</i> TX0012	422729710	3
<i>Enterococcus faecalis</i> TX1342	422701955	3
<i>Eubacterium dolichum</i> DSM 3991	160915782	3
<i>Eubacterium rectale</i> ATCC 33656	238924075	3
<i>Eubacterium ventriosum</i> ATCC 27560	154482474	3
<i>Facklamia hominis</i> CCUG 36813	406671118	3
<i>Lactobacillus farciminis</i> KCTC 3681	336394701	3
<i>Listeriaceae bacterium</i> TTU M1-001	381184145	3
<i>Staphylococcus aureus</i> subsp. aureus	403411236	3
<i>Staphylococcus lugdunensis</i> M23590	315659848	3
<i>Streptococcus anginosus</i> 1_2_62CV	319939170	3
<i>Streptococcus gallolyticus</i> UCN34	288905632	3
<i>Streptococcus gordonii</i> str. Challis substr. CH1	157150687	3

<i>Streptococcus infantarius</i> ATCC BAA-102	3	171779984
<i>Streptococcus macedonicus</i> ACA-DC 198	3	374338350
<i>Streptococcus mitis</i> ATCC 6249	3	306829274
<i>Streptococcus mutans</i> NLM15	3	449203378
<i>Streptococcus mutans</i> NLM15	3	450064617
<i>Streptococcus mutans</i> NLM18	3	449151037
<i>Streptococcus mutans</i> NLM18	3	450133520
<i>Streptococcus mutans</i> ST1	3	449228751
<i>Streptococcus mutans</i> ST1	3	450114718
<i>Streptococcus mutans</i> U2A	3	449232458
<i>Streptococcus mutans</i> U2A	3	450125471
<i>Streptococcus oralis</i> SK1074	3	418974877
<i>Streptococcus oralis</i> SK313	3	417940002
<i>Streptococcus parasanguinis</i> F0449	3	419799964
<i>Streptococcus pasteurianus</i> ATCC 43144	3	336064611
<i>Streptococcus salivarius</i> JIM8777	3	387783792
<i>Streptococcus salivarius</i> PS4	3	419707401
<i>Streptococcus</i> sp. BS35b	3	401684660
<i>Streptococcus</i> sp. C150	3	322372617
<i>Streptococcus</i> sp. GMD6S	3	406576934
<i>Streptococcus suis</i> 89/1591	3	223932525
<i>Streptococcus suis</i> D9	3	386584496
<i>Streptococcus suis</i> ST3	3	330833104
<i>Streptococcus thermophilus</i> CNRZ1066	3	55822627
<i>Streptococcus thermophilus</i> JIM 8232	3	386344353
<i>Streptococcus thermophilus</i> LMD-9	3	116627542
<i>Streptococcus thermophilus</i> LMG 18311	3	55820735
<i>Streptococcus thermophilus</i> MN-ZLW-002	3	387909441
<i>Streptococcus thermophilus</i> MTCC 5460	3	445374534
<i>Streptococcus thermophilus</i> ND03	3	386086348
<i>Streptococcus vestibularis</i> ATCC 49124	3	322517104
<i>Anaerophaga</i> sp. HS1	4	371776944
<i>Anaerophaga thermohalophila</i> DSM 12881	4	346224232
<i>Bacteroides coprophilus</i> DSM 18228	4	224026357
<i>Bacteroides coprosuis</i> DSM 18011	4	333031006
<i>Bacteroides dorei</i> DSM 17855	4	212694363
<i>Bacteroides eggerthii</i> 1_2_48FAA	4	317474201
<i>Bacteroides faecis</i> 27-5	4	380696107

<i>Bacteroides fluxus</i> YIT 12057	4	329965125
<i>Bacteroides nordii</i> CLO2T12C05	4	393788929
<i>Bacteroides</i> sp. 20_3	4	301311869
<i>Bacteroides</i> sp. D2	4	383115507
<i>Bacteroides uniformis</i> CLO3T00C23	4	423303159
<i>Bacteroides vulgatus</i> CLO9T03C04	4	423312075
<i>Bergeyella zoohelcum</i> ATCC 43767	4	423317190
<i>Capnocytophaga gingivalis</i> ATCC 33624	4	228473057
<i>Capnocytophaga</i> sp. CM59	4	402830627
<i>Capnocytophaga</i> sp. oral taxon 324 str. F0483	4	429756885
<i>Capnocytophaga</i> sp. oral taxon 326 str. F0382	4	429752492
<i>Capnocytophaga</i> sp. oral taxon 412 str. F0487	4	393778597
<i>Chryseobacterium</i> sp. CF314	4	399023756
<i>Fibrobacter succinogenes</i> subsp. <i>succinogenes</i> S85	4	261414553
<i>Flavobacteriaceae bacterium</i> S85	4	372210605
<i>Flavobacterium columnare</i> ATCC 49512	4	365960762
<i>Fluviicola taffensis</i> DSM 16823	4	327405121
<i>Ignavibacterium album</i> JCM 16511	4	385811609
<i>Mucilaginibacter paludis</i> DSM 18603	4	373954054
<i>Myroides odoratus</i> DSM 2801	4	374597806
<i>Ornithobacterium rhinotracheale</i> DSM 15997	4	392391493
<i>Prevotella bivia</i> JCVIHMPO10	4	282858617
<i>Prevotella buccae</i> ATCC 33574	4	315607525
<i>Prevotella nigrescens</i> ATCC 33563	4	340351024
<i>Prevotella</i> sp. MSX73	4	402307189
<i>Prevotella timonensis</i> CRIS 5C-B1	4	282881485
<i>Prevotella veroralis</i> F0319	4	260592128
<i>Sphingobacterium spiritivorum</i> ATCC 33861	4	300771242
<i>Weeksella virosa</i> DSM 16922	4	325955459
<i>Acidovorax avenae</i> subsp. <i>avenae</i> ATCC 19860	5	326315085
<i>Alicyciphilus denitrificans</i> BC	5	319760940
<i>Alicyciphilus denitrificans</i> K601	5	330822845
<i>Azospirillum</i> sp. B510	5	288957741
<i>Bradyrhizabium</i> sp. BTAi1	5	148255343
<i>Candidatus Puniceispirillum marinum</i> IMCC1322	5	294086111
<i>Dinoroseobacter shibae</i> DFL 12	5	159042956
gamma proteobacterium HdN1	5	304313029

<i>Nitrobacter hamburgensis</i> X14	92109262	5
<i>Nitrosomonas</i> sp. AL212	325983496	5
<i>Ralstonia syzygii</i> R24	344171927	5
<i>Rhodovulum</i> sp. PH10	402849997	5
<i>Sphingobium</i> sp. AP49	398385143	5
<i>Sphingomonas</i> sp. S17	332188827	5
<i>Verminephrobacter eiseniae</i> EF01-2	121608211	5
<i>Bacteroides fragilis</i> 638R	375360193	6
<i>Bacteroides fragilis</i> NCTC 9343	60683389	6
<i>Bacteroides</i> sp. 2_1_16	265767599	6
<i>Bacteroides</i> sp. 3_1_19	298377533	6
<i>Bacteroides</i> sp. D2	383110723	6
<i>Bacteroidetes</i> oral taxon 274 str. F0058	298373376	6
<i>Barnesiella intestinihominis</i> YIT 11860	404487228	6
<i>Belliella baltica</i> DSM 15883	390944707	6
<i>Bergeyella zoohelcum</i> CCUG 30536	406673990	6
<i>Capnocytophaga canimorsus</i> Cc5	340622236	6
<i>Capnocytophaga ochracea</i> DSM 7271	256819408	6
<i>Capnocytophaga</i> sp. oral taxon 329 str. F0087	332882466	6
<i>Capnocytophaga</i> sp. oral taxon 335 str. F0486	420149252	6
<i>Capnocytophaga</i> sp. oral taxon 380 str. F0488	429748017	6
<i>Capnocytophaga sputigena</i> Capno	213962376	6
<i>Flavobacterium psychrophilum</i> JIPO2/86	150025575	6
<i>Galibacter</i> sp. ck-12-15	408370397	6
<i>Indibacter alkaliphilus</i> LW1	404451234	6
<i>Joostella marina</i> DSM 19592	386818981	6
<i>Kordia algicida</i> OT-1	163754820	6
<i>Marinilabilia</i> sp. AK2	410030899	6
<i>Myroides injenensis</i> M09-0166	399927444	6
<i>Niabella soli</i> DSM 19437	374372722	6
<i>Parabacteroides johnsonii</i> DSM 18315	218258638	6
<i>Parabacteroides</i> sp. D13	256840409	6
<i>Porphyromonas</i> sp. oral taxon 279 str. F0450	402847315	6
<i>Prevotella histicola</i> F0411	357042839	6
<i>Prevotella intermedia</i> 17	387132277	6
<i>Prevotella nigrescens</i> F0103	445119230	6
<i>Prevotella oralis</i> ATCC 33269	323344874	6
<i>Prevotella</i> sp. oral taxon 306 str. F0472	383811446	6

<i>Riemerella anatipestifer</i> RA-CH-1	407451859	6
<i>Riemerella anatipestifer</i> RA-GD	386321727	6
<i>Zunongwangia profunda</i> SM-A87	295136244	6
<i>Actinomyces coleocanis</i> DSM 15436	227494853	7
<i>Actinomyces georgiae</i> F0490	420151340	7
<i>Actinomyces naeslundii</i> str. Howell 279	400293272	7
<i>Actinomyces</i> sp. ICM47	396585058	7
<i>Actinomyces</i> sp. oral taxon 175 str. F0384	343523232	7
<i>Actinomyces</i> sp. oral taxon 181 str. F0379	429758968	7
<i>Actinomyces</i> sp. oral taxon 848 str. F0332	269219760	7
<i>Actinomyces turicensis</i> ACS-279-V-Co14	405979650	7
<i>Bifidobacterium dentium</i> Bd1	283456135	7
<i>Bifidobacterium longum</i> DJO10A	189440764	7
<i>Bifidobacterium longum</i> subsp. longum 2-2B	419852381	7
<i>Bifidobacterium longum</i> subsp. longum KACC 91563	384200944	7
<i>Bifidobacterium</i> sp. 12_1_47BFAA	317482066	7
<i>Corynebacterium accolens</i> ATCC 49725	227502575	7
<i>Corynebacterium accolens</i> ATCC 49726	306835141	7
<i>Corynebacterium diptheriae</i> 241	375289763	7
<i>Corynebacterium diptheriae</i> 31A	376283539	7
<i>Corynebacterium diptheriae</i> BH8	376286566	7
<i>Corynebacterium diptheriae</i> bv. intermedium str. NCTC 5011	419861895	7
<i>Corynebacterium diptheriae</i> C7 (beta)	376289243	7
<i>Corynebacterium diptheriae</i> HC02	376292154	7
<i>Corynebacterium diptheriae</i> NCTC 13129	38232678	7
<i>Corynebacterium diptheriae</i> VA01	376256051	7
<i>Corynebacterium matruchotii</i> ATCC 14266	305681510	7
<i>Corynebacterium matruchotii</i> ATCC 33806	225021644	7
<i>Gardnerella vaginalis</i> 1500E	415717744	7
<i>Gardnerella vaginalis</i> 284V	415703177	7
<i>Gardnerella vaginalis</i> 5-1	298252606	7
<i>Mobiluncus curtisii</i> subsp. holmesii ATCC 35242	315656340	7
<i>Mobiluncus mulieris</i> 28-1	269977848	7
<i>Mobiluncus mulieris</i> FB024-16	307700167	7
<i>Scardovia inopinata</i> F0304	294790575	7
<i>Actinomyces</i> sp. oral taxon 180 str. F0310	315605738	8

<i>Gluconacetobacter diazotrophicus</i> PAI 5	209542524	8
<i>Gluconacetobacter diazotrophicus</i> PAI 5	162147907	8
<i>Methylocystis</i> sp. ATCC 49242	323139312	8
<i>Methylosinus trichosporium</i> OB3b	296446027	8
<i>Rhodopseudomonas palustris</i> BisB18	90425961	8
<i>Rhodopseudomonas palustris</i> BisB5	91975509	8
<i>Tistrella mobilis</i> KA081020-065	389874754	8
<i>Mycoplasma canis</i> PG 14	384393286	9
<i>Mycoplasma canis</i> PG 14	419703974	9
<i>Mycoplasma canis</i> UF31	384937953	9
<i>Mycoplasma canis</i> UF33	419704625	9
<i>Mycoplasma canis</i> UFG1	419705269	9
<i>Mycoplasma canis</i> UFG4	419705920	9
<i>Mycoplasma cynos</i> C142	433625054	9
<i>Mycoplasma gallisepticum</i> NC95_13295-2-2P	401767318	9
<i>Mycoplasma gallisepticum</i> NY01_2001.047-5-1P	401768851	9
<i>Mycoplasma gallisepticum</i> str. F	284931710	9
<i>Mycoplasma gallisepticum</i> str. F	385326554	9
<i>Mycoplasma gallisepticum</i> str. R(low)	294660600	9
<i>Mycoplasma synoviae</i> 53	71894592	9
<i>Mycoplasma synoviae</i> 53	144575181	9
<i>Prevotella buccalis</i> ATCC 35310	282878504	10
<i>Prevotella ruminicola</i> 23	294674019	10
<i>Prevotella stercorea</i> DSM 18206	359406728	10
<i>Prevotella tanneriae</i> ATCC 51259	258648111	10
<i>Prevotella timonensis</i> CRIS 5C-B1	282880052	10
<i>Burkholderiales bacterium</i> 1_1_47	303257695	11
<i>Parasutterella excrementihominis</i> YIT 11859	331001027	11
<i>Sutterella wadsworthensis</i> 3_1_45B	319941583	11
<i>Elusimicrobium minutum</i> Pei191	187250660	12
<i>Sphaerochaeta globus</i> str. Buddy	325972003	12
uncultured Termite group 1 bacterium phylotype Rs-D17	189485059	12
<i>Flavobacterium branchiophilum</i> FL-15	347536497	13
<i>Flavobacterium columnaire</i> ATCC 49512	365959402	13
<i>Odoribacter laneus</i> YIT 12061	374384763	13
<i>Prevotella denticola</i> CRIS 18C-A	325859619	14
<i>Prevotella micans</i> F0438	373501184	14
<i>Prevotella</i> sp. C561	345885718	14

<i>Francisella tularensis</i> subsp. <i>tularensis</i> WY96-3418	134302318	15
<i>Francisella</i> cf. <i>novicida</i> 3523	387824704	16
<i>Francisella</i> cf. <i>novicida</i> Fx1	385792694	16
<i>Francisella novicida</i> FTG	208779141	16
<i>Francisella novicida</i> GA99-3548	254374175	16
<i>Francisella novicida</i> U112	118497352	16
<i>Francisella tularensis</i> subsp. <i>novicida</i> GA99-3549	254372717	16
<i>Wolinella succinogenes</i> DSM 1740	34557932	17
<i>gamma proteobacterium</i> HTCC5015	254447899	18
<i>Legionella pneumophila</i> 130b	307608922	19
<i>Legionella pneumophila</i> str. Paris	54296138	19
<i>Mycoplasma ovipneumoniae</i> SC01	363542550	20
<i>Streptobacillus moniliformis</i> DSM 12112	269123826	21
<i>Mycoplasma mobile</i> 163K	47458868	22
<i>Alcanivorax</i> sp. W11-5	407803669	23
<i>Caenispirillum salinarum</i> AK4	427429481	23
<i>Rhodospirillum rubrum</i> ATCC 11170	83591793	23
<i>Treponema</i> sp. JC4	384109266	23
<i>Ruminococcus albus</i> 8	325677756	24
uncultured delta proteobacterium HF0070_07E19	297182908	24
<i>Acidothermus cellulolyticus</i> 11B	117929158	25
<i>Nitratifactor saisuginis</i> DSM 16511	319957206	26
<i>Akkermansia muciniphila</i> ATCC BAA-835	187736489	27
<i>Parvibaculum lavamentivorans</i> DS-1	154250555	28



### Example 6

**[0001]** Phylogenetic clustering of Cas9 defines dual-RNA:Cas9 exchangeability.

**[0002]** As described above, clustering of Cas9 orthologs correlates with the ability to substitute for the RNA-stabilizing role of *S. pyogenes* Cas9 in tracrRNA:pre-crRNA processing by RNase III *in vivo* (Figure 4B). The exchangeability between Cas9 and dual-RNA in closely related CRISPR-Cas systems was investigated at the level of DNA interference.

**[0003]** Plasmid cleavage assays were performed using *S. pyogenes* Cas9 complexed with dual-RNAs from selected CRISPR-Cas systems representative of the clustering of the type II CRISPR-Cas systems. As shown in Figure 6A (upper panel), *S. pyogenes* Cas9 can cleave target DNA in the presence of dual-RNAs from *S. mutans* and *S. thermophilus*\* (type II-A, yellow subcluster), but not from any other tested species. The same result was observed when the dual-RNA from *S. pyogenes* was incubated with Cas9 orthologs from different bacteria (Figure 6A, lower panel). Cleavage assays were also performed with all Cas9 orthologs incubated with cognate and non-cognate dual-RNAs on their PAM-specific plasmid DNA. Only the combinations of Cas9 and dual-RNA within the same type II subcluster conferred dsDNA cleavage activity (Figure 6B, Supplementary Figure S10). More striking was the gradient of activity dependent on how closely related the species are in the corresponding type II group. This effect can be observed for *C. jejuni* Cas9 that is able to cleave DNA in the presence of dual-RNA from *P. multocida* and *N. meningitidis*, but not as efficient as with its own RNA (type II-C, blue subcluster). This finding is in good agreement with the phylogenetic tree of Cas9 (Figure 1A) showing that all three Cas9 orthologs belong to type II-C but *C. jejuni* Cas9 clusters more distantly from *P. multocida* and *N. meningitidis* Cas9. This effect was even greater for *S. thermophilus*\*\* Cas9 which belongs to type II-A together with *S. pyogenes*, *S. mutans* and *S. thermophilus*\*. However, none of the dual-RNAs from the three latter loci could direct DNA cleavage by *S. thermophilus*\*\* Cas9. This result supports the recent findings demonstrating the lack of exchangeability between Cas9 from CRISPR1 and CRISPR3 of *S. thermophilus* DGCC7710 with regard to dual-RNA binding (17). Cas9 and tracrRNA:crRNA interchangeability is contemplated to directly result from Cas9 co-evolution with dual-RNA and follows the Cas9 phylogeny that may differ from the phylogeny of the respective bacterial species due to horizontal transfer.

**[0004]** Thus, to investigate the interchangeability between type II subgroups at the level of DNA interference, the PAMs specific for each of the 8 selected Cas9 orthologs (28) were determined. By aligning potential crRNA-targeted sequences, conserved motifs adjacent to the protospacers in all selected species were identified. These motifs were then shown to be essential for DNA interference activity of the cognate dual-RNA:Cas9 complex *in vitro*. The interchangeability between dual-RNA and Cas9 from different subclusters was tested using plasmid cleavage assays. Only closely related Cas9

proteins can exchange their cognate dual-RNAs and still exert cleavage activity when using the Cas9 specific PAM. The specificity of Cas9 towards dual-RNAs is highly sensitive to the Cas9 sequence relatedness. This sensitivity is observed with Cas9 from *C. jejuni* that displays full cleavage activity with its cognate dual-RNA but reduced activity with dual-RNAs from *N. meningitidis* or *P. multocida* which belong to different subclusters of type II-C. It is contemplated that Cas9 possesses specificity for the secondary structure of dual-RNAs, given that bioinformatics predictions suggest similar structures of repeat:antirepeat duplexes in closely related CRISPR-Cas systems (Supplementary Figure S12).

**[0005]** While the present invention has been described in terms of specific embodiments, it is understood that variations and modifications will occur to those skilled in the art. Accordingly, only such limitations as appear in the claims should be placed on the invention.

**Documents Cited**

**[0006]** All publications and patents cited in this specification are herein incorporated by reference as if each individual publication or patent were specifically and individually indicated to be incorporated by reference and are incorporated herein by reference to disclose and describe the methods and/or materials in connection with which the publications are cited. The citation of any publication is for its disclosure prior to the filing date and should not be construed as an admission that the present invention is not entitled to antedate such publication by virtue of prior invention. Further, the dates of publication provided may be different from the actual publication dates which may need to be independently confirmed.

1. Cho, S.W., Kim, S., Kim, J.M. and Kim, J.S. (2013) Targeted genome engineering in human cells with the Cas9 RNA-guided endonuclease. *Nat. Biotechnol.*, **31**, 230-232.
2. Cong, L., Ran, F.A., Cox, D., Lin, S., Barretto, R., Habib, N., Hsu, P.D., Wu, X., Jiang, W., Marraffini, L.A. *et al.* (2013) Multiplex genome engineering using CRISPR/Cas systems. *Science*, **339**, 819-823.
3. DiCarlo, J.E., Norville, J.E., Mali, P., Rios, X., Aach, J. and Church, G.M. (2013) Genome engineering in *Saccharomyces cerevisiae* using CRISPR-Cas systems. *Nucleic Acids Res.*, **41**, 4336-4343.
4. Friedland, A.E., Tzur, Y.B., Esvelt, K.M., Colaiacovo, M.P., Church, G.M. and Calarco, J.A. (2013) Heritable genome editing in *C. elegans* via a CRISPR-Cas9 system. *Nat. Methods*, **10**, 741-743.
5. Gratz, S.J., Cummings, A.M., Nguyen, J.N., Hamm, D.C., Donohue, L.K., Harrison, M.M., Wildonger, J. and O'Connor-Giles, K.M. (2013) Genome engineering of *Drosophila* with the CRISPR RNA-guided Cas9 nuclease. *Genetics*, **194**, 1029-1035.
6. Hwang, W.Y., Fu, Y., Reyon, D., Maeder, M.L., Tsai, S.Q., Sander, J.D., Peterson, R.T., Yeh, J.R. and Joung, J.K. (2013) Efficient genome editing in zebrafish using a CRISPR-Cas system. *Nat. Biotechnol.*, **31**, 227-229.
7. Jiang, W., Bikard, D., Cox, D., Zhang, F. and Marraffini, L.A. (2013) RNA-guided editing of bacterial genomes using CRISPR-Cas systems. *Nat. Biotechnol.*, **31**, 233-239.
8. Mali, P., Yang, L., Esvelt, K.M., Aach, J., Guell, M., DiCarlo, J.E., Norville, J.E. and Church, G.M. (2013) RNA-guided human genome engineering via Cas9. *Science*, **339**, 823-826.
9. Shen, B., Zhang, J., Wu, H., Wang, J., Ma, K., Li, Z., Zhang, X., Zhang, P. and Huang, X. (2013) Generation of gene-modified mice via Cas9/RNA-mediated gene targeting. *Cell Res.*, **23**, 720-723.
10. Wang, H., Yang, H., Shivalila, C.S., Dawlaty, M.M., Cheng, A.W., Zhang, F. and Jaenisch, R. (2013) One-step generation of mice carrying mutations in multiple genes by CRISPR/Cas-mediated genome engineering. *Cell*, **153**, 910-918.
11. Jinek, M., East, A., Cheng, A., Lin, S., Ma, E. and Doudna, J. (2013) RNA-programmed genome editing in human cells. *eLIFE*, **2**, e00471.
12. Li, J.F., Norville, J.E., Aach, J., McCormack, M., Zhang, D., Bush, J., Church, G.M. and Sheen, J. (2013) Multiplex and homologous recombination-mediated genome editing in *Arabidopsis* and *Nicotiana benthamiana* using guide RNA and Cas9. *Nat. Biotechnol.*, **31**, 688-691.

13. Nekrasov, V., Staskawicz, B., Weigel, D., Jones, J.D. and Kamoun, S. (2013) Targeted mutagenesis in the model plant *Nicotiana benthamiana* using Cas9 RNA-guided endonuclease. *Nat. Biotechnol.*, **31**, 691-693.
14. Jinek, M., Chylinski, K., Fonfara, I., Hauer, M., Doudna, J.A. and Charpentier, E. (2012) A programmable dual-RNA-guided DNA endonuclease in adaptive bacterial immunity. *Science*, **337**, 816-821.
15. Chylinski, K., Le Rhun, A. and Charpentier, E. (2013) The tracrRNA and Cas9 families of type II CRISPR-Cas immunity systems. *RNA Biol.*, **10**, 726-737.
16. Deltcheva, E., Chylinski, K., Sharma, C.M., Gonzales, K., Chao, Y., Pirzada, Z.A., Eckert, M.R., Vogel, J. and Charpentier, E. (2011) CRISPR RNA maturation by trans-encoded small RNA and host factor RNase III. *Nature*, **471**, 602-607.
17. Karvelis, T., Gasiunas, G., Miksys, A., Barrangou, R., Horvath, P. and Siksnys, V. (2013) crRNA and tracrRNA guide Cas9-mediated DNA interference in *Streptococcus thermophilus*. *RNA Biol.*, **10**, 841-851.
18. Garneau, J.E., Dupuis, M.E., Villion, M., Romero, D.A., Barrangou, R., Boyaval, P., Fremaux, C., Horvath, P., Magadan, A.H. and Moineau, S. (2010) The CRISPR/Cas bacterial immune system cleaves bacteriophage and plasmid DNA. *Nature*, **468**, 67-71.
19. Magadan, A.H., Dupuis, M.E., Villion, M. and Moineau, S. (2012) Cleavage of phage DNA by the *Streptococcus thermophilus* CRISPR3-Cas system. *PLoS One*, **7**, e40913.
20. Haft, D.H., Selengut, J., Mongodin, E.F. and Nelson, K.E. (2005) A guild of 45 CRISPR-associated (Cas) protein families and multiple CRISPR/Cas subtypes exist in prokaryotic genomes. *PLoS Comput. Biol.*, **1**, e60.
21. Makarova, K.S., Grishin, N.V., Shabalina, S.A., Wolf, Y.I. and Koonin, E.V. (2006) A putative RNA-interference-based immune system in prokaryotes: computational analysis of the predicted enzymatic machinery, functional analogies with eukaryotic RNAi, and hypothetical mechanisms of action. *Biol. Direct*, **1**, 7.
22. Gasiunas, G., Barrangou, R., Horvath, P. and Siksnys, V. (2012) Cas9-crRNA ribonucleoprotein complex mediates specific DNA cleavage for adaptive immunity in bacteria. *Proc. Natl. Acad. Sci. U. S. A.*, **109**, E2579-2586.
23. Sapranaukas, R., Gasiunas, G., Fremaux, C., Barrangou, R., Horvath, P. and Siksnys, V. (2011) The *Streptococcus thermophilus* CRISPR/Cas system provides immunity in *Escherichia coli*. *Nucleic Acids Res.*, **39**, 9275-9282.
24. Mali, P., Aach, J., Stranges, P.B., Esvelt, K.M., Moosburner, M., Kosuri, S., Yang, L. and Church, G.M. (2013) Cas9 transcriptional activators for target specificity screening and paired nickases for cooperative genome engineering. *Nat Biotechnol.*, **31**, 833-838.
25. Ran, F.A., Hsu, P.D., Lin, C.Y., Gootenberg, J.S., Konermann, S., Trevino, A.E., Scott, D.A., Inoue, A., Matoba, S., Zhang, Y. *et al.* (2013) Double nicking by RNA-guided CRISPR Cas9 for enhanced genome editing specificity. *Cell*, **154**, 1380-1389.
26. Deveau, H., Barrangou, R., Garneau, J.E., Labonte, J., Fremaux, C., Boyaval, P., Romero, D.A., Horvath, P. and Moineau, S. (2008) Phage response to CRISPR-encoded resistance in *Streptococcus thermophilus*. *J. Bacteriol.*, **190**, 1390-1400.
27. Horvath, P., Romero, D.A., Coute-Monvoisin, A.C., Richards, M., Deveau, H., Moineau, S., Boyaval, P., Fremaux, C. and Barrangou, R. (2008) Diversity, activity, and evolution of CRISPR loci in *Streptococcus thermophilus*. *J. Bacteriol.*, **190**, 1401-1412.
28. Mojica, F.J., Diez-Villasenor, C., Garcia-Martinez, J. and Almendros, C. (2009) Short motif sequences determine the targets of the prokaryotic CRISPR defence system. *Microbiology*, **155**, 733-740.

29. Bikard, D., Jiang, W., Samai, P., Hochschild, A., Zhang, F. and Marraffini, L.A. (2013) Programmable repression and activation of bacterial gene expression using an engineered CRISPR-Cas system. *Nucleic Acids Res.*, **41**, 7429-7437.
30. Qi, L.S., Larson, M.H., Gilbert, L.A., Doudna, J.A., Weissman, J.S., Arkin, A.P. and Lim, W.A. (2013) Repurposing CRISPR as an RNA-guided platform for sequence-specific control of gene expression. *Cell*, **152**, 1173-1183.
31. Charpentier, E. and Doudna, J.A. (2013) Biotechnology: Rewriting a genome. *Nature*, **495**, 50-51.
32. Horvath, P. and Barrangou, R. (2013) RNA-guided genome editing a la carte. *Cell Res.*, **23**, 733-734.
33. van der Oost, J. (2013) Molecular biology. New tool for genome surgery. *Science*, **339**, 768-770.
34. Hou, Z., Zhang, Y., Propson, N.E., Howden, S.E., Chu, L.F., Sontheimer, E.J. and Thomson, J.A. (2013) Efficient genome engineering in human pluripotent stem cells using Cas9 from *Neisseria meningitidis*. *Proc. Natl. Acad. Sci. U. S. A.*, **110**, 15644-15649.
35. Sambrook, J., Fritsch, E.F. and Maniatis, T. (1989) *Molecular Cloning: a Laboratory Manual*. 2nd edn. Cold Spring Harbor, N. Y. ed. Cold Spring Harbor Laboratory Press.
36. Caparon, M.G. and Scott, J.R. (1991) Genetic manipulation of pathogenic streptococci. *Methods Enzymol.*, **204**, 556-586.
37. Kirsch, R.D. and Joly, E. (1998) An improved PCR-mutagenesis strategy for two-site mutagenesis or sequence swapping between related genes. *Nucleic Acids Res.*, **26**, 1848-1850.
38. Siller, M., Janapatla, R.P., Pirzada, Z.A., Hassler, C., Zinkl, D. and Charpentier, E. (2008) Functional analysis of the group A streptococcal *luxS/AI-2* system in metabolism, adaptation to stress and interaction with host cells. *BMC Microbiol.*, **8**, 188.
39. Mangold, M., Siller, M., Roppenser, B., Vlamincx, B.J., Penfound, T.A., Klein, R., Novak, R., Novick, R.P. and Charpentier, E. (2004) Synthesis of group A streptococcal virulence factors is controlled by a regulatory RNA molecule. *Mol. Microbiol.*, **53**, 1515-1527.
40. Herbert, S., Barry, P. and Novick, R.P. (2001) Subinhibitory clindamycin differentially inhibits transcription of exoprotein genes in *Staphylococcus aureus*. *Infect. Immun.*, **69**, 2996-3003.
41. Pall, G.S. and Hamilton, A.J. (2008) Improved northern blot method for enhanced detection of small RNA. *Nat. Protoc.*, **3**, 1077-1084.
42. Urban, J.H. and Vogel, J. (2007) Translational control and target recognition by *Escherichia coli* small RNAs *in vivo*. *Nucleic Acids Res.*, **35**, 1018-1037.
43. McClelland, M., Hanish, J., Nelson, M. and Patel, Y. (1988) KGB: a single buffer for all restriction endonucleases. *Nucleic Acids Res.*, **16**, 364.
44. Makarova, K.S., Haft, D.H., Barrangou, R., Brouns, S.J., Charpentier, E., Horvath, P., Moineau, S., Mojica, F.J., Wolf, Y.I., Yakunin, A.F. *et al.* (2011) Evolution and classification of the CRISPR-Cas systems. *Nat. Rev. Microbiol.*, **9**, 467-477.
45. Altschul, S.F., Madden, T.L., Schaffer, A.A., Zhang, J., Zhang, Z., Miller, W. and Lipman, D.J. (1997) Gapped BLAST and PSI-BLAST: a new generation of protein database search programs. *Nucleic Acids Res.*, **25**, 3389-3402.
46. Wheeler, D. and Bhagwat, M. (2007) BLAST QuickStart: example-driven web-based BLAST tutorial. *Methods Mol. Biol.*, **395**, 149-176.
47. Edgar, R.C. (2004) MUSCLE: multiple sequence alignment with high accuracy and high throughput. *Nucleic Acids Res.*, **32**, 1792-1797.
48. Soding, J., Biegert, A. and Lupas, A.N. (2005) The HHpred interactive server for protein homology detection and structure prediction. *Nucleic Acids Res.*, **33**, W244-248.

49. Price, M.N., Dehal, P.S. and Arkin, A.P. (2010) FastTree 2--approximately maximum-likelihood trees for large alignments. *PLoS One*, **5**, e9490.
50. Bernhart, S.H., Tafer, H., Muckstein, U., Flamm, C., Stadler, P.F. and Hofacker, I.L. (2006) Partition function and base pairing probabilities of RNA heterodimers. *Algorithms Mol. Biol.*, **1**, 3.
51. Hofacker, I.L., Fekete, M. and Stadler, P.F. (2002) Secondary structure prediction for aligned RNA sequences. *Journal of molecular biology*, **319**, 1059-1066.
52. Darty, K., Denise, A. and Ponty, Y. (2009) VARNA: Interactive drawing and editing of the RNA secondary structure. *Bioinformatics*, **25**, 1974-1975.
53. Bhaya, D., Davison, M. and Barrangou, R. (2011) CRISPR-Cas systems in bacteria and archaea: versatile small RNAs for adaptive defense and regulation. *Annu. Rev. Genet.*, **45**, 273-297.
54. Zhang, Y., Heidrich, N., Ampattu, B.J., Gunderson, C.W., Seifert, H.S., Schoen, C., Vogel, J. and Sontheimer, E.J. (2013) Processing-independent CRISPR RNAs limit natural transformation in *Neisseria meningitidis*. *Mol. Cell*, **50**, 488-503.
55. Takeuchi, N., Wolf, Y.I., Makarova, K.S. and Koonin, E.V. (2012) Nature and intensity of selection pressure on CRISPR-associated genes. *J. Bacteriol.*, **194**, 1216-1225.
56. Makarova, K.S., Aravind, L., Wolf, Y.I. and Koonin, E.V. (2011) Unification of Cas protein families and a simple scenario for the origin and evolution of CRISPR-Cas systems. *Biol. Direct.*, **6**, 38.
57. Barrangou, R., Fremaux, C., Deveau, H., Richards, M., Boyaval, P., Moineau, S., Romero, D.A. and Horvath, P. (2007) CRISPR provides acquired resistance against viruses in prokaryotes. *Science*, **315**, 1709-1712.
58. Sun, W., Li, G. and Nicholson, A.W. (2004) Mutational analysis of the nuclease domain of *Escherichia coli* ribonuclease III. Identification of conserved acidic residues that are important for catalytic function *in vitro*. *Biochemistry*, **43**, 13054-13062.
59. Sun, W., Jun, E. and Nicholson, A.W. (2001) Intrinsic double-stranded-RNA processing activity of *Escherichia coli* ribonuclease III lacking the dsRNA-binding domain. *Biochemistry*, **40**, 14976-14984.

### Claims

We claim:

1. A single-molecule guide RNA comprising:  
a DNA-targeting segment and a protein-binding segment wherein the protein-binding segment comprises a tracrRNA set out in Supplementary Table S5.
2. The single-molecule guide RNA of claim 1 wherein the protein-binding segment comprises a CRISPR repeat set out in Supplementary Table S5 that is the cognate CRISPR repeat of the tracrRNA of the protein-binding segment.
3. The single-molecule guide RNA of claim 1 or 2 wherein the DNA-targeting segment further comprises RNA complementary to a protospacer-like sequence in a target DNA 5' to a PAM sequence.
4. The single-molecule guide RNA of claim 3 wherein the tracrRNA and CRISPR repeat are respectively at least 80% identical to the *C. jejuni* tracrRNA and CRISPR repeat set out in Supplementary Table S5 and wherein the PAM sequence is NNNNACA.
5. The single-molecule guide RNA of claim 4 or 8 wherein the RNA complementary to a protospacer-like sequence is RNA complementary to the target sequences set out in one of SEQ ID NOs: 801-973, 1079-1222, 1313-1348, 1372-1415, 1444-1900, 2163-2482 or 2667-2686.
6. A single-molecule guide RNA comprising:  
a DNA-targeting segment and a protein-binding segment, wherein the protein-binding segment comprises a tracrRNA at least 80% identical over at least 20 nucleotides to a tracrRNA set out in Supplementary Table S5.
7. A single-molecule guide RNA comprising:  
a DNA-targeting segment and a protein-binding segment, wherein the protein-binding segment comprises a tracrRNA at least 80% identical over at least 20 nucleotides to a tracrRNA set out in Supplementary Table S5, a CRISPR repeat at least 80% identical to a CRISPR repeat set out in Supplementary Table S5, or both.
8. The single-molecule guide RNA of claim 7 wherein the tracrRNA and CRISPR repeat are respectively at least 80% identical to the *C. jejuni* tracrRNA and CRISPR repeat set out in Supplementary Table S5 and wherein the PAM sequence is NNNNACA.

9. The single-molecule guide RNA of claim 1 or 6 comprising a linker between the DNA-targeting segment and the protein-binding segment.

10. A DNA encoding a single-molecule guide RNA comprising:  
a DNA-targeting segment and a protein-binding segment, wherein the protein-binding segment comprises a tracrRNA set out in Supplementary Table S5.

11. A DNA encoding a single-molecule guide RNA comprising:  
a DNA-targeting segment and a protein-binding segment, wherein the protein-binding segment comprises a tracrRNA at least 80% identical over at least 20 nucleotides to a tracrRNA set out in Supplementary Table S5, a CRISPR repeat at least 80% identical to a CRISPR repeat set out in Supplementary Table S5, or both.

12. A vector comprising a DNA encoding a single-molecule guide RNA comprising:  
a DNA-targeting segment and a protein-binding segment wherein the protein-binding segment comprises a tracrRNA set out in Supplementary Table S5.

13. A vector comprising a DNA encoding a single-molecule guide RNA comprising:  
a DNA-targeting segment and a protein-binding segment, wherein the protein-binding segment comprises a tracrRNA at least 80% identical over at least 20 nucleotides to a tracrRNA set out in Supplementary Table S5, a CRISPR repeat at least 80% identical to a CRISPR repeat set out in Supplementary Table S5, or both.

14. A cell comprising a DNA encoding a single-molecule guide RNA comprising:  
a DNA-targeting segment and a protein-binding segment, wherein the protein-binding segment comprises a tracrRNA set out in Supplementary Table S5.

15. A cell comprising a DNA encoding a single-molecule guide RNA comprising:  
a DNA-targeting segment and a protein-binding segment, wherein the protein-binding segment comprises a tracrRNA at least 80% identical over at least 20 nucleotides to a tracrRNA set out in Supplementary Table S5, a CRISPR repeat at least 80% identical to a CRISPR repeat set out in Supplementary Table S5, or both.

16. A double-molecule guide RNA comprising:  
a targeter-RNA and an activator-RNA complementary thereto, wherein the activator-RNA comprises a tracrRNA set out in Supplementary Table S5, and



wherein the guide RNA comprises a modified backbone, a non-natural internucleoside linkage, a nucleic acid mimetic, a modified sugar moiety, a base modification, a modification or sequence that provides for modified or regulated stability, a modification or sequence that provides for subcellular tracking, a modification or sequence that provides for tracking, or a modification or sequence that provides for a binding site for a protein or protein complex.

17. The double-molecule guide RNA of claim 16, wherein the targeter-RNA comprises a CRISPR repeat set out in Supplementary Table S5 that is the cognate CRISPR repeat of the tracrRNA of the protein-binding segment.

18. The double-molecule guide RNA of claim 16 or 17 wherein the targeter-RNA further comprises RNA complementary to a protospacer-like sequence in a target DNA 5' to a PAM sequence.

19. The double-molecule guide RNA of claim 18 wherein the tracrRNA and CRISPR repeat are respectively at least 80% identical to the *C. jejuni* tracrRNA and CRISPR repeat set out in Supplementary Table S5 and wherein the PAM sequence is NNNNACA.

20. The double-molecule guide RNA of claim 19 or claim 23 wherein the RNA complementary to a protospacer-like sequence is RNA complementary to the target sequences set out in one of SEQ ID NOs: 801-973, 1079-1222, 1313-1348, 1372-1415, 1444-1900, 2163-2482 or 2667-2686.

21. A double-molecule guide RNA comprising:  
a targeter-RNA and a activator-RNA, wherein the activator-RNA comprises a tracrRNA at least 80% identical over at least 20 nucleotides to a tracrRNA set out in Supplementary Table S5.

22. The double-molecule guide RNA of claim 21, wherein the targeter-RNA comprises a CRISPR repeat set out in Supplementary Table S5, the cognate CRISPR repeat of the tracrRNA of the activator-RNA set out in Supplementary Table S5, or a CRISPR repeat at least 80% identical to a CRISPR repeat set out in Supplementary Table S5.

23. The double-molecule guide RNA of claim 21 wherein the tracrRNA and CRISPR repeat are respectively at least 80% identical to the *C. jejuni* tracrRNA and CRISPR repeat set out in Supplementary Table S5 and wherein the PAM sequence is NNNNACA.

24. The double-molecule guide RNA of claim 16 or 21 comprising a linker between the targeter-RNA and the activator-RNA.

25. A DNA encoding a double-molecule guide RNA comprising:  
a targeter-RNA and a activator-RNA complementary thereto, wherein the activator-RNA comprises a tracrRNA set out in Supplementary Table S5.
26. A DNA encoding a double-molecule guide RNA comprising:  
a targeter-RNA and a activator-RNA complementary thereto, wherein the activator-RNA comprises a tracrRNA at least 80% identical over at least 20 nucleotides to a tracrRNA set out in Supplementary Table S5, a CRISPR repeat at least 80% identical to a CRISPR repeat set out in Supplementary Table S5, or both.
27. A vector comprising a DNA encoding a double-molecule guide RNA comprising:  
a targeter-RNA and a activator-RNA complementary thereto, wherein the activator-RNA comprises a tracrRNA set out in Supplementary Table S5.
28. A vector comprising a DNA encoding a double-molecule guide RNA comprising:  
a targeter-RNA and a activator-RNA complementary thereto, wherein the activator-RNA comprises a tracrRNA at least 80% identical over at least 20 nucleotides to a tracrRNA set out in Supplementary Table S5, a CRISPR repeat at least 80% identical to a CRISPR repeat set out in Supplementary Table S5, or both.
29. A cell comprising a DNA encoding a double-molecule guide RNA comprising:  
a targeter-RNA and a activator-RNA complementary thereto, wherein the activator-RNA comprises a tracrRNA set out in Supplementary Table S5.
30. A cell comprising a DNA encoding a double-molecule guide RNA comprising:  
a targeter-RNA and a activator-RNA complementary thereto, wherein the activator-RNA comprises a tracrRNA at least 80% identical over at least 20 nucleotides to a tracrRNA set out in Supplementary Table S5, a CRISPR repeat at least 80% identical to a CRISPR repeat set out in Supplementary Table S5, or both.
31. A method for manipulating DNA in a cell, comprising contacting the DNA with a Cas9 ortholog-guideRNA complex, wherein the complex comprises:  
(a) a *C. jejuni* Cas9 endonuclease heterologous to the cell or an endonuclease with an activity portion at least 90% identical to the activity portion of the *C. jejuni* Cas9 endonuclease, and a guide RNA targeting the complex to a protospacer-like sequence in the DNA 5' to the PAM sequence NNNNACA;  
(b) a *P. multocida* Cas9 endonuclease heterologous to the cell or an endonuclease with an activity portion at least 90% identical to the activity portion of the *P. multocida* Cas9 endonuclease, and a

guide RNA targeting the complex to a protospacer-like sequence in the DNA 5' to the PAM sequence GNNNCNNA or NNNNC;

(c) an *F. novicida* Cas9 endonuclease heterologous to the cell or an endonuclease with an activity portion at least 90% identical to the activity portion of the *F. novicida* Cas9 endonuclease, and a guide RNA targeting the complex to a protospacer-like sequence in the DNA 5' to the PAM sequence NG;

(d) an *S. thermophilus*\*\* Cas9 endonuclease heterologous to the cell or an endonuclease with an activity portion at least 90% identical to the activity portion of the *S. thermophilus*\*\* Cas9 endonuclease, and a guide RNA targeting the complex to a protospacer-like sequence in the DNA 5' to the PAM sequence NNAAAAW;

(e) an *L. innocua* Cas9 endonuclease heterologous to the cell or an endonuclease with an activity portion at least 90% identical to the activity portion of the *L. innocua* Cas9 endonuclease, and a guide RNA targeting the complex to a protospacer-like sequence in the DNA 5' to the PAM sequence NGG; or

(f) an *S. dysgalactiae* Cas9 endonuclease heterologous to the cell or an endonuclease with an activity portion at least 90% identical to the activity portion of the *S. dysgalactiae* Cas9 endonuclease, and a guide RNA targeting the complex to a protospacer-like sequence in the DNA 5' to the PAM sequence NGG.

32. The method of claim 31 wherein the cell is a bacterial cell, a fungal cell, an archaea cell, a plant cell or an animal cell.

33. The method of claim 31 wherein the guide RNA is a single-molecule guide RNA.

34. The method of claim 31 wherein the guide RNA is a double-molecule guide RNA.

35. The method of claim 31 wherein the endonuclease is a nickase.

36. The method of claim 31 wherein the endonuclease comprises a mutation corresponding to *S. pyogenes* E762A, HH983AA or D986A.

37. The method of claim 31 wherein the endonuclease is a dead mutant/DNA binding protein.

38. The method of claim 31 wherein the protospacer-like sequence targeted is in a CCR5, CXCR4, KRT5, KRT14, PLEC or COL7A1 gene.

39. The method of claim 31 wherein the protospacer-like sequence is in a chronic granulomatous disease (CGD)-related gene CYBA, CYBB, NCF1, NCF2 or NCF4.

40. The method of claim 31 wherein the protospacer-like sequence targeted is in, or is up to 1000 nucleotides upstream of, a gene encoding B-cell lymphoma/leukemia II A (BCL11A) protein, an erythroid enhancer of BCL11A or a BCL11A binding site.

41. The method of claim 31 wherein the endonuclease and the guide RNA are introduced to the cell by the same or different recombinant vectors encoding the endonuclease and the guide RNA.

42. The method of claim 31 wherein at least one recombinant vector is a recombinant viral vector.

43. A recombinant vector encoding:

(a) a guide RNA, wherein the guide RNA comprises a DNA-targeting segment complementary to a protospacer-like sequence in the DNA 5' to the PAM sequence NNNNACA; and

(b) s *C. jejuni* Cas9 endonuclease or an endonuclease with an activity portion at least 90% identical to the activity portion of the *C. jejuni* Cas9 endonuclease.

44. A recombinant vector encoding:

(a) a guide RNA, wherein the guide RNA comprises a DNA-targeting segment complementary to a protospacer-like sequence in the DNA 5' to the PAM sequence GNNNCNNA or NNNNC; and

(b) a *P. multocida* Cas9 endonuclease or an endonuclease with an activity portion at least 90% identical to the activity portion the *P. multocida* Cas9 endonuclease.

45. A recombinant vector encoding:

(a) a guide RNA, wherein the guide RNA comprises a DNA-targeting segment complementary to a protospacer-like sequence in the DNA 5' to the PAM sequence NG; and

(b) a *F. novicida* Cas9 endonuclease or an endonuclease with an activity portion at least 90% identical to the activity portion of the *F. novicida* Cas9 endonuclease.

46. A recombinant vector encoding:

(a) a guide RNA, wherein the guide RNA comprises a DNA-targeting segment complementary to a protospacer-like sequence in the DNA 5' to the PAM sequence NNAAAAG; and

(b) a *S. thermophilus*\*\* Cas9 endonuclease or an endonuclease with an activity portion at least 90% identical to the activity portion of the *S. thermophilus*\*\* Cas9 endonuclease.

47. A recombinant vector encoding:

(a) a guide RNA, wherein the guide RNA comprises a DNA-targeting segment complementary to a protospacer-like sequence in the DNA 5' to the PAM sequence NGG; and

(b) a *L. innocua* Cas9 endonuclease or an endonuclease with an activity portion at least 90% identical to the activity portion of the *L. innocua* Cas9 endonuclease.

48. A recombinant vector encoding:

(a) a guide RNA, wherein the guide RNA comprises a DNA-targeting segment complementary to a protospacer-like sequence in the DNA 5' to the PAM sequence NGG; and

(b) a *S. dysgalactiae* Cas9 endonuclease or an endonuclease with an activity portion at least 90% identical to the activity portion of the *S. dysgalactiae* Cas9 endonuclease.

49. The recombinant vector of claim 43, 44, 45, 46, 47 or 48 wherein the recombinant vector is a recombinant viral vector.

50. A modified Cas9 endonuclease comprising one or more mutations corresponding to *S. pyogenes* mutation E762A, HH983AA or D986A.

51. The modified Cas 9 endonuclease of claim 50 further comprising one or more mutations corresponding to *S. pyogenes* mutation D10A, H840A, G12A, G17A, N854A, N863A, N982A or A984A.

52. A method for manipulating DNA in a cell, comprising contacting the DNA with a Cas9 ortholog-guide RNA complex, wherein the complex comprises:

(a) a Cas9 endonuclease heterologous to the cell and

(b) a cognate guide RNA of the Cas9 endonuclease comprising a tracrRNA set out in Supplementary Table S5 or a guide RNA comprising a tracrRNA at least 80% identical to a cognate tracrRNA set out in Supplementary Table S5 over at least 20 nucleotides.

53. The method of claim 52 wherein the cell is a bacterial cell, a fungal cell, an archaea cell, a plant cell or an animal cell.

54. The method of claim 52 wherein the guide RNA is a single-molecule guide RNA.

55. The method of claim 52 wherein the guide RNA is a double-molecule guide RNA.

56. The method of claim 52 wherein the endonuclease is a nickase.

57. The method of claim 52 wherein the endonuclease comprises a mutation corresponding to S pyogenes mutations E762, HH983AA or D986A.

58. The method of claim 52 wherein the endonuclease is a dead mutant/DNA binding protein.

59. The method of claim 52 wherein the protospacer-like sequence targeted is in a CCR5, CXCR4, KRT5, KRT14, PLEC or COL7A1 gene or a sequence up to 1000 nucleotides upstream of the gene.

60. The method of claim 52 wherein the protospacer-like sequence is in a chronic granulomatous disease (CGD)-related gene CYBA, CYBB, NCF1, NCF2 or NCF4 or a sequence up to 1000 nucleotides upstream of the gene.

61. The method of claim 52 wherein the protospacer-like sequence targeted is in, or is up to 1000 nucleotides upstream of, a gene encoding B-cell lymphoma/leukemia IIA (BCL11A) protein, an erythroid enhancer of BCL11A or a BCL11A binding site.

62. The method of claim 52 wherein the endonuclease and the guide RNA are introduced to the cell by the same or different recombinant vectors encoding the endonuclease and the guide RNA.

63. The method of claim 52 wherein at least one recombinant vector is a recombinant viral vector.

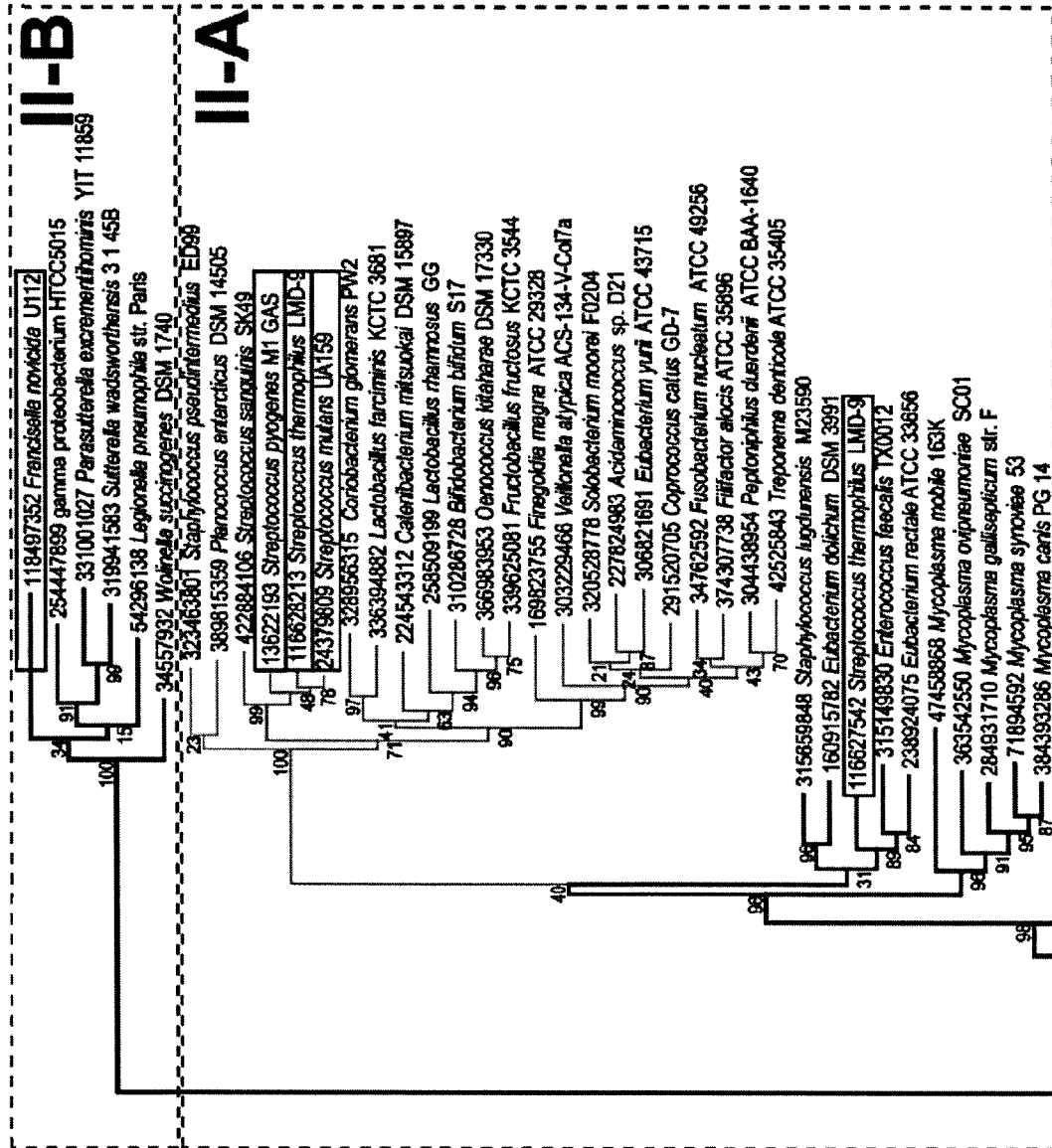


Figure. 1A



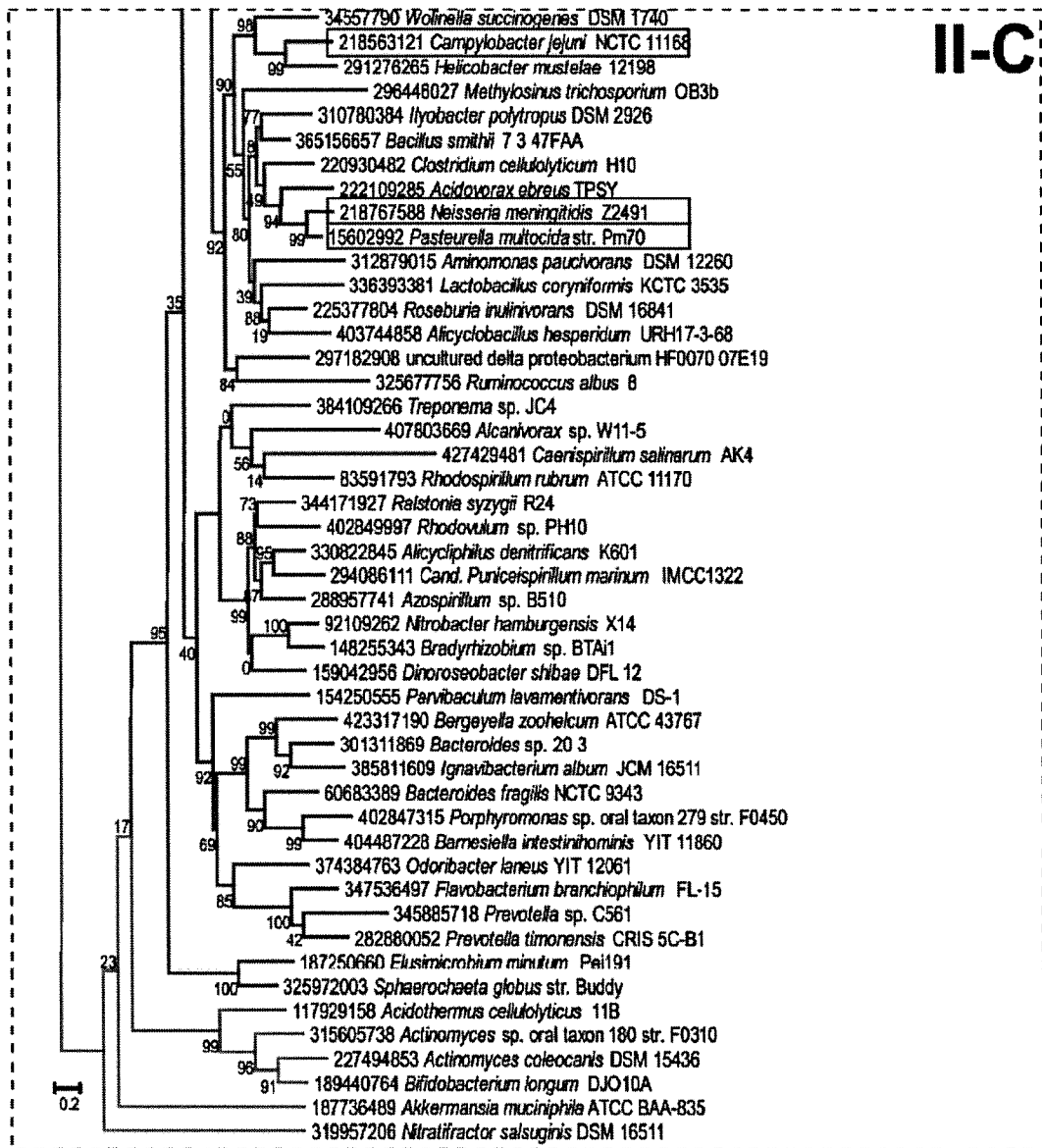


Figure. 1A  
(Continued)

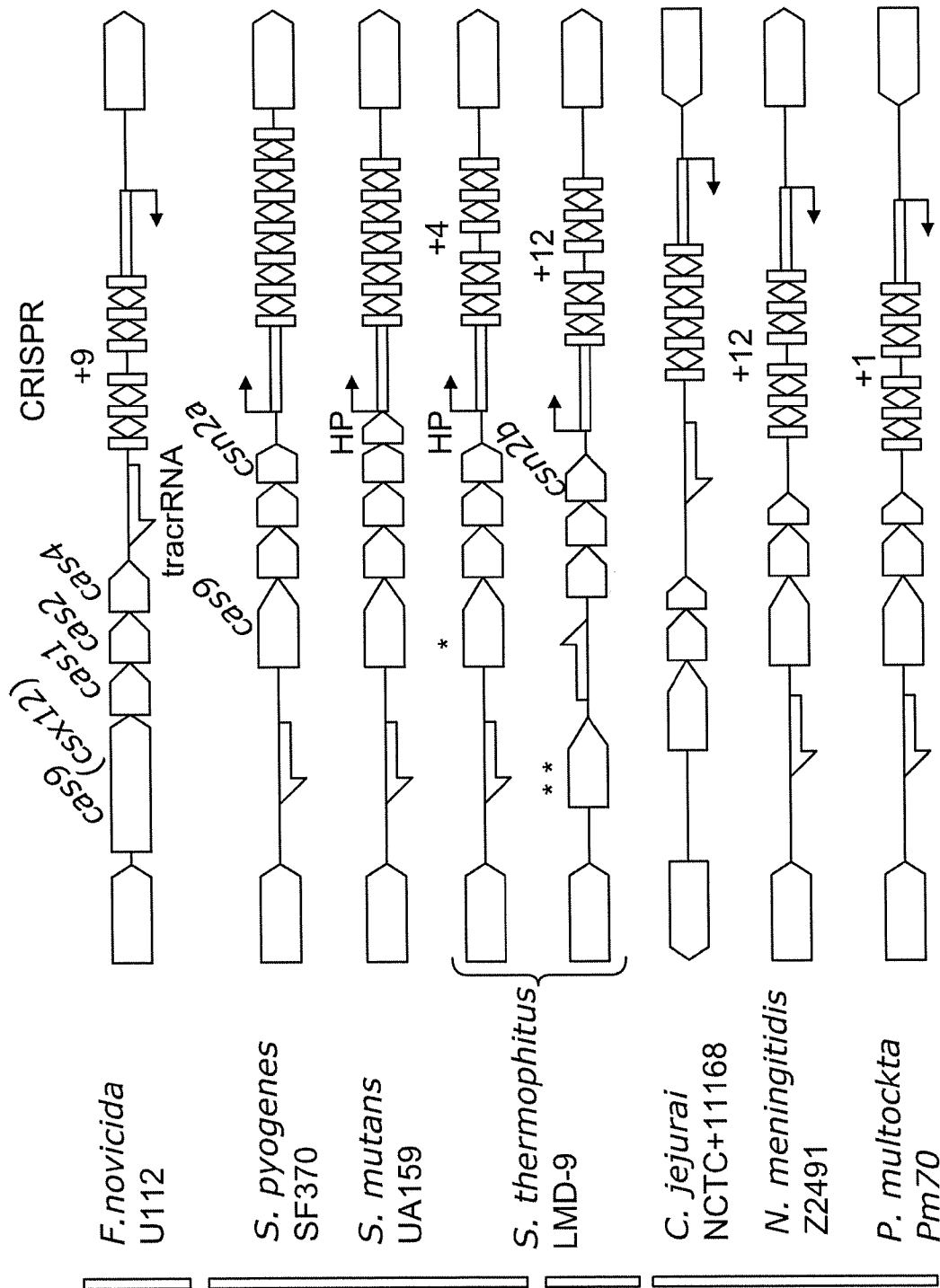


Figure 1B

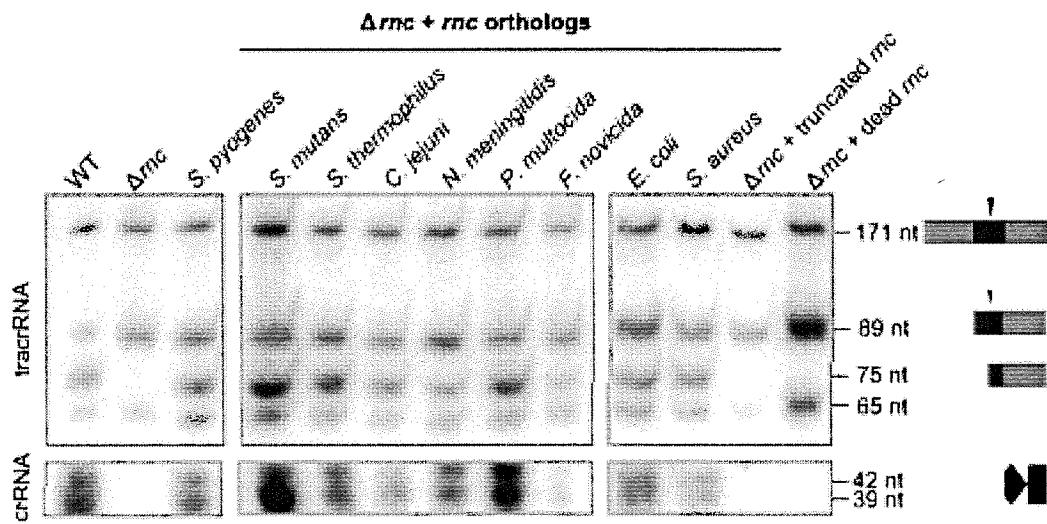


Figure 2

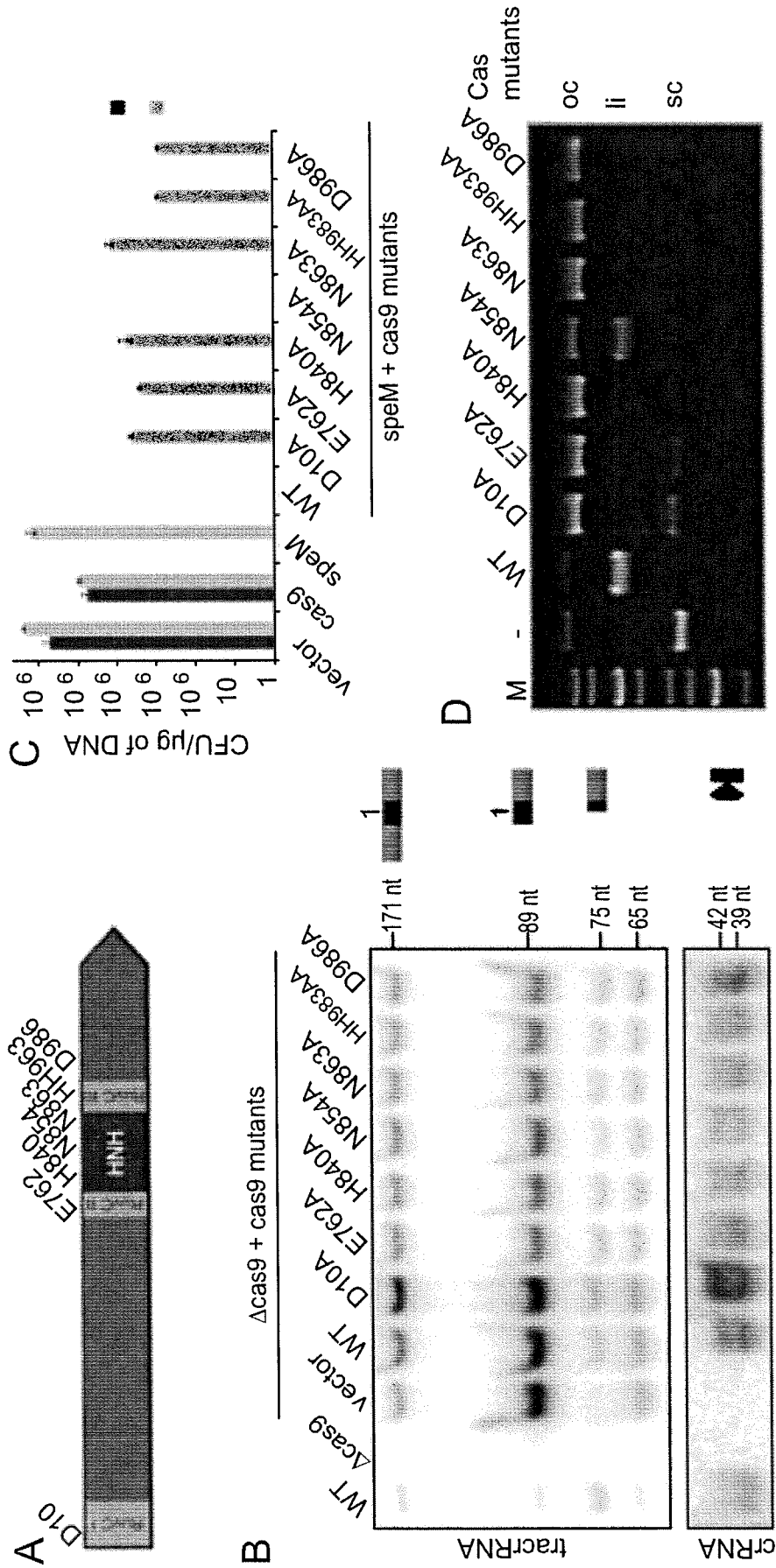


Figure 3

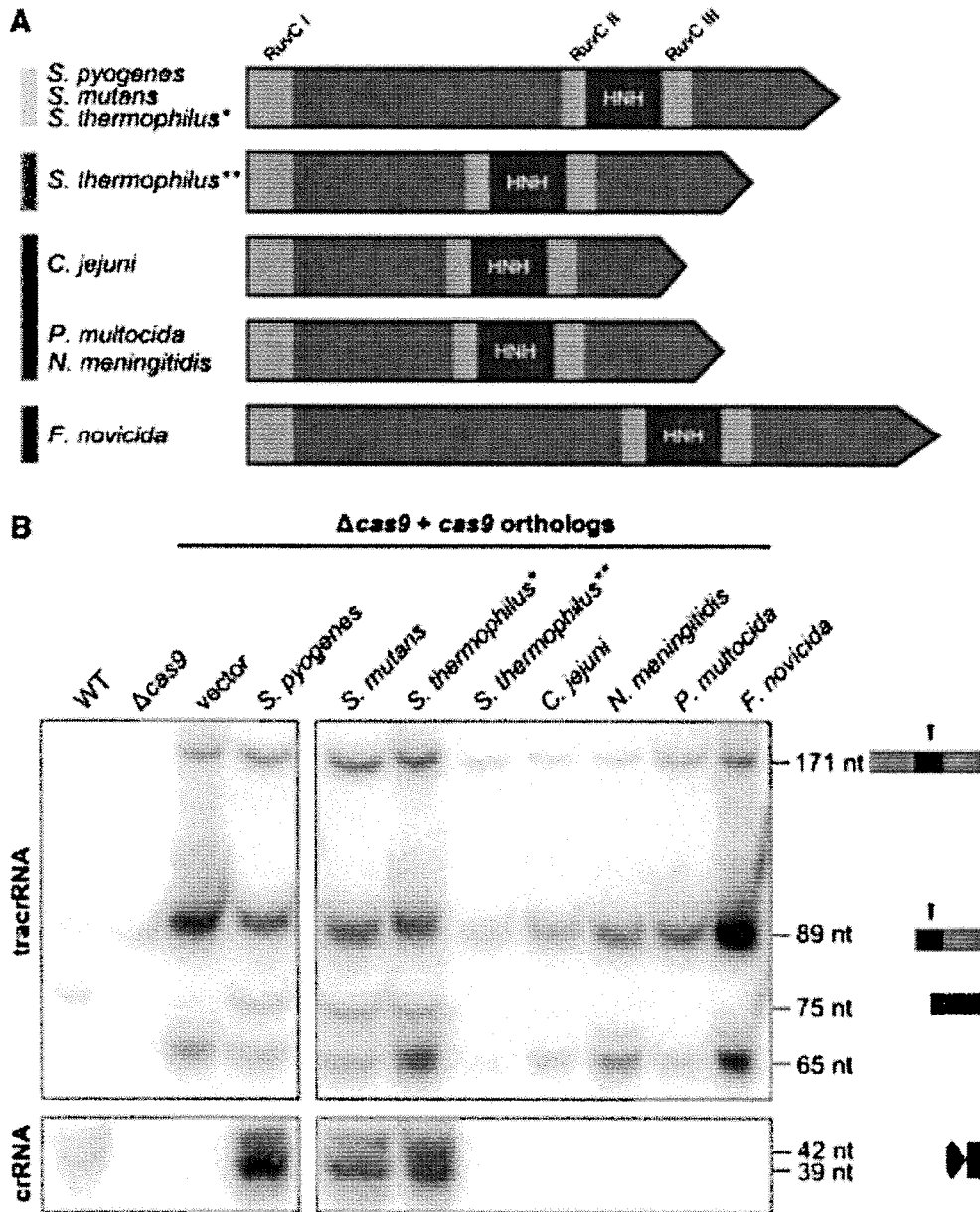


Figure 4

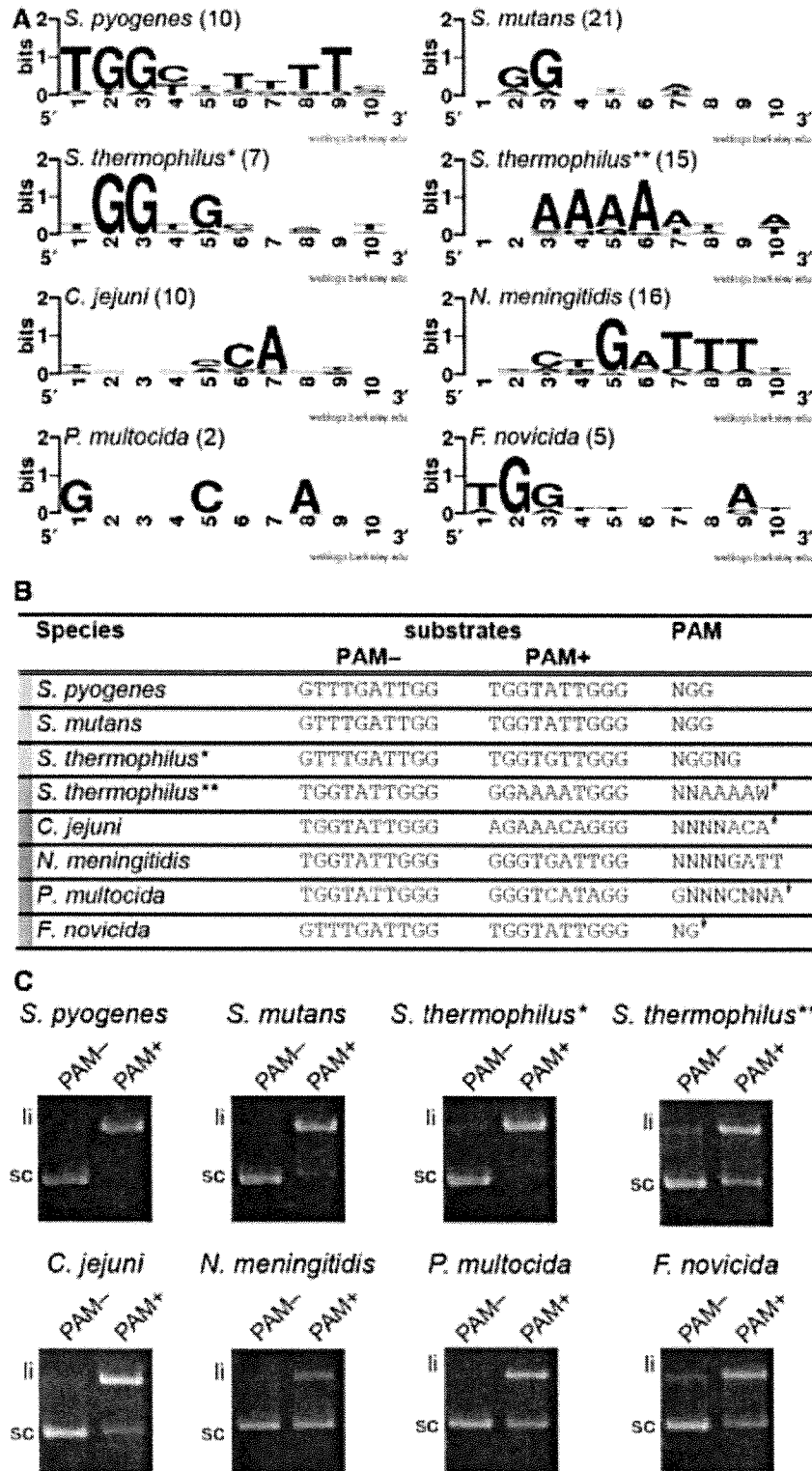


Figure 5

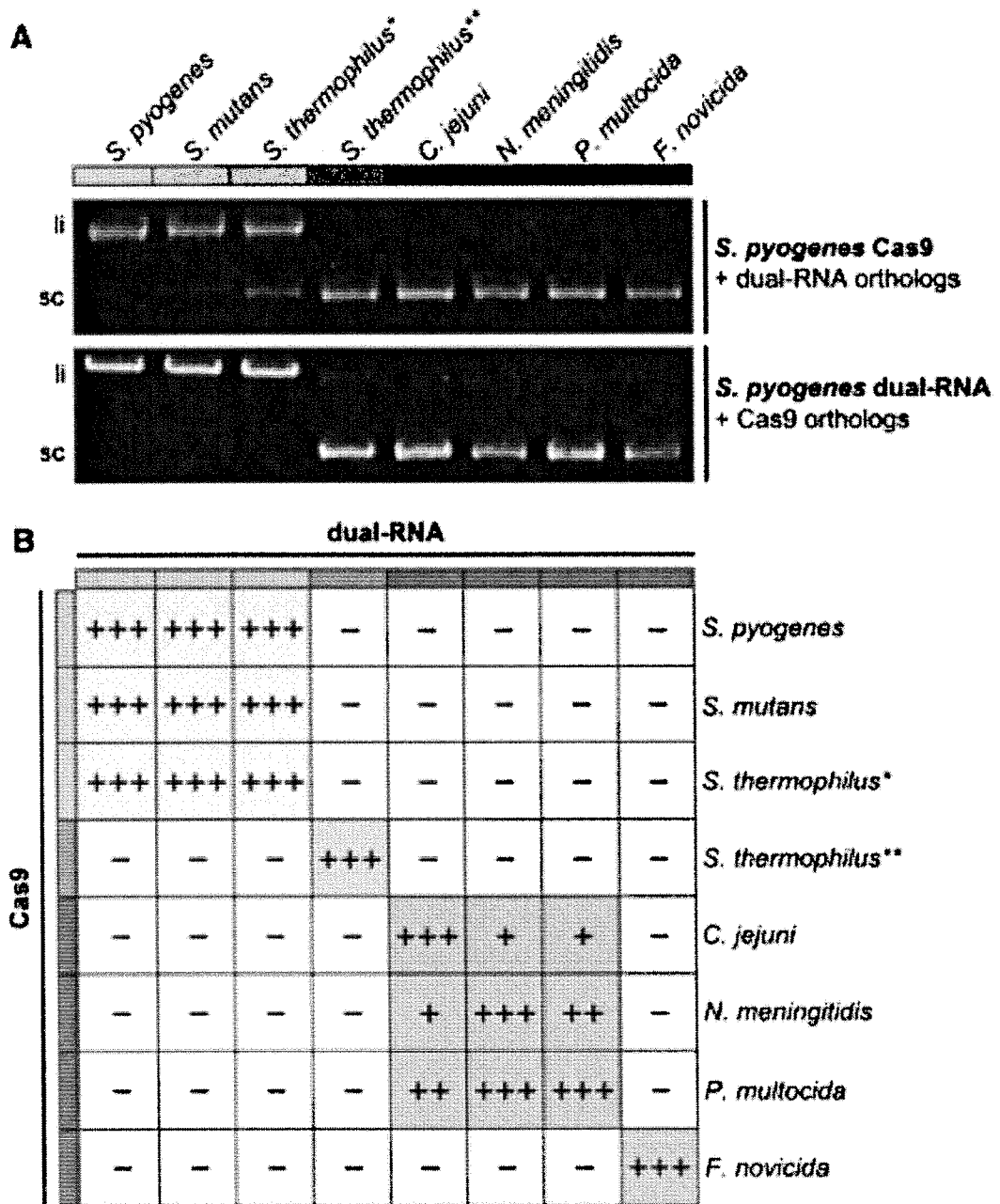
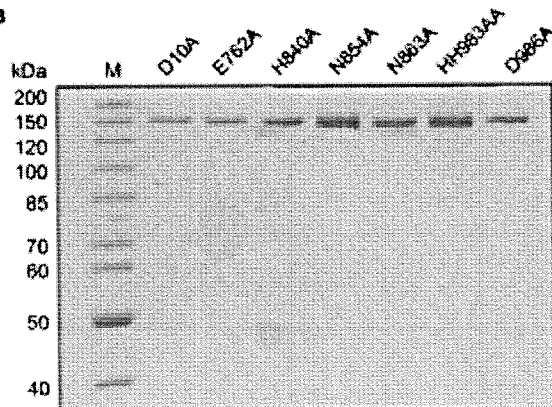


Figure 6

A

Strain <sup>a</sup>	GI accession number <sup>b</sup>	Amino acids (including purification-tag)	Molecular weight [kDa] (including purification-tag)	$\epsilon$ [M <sup>-1</sup> cm <sup>2</sup> ]
<i>S. pyogenes</i> SF370	13622193	1368 (1392)	158.4 (161.2)	110470
<i>S. mutans</i> UA159	24379809	1345 (1369)	156.8 (159.4)	114190
<i>S. thermophilus</i> * LMD-9	116628213	1388 (1412)	161.0 (163.8)	128390
<i>S. thermophilus</i> ** LMD-9	116627542	1121 (1145)	129.4 (132.2)	103860
<i>C. jejuni</i> NCTC 11168	218583121	984 (1008)	114.9 (117.7)	78640
<i>N. meningitidis</i> A Z2491	218767588	1082 (1106)	124.3 (127.1)	101950
<i>P. multocida</i> Pm70	15602992	1056 (1080)	121.8 (124.6)	121460
<i>F. novicida</i> U112	118497352	1629 (1653)	190.4 (193.2)	189080

B



C

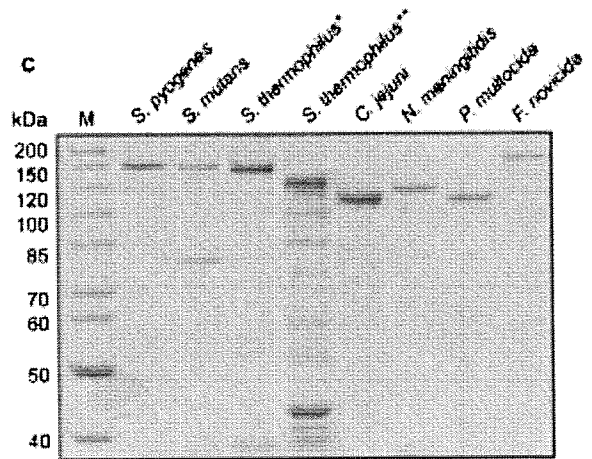


Figure S1



NYRIGLDVGLNSIGFCVAV-----EVDQHD-  
 DYSIGLDMGTSSVGVAVT-----D-----ERGTLL-  
 MYTIGLDIGTNSVGYAVT-----D-----PSYHLI-  
 DYFVGLDIDGINSVGVAVT-----N-----TSYELL-  
 DYIIGFDIGTNSVGVAVT-----D-----LDYNNV-  
 EYIIGLDVGTNSVGVAVT-----D-----SQYNLC-  
 NYIIGLDVGTSSVGVAVT-----D-----TDYNNV-  
 EYFGLDMGTGSLGVAVT-----D-----STYQVM-  
 DYFGLDVGDTGSGVAVT-----D-----TDYKLI-  
 EYIIGLDIGTASVGVAVT-----D-----ESYNIP-  
 DYCIGLDLGTGSGVAVT-----D-----MNHRLM-  
 PYSIGLDIGTNSVGVAVT-----TDNYKVPKMKVLCNTPSK-  
 PYSIGLDIGTNSVGVAVT-----TDDYKVPKMKVLCNTPDK-  
 KYSIGLDIGTNSVGVAVI-----TDEYKVPKMKVLCNTPDR-  
 DYAISLDIGNASVGVSAF-----T-----PNYRLV-  
 DXSVGLDIDGTSSVGVAAI-----D-----NKYHLI-  
 NYSIGLDIGVSSVGVAVV-----T-----EDYRVFAFKFKVLCNTEK-  
 VYVGLDIDGTGSGVAVL-----D-----ENGLLA-  
 QYVIGLDVGTNSVGVAVT-----D-----TSYNLL-  
 PYNIGLDIDGTSSVGVAVT-----N-----DNYDLL-  
 PLYLSLDIDGTGSGVAVT-----D-----KGFNVL-  
 NYTIGLDIGTNSVGVAVI-----D-----ENYKIL-  
 PYGIGLDIGTNSVGVAVV-----D-----ANSHLL-  
 KYIIGLVPVTNSVGVAVT-----D-----EFYNNI-  
 -----EEEE-----EE-----  
 KVVGLDLGLIASVGVCLT-----D-----ISQKED-  
 EYVIGLDLGVSGVAVI-----D-----DN-----  
 KIVIGFDLGVASVGVSTIV-----NA-----  
 NITIGFDLGIASIGWAI-----DS-----  
 KVTLGFDLGIASVGVAVI-----DS-----DK-----  
 MYSIGLDLGISSVGVSVI-----NK-----DY-----  
 DAVLGLDIDGTGSGVGL-----NK-----DL-----  
 RAVLGLDIDGTGSGVGLI-----NK-----DL-----  
 NYVGLDVGIGSISGVAVI-----NI-----QNNRQD-  
 QYRIGLDIGTNSVGVAVL-----NK-----KTDKNG-  
 GYRIGLDVGTGTSVGYAVL-----NK-----AYINKE-  
 KYSIGLDLGIASVGVSVI-----N-----LPKSIQDY-  
 KKIVGLDLGTNSIGWALI-----S-----EAENKD-  
 KKVLGLDIDGTNSIGWALI-----S-----EAENKD-  
 KKILSVLDIGTNSIGWALI-----QET-----GK-----  
 PYRIGLDLGVSGIGFAIVS-----MEE-----MDGNVL-  
 PYSIGLDLGVSGIGVAVA-----A-----YDFLKK-  
 MRVVLGDLGDLGIASIGWALIE-----IEE-----SNRQEL-

Actinomyces coelicoccus DSM 15436#  
 Coriobacterium glomerans PW2  
 Acidaminococcus sp. D21  
 Veillonella atypica ACS-134-V-Col7a  
 Fusobacterium nucleatum ATCC 49256  
 Filifactor alocis ATCC 35896  
 Solobacterium moorei F0204  
 Coprococcus catus GD-7  
 Treponema denticola ATCC 35405  
 Peptoniphilus dherdenii ATCC BRA-1640  
 Catenibacterium mitsuokai DSM 15897  
 Streptococcus thermophilus LMD-9  
 Streptococcus mutans UAL159  
 Streptococcus pyogenes SF370  
 Bifidobacterium bifidum S17  
 Oenococcus kitaharæ DSM 17330  
 Streptococcus sanguinis SK49  
 Fructobacillus fructosus KCTC 3544  
 Eubacterium yuzii ATCC 43715  
 Lactobacillus farciminis KCTC 3681  
 Staphylococcus pseudintermedius ED99  
 Planococcus antarcticus DSM 14505  
 Lactobacillus rhamnosus GG  
 Finegoldia magna ATCC 29328  
 227501312  
 Mycoplasma mobile 163K  
 Mycoplasma gallisepticum str. F  
 Mycoplasma synoviae 53  
 Mycoplasma ovipneumoniae SC01  
 Mycoplasma canis PG 14  
 Eubacterium rectale ATCC 33656  
 Enterococcus faecalis TX0012  
 Streptococcus thermophilus LMD-9  
 Staphylococcus lugdunensis M23590  
 Eubacterium dolichum DSM 3991  
 Ruminococcus albus 8  
 Roseburia inulinivorans DSM 16841  
 Lactobacillus coryniformis KCTC 3535  
 Lijobacterium polytropus DSM 2926  
 Bacteroides sp. 20 3  
 Ignavibacterium album JCM 16511  
 Bacteroides fragilis NCTC 9343  
 Mitratrifractor salunginis DSM 16511  
 Elusimicrobium minutum Pe191  
 Sphaerochaeta globus str. Buddy  
 Methylobacillus trichosporium OB3b

227494853  
 328956315  
 227824983  
 303229466  
 34762592  
 374307738  
 320528778  
 291520705  
 42525843  
 304438954  
 224543312  
 116628213  
 24379809  
 13622193  
 310286728  
 366983953  
 422884106  
 339825081  
 306821691  
 336394882  
 323463801  
 389815359  
 258509199  
 169823755  
 Jnet  
 47458868  
 284931710  
 71894592  
 363542550  
 384393286  
 238924075  
 315149830  
 116627542  
 315659848  
 160915782  
 325677756  
 225377804  
 336393381  
 310780384  
 301311869  
 385811609  
 60683389  
 319957206  
 187250660  
 325972003  
 296446027

Figure S2

---PTPLGTLN-LSVYRHDAIGIDPNGKXT---NTTFLAMSGVARTRRLRFLRKRRLAALDRFIEAQ---GNTLPEADYKDP---  
 ---HFKRKPT-WGSRLEFEAQTA---VARRP-RGQRVYRVRWRKLDLQCLQEQMO-QAEDFFTRLRQOS-RLLEDRAAEHADY---RWFLENDCKPTEK---  
 ---KFKGPEM-WGARVFAQNSA---ERRSF-RTSRRRLDRQQRVKVQVIFAPVISP-IDRFFTRILHES-ALMVEDVAETD---KHIFENDPYTK---  
 ---KPFHKKM-WGSRLEFEQSAV---TRRGF-RSMRRRLRERKRLKLELEFADAMQO-VDSFTFIRLHES-KYHYEDKTTGHSS---KHILFIDEDYDQ---  
 ---RFNKDM-WGSRLEFEAKTA---ERRVQ-WNSRRRLRKRKWRLLLEIFSEIMK-IDSNFFRRLKES-SIMLEDKSKE---KFTLEFDDNYKY---  
 ---KFKKDM-WGIRLFFSANTAK---DRRLO-RGARRRERKRRKIDILLQEFIFESCK-IDFTFFIKLHES-RLHLEDKSDF---KPLFLKEDYSOI---  
 ---KFKRSM-WGARLFEASTAE---ERRTH-RGNRRRLRARKYKILLQEFIFESCK-IDFTFFIRLHES-RLHLEDKSDF---KPLLEFNFNTDK---  
 ---RKHGKAL-WGRLPESASTAE---ERRMP-RTARRRULDRNWRIOVLQEFIFESLTK-VDPFFVRLHES-KYVPEKRAEGCPEL---PVALLVDDNYTK---  
 ---KARADL-WGMRCEFAETA---VRLHL-RGARRLERKRRKILLQEFIFESLTK-VDEGFFQRMKES-PFAEDKTLIQ---ENTLENDKDFADK---  
 ---KFKGKM-WGVRLEFDKATA---ERRTQ-RGSRRLNRKRERINLLQDLAFATLSK-VENFTLRLHNS-DLYREKDKLKS---KYTLFNDKDFKOR---  
 ---KFKGKHL-WGSRLEFSNAETA---NRRAS-RSIRRRYKRRERIRILLRALLQDMVLE-IDPFFTRLEHT-SFLDEEKAKYLGTDYKO---NYNLFIDEDVNY---  
 ---KFKKML-LGVLFFDSGITAE---GRRLK-RTARRRYTRRRNRLLYQEFIFESMGK-VDDSFTRULEDS-FIVTEDKREE---RHPFNGLEEEVVK---  
 ---ESIKKML-IGALLPDSGETAE---ATRLK-RTARRRYTRRKNRILYQEFIFESMGK-VDDSFTRULEES-FIVTEDKREE---RHPFNGIVDEVA---  
 ---RAKGHEL-IGVRLFPDADTA---SRRVA-RTTRRYRNRWRRLRLLDLPQALSE-IDSEFLARRKYS-WVHEDENWADQWY---GSLVEDSNEQDKR---  
 ---RAKSNL-IGVRLPDSAVTAE---KRGY-RTTRRLSRNWRRLRLLNDIFAGPLTDGDNELARLKY-SVHPEQOSQAHFA---AGLEFDSKQDKD---  
 ---EKIKKML-IGSTFVSAQPAK---GTRVF-RVNRRLIDRNRHITVLRDIFQKLEK-VDNKFRRLDESFRVLGDSSELOIK---OPFFGDKLETA---  
 ---RAKGNL-WGVRLEFDKATA---DRRGF-RTTRRLSRNWRRLRLLDELFAELNE-IDSSFFRRLKYS-YVHPEENKAHY---GSLFFTEETKK---  
 ---RAKGNL-WGARLFEKATA---ERRTK-RTSRRSREKAKKALFADALNR-VDESFFTRILEESFTLDRSENWRQY---FLFNDAFTDKD---  
 ---NKKKML-WGVRLEFEAQAK---ETRLN-RSIRRRYKRRKRNINWEIFSEELAK-IDSEFLIRLQNS-WSKKDPDKRDKY---NLFIDGPTDKE---  
 ---KVEDDA-LGVVLFDAALTAQ---ERROF-RTSRRRNRRIKRLGLLOELLAPVQ---NENFY-QORQAMKNDWDFKNSL---SEVLSFGYTES---  
 ---NYNRRHA-FGVEEFESAESA---GRRLK-RGNRRYKRRKRLQQLQELFOSYIT---DSGFFSKYDSQFVWANN-EFENRSL---TEVLSLAISS---  
 ---RLKGETA-IGARLFEQOSAA---DRGSS-RTTRRLSRNWRRLRLLSFLRDFPAPHTK-IDPFFTRKQKYS-EISPKDKRF---KYEKLEFNDRTDA---  
 ---RAKGNL-WGVRLEFDKATA---NTRIF-RSGRRNRDRGMRQJLRFIFEDFELTK-VDKDFYDRLDES-KFWAEDKYSVG---KYSLENDKNSDK---  
 ---E---E---E---E---E---HHH---HHHHHHHHHHHHHHHHHH---HH---HH---H---  
 ---NKFFLIL-LGVRLEFETVDDSDK---LLNFRTRK-RGQRNRRLFRTRKDFIKYLLDN-NI-ILEFDKPKIL-VRNFIKY---INPFSKLELKYK---  
 ---ETNIIHH-LGSRLEFSOAKTAE---DRSP-RGYRLLRERKYLKRFVNLWKNSY-F---GFKKEDILNMYQ---  
 ---ETKEVID-LGVRLESEPEKAD---YRRAK-RTTRRLRKRKSKREKHLKLNVAEI-F---GLQSSNEILNMYK---  
 ---TYSKILD-WGRTFEERTAN---ERRAF-RSTRRLRERKAYNRQFIMLILKYDL-F---ELKNTSDIORANKKOTENYKILISFFEYI---  
 ---ETWQYK-LGSRLEFDAPTNL---ERRTQ-RGTRRLRKRKYNQKFNLV-KRTEV-F---GLSSEALENFRF---  
 ---ESETVIE-AGSRIFPEASADQ---LRDM-RGAKRNRRLKTRINDFKWEMN---MUSIPOP---  
 ---RFGWID-LGVRLEFSKNSKZL---ERRTN-RGRRLLRKRKRRVRLNLEFES---AVGFVEDKSLKNS---  
 ---VYGEIHH-KNSRIFPAQOENL---VRRTN-RGRRLLRKRKRRVRLNLEFES---GLIT---  
 ---ETKNIID-AGVRLPEANVENN---GRSK-RGSRLLRKRRIERLERVKKLEEDY---NLLDQ---  
 ---DESEIVD-YGVRLEKEGHAENE---TRTK-RGRRLLRKRRRVREDMLELQAA---GLIS---  
 ---EKKRIED-FVVRIFKSGEIQEKNSN---SRASQCHRS-RGLRLLRKRKRRKRLKLNLSLI---GLTSEKIDYVEE---  
 ---EVRITD-LGVRIFDVAENFKNG---DALAAPRDA-RTTRRLRKRKRRERIEKFLQENG---LHESMRERYK---  
 ---LRYKILT-LDSVIVPRAENPQTG---ASLAEPRIK-RGLRRLRKRKTRKQRTQQLFHS---GLLSKPELEQILA---  
 ---KDERIED-WGRTIFKQAINPEDG---SSLASRREK-RGSRRLRKRKRRERIEKFLQENG---GAVKNETEIKYK---  
 ---ELYGIEA-CGSRILPMDAAILGNEFDKNS---ISQADRTSY-RGIRLLRERHLLRERHLLDGLL-GF-LPHYSDSLNRY-GKFLNDIEC---KLPWKVDETSYKFIQESFKEMLANPTEHPLIAN---  
 ---GKGRLEW-LTGRVPLDADYMAFIDGKGLPVQ-ITPAKRRQK-RGSRLLRKRKRRERIEKFLQENG-ITW-LPDEFILMPKRIKETSIPGKFSRISD---YVPISDSYRFFYFEGFPENEIEQVLEEINFARKTK---  
 ---ERSSIVK-LGVRNPLIVDELTPNEKGS---ITWADRTLK-RGNRRLRKRKRRERIEKFLQENG---KLITDETIJSE---  
 ---DLRYCLD-NSVVMKNNPVDKSG---ESSOSTISTQ-KMRRLRKRKRRERIEKFLQENG---GLDYSEFMKIND---  
 ---LPEKELM-VGSRIFKASGAA---DRKLS-RGQRNRNRTRERMYLWVLAEQ-KL---ALPVPADLDRKEN---SSGEETSARR---  
 ---QPSDILVF-VSRIFIPSTIGAA---ERRQK-RGQRNGLRFRANRKLFLAKLAER-NL---EQDVPDARLRF---  
 ---SQGTIIG-AGTWMEFAFEKTOAGA---KLKSEQRRTF-RGQRVYRVRWRKRRVRLNLEFES---MLSYS---  
 ---SQGTIIG-AGTWMEFAFEKTOAGA---KLKSEQRRTF-RGQRVYRVRWRKRRVRLNLEFES---GLLPSRDLAKQ---

Figure S2 (Continued)

347536497	<i>Flavobacterium branchiophillum</i> FL-15	AKILGLDLGTFNSIGRAV-----VERENI
345885718	<i>Prevotella</i> sp. C561	QKVLGLDLGTFNSIGSAV-----RNLDSL
282880052	<i>Prevotella timonensis</i> CRIS 5C-B1	KRILGLDLGTFNSIGRAV-----VMDDEHA
312879015	<i>Aminomona peucivora</i> DSM 12260	-----MIGHRVGGCLF--D-----DHWTFN
294086111	<i>Candidatus Puniceispirillum marzinum</i> IMCC1322	MRRLGLDLGTFNSIGWCLL-----DLGDDG
330822845	<i>Alicyclophillus denitrificans</i> K601	KYRLALDLGTFNSIGWALF-----RLDACN
344171927	<i>Ralstonia syzygii</i> R24	QHRWELDLGTFNSIGRAV-----ALIEG
159042956	<i>Dinoroseobacter shibae</i> DFL 12	-MRGLDLGTFNSIGWMLY--ETD-----GAGSDA
83591793	<i>Rhodospirillum rubrum</i> ATCC 11170	FWILGLDLGTFNSIGRAVF--SCE-----EKGPP
288957741	<i>Azospirillum</i> sp. B510	PYRLSFDLGFNSIGWGLL-----NLDROG
427429481	<i>Caenispirillum salinarum</i> AK4	RWSLALDIGERSIGRAVA-----EYDAEG
92109262	<i>Nitrobacter hamburgensis</i> X14	LYRLGLDLGTFNSIGWTFV--HLE-----KRGDRH
148255343	<i>Bradyrhizobium</i> sp. BTall	AYRLGVDLGANSISGNFVW--HLD-----DHG
34557790	<i>Wolinella succinogenes</i> DSM 1740	ERILGVDDLGISSIGWAIV--E-----YDKDDE
218563121	<i>Campylobacter jejuni</i> NCTC 11168	ARILAFDGISSIGRAFS-----E-----E
Unet	218563121	-EEEEEE-----EEEEEE
291276265	<i>Helicobacter mustelae</i> 12198	IRTLGLDLGTFNSIGRAV--EGE-----YTDKGL
222109285	<i>Acidovorax ebreus</i> TPSY	QHVFLDLGTFNSIGWAIL-----NL
365156657	<i>Bacillus smithii</i> 7 3 47FA	NYKMLDLGTFNSIGRAV-----DK
220930482	<i>Clostridium cellulolyticum</i> H10	EYTLGLDLGTFNSIGWAIL-----DYSSTQ
297182908	uncultured delta proteobact. HF0070 07E19	ERIFGFDIGTFNSIGFSVI-----EID-ED
154250555	<i>Parvibaculum lavamentivorans</i> DS-1	NYILGLDLGTFNSIGRAVW-----EIN-EN
218767588	<i>Neisseria meningitidis</i> Z2491	SLTFSPDGIYASIGRAVI-----ASASHDD
15602992	<i>Pasteurella multocida</i> str. Pm70	RYRVGIDVGRSISGFCAV-----EVDDED
187736489	<i>Akkermansia muciniphila</i> ATCC BAA-835	TRVLGVGVGRSISGLAAV-----SPEED
315605738	<i>Actinomyces</i> sp. oral taxon 180 str. F0310	RYRIGIDVGNISVGLAAV-----EVSDEN
117929158	<i>Acidothermus cellulolyticus</i> 11B	AYRLGLDLGTFNSIGRAV-----ALEKDESG
189440764	<i>Bifidobacterium longum</i> DJO10A	KYRVGLDLGTFNSIGRAV-----SMDEQ
403744858	<i>Alicyclobacillus hesperidum</i> URH17-3-68	RYRVGLDLGTFNSIGRAV-----ERNLE
407803669	<i>Alcanivorax</i> sp. W11-5	KHILGLDLGTFNSIGWALI-----AIDAQ
423317190	<i>Bergeyella zoohelcum</i> ATCC 43767	KHVLGLDLGTFNSIGWALI-----RENS
402847315	<i>Porphyromonas</i> sp. oral taxon 279 str. F0450	KNILGLDLGTFNSIGRAV-----DQ
404487228	<i>Barnesiella intestinihominis</i> YIT 11860	ETTLGLDLGTFNSIGLAIV-----SIDKD
374384763	<i>Odoribacter lanens</i> YIT 12061	KWRLGLDLGTFNSIGNSVF-----RTGPGVFGF
384109266	<i>Treponema</i> sp. JCA	EEEEEE-----EEEEEE
402849997	<i>Rhodovulum</i> sp. PH10	IIGVGLDLGTFNSIGWTFIT-----SHPDSE
Unet	331001027	VSPISVDLGGKNTGFFSF-----T
331001027	<i>Parasutterella excrementihominis</i> YIT 11859	ISPICIDLGGKFTGVCIS-----HLEFAFEL
34557932	<i>Wolinella succinogenes</i> DSM 1740	SCSISIDMGAKYTGTFYA-----LEDRE-EL
54296138	<i>Legionella pneumophila</i> str. Paris	ISPIAIDLGAFTGVALY-----OYLEGA
319941583	<i>Sutterella wadsworthensis</i> 3 1 45B	ILPIAIDLGVKNTGVFSA-----FYQKGTSL
254447899	gamma proteobact. HTCC5015	-----E-----E
118497352	<i>Francisella novicida</i> UL12	-----E-----E
Motifs		-----E-----E
informative positions		-----E-----E

RuvC\_I  
 \*\*\*\*\*

Figure S2 (Continued)



227494853	<i>Actinomyces coleocanis</i> DSM 15436#	---YTFWL---VRAKLAQTFP---
328956315	<i>Coriobacterium glomerans</i> FW2	---DYIOREFTLYH---VR-SWLM---
227824983	<i>Acidaminococcus</i> sp. D21	---EYSDYPTIEH---LI-VDIM---
303229466	<i>Veillonella atypica</i> ACS-134-V-Col7a	---DFTEYPTIYH---LR-KDL---
34762592	<i>Fusobacterium nucleatum</i> ATCC 49256	---DFYQYPTIFH---LR-DELI---
374307738	<i>Filifactor slocis</i> ATCC 35896	---EYKKEFTIIE---LR-KHLI---
320528778	<i>Solobacterium moorei</i> F0204	---DYLKRYPTIYH---LR-SDLI---
291520705	<i>Coprococcus catus</i> GD-7	---NYKDYPTIYH---LR-KMLM---
42525843	<i>Treponema denticola</i> ATCC 35405	---TYHKAYPTIHS---LI-KAWI---
304438954	<i>Peptoniphilus duerdenii</i> ATCC BAA-1640	---DYKRYPTIIEH---LI-MDLI---
224543312	<i>Catenibacterium mitoukai</i> DSM 15897	---TYHKYPTIYH---LR-KALC---
116628213	<i>Streptococcus thermophilus</i> IMD-9	---YEDDEPTIYH---LR-KYLA---
24379809	<i>Streptococcus mutans</i> UA159	---YHENFTIYH---LR-QYLA---
13622193	<i>Streptococcus pyogenes</i> SF370	---YHEKYPYIYH---LR-KKLV---
310286728	<i>Bifidobacterium bifidum</i> S17	---FYKRYPTIYH---LR-KALM---
365983953	<i>Oenococcus oenas</i> DSM 17330	---FYKRYPTIYH---LR-LALM---
422884106	<i>Streptococcus sanguinis</i> SK49	---FYKRYPTIYH---LR-KHLA---
339625081	<i>Fructobacillus fructosus</i> KCTC 3544	---FERSYPTIYH---LR-QELM---
308821691	<i>Eubacterium yurii</i> ATCC 43715	---YYSKYPTIYH---LR-SALI---
336394882	<i>Lactobacillus farciminis</i> KCTC 3681	---YREKFTIYH---LR-KALI---
323463801	<i>Eubacterium pseudintermedium</i> ED99	---NKYPTIYH---LQ-EALI---
389815359	<i>Planococcus antarcticus</i> DSM 14505	---RFYEDYPSMYH---LR-LHLM---
259509199	<i>Lactobacillus rhamnosus</i> GG	---QYFKKFTIYH---LR-KYLM---
169823755	<i>Fingoldia magna</i> ATCC 29328	---HHH---HHH---HHH---HHH---
Unset	227501312	---SVTALFIGFEN---LR-KAAI---
47458868	<i>Mycoplasma mobile</i> 163K	---EOKLENTVLM---LK-SEAL---
284931710	<i>Mycoplasma gallisepticum</i> str. F	---DQSSKRYNIAK---LK-IMAL---
71894592	<i>Mycoplasma synoviae</i> 53	---KKAARKSNMILS---VK-VKAI---
363542550	<i>Mycoplasma ovipneumoniae</i> SC01	---ELSKRYPAIE---LK-TKAL---
384393286	<i>Mycoplasma canis</i> PG 14	---KSTKIVG---LK-VRAI---
238924075	<i>Eubacterium rectale</i> ATCC 33656	---CPYQ---LR-VAGL---
315149830	<i>Enterococcus faecalis</i> TX0012	---DFTKISIMLNFYQ---LR-VKGL---
116627542	<i>Streptococcus thermophilus</i> IMD-9	---SQIFQSTFPIA---LR-VKGL---
315659848	<i>Staphylococcus lugdunensis</i> M23590	---TSFHLNPFYD---VA-VAGL---
160915782	<i>Eubacterium dolichum</i> DSM 3991	---TADNRFVIG---LR-MKGL---
325677756	<i>Ruminococcus albus</i> 8	---GNLFDVIG---LR-YEGL---
223377804	<i>Roseburia inulinivorans</i> DSM 16841	---FFQAKYSVYE---LR-VAGL---
336393381	<i>Lactobacillus coryniformis</i> KCTC 3535	---NAYLESFWE---LR-ANSL---
310780384	<i>Lilybacter polytropus</i> DSM 2926	---NKAVFYDWTIYI---LR-KKAL---
301311859	<i>Bacteroides</i> sp. 20 3	---LPEWVVYI---LR-KKAI---
389811609	<i>Ignavibacterium albus</i> JCM 16511	---NGRUTTFETIR---LR-AKAV---
60683389	<i>Bacteroides fragilis</i> NCTC 9343	---FAMNFIKRNWQ---LR-AVDAM---
319957206	<i>Nitratifactor salunginis</i> DSM 16511	---FLGDVIGQDIYE---LR-VKSL---
187250660	<i>Elusimicrobium minutum</i> Feil191	---EDAVVRANPFYE---LR-LAGL---
325972003	<i>Sphaerochaeta globus</i> str. Buddy	---FGLDFWR---LR-AEAL---
296446027	<i>Methylosinus trichosporium</i> CBS3b	

Figure S2 (Continued)

```

--IRDEN--DLH--EKLAVRHLIARHRCWRSFVVP
--ETDEQ--ADI--RUYVALAHNIIVHRGNFLREGD
--ESSEK--HDP--RUYVALAVAMVAHRGHFLNEVD
--MENGU--DDI--RULFLAVHHLIKYRGNFLYEGA
--KNPEK--KDI--RAYVALAHSIIFKSRGHFLFEGQ
--ESSEK--QDI--RAYVALAHNIILKTRGHFLLDGD
--BNSTE--HDI--RUYVALAHHLIKYRGHFLYDNS
--ETTEI--FDI--RUYVYVLEHMKRCHFLFLSGD
--ENRVA--PDP--RAYVALACHNIILKTRGHFLFEGD
--EDEGK--KDI--RUYVALACHYLLANRGHFLFEGQ
--ESTEK--ADP--RAYVALAHHIVKVRGNFLYEGQ
--DSTYK--ADL--RUYVALAHMLIKYRGHFLIEGK
--DNPEK--VDL--RUYVALAHILIKRCHFLIEGK
--DSTDK--ADI--RAYVALAHMLIKRCHFLIEGD
--EDDSQ--HDI--REYVALAHBNVYKRGHFLVEGT
--NDQK--HDI--RUV--HLVYKRGHFLIEGD
--DANSFPADI--REVVALSHILIKYRGHFLLDK
--QFNNK--FDI--REYVALAHHLIKYRGHFLSSQK
--NSDEK--FDV--RUYVALAILHLFHRGHFLNASE
--LNDK--ADI--RAYVALAHNIILKTRGHFLYEQ
--LADK--FDP--ELIYVALYHLVYKRGHFLFDEL
--ESMK--MDL--RUYVALAHNIILKTRGHFLFQGM
--THYK--ADP--REYVALAHHLIKRGHFLFPGA
--ESEK--VDI--RYZVALAKOMKRGHFLFDGQ
--H--H--HHHHHHHHHH
--MEYK--LAK--SELVALYVYLSLRGAFVFNIPK
--NAN--IDP--KALSWILLEDYLANRGHFLYEDAK
--KEE--IKP--SELVALMILRDYQKRGYFYFN
--DSK--IEK--LQDLWILLEDYLANRGHFLYLESE
--SOE--VCP--DELAWILLEDYLANRGHFLYDEK
--TPE--ITL--DEYVALIYSLKRGHFLYLEDALDDI
--TLP--LSR--GELVALYVYLSLRGAFVFNIDSV
--TBE--LSH--SELVALAKMVRGHFLYLDAS
--SEA--LSK--DEYVALAHMLIKRGIHIDVIDS
--NER--LNG--SELVALAHMLIKRGSVETIEDEAK
--SEK--LTP--SELVALAHMLIKRGSVETIEVVDIED
--DRK--LKD--SELVALAHMLIKRGSVETIEDEAK
--DRR--LWV--SELVALYVYLSLRGAFVFNIDSV
--EAK--ISH--SELVALAHMLIKRGSVETIEDEAK
--FOK--ISK--SELVALAHMLIKRGSVETIEDEAK
--LKP--TYK--SELVALAHMLIKRGSVETIEDEAK
--TBE--ISL--SELVALAHMLIKRGSVETIEDEAK
--KRP--LSP--SELVALAHMLIKRGSVETIEDEAK
--DER--LSL--SELVALAHMLIKRGSVETIEDEAK
--NEQ--LTL--SELVALAHMLIKRGSVETIEDEAK
--DRI--LGP--VELVALGELIARHRCWRSFVFNIDSV

```

Figure S2 (Continued)

295446027	<i>Methylophilus trichosporium</i> QB3b	-----
347536897	<i>Flavobacterium branchiophilum</i> FL-15	-----
345885718	<i>Prevotella</i> sp. C561	-----
28280052	<i>Prevotella timonensis</i> CRIS 5C-81	-----
312879015	<i>Aminomonas paucivorans</i> DSM 12260	-----
294086111	<i>Candidatus Funticispirillum marinum</i> IMCC1322	-----
330822845	<i>Alicyclophilus denitrificans</i> K601	-----
344171927	<i>Ralstonia snyderii</i> R24	-----
159042956	<i>Dinoroseobacter shibae</i> DEL 12	-----
83591793	<i>Rhodospirillum rubrum</i> ATCC 11170	-----
28957741	<i>Azospirillum</i> sp. B510	-----
427429481	<i>Casnispirillum salinarum</i> AK4	-----
92109262	<i>Nitrobacter hamburgensis</i> X14	-----
148255343	<i>Bradyrhizobium</i> sp. BT411	-----
34557790	<i>Molinitella succinogenes</i> DSM 1740	-----
218563121	<i>Campylobacter jejuni</i> WCFC 11168	-----
	<i>Ureth</i>	-----
291276265	<i>Halicobacter mustelae</i> 12198	-----
222109285	<i>Acidovorax ebreus</i> TFSY	-----
365156657	<i>Bacillus smithii</i> 7 3 47FA	-----
220930482	<i>Clostridium cellulolyticum</i> H10	-----
297182908	uncultured delta proteobact. HF0070 07E19	-----
154250555	<i>Parvibaculum lavamentivorans</i> DS-1	-----
218767588	<i>Neisseria meningitidis</i> Z2491	-----
15602992	<i>Pasteurella multocida</i> str. Pm70	-----
187736489	<i>Akkermansia muciniphila</i> ATCC BAA-835	-----
315606738	<i>Actinomyces</i> sp. oral taxon 180 str. F0310	-----
117929158	<i>Acidothermus cellulolyticus</i> 11B	-----
189440764	<i>Bifidobacterium longum</i> DJO10A	-----
403744858	<i>Alicyclobacillus hesperidum</i> URH17-3-58	-----
407803669	<i>Alicyclovax</i> sp. W11-5	-----
423317190	<i>Bergeyella zoohelcum</i> ATCC 43767	-----
402847315	<i>Periphyromonas</i> sp. oral taxon 279 str. F0450	-----
404487228	<i>Barnesiella intestinihominis</i> YIT 11860	-----
374384763	<i>Odoribacter laneus</i> YIT 12061	-----
384109266	<i>Tropozema</i> sp. Jc4	-----
402849997	<i>Rhodovulum</i> sp. PH10	-----
	<i>Ureth</i>	-----
331001027	<i>Parasutterella excrementihominis</i> YIT 11859	-----
34557932	<i>Molinitella succinogenes</i> DSM 1740	-----
54296138	<i>Legionella pneumophila</i> str. Paris	-----
319941583	<i>Sutterella wadsworthensis</i> 3 1 458	-----
294447899	gamma proteobact. HTCC5015	-----
118497352	<i>Francisella novicida</i> U112	-----
	<i>Motifs</i>	-----
	<i>Informative positions</i>	-----

Figure S2 (Continued)

```

---KHK---VSL---FELGRALFHLNQRGFLSNLLDQASGILIEHCPLKIAIVEDLISIDELSTNLTDPYFEGILD
---TRQ---FDTEOPLER---YKLRALYELAQHGFSSKGGKTLSCQETNSPFSYTD
---HKK---LDLTFVADR---YTLGRALYELAQHGFSSKGGKTLSCQETNSPFSYTD
---DRK---LSF---FENVVLYELTKRGFQS
---DEA---IDF---YEMGRALFHLNQRGFKS
---QRA---LLP---GEFARALFHLNQRGFKS
---DQA---LTL---PEFARALFHLNQRGFKS
---DEF---LPL---FELGRALFHLNQRGFKS
---SRF---LAP---DALWALALHLNQRGFKS
---RRR---LEP---FELGRALFHLNQRGFKS
---RRF---LDG---PELKVLSHMAHARGTRALAEIQ
---YDA---LPA---HVVGRALFHLNQRGFKS
---DEY---LPL---HVVGRALFHLNQRGFLA
---YRL---LKG---DELARVLHLNQRGFKS
---MEL---LSK---QDFARVLHLNQRGYDD
---HH---HHHHHHHHHH
---YRV---LDK---FELARVLHLNQRGYDD
---DRJ---LTP---LEMARVYHCKCKRGFEM
---ERR---LNN---DELARVLHLNQRGFKS
---SRI---LKP---YELAVLHLNQRGFKS
---ERR---LTK---QKLAVALFHLVNRGTFP
---EEG---LSA---YEFGRALYELAQHGFAG
---DRK---LTP---LENSAVLHLNQRGYLS
---ERR---LSA---LENSAVLHLNQRGYLS
---EGHRT---LAP---LELMEVLSMAYANRGYIN
---LEDET---ERR---CCLSVAMAHARHCWMSFSK
---VYDET---ERR---KLLGVAVSHMAHARGCWMSPTT
---IEDDE---LRR---ESIATALRMAHARGWMS
---RC---LTA---FELARVLHLNQRGFKS
---ER---LEL---COLARVLNQRGFKS
---ER---VGL---EDLGRITLYKMQLRGAGGSLRPERKEDIPEEQSDKKNRSLFASKIVLGRQPELIFK
---RR---LTL---FELGRVLCHELNQRGYRS
---QR---LEL---MELGRVLSHMLNQRGYKS
---ED---VTR---FELGRILYOMQRGFLS
---EK---LEP---YELGRALFHLNQRGFKS
---ES---LTP---HEMGRVIFHLNQRGFKS
---HH---HHHHHHHHHH---EEZ---E
---APEE---REMLRALSCYLAKRGYARTAE---FDTVSLDELDSVSSASFTFTFN---DSEFLNIGMEAIANSFE---HHHHHHHH---HH
---TDEE---MARETALCHYLANRGYTYVYDD---LDEKKDALSDDL---KELSRKLSQSIDRSDVDEFLQIAS---TTKALN---KELSGOE
---EKV---MKGREALSGELNQRGSRPWADG---IDEXYKDKETTLLL---KELLS---ESEHNFIDM---FLQKMQ---DYKNGF
---DKDTQOAISETLNRGFSYI---TEPLAKFINGLAKRGYTYI---SEEVDESMNVSLPFSMMPDFYFN---SSAPLEQLAKLSDEN---NVEKTL---GDPNIPA
---DKDTQOAISETLNRGFSYI---TDCYSBYVLIYVPEQVKALINDIFDNYNGEDDLDLSYKLAPEQSKISEIYNKLMQKILEFKMLCLTDIKDKKVSFKTLX---KLVYER---AEGKIPS

```

Figure S2 (Continued)



227494853	<i>Actinomyces collocanis</i> DSM 15436#	VRSLRVQPPR
328956315	<i>Coriobacterium glomerans</i> PW2	SLSAKARPKALNHLRETRLVSSRGRFECSTADNGSLAMLTHTPDISPS
227824983	<i>Acidaminococcus</i> sp. D21	KDMIGVLSFDAPYFEFLAFTLUNGVSFWVCSKALQATLISRRNSVMD
303229466	<i>Veillonella atypica</i> ACS-134-V-Col17a	TFNSMAF--TFEDVLQAVNITENCFTDMSATSSISNLMESGKTKSKAKATERLV
34762592	<i>Fusobacterium nucleatum</i> ATCC 49256	NIKKEIKPETHYNNIISPLEDNGINKSIDKDNIEKLEKLIICDGGKIGDM
374307738	<i>Filifactor allocis</i> ATCC 35896	IQSAKQRPILDTFLSISQEQNLVSLSENQDEYEFELKMRSTAKSEK
320528778	<i>Solobacterium moorei</i> F0204	ANGVKTLDKAVSDFEELNENDISENIEKKEEIVLSDKHITREKKI
291520705	<i>Coprococcus catus</i> GD-7	ISQIKKSKSTFQQLQNICQKELKWHISLDDAAIQFVHVLKDRNITRS
42525843	<i>Treponema denticola</i> ATCC 35405	FDSEMFDSIQALFEYREMEVDIDADSQKVEIKDSSLENSKQSR
304338954	<i>Peptoniphilus churdenii</i> ATCC BAA-1640	KFDKNSFDKSINDLKHLEDEYNDLISFNNEDLIEITDTLANKINKKELNIVG
224543312	<i>Catenibacterium mitsukoi</i> DSM 15897	KFNHDSNIEKISDIFTQFTSENNIPTYEDDEKKNLEILELAKPISKKAQVDEVTI
116628213	<i>Streptococcus thermophilus</i> IMD-9	FNSRNDIQKNFQDFLTYNALSFDLSLENSAQLEEVKDKISKLEKLU
24379809	<i>Streptococcus mutans</i> UA159	FDTRNDVQRLEFQFLAVYDITFENSISLQFNQVVEITDKISKAKC
13622193	<i>Streptococcus pyogenes</i> SF370	INPNSVDKLFQDVVYTYQLFENPINSAGVDKAILERLUSKERRLE
310286728	<i>Bifidobacterium bifidum</i> S17	LESSMAFKEDELLKILGRITRYEMSEGEQNSDIDEDENKLVAPANGQADALCATR
366983953	<i>Ornithobacterium kitaharae</i> DSM 17330	VKADSAF--DVHTFADAIQRYAESNNSDENLGGKIDKDKLSAALDKGSSASGRAE
422884106	<i>Streptococcus sanguinis</i> SK49	INP--NNINMNSMIDET--ESQVDFKLSIDES--KMLADIFKRSSE
339625091	<i>Fructobacillus fructosus</i> KCTC 3544	KIT-IGSTYNPEDLANAI--EYVADKGGLSWELMNPQOL--TEIISGEAGYG
306821691	<i>Eubacterium yurii</i> ATCC 43715	AGD--GDIQGDVYFM--DIVESECFEIELEPRTINI--DNEKILSQK
336394882	<i>Lactobacillus farciminis</i> KCTC 3681	KFN--ISMLNNLSKRAL--ELMQLIKYDILSFPDCCDM--NHLISDILIGRG
323463801	<i>Staphylococcus pseudintermedius</i> ED99	KLEMLNKNWHEDEVELI--EYENLNNIKMLDYKET--KVIYELIADNE
389913359	<i>Planococcus antarcticus</i> DSM 14505	WSEASAKEDDOLLELV--TYVAELKLSPLDLSSESON--KAATLILMNRN
258509199	<i>Lactobacillus rhamnosus</i> GG	AKDFNTKVDLEDFPALTAAYAVYDIELEGFDLAKADDFKALLDEQATPSDTOR
169823755	<i>Finergoldia magna</i> ATCC 29328	ISEVTDKPKAEQLILLINDLKLKLEZELMDSIFELLADVWNEKRTDKNNIKLELII
Unet	227501312	-----BHHHHHHHHHHHH-----HSHHH--BHHHH-----BHHHHHHHHHHHH
47458868	<i>Mycoplasma mobile</i> 163K	-----
284931710	<i>Mycoplasma gallisepticum</i> str. F	-----
71894592	<i>Mycoplasma synoviae</i> 53	-----
363542550	<i>Mycoplasma ovipneumoniae</i> SC01	-----
384393286	<i>Mycoplasma canis</i> PG 14	-----
23824075	<i>Eubacterium rectale</i> ATCC 33656	-----
315149830	<i>Enterococcus faecalis</i> TX0012	-----
116627542	<i>Streptococcus thermophilus</i> IMD-9	-----
315659848	<i>Staphylococcus lugdunensis</i> M23590	-----
160915782	<i>Subacterium dolichum</i> DSM 3991	-----
325677756	<i>Ruminococcus albus</i> 8	-----
225377804	<i>Roseburia inulinivorans</i> DSM 16841	-----
336393381	<i>Lactobacillus coryniformis</i> KCTC 3535	-----
310780384	<i>Illybacter polytropus</i> DSM 2926	-----
301311889	<i>Bacteroides</i> sp. 20 3	-----
385811609	<i>Ignavibacterium album</i> JCM 16511	-----
60683389	<i>Bacteroides fragilis</i> NCTC 9343	-----
319957206	<i>Nitratifactor salsauginis</i> DSM 16511	-----
187250660	<i>Elusimicrobium minutum</i> Peil191	-----
325972003	<i>Sphaerochaeta globus</i> str. Buddy	-----

Figure S2 (Continued)



296446027	<i>Methylosinus trichosporium</i> OR3b	-----
347536497	<i>Flavobacterium branchiophilum</i> FL-15	-----
345885718	<i>Prevotella</i> sp. C561	-----
282880052	<i>Prevotella timonensis</i> CRIS 5C-B1	-----
312879015	<i>Aminomonas paucivorans</i> DSM 12260	-----
294086111	<i>Candidatus Paucispirillum marinum</i> IMCC1322	-----
330822845	<i>Alicyclophilius denitrificans</i> K601	-----
344171927	<i>Ralstonia sylvatica</i> N24	-----
159042956	<i>Rhodospirillum rubrum</i> DFL 12	-----
83591793	<i>Rhodospirillum rubrum</i> ATCC 11170	-----
288957741	<i>Azospirillum</i> sp. B510	-----
427429481	<i>Caenospirillum salinarum</i> AK4	-----
92109262	<i>Nitrobacter hamburgensis</i> X14	-----
14825343	<i>Bradyrhizobium</i> sp. BTM11	-----
34557790	<i>Molinitella succinogenes</i> DSM 1740	-----
218563121	<i>Campylobacter jejuni</i> NCTC 11168	-----
	<i>Janet</i>	-----
291276265	<i>Helicobacter mustalae</i> 12198	-----
222109285	<i>Acidovorax ebreus</i> TPSY	-----
365156657	<i>Bacillus smithii</i> 7 3 ATFAA	-----
220930482	<i>Clostridium cellulolyticum</i> H10	-----
297182908	uncultured delta proteobact. HF0070 07819	-----
154250555	<i>Parvibaculum lavamentivorans</i> DS-1	-----
218767588	<i>Neisseria meningitidis</i> Z2491	-----
15602992	<i>Pasteurella maltocida</i> str. Fm70	-----
187736489	<i>Akkermansia muciniphila</i> ATCC BAA-835	-----
315605738	<i>Actinomyces</i> sp. oral taxon 180 str. F0310	-----
117929158	<i>Acidothermus cellulolyticus</i> 118	-----
189440764	<i>Bifidobacterium longum</i> DJ010A	-----
403744858	<i>Alicyclobacillus hesperidum</i> URH17-3-68	-----
407803669	<i>Alicyclobacillus</i> sp. W11-5	-----
423317190	<i>Bergeyella soehelcum</i> ATCC 43767	-----
402847315	<i>Porphyromonas</i> sp. oral taxon 279 str. F0450	-----
404487228	<i>Barnesiella intestinihominis</i> YIT 11860	-----
374384763	<i>Odoribacter lanens</i> YIT 12061	-----
384109266	<i>Treponema</i> sp. JC4	-----
402849997	<i>Rhodovulum</i> sp. PH10	-----
	<i>Janet</i>	-----
331001027	<i>Parasutterella excrementihominis</i> YIT 11859	-----
34557932	<i>Molinitella succinogenes</i> DSM 1740	-----
54296138	<i>Legionella pneumophila</i> str. Paris	-----
319941583	<i>Sutterella wadsworthensis</i> 3 1 45B	-----
254447899	gamma proteobact. HTCC5015	-----
118497352	<i>Francisella novicida</i> U112	-----
	<i>Motifs</i>	-----
	informative positions	-----

Figure S2 (Continued)



227494853	<i>Actinomyces coleocanis</i> DSM 154364	LRFRREKTDTPENKPKCE
328956315	<i>Coriobacterium glomerans</i> FW2	YRIGEDLALAKVAVIYAPDQIENFFSCTATPGIYDAQAQGYATNML
227824983	<i>Acidaminococcus</i> sp. D21	YQEHHDHFOQAYFYVYLAKEYDDIFRANVDSYTTWYVYSYVEVAGTFLP
303229466	<i>Veillonella atypica</i> ACS-134-V-Col7a	FWRKHEHIVLAKSLAKLURVYVINEFASDKAGLRVYVYIQGCTE
34762592	<i>Fusobacterium nucleatum</i> ATCC 49256	YSEHKDHEHLYTTRKYKNEYDELFRDNNYYPAYIGLREKQ
374307738	<i>Filifactor aloocis</i> ATCC 35896	YEHANLKLADILLAYCDEYENMEDEKRSAGSYAVVYGLKMK
320528778	<i>Solobacterium moorei</i> F0204	FNKHDMHLYKTRHEHDEYKVIFFSPTTEKRYAAVSGTNS
291520705	<i>Coprococcus catus</i> GD-7	YQHQADLAKVYQYKEDYKVVYVDFRKLNNYSVIGCTKIKNGKVVDA
42525843	<i>Treponema denticola</i> ATCC 35405	YRKHDTHTKLVVDFRPFQYKRVFGYMKRKNMNYSGVGE
304438954	<i>Peptoniphilus dwardenii</i> ATCC BAA-1640	FRRHGHATKTRILIKKYLSPKANLFRKSNIDNYVAYSNITSNKTKK
224543312	<i>Catenibacterium mitisookai</i> DSM 15897	YNHHDHAKQDCIDNNVFNKYTFMFRDSESGEYTYLNR
116628213	<i>Streptococcus thermophilus</i> IMD-9	YNEHAKHLAKKEYIRNLSLKYNEVFRDPTANGIAGYIDGATGSDFFVYL
24379809	<i>Streptococcus mutans</i> UA159	YNEHQDIAQMLQFTRQSLDXYNEVSDVSDGYLSDYIDGKTWQAFYVL
13622193	<i>Streptococcus pyogenes</i> SF370	YDEHRQDITLKAUVQQLRERKVKHIFPQSKNGIAGYIDGAGSERFYAFI
310286728	<i>Bifidobacterium bifidum</i> S17	YAHRAAMNLIKIQLRTEKEDIDENYGLVAGKIDS
366983953	<i>Oenococcus oeni</i> DSM 17330	FYEHQADWEYIASHIRAKN---ACNGHYEKSRKFTG
422884106	<i>Streptococcus sanguinis</i> SK49	YFERRDIOREK-FFLKMLSEQYLDLIF---GRKQNGEVDKSTAGY
339625081	<i>Fructobacillus fructosus</i> KCTC 3544	YDHRHREKLSLAKQAGLHHRFSELY
306821691	<i>Bubacterium yurii</i> ATCC 43715	YEMHDIKCKRDSMHTREAVEQ---SRQAYDYIINKPYGT
336394882	<i>Lactobacillus farciminis</i> KCTC 3681	YDKTRNLKQVADVYKASTQPKLVS-SKSLKQYDAPTEMDYFSS
323463801	<i>Staphylococcus pseudintermedius</i> ED99	FAGYQKMASLNLQDRT-FEENVYRSYFIS-DEMSREYQKSHAVFLCE
389815359	<i>Planococcus antarcticus</i> DSM 14505	YQGHKHELEAYVYAKNDHELAKTIGLYDRVINGDMLKPELSDVYKALAK
258509199	<i>Lactobacillus zhamosus</i> CG	YDEHRQDITLKAUVQQLRERKVKHIFPQSKNGIAGYIDGAGSERFYAFI
169623755	<i>Finnegoldia magna</i> ATCC 29328	HHHHHHHHHHHHHH---HHH-HHHHHHHHH
Unet	227501312	YKISGKIRST
47458868	<i>Mycoplasma mobile</i> 163X	YAGLIDSKEDMDLSEEL
284931710	<i>Mycoplasma gallisepticum</i> str. F	LDKAKORAKEDSESDAKKES
71894592	<i>Mycoplasma synoviae</i> 53	-----FYDL
363542550	<i>Mycoplasma ovipneumoniae</i> SC01	-----FYSEKMDKIDKEA
384393286	<i>Mycoplasma canis</i> PG 14	YAGSQILMEKEVLD
238924075	<i>Bubacterium rectale</i> ATCC 33656	LNENNVKUGINACNY
315149830	<i>Enterococcus faecalis</i> TX0012	YQYCGIACDFTV-EDGKHKR
116627542	<i>Streptococcus thermophilus</i> IMD-9	MNEGVRG
315659848	<i>Staphylococcus lugdunensis</i> M23590	LNFWNGING
160915782	<i>Subactarum dolichus</i> DSM 3991	FDEPNVYKRFHNAAR
325677756	<i>Ruminococcus albus</i> 8	RFCEPNRKGTY
225377804	<i>Roseburia inulinivorans</i> DSM 16841	YNDKRNKGYIDG
336393381	<i>Lactobacillus cozymiformalis</i> KCTC 3535	FNTVFNKAGS
310780384	<i>Ilyobacter polytropus</i> DSM 2926	LEADGSPKQKREGLRSPRADDWFLIKKYEADIKL
301311869	<i>Bacteroides</i> sp. 20 3	LS-KGFKQIKPD---LPEKEDVACTPADNPRC
385811609	<i>Ignavibacterium album</i> JCM 16511	YTERHAKGKVEQREHHEGEGYSHRDEAKREHLQWRVNGLRBAL
60663389	<i>Bacteroides fragilis</i> KCTC 9343	YKHDD
187250660	<i>Mitratifactor salmuginis</i> DSM 16511	FFEKSHKRSKISWANN
325972003	<i>Elusimicrobium minutum</i> Peil91	FNTNGLICITANSESVAISK
296446027	<i>Sphaerobactera globus</i> str. Buddy	QPTFNKASLIVRFRNRGEG

Figure S2 (Continued)



DDALLRE-----LGCKLMSDWAELKRIKIOGL-----  
 KRDADKL-----KRDADKL-----KRDADKL-----KRDADKL-----  
 TOCGKDA-----TOCGKDA-----TOCGKDA-----TOCGKDA-----  
 HTVSDG-----HTVSDG-----HTVSDG-----HTVSDG-----  
 NYENGVST-----NYENGVST-----NYENGVST-----NYENGVST-----  
 TETDKD-----TETDKD-----TETDKD-----TETDKD-----  
 NDEQEDI-----NDEQEDI-----NDEQEDI-----NDEQEDI-----  
 KENAEK-----KENAEK-----KENAEK-----KENAEK-----  
 KODEKG-----KODEKG-----KODEKG-----KODEKG-----  
 NESDEYTV-----NESDEYTV-----NESDEYTV-----NESDEYTV-----  
 KENREK-----KENREK-----KENREK-----KENREK-----  
 AKMKER-----AKMKER-----AKMKER-----AKMKER-----  
 ADMQDR-----ADMQDR-----ADMQDR-----ADMQDR-----  
 KONREK-----KONREK-----KONREK-----KONREK-----  
 IDAFKED-----IDAFKED-----IDAFKED-----IDAFKED-----  
 KDTYOEN-----KDTYOEN-----KDTYOEN-----KDTYOEN-----  
 LENK-----LENK-----LENK-----LENK-----  
 LDYFEKE-----LDYFEKE-----LDYFEKE-----LDYFEKE-----  
 KEKERN-----KEKERN-----KEKERN-----KEKERN-----  
 KENK-----KENK-----KENK-----KENK-----  
 TDEN-----TDEN-----TDEN-----TDEN-----  
 TIEM-----TIEM-----TIEM-----TIEM-----  
 APNVEAH-----APNVEAH-----APNVEAH-----APNVEAH-----  
 MKKEFTK-----MKKEFTK-----MKKEFTK-----MKKEFTK-----  
 E-----E-----E-----E-----  
 EKFAME-----EKFAME-----EKFAME-----EKFAME-----  
 PEKFKEE-----PEKFKEE-----PEKFKEE-----PEKFKEE-----  
 TESKKEE-----TESKKEE-----TESKKEE-----TESKKEE-----  
 PEFSEK-----PEFSEK-----PEFSEK-----PEFSEK-----  
 EYFIEE-----EYFIEE-----EYFIEE-----EYFIEE-----  
 TDFPCDG-----TDFPCDG-----TDFPCDG-----TDFPCDG-----  
 TDWIAL-----TDWIAL-----TDWIAL-----TDWIAL-----  
 TDFFINR-----TDFFINR-----TDFFINR-----TDFFINR-----  
 DENFINK-----DENFINK-----DENFINK-----DENFINK-----  
 SNEKSA-----SNEKSA-----SNEKSA-----SNEKSA-----  
 DDKYIAK-----DDKYIAK-----DDKYIAK-----DDKYIAK-----  
 TEGLEEA-----TEGLEEA-----TEGLEEA-----TEGLEEA-----  
 TDFEYAT-----TDFEYAT-----TDFEYAT-----TDFEYAT-----  
 TDVYLEK-----TDVYLEK-----TDVYLEK-----TDVYLEK-----  
 DRDHYIA-----DRDHYIA-----DRDHYIA-----DRDHYIA-----  
 NMINKE-----NMINKE-----NMINKE-----NMINKE-----  
 TRELKKE-----TRELKKE-----TRELKKE-----TRELKKE-----  
 PEEQVSEL-----PEEQVSEL-----PEEQVSEL-----PEEQVSEL-----  
 LTBEEYIEK-----LTBEEYIEK-----LTBEEYIEK-----LTBEEYIEK-----  
 SDEYCDR-----SDEYCDR-----SDEYCDR-----SDEYCDR-----  
 TADIQTA-----TADIQTA-----TADIQTA-----TADIQTA-----

Figure S2 (Continued)



MIDGKXVILGQVYISLY---HOKAIR---WQYTRREHYLSEFITCKVQIGQDIME---EKKINEX  
MSEHNLIYVGAFAQLED---EGYVRNNDYRAIRSEFQHELEIFIKFQ---DELIV---ESELIER  
MEKACCEYLGDYFKLYDACCURAIR---QRYVDRNKYQREFDALICEQO---Z  
YSHYLSRSHLLEERKULFOSRYLGPFA---SENLEEA  
YKFFDRVYKQGFDTYWMQAQAFNSLY---IEAAREE  
YDLYIDRANKQGFDTALMAQAQAFNSLY---HEARAD  
YKLYIARWILAQFDALWASQRFNAFYL---ADAARD  
YDFYDRRHIEEFHKLMAQCAHPEL---TETIRD  
GVAPTRALLAELAKIVASQAYVGL---DMPAV  
YFFPARNGEAEFDYWMQAQAFNSLY---TAEAREI  
GVNPTRALIQGFDAIRAIQAPRHFDL---FWDSE  
---RLP---GDVILPRADAFHCVBRARATITGKASDYPTRMIRDEFNATMAGQSAHHATI---TDEARTE  
YEYFFTRAVDAEFAEALMAQAPHFPM---TAAAHDT  
YEIISIRHDLAVEVEALFVACQEMRSTIA---TDALKAA  
YERLQSFYADLAKLALFKAGREFGFSF---SKNFESE  
---HHHHHHHHHH  
YARCVERSELREKATYFIOQELKSPV---NEELIYK  
YDKALSRVILGELALMFAFORLGNPH---ASDFEK  
YBNLARDUENELKLFKAGREFNPFV---TEBLEK  
YIHTYAREHLMELKYFSIQKLGSPV---TEKLEHD  
YSHMANKVFERALOLLAFORQOSE-I---SKOFKFT  
KGIHARWVAEVEFERLWVQSKFBALK---SEEMRAE  
YSHYFRDLQNLIDLFKQKGFSPV---SGLKEG  
YHTFERLLAKLMLAFACQHFQPHK---KEHIQCY  
LMTAFPELLIVEKVRILGSAFLIPGL---TALIEL  
LEGVYQSHVAELAQICRTQRY---SETTEK  
FETLRQSDVLMELACLADVQGL---FEDVBSN  
LFVRLMOERAMELKOIFVQRVY---ADENKPL  
RHTVSRQVIVDVRKLIFAQORAL---GMDVNSELEDS  
RMTALRHEQIEHEFERLWATQSOF---HDVMDHGKKE  
NWTLLRARVSEFEALWNEGVYH---YPELEMLDKKTL  
DQIFSRQCTIDEDYDQIMAVQKRV---YFDLLTDEFIRM  
---FRQAYVEFDCIMWCQRKF---YFDLLTDEFIR  
ARVTRDNYINREPELLWQRQAGHLGLAHEQATRNIF  
---TYPTREMYKEEFELRSQEKY---YPOVMDIDLXKA  
---ATYPTREVRVAFERLWATQSOF---APDILLTDEFIR  
---HEE---HEE  
---HHHHHH  
SGDVLVAGLDEDRATIPFVEDQNRPEPDEDCTLYMFKALSSCYG  
KE--IMELLTYTFEVMETIFFYDMMNRGAVKCTTLRLAHEZYLDKEL  
SQDYISILAKTFFPELTIFFYEARTWYKSKKDCSLLAPKELNELY  
SEDIIETICTHDFNRTIFFYEDQNRPEPDEDCTLYMFKALSSCYG  
AP--LLEKLSLSAULTIFFYEDQNRPEPDEDCTLYMFKALSSCYG  
---VTMAGLVDYALDELDFCSGIFPYLDMMNRKPEKCSLLMFKFLDNQY

Figure S2 (Continued)



227494853	<i>Actinomyces coleocanis</i> DSM 15436#	I
328956315	<i>Coriobacterium glomerans</i> PW2	V
227824983	<i>Acidaminococcus</i> sp. D21	I
303229466	<i>Veillonella atypica</i> ACS-134-V-Col17a	F
34762592	<i>Eusobacterium nucleatum</i> ATCC 49256	K
374307738	<i>Filifactor aloccis</i> ATCC 35896	Y
320528778	<i>Solobacterium moorei</i> F0204	S
291520705	<i>Coprococcus catus</i> GD-7	I
42525843	<i>Treponema denticola</i> ATCC 35405	L
304438954	<i>Peptoniphilus duerdenii</i> ATCC BAA-1640	K
224543312	<i>Catenibacterium mitsuokai</i> DSM 15897	L
116628213	<i>Streptococcus thermophilus</i> IMD-9	I
24379809	<i>Streptococcus mutans</i> UA159	I
13622193	<i>Streptococcus pyogenes</i> SF370	I
310286728	<i>Bifidobacterium bifidum</i> S17	G
366983953	<i>Oenococcus kitaharae</i> DSM 17330	SHE
422884106	<i>Streptococcus sanguinis</i> SK49	I
339625081	<i>Fructobacillus fructosus</i> KCTC 3544	D
306821691	<i>Eubacterium yurii</i> ATCC 43715	I
336394882	<i>Lactobacillus farciminis</i> KCTC 3681	I
323463801	<i>Staphylococcus pseudintermedius</i> ED99	I
389815359	<i>Planococcus antarcticus</i> DSM 14505	R
258509199	<i>Lactobacillus rhamnosus</i> GG	I
169823755	<i>Finegoldia magna</i> ATCC 29328	H
Jnet	227501312	E
47458868	<i>Mycoplasma mobile</i> 163K	E
284931710	<i>Mycoplasma gallisepticum</i> str. F	Y
71894592	<i>Mycoplasma synoviae</i> 53	Y
363542550	<i>Mycoplasma ovipneumoniae</i> SC01	F
384393286	<i>Mycoplasma canis</i> PG 14	F
238924075	<i>Eubacterium rectale</i> ATCC 33656	Y
315149830	<i>Enterococcus faecalis</i> TX0012	F
116627542	<i>Streptococcus thermophilus</i> LMD-9	Y
315659848	<i>Staphylococcus lugdunensis</i> M23590	Y
160915782	<i>Eubacterium dolichum</i> DSM 3991	I
325677756	<i>Ruminococcus albus</i> 8	I
225377804	<i>Roseburia inulinivorans</i> DSM 16841	Y
336393381	<i>Lactobacillus coryniformis</i> KCTC 3535	Y
310780384	<i>Ilyobacter polytropus</i> DSM 2926	Y
301311869	<i>Bacteroides</i> sp. 20 3	Y
385811609	<i>Ignavibacterium album</i> JCM 16511	CL
60683389	<i>Bacteroides fragilis</i> NCTC 9343	NE
319957206	<i>Nitratifactor salmuginis</i> DSM 16511	LYPNE
187250660	<i>Elusimicrobium minutum</i> Pe1191	RKLATVLYPSQSKF
325972003	<i>Sphaerochaeta globus</i> str. Buddy	IL
296446027	<i>Methylosinus trichosporium</i> OR3h	PRNSISI
		FGPKKEFEN

Figure S2 (Continued)

IELVPAADSPKGA-----SCEIIVGYDV-----LPGQEGK-----RR-----AEKAHP  
 SLVSRIPYYVGPL-----YLQOE-P-----VLQOE-P-----P-KSSL  
 SNQDKLISIMFRIPYYVGPL-----ERKAGKI YPWN FNDKVDLQKSEAFIRMTMCTI-----VL-----P-LDSL  
 SVTEKILMELFRIPYYVGPL-----FSAVRKQAGRVIPWN FEKIDREKSAAFKMLTKCTI-----VL-----P-KSSL  
 DKLKTFRFRIPYYVGPL-----SHVVRKEGKI L PWN FEQKVDIEKSAEAFIKRMINKCTI-----VL-----P-KDSF  
 SVVOKIESIFRFRIPYYVGPL-----VMMVRKFGREDRI YPWN MEEI IDFEKSNENFIRMTMCTI-----VL-----P-KHSL  
 VVDKIKLIFKFKIPYYVGPL-----ASDEKIPWN FSNVIDDKTAHEFANRLIGRCTI-----VL-----P-MDSL  
 QOOLEFRFRIPYYVGPL-----FVWSVARSDARI YPWN FTEVIDVESAEEKFIRRMINKCTI-----VL-----P-KDSL  
 SHSEKIIMLIFRFRIPYYVGPL-----NDMKHKEFFPD-----VLVGE-S-----P-KSSL  
 DKVESIMEFRIPYYVGPL-----NPSKYAWIK-----VLKBE-K-----P-KHSL  
 LSIIFRFRIPYYVGPL-----NETSEHAWIK-----YFPDE-E-----P-KHSL  
 EKILIFRFRIPYYVGPL-----ARGNSDEAWSI-----YLPGE-K-----P-KHSL  
 EKILIFRFRIPYYVGPL-----ARGNSDEAWIS-----YLPNQ-K-----P-KHSL  
 EKILIFRFRIPYYVGPL-----ARGNSREAWMT-----NLPNE-K-----P-KHSL  
 KQINKIBELFRFRIPYYVGPL-----VVPEDLQKSDNSENHVMV-----YLVGE-P-----P-KHSL  
 LNKITALINFRFRIPYYVGPL-----VEEQKTIADGKNIPDPINEMV-----TL-----P-KHSL  
 DKLILSILIFKIPYYVGPL-----AGNSREAWIKRATSSDILDDNDEDTNGKIRPWN YOKL INWDETROAFITNLI GNDI-----TL-----P-KHSL  
 GQNKIELEFRFRIPYYVGPL-----YKDGRI L PWN FEKVDVKSKEEFIRRMINKCTI-----TL-----P-QNSL  
 TVSEMI IOLFEQIPYYVGPL-----DKNEKONKANSWAK-----YISDE-H-----P-KHSL  
 EKLSILSIFRFRIPYYVGPL-----QSAEKNPANME-----YLVGE-P-----P-KHSL  
 TEKVQILAFRFRIPYYVGPL-----VKDITQSPFSWVZ-----YLVNE-D-----P-KHSL  
 KMPYQDELHFRFRIPYYVGPL-----ENVFAMWRDPG-----YLVGE-D-----P-KHSL  
 EKIRYTFRFRIPYYVGPL-----NDIYHNKNG-----YLVPE-T-----P-KSSI  
 HHHHHH-----E-----HHHHHHHHHH-----P-KHSL  
 KSFFRIRNYSRSGPNEKSFYGL-----YANENG-----YDKKL-Y-----P-KNSF  
 ESLFVVRNYSRSGSINSVSYGL-----YHDEKCKVYQYNNIDKTI GKN-----IFPDE-Y-----P-KNSP  
 LKLFNYSVRPNKPGSKNSRTAYGV-----FSTD IDPETHKFKDYSNIMDKTI GKS-----RA-----P-KNL P  
 LNLFTSVRDYANGRSGSEHSASEYGI-----FOK-----P-FVEE-N-----P-VNYP  
 KKFSTFRDISRSGSDNMPSPYGI-----PGEF-GDNGQG-GRYEHIWDKTI GKS-----IFPNE-Q-----P-KYLP  
 MLFNKRKYIYEGPNEKSRDYGR-----FYTKLDANGVITEDNIEFKLI GKS-----YVPE-I-----A-AASY  
 VQPGIAEAGLIYRKRKYIYHGFNEANSRPGYGR-----WSDFOKTGEPAVNI FDKLI GKD-----FOGEL-----RA-----S-GLSL  
 LEILTEKRYIYHGFNEKSRDYGR-----YRTSGETLDNIEGLI GKCTI-----YVPE-F-----A-KASY  
 LEIVMRRYIYEGPNEKSRDYGR-----PKAWYETIMGHCTI-----YFPDELR-----SV-----KYAY  
 IYIISKRMYIDRGGPLSPYGR-----YVFGQKEPIDLIEKMGKCS-----LEPNE-P-----P-KLAY  
 KDI IFAQRDFEIGPKNERFRPTG-----YIDSLGKCO-----FFKQO-E-----RG-----S-RFTV  
 VEIMTSORSFDMGGLQPDGKPS-----PYAMEGFDVYKCTI-----FEKDE-Y-----RA-----P-KATY  
 LIGDMRAGIQAQRDFEGRGAGYAGDQ-----IKRMVGDID-----PEPTE-D-----RA-----A-KATY  
 IEVNSQRNFDGPKSPYSEIGQ-----TAKMIGNCTI-----FESSE-K-----RT-----A-KNTW  
 RDFCHUFDIIPYORPLKS-----KKSLLDNCP-----YEENR-YI-----KKSLLDNCP-----DRESGEIKHA-SIKCIA-KSHP  
 SDVLHTISEDIYIYORLKS-----QKSLISECR-----YEKAK-GI-----D-----GEL-Y-GLKCI P-KSFP  
 RDII IYIYORLKS-----OKGLIGFCH-----PESRQIEV-----DIDGKKIKVGNRVIS-RSFP  
 DELKACITDQEMPTID-----ESLFGKCTI-----FYKDE-I-----AA-----P-AYSY  
 TKAIGYSEK-----LIPESGFCP-----YLKDE-K-----P-ASHK  
 VSAILFEMEK-----IVPEAGCCP-----YFPDE-K-----P-ASHK  
 TKAAPFORPLQD-----SEKIVGFCP-----PEVDE-K-----P-KRCY

Figure S2 (Continued)

347536497	<i>Flavobacterium branchiophilum</i> FL-15	K
345885718	<i>Prevotella</i> sp. C561	L
282880052	<i>Prevotella timonensis</i> CRIS 5C-B1	L
312879015	<i>Aminomonas paucivorans</i> DSM 12260	F
294086111	<i>Candidatus Punicispirillum marinum</i> IMCC1322	L
330822845	<i>Allicyclophilius denitrificans</i> K601	L
344171927	<i>Ralstonia syzygii</i> R24	L
159042956	<i>Dinoroseobacter shibae</i> DFL 12	L
83591793	<i>Rhodospirillum rubrum</i> ATCC 11170	T
288957741	<i>Azospirillum</i> sp. B510	L
427429481	<i>Caenospirillum salinarum</i> AK4	L
92109262	<i>Nitrobacter hamburgensis</i> X14	I
148255343	<i>Bradyrhizobium</i> sp. BTA11	I
34557790	<i>Wolinella succinogenes</i> DSM 1740	Y
218563121	<i>Campylobacter jejuni</i> NCTC 11168	V
Jnet	218563121	H
291276265	<i>Helicobacter mustelae</i> 12198	LLGNP
222109285	<i>Acidovorax ebreus</i> TPSY	L
365156657	<i>Bacillus smithii</i> 7 3 47FAA	Y
220930482	<i>Clostridium cellulolyticum</i> H10	F
297182908	uncultured delta proteobact. HF0070 07E19	Y
154250555	<i>Parvibaculum lavamentivorans</i> DS-1	I
218767588	<i>Neisseria meningitidis</i> Z2491	I
15602992	<i>Pasteurella multocida</i> str. Pm70	M
187736489	<i>Arkermansia muciniphila</i> ATCC BAA-835	I
315605738	<i>Actinomyces</i> sp. oral taxon 180 str. F0310	I
117929158	<i>Acidothermus cellulolyticus</i> 11B	V
189440764	<i>Bifidobacterium longum</i> DJ010A	F
403744858	<i>Allicyclobacillus hesperidum</i> URH17-3-68	Y
407803669	<i>Alicanivorax</i> sp. W11-5	I
423317190	<i>Bergeyella zoohelcum</i> ATCC 43767	I
402847315	<i>Porphyromonas</i> sp. oral taxon 279 str. F0450	I
404487228	<i>Barnesiella intestinihominis</i> YIT 11860	I
374384763	<i>Odoribacter laneus</i> YIT 12061	L
384109266	<i>Treponema</i> sp. JCA	I
402849997	<i>Rhodoculum</i> sp. PH10	I
Jnet	331001027	I
331001027	<i>Parasutterella excrementihominis</i> YIT 11859	I
34557932	<i>Wolinella succinogenes</i> DSM 1740	I
54296138	<i>Legionella pneumophila</i> str. Paris	I
319941583	<i>Sutterella wadsworthensis</i> 3 1 45B	I
254447899	gamma proteobact. HTCC5015	I
118497352	<i>Francisella novicida</i> U112	I

Motifs  
informative positions

Figure S2 (Continued)

```

FDGLAKDLYKAIFFQRLPKS-----OKGLIGKCS-----FEKSK-S-----RC-----A-ISHP
ISE--KKNWGTIFYQRLPKS-----QRGNVQKCT-----LERSK-P-----A-IGHP
SSELIEDIQRAIFFQRLPKS-----QRGVGRCT-----FERGK-P-----RC-----A-DSHP
-----LELVAFQNFASGED-----IRNKAGHCS-----LEPDQ-I-----RA-----P-RRSA
-----KEIVFQRL-----KQEVGRCT-----FLSDE-D-----RI-----S-KALP
-----KDTLLQRL-----RVPKGRCT-----LLPEE-E-----RA-----P-LALP
-----RAILLQRL-----LFPVQKCF-----LEPNQ-P-----RV-----A-AALP
-----PEKIFQRL-----KEPEVGLCLFSGHGVPPK-P-----RL-----P-KAHP
-----RVLDQRL-----RSKGAGPCA-----FLPGE-D-----RA-----L-RALP
-----RRIFQRL-----KPPVGRCT-----LYPDD-G-----RA-----P-RALP
-----RRVLDQRL-----AVPAPFCL-----ELELR-----KETFQERTIYREADRG-----L-TVDF
-----CHEIYQRL-----KPAIVGKCT-----LDPAT-PP-----P-WSHF
-----REALFSQRA-----KRFSGKCS-----LDPATVSD-----DVGDF-----RC-----A-WSHF
-----RELAFVQRL-----LEKMGHCT-----YPER-R-----RA-----P-KSAP
-----LSVAFYGRALKD-----FSLVGNCS-----FTDE-K-----RA-----P-KNSP
-----HHHHHH-----H-----H-----
DAQSKQERGLIFYQRLPKS-----FQDKIGKCS-----HIKKGNS-----PY-----C-KHAP
ILGDGDRKSGLEWQRLPKS-----LLKMLGKCT-----FEKGE-Y-----RA-----P-KASF
-----LNIWSSQRFASKED-----LEKVGFCF-----FEKKE-K-----RA-----P-KATY
-----LNIWGFQRFASKED-----ILSKVGKCT-----LKER-L-----RA-----P-TSY
-----LDVLMQSGR-----SPRLGNCS-----LIPSE-L-----RA-----P-SSAP
-----SDTYQRLVFW-----RNTLGEGR-----EMGR-P-----LC-----P-KGSW
-----ETLLMQRFALSGDA-----VQRMGHCT-----FERAE-P-----KA-----A-KNY
-----TELLMQRFALSGEA-----ILKMLGKCT-----HEKNE-F-----KA-----A-KHY
-----VLSFESKEPAPS-----LLFQLIREFDNRLISRCFVTAQVYEAELKKG-----NSEQSAREAEKLSKVPFANCP
-----IDAVFCQRLPV-----BARQERVGLDELQ-----LALDPAKQ-----PR-----A-RAHP
-----RSVYAVSPKGS-----AEQVQGD-----LAPQ-A-----RA-----L-KASL
DDPGGSDSYGSGVSPDGVR-----QSIYRMVGSCT-----FETGE-----RA-----KRAP-RSSY
-----RNALFFQRLPKS-----PADKVGRC-----LQTNL-----FRAP-RAQI
KGLKYLIKQVVFYQRLPKD-----QSHLSDCH-----YEPNE-----KALA-KSH
-----RDEIYQRLPKS-----CKHIVSLCE-----FEKQERM-----RVQDDGKGGWQIVERRVAFGPKVAP-KSSP
-----EQLKFKNESVLFQRLPKS-----CKYIVSRCE-----FEKRE-----YLNAGKTEAGPKVSP-RTSP
-----FYQRLPK-----QKSLSKCV-----FEGRN-----FYDFVQKWIITAGTTPAP-LSHP
-----AGILFRQDLA-----PQHGICY-----YENDK-----ERTF-KAMP
-----HHHHHHHH-----HH-----HHHHHH-----E-----EEE-----HHH
EKWKSWANKE-----AGAYPLTETETEILKNTDRKSR-----E-----IRKSDVLPDSYRLAY
FNWDLAKRL-----MEKFNDDLADSTVKGYS-----EDST
FNWRLIPGI-----IDAHFFLEKDEHEHKLRDKR-----IISPSKQDEKRSY
EINKTWSARL-----TSAFTLAPAEAILERSYDRKSR-----VAVNGHEPLFLAYQLSY
FKWKEVQGLVK-QNDNAYLNENVT-----FKWKEVQGLVK-QNDNAYLNENVT-----VAVNGHEPLFLAYQLSY
FNWQYLQELKQLQSIQNYLDSFTDLKVLKSSKDPF--Y-----FKSSNGQIASGQRDYKDLDR

```

Figure S2 (Continued)

AF-QRYRIAS-IVSNIRI-----RHLSGADERLDVETQRRVFEYLIN  
 LY-EEFCVIN-ELNGAHM-----SIDGDDHRFADADREGLLEKLFK  
 LY-EKFMILN-ETNIRI-----DGYPIVDVKQVQVGLFEK  
 LY-SEFMILN-ELNVRIL-----DGKALAQGYKQHLIDSIFK  
 LY-SYITIM-ELMKVQV-----NDEFLENNRKRKIDELFEK  
 LY-SYVWILN-ELNWKV-----RCKKLPSTSLKQVFDLFE  
 LY-SYVWILN-ELNPIKV-----NGKALPVEVKQAIYTDLFE  
 VY-SFVWILN-ELNLRIL-----NGEKI SVELKQRIYSELFC  
 LY-SYVTVILN-ETNLIQI-----IIDKRNICDILKOKIYEDLFEK  
 LY-NEFMILN-ELNNIKV-----NEFLI TEEMKKAIFYSELFEK  
 IV-SKVEVYN-ELANKIRV-----DDKLEVDVKNDIYNELFM  
 LY-ETFMVYN-ELTKVRF-----IAESMDYQFOLDSOKKDIVRLEFK  
 LY-EASTVYN-ELTKVKY-----KTEQ-GKTAFFDANKQELFDGVFK  
 LY-EYFTVYN-ELTKVKY-----VTEGMRKPAFISGEQKKAIVDLLEFK  
 LY-QEVEVIN-ELNVRIL-----SVRTGNWDRGRMRILGREETILCORLFM  
 LY-QEVDVILQ-ELANNIRV-----SGRRLDIRAKQDAFEHLFK  
 LY-EVWMLQV-ELTRVKY-----KDKYG-----KAHFPSSELRONINGLFEK  
 RY-QLFTVILN-ELNWKV-----NGKPKDSTK KADLINDLFEK  
 LY-EKFMILN-ETNIRI-----DCKKPI SVEAKQKIYNDLFEV  
 IV-QEVEVIN-ELNNIRI-----TENLK-----TNPIGSRILVETNORLYNELFK  
 LY-QEVEVIN-ELNATQI-----RLQTFD-----PANKRYRMPQIKLFAVEHLFK  
 TV-EFEVEVIN-ELNGIGL-----RTTGA-----ESDFRRLSYEMKCKWIIDNFK  
 LY-QEVEVIN-ELNVRIL-----NNECLGTDQKQRLREVEFK  
 LY-SYVWILN-ELNNIRI-----NGKPLDIDVKLQLELFEK  
 HH-HHHHHHH-H-----EEE-----HHHHHHHHHHH  
 SA-KVVDIYN-KLTDWKH-----KNEVYSERLKRKILILSREIN  
 TA-MIFNEIN-ELSTIKS-----YSIYLQGFNIOEFKAYLANKLLDILLI  
 SA-LLENLQV-ELCTIKN-----EPTFKKNWMLNAEQKSEILKVFVTELEF  
 SY-EIFNMLN-QLINLST-----DLKT-----TNKTIWQLSSNDRNEILLDELKVKBE  
 SA-LLENFILN-ELANIRL-----YSTD-----KKNLQPLWKLSSVDKRLNILLNLFNPI  
 TA-QEYVWILN-DLANNIT-----NGRLEENERHEIVERIKS  
 SA-QQYNMLN-DLNLKAI-----DGEVPLSSEQSEYILTELM  
 TA-OEFNMLN-DLANNIV-----PTETKILSKQKQKQIINYVKV  
 SA-DLENALN-DLANNIV-----QBDGLSKLEYHERKYHLENVK  
 SA-ELENMLN-DLANNISL-----EGEKLTSEQKAMLIKIVHE  
 IA-DIYAVN-VLSQYIY-----TNWRSSEVFTSPANDLINSALK  
 TA-ELEVALQ-KINHTKI-----IDFGTGRFFSEERKTIIGLALS  
 TF-QEFLNLIQ-KMISLNY-----QNTYGDYHTLNGLDKQAILDAVFA  
 SG-EREVFLQ-KENFRIL-----VELSKRPLTEERDIVERKEVIL  
 LY-QEFRILWQ-FIVNIRI-----YKRETDVDTVQELLIP  
 LY-QEFRILWQ-DIHNIRV-----IRKESVTVNGKXKINI  
 LF-QEFLKIQV-ILANNIKV-----YVYVGVKHKRKRKIKENYSALFELNDAPQELMGRRCOEKELLAGELFI  
 LY-DIYRLYK-KLADINI-----DCXEVTQEDREKYLEWKEKLIA  
 LN-EERLWE-TLNNARY-----SDPIVDIVTGEITGYEYKQFTKQKLPDYLLT  
 LN-EERLWE-ALNNARI-----KNEMQEGAAKRYQSASES

Actinomyces coleocanis DSM 15436†  
 Coriobacterium glomerans PM2  
 Acidaminococcus sp. D21  
 Veillonella atypica ACS-134-V-Col17a  
 Fusobacterium nucleatum ATCC 49256  
 Filifactor aloocis ATCC 35896  
 Solobacterium moorei F0204  
 Coprococcus catus GD-7  
 Treponema denticola ATCC 35405  
 Peptoniphilus duerdenii ATCC BAA-1640  
 Catenibacterium mitsuokai DSM 15897  
 Streptococcus thermophilus IMD-9  
 Streptococcus mutans UM159  
 Streptococcus pyogenes SF370  
 Bifidobacterium bifidum S17  
 Oenococcus oeni DSM 17330  
 Streptococcus sanguinis SK49  
 Fructobacillus fructosus KCTC 3544  
 Eubacterium yurii ATCC 43715  
 Lactobacillus farciminis KCTC 3681  
 Staphylococcus pseudintermedius ED99  
 Planococcus antarcticus DSM 14505  
 Lactobacillus zhambosus GC  
 Finegoldia magna ATCC 29328  
 227501312  
 Mycoplasma mobile 163K  
 Mycoplasma gallisepticum str. F  
 Mycoplasma synoviae 53  
 Mycoplasma ovipneumoniae SC01  
 Mycoplasma canis PG 14  
 Eubacterium rectale ATCC 33656  
 Enterococcus faecalis TK0012  
 Streptococcus thermophilus LMD-9  
 Staphylococcus lugdunensis M23590  
 Eubacterium dolichum DSM 3991  
 Ruminococcus albus 8  
 Roseburia inulinivorans DSM 16841  
 Lactobacillus coryniformis KCTC 3635  
 Ilyobacter polytropus DSM 2926  
 Bacteroides sp. 20 3  
 Ignaviobacterium album JCM 16511  
 Bacteroides fragilis NCTC 9343  
 Nitratifactor saluginis DSM 16511  
 Elusimicrobium minutum Peil191  
 Sphaerochaeta globus str. Buddy

227494853  
 328956315  
 227824983  
 303229466  
 34762592  
 374307738  
 320528778  
 291520705  
 42525843  
 304438954  
 224543312  
 116628213  
 24379809  
 13622193  
 310286728  
 366983953  
 422884106  
 339625081  
 306821691  
 336394882  
 323463801  
 399815359  
 258509199  
 169823755  
 Jnet  
 47458868  
 284931710  
 71894592  
 363542550  
 384393286  
 238924075  
 315149830  
 116627542  
 315659848  
 160915782  
 325677756  
 225377804  
 336393381  
 310780384  
 301311869  
 385811609  
 60683389  
 319957206  
 187250660  
 325972003

Figure S2 (Continued)

AKPTAD IT WSD VA BEIGVER NLLMTRATQF ADGERASAKPPVDVT NVAFATCKIKPKKEMWLNADYEARCVMSALSJH  
R KRT VS YGD VA GMEERERNOIGARVC GOGEKGFESKLSY IF FCK DVFKVRLQES GOGEKGFESKLSY IF FCK DVFKVRLQES  
K RR VT VKD IQ NLLSLG ALDHKG LFGDITHSNVNTY HH FK SLMERGV LFGDITHSNVNTY HH FK SLMERGV  
QDHK MT KR IE LFLKDN YTKKHP EYLAVNQI AMRTE LAGIKDFNSVYVYTKFKD LAGIKDFNSVYVYVYTKFKD  
E NK VS EKK FK EYLAVNQI AMRTE LAGIKDFNSVYVYVYTKFKD LAGIKDFNSVYVYVYTKFKD  
N KSK VT GKN LL EYLAVNQI AMRTE LAGIKDFNSVYVYVYTKFKD LAGIKDFNSVYVYVYTKFKD  
NSKK VT RKS TY EYLAVNQI AMRTE LAGIKDFNSVYVYVYTKFKD LAGIKDFNSVYVYVYTKFKD  
K YK VT RKK LB RYLAV EYLAVNQI AMRTE LAGIKDFNSVYVYVYTKFKD LAGIKDFNSVYVYVYTKFKD  
K YK VT OKQ IS EYLAVNQI AMRTE LAGIKDFNSVYVYVYTKFKD LAGIKDFNSVYVYVYTKFKD  
T KKK VT LRA VS EYLAVNQI AMRTE LAGIKDFNSVYVYVYTKFKD LAGIKDFNSVYVYVYTKFKD  
K NKT VT EKK LK EYLAVNQI AMRTE LAGIKDFNSVYVYVYTKFKD LAGIKDFNSVYVYVYTKFKD  
D KRK VT DKD TI EYLAVNQI AMRTE LAGIKDFNSVYVYVYTKFKD LAGIKDFNSVYVYVYTKFKD  
V YK VT KDK LM EYLAVNQI AMRTE LAGIKDFNSVYVYVYTKFKD LAGIKDFNSVYVYVYTKFKD  
T NRK VT VKQ LK EYLAVNQI AMRTE LAGIKDFNSVYVYVYTKFKD LAGIKDFNSVYVYVYTKFKD  
K QGT VT KRT AE EYLAVNQI AMRTE LAGIKDFNSVYVYVYTKFKD LAGIKDFNSVYVYVYTKFKD  
V OKT VS ATN LK EYLAVNQI AMRTE LAGIKDFNSVYVYVYTKFKD LAGIKDFNSVYVYVYTKFKD  
NN SRK VN AKS LI EYLAVNQI AMRTE LAGIKDFNSVYVYVYTKFKD LAGIKDFNSVYVYVYTKFKD  
A KKT VS LSA LK EYLAVNQI AMRTE LAGIKDFNSVYVYVYTKFKD LAGIKDFNSVYVYVYTKFKD  
K GKK VS QKD IK EYLAVNQI AMRTE LAGIKDFNSVYVYVYTKFKD LAGIKDFNSVYVYVYTKFKD  
K YK VT VKK LT EYLAVNQI AMRTE LAGIKDFNSVYVYVYTKFKD LAGIKDFNSVYVYVYTKFKD  
K YK VT HSK FL EYLAVNQI AMRTE LAGIKDFNSVYVYVYTKFKD LAGIKDFNSVYVYVYTKFKD  
Q YK VS TRK LL EYLAVNQI AMRTE LAGIKDFNSVYVYVYTKFKD LAGIKDFNSVYVYVYTKFKD  
R HSS VT IKQ VA EYLAVNQI AMRTE LAGIKDFNSVYVYVYTKFKD LAGIKDFNSVYVYVYTKFKD  
K KTK VT LKS JR EYLAVNQI AMRTE LAGIKDFNSVYVYVYTKFKD LAGIKDFNSVYVYVYTKFKD  
HHR HH HH EYLAVNQI AMRTE LAGIKDFNSVYVYVYTKFKD LAGIKDFNSVYVYVYTKFKD  
KDSK AVEKILKEENIKET NLSJAY NKDDNINLPINAVHSILT IPKHLINFNENLIS NENDLSKIMSFYKQOSEKLFVFNKESYEINQNNVLIHIFDA  
KFNGBP IDARQFKUREETAEI GKETIKDVENEBEKLEKEDHWKLGKLANE NGKI QYNDLSLAKFVKHLKQLKDFLEDOYATLDKINFLQSLVYLKHLRYSNR  
NRKDKY SDKKFNKLDKIK KYLLNEA LENENLNEKI LKNRDLNDT VLGKGVYKESKNATADAALAEFSSIKPLVYVYVYTKFKD LAGIKDFNSVYVYVYTKFKD  
KART ISILKNEIKRTI IKDFEPEKSDIDDQD TIEGRKI KEPEPT KLEVTXHLIATYSHSSD NWLNTNN ILEFLPY  
SEK KKL TSNINDIVKESI KSIM IS VEDIDMIKDEWAGKE PNVYVGLSGINI EESAK ENKPKFQDLKILNVLINLNDVGIKFEKDRND IKNLEL  
S NT IN MR KIISDCMGENIDDFAGAR KEPLSGNR KILGVR KRLSGNR KILGVR KRLSGNR KILGVR KRLSGNR  
KEPTR FG VND VV KILGVR KRLSGNR KILGVR KRLSGNR KILGVR KRLSGNR KILGVR KRLSGNR  
E KA MG PAKLFKYA KILGVR KRLSGNR KILGVR KRLSGNR KILGVR KRLSGNR KILGVR KRLSGNR  
QAKP T LAQ IA NEINVN PEDIKYR KILGVR KRLSGNR KILGVR KRLSGNR KILGVR KRLSGNR  
K GK IT PKO LA KEVGS LEQIRGFR KILGVR KRLSGNR KILGVR KRLSGNR KILGVR KRLSGNR  
N GS MD KRE LK AKSYHLDIS KILGVR KRLSGNR KILGVR KRLSGNR KILGVR KRLSGNR  
S KE LK YGT IR KKNIDP SILKFNINYS AKRGEPEEERVIDTEKAKF AKRGEPEEERVIDTEKAKF  
KAEP TNYKPFDFGE LR YEK LR EWLNEKEIDQKAFYPP BARFNLVNYGSLQTKEL EYVEKTYRFBVFRAY HDLVKVLPEENWQ  
K KE YV LK EWLNEKEIDQKAFYPP BARFNLVNYGSLQTKEL EYVEKTYRFBVFRAY HDLVKVLPEENWQ  
T EAD YINENLEKLR YINENLEKLR YINENLEKLR YINENLEKLR YINENLEKLR YINENLEKLR YINENLEKLR  
D ETQL YINENLEKLR YINENLEKLR YINENLEKLR YINENLEKLR YINENLEKLR YINENLEKLR YINENLEKLR  
R DK MT KSE VL KLLFDPQELDNFTIDGNKTY ELENKDSLSKEDLILSILNINSIGIKSK KEETTRHLNFANRKLKCNETHS RYRKYFKLGDGEYLLN  
QGNLAK IT HKD LR KILGLAP BOKIFGED ERIV ALFOYSKMIEMSGH EPVDFKPVKWEYIKAVFDLLNWNVDILGFN SNEELDN  
S SE LT PAQ TK KILGLK NTNPFEDI GCKKEPTVFFFLAD LAKEKELFASLOKHPD ALQIFRE  
DEORE IL FHI AR SCDITPKLVQKEFPALKTSII VLQCKEAKIOLAGE RFRLEEKSFWRLL RFRLEEKSFWRLL

Figure S2 (Continued)

A-AK VS-FTA-IR-KKLIPE-TTVFVGVK-ADESKLDVARSCK-AAECTARLSRVIVDALGELAWCALIC-----SPEKLDK  
K-ND-FN-FDV-LA-RELIEK-GSSFGFYSSKNDFF-YWFRXKPTDVAACQVAASLKNALGEDWK-TKSFYKQRTINSKEQVSRVTDY  
R-TE-FK-FED-I-KELIEK-RLGHFYSYNDKT-----INYKDSIVSAGCPITARPRKMLGEWE-----SFRVSGQKERQAHSKNNISFHRVSY  
K-EN-FD-FED-LA-KALACK-K-NYAMWHDKEERA-YKFRYMTQVGFVGGPTIAQKLSIFGDWDK-TGLAETYLLOKNGSKSLQEMVD-  
Q-EK-VT-HKT-IR-RHLEIPE-EWFLTGLPYHRS-----GDRAEKLEFVHLAGI-----HEIRKALDKGPPDPAWDLIRS-----RRDLLDS  
K-KK-LT-FKA-MR-AILKQGV-VDYPVGNL-----AALARKELFCAAWSCF-----IMRDAKMLIGSAMDKL-----DEEQDS  
K-AK-LS-FEQ-IR-KLKLSC-SVQFN-----LEDARNTLKNMATS-----AALARKELFCAAWSCF-----DEEQDS  
R-PK-CG-FDM-LR-KIVE-GANKEAYF-----TIESERRK-ELKGC-----DTAAKLAQVNALCTRQWAL-----SLDEQR  
KKPTK-SLSSMVK-LPA-LA-KVLRDGERFTLETGVD-----AIACDPLRASPAHP-----DRCPRWLILDADAQW-----EVISR  
AREVE-WPA-LR-RALGLAR-GVAKFAETE-----RNGAKOARGTAGNL-----TEALLAPLIPGWSGW-----DLDRQR  
RPPKARPKGKVSVEK-IR-GHLELPP-GTGFSL-----ESDRPELLGDEIGA-----RIAPAPFGWTAL-----PLEEQDA  
DPDGLAARDGR-IPYTR-LR-KLICYDNS-FVCFAG-----ERDTSGGEGTIVTFD-----FLMARWIDGWDL-----FLKARSL  
N-RE-VK-FDK-IR-TLKIPA-EARENL-----ESDRRAALDGDQYA-----ARLSDKGF-NKAWRCF-----PBERQIA  
T-DQ-LS-FDE-IR-GHGLPS-DARENL-----ESDRRHILKGDATG-----KILLSARRHF-CPAMHRR-----SLDRQID  
R-EK-VE-FRH-IR-KFLDLS-NEIFKGLHYKPKTAK-----KREATLFDNEPIEL-EPDKVPAKKAALSIRGAALKREALGNEFFYGRFVALGKHAD  
N-GT-LT-YKQ-TK-KLGLSD-DYEFKG-----KRGTYFIEFKYKEF-IK-----ALGEHNL-----SQDDLNE  
HHH-HH-HH-HH-HH-HH-----HHH-HH-HH-HH-HH-HH-----H-----HHHHH  
EAQN-EXCLT-YSK-LK-LLDLPS-DFEFLGLDYS-GHKEKAVFLSFT-FKLNKITO-----FKLNKITO-----DRKTQK  
TETSQ-YKT-IR-NAFKAGLWGDVRFGLAYPSQAIDARK-TKPEDQFVVKLPAW-----HELRKAFAAGHGAALWOQISTPALDGDPTLLDQ  
K-NK-MT-YD-IR-KLNLS-DIHFKGLDYDKSS-LRQENIRFLEDSY-----LRQENIRFLEDSY-----HKTRKCIENVYKDGIRMF-----NETDIDT  
K-NE-IK-YSE-IR-KLDIEP-EILKAHNLTKNP-SGNSKAFYEMKSY-----KLNKSTLPTDINCKLHS-----NKNESLN  
R-ST-SS-YWQ-IR-RDFQIP-EYFNLVNYERDP-----DVLDQBYLQOQERKTLA-----NETRNWQLEKILITGHP-----IQTLE  
Q-AS-MS-WPVSALK-ALYKQGFPAKSLKFNL-----ALYKQGFPAKSLKFNL-----ELGSEKLLGNLGA-KLADMFG-PDMFAHPRKQEIHRHAVHRLWAADYGETPDKKR  
K-SK-LT-YAQ-AR-KLJLED-TAFKGLYKXDN-----AEASTLMEKAY-----HALSRALEKEGLKDKKSLNL-----SELOQE  
K-SK-LT-YAQ-VR-KLGLSE-QALFKHLYSKEN-----AESATPMEKAW-----HAIKKALEMQGLKOTWODLAK-----KEDLLE  
E-CK-LT-KAS-LE-KALSSRLKETEINVSNYF-TLRPDSSEALYIMPAVEVLRQSGIGLILSPSVXRI-----TVVMSASVDWLADWMDCANDESRGHMIDAIN  
HTSES-PT-WDD-VA-RKLEVR-HRUGSSRA5-LETCGLAYPPVDT-----LETCGLAYPPVDT-----LETCGLAYPPVDT-----LETCGLAYPPVDT  
QTERS-LT-WSD-IA-LEILKLPN-ESDLSVPE-----EDGSSLAYSQEAPP-DETSARIAEFAIARNRKKIPTFAQWQBQ-----DKTSSD  
PSSED-IT-WSD-IC-DFLGFK-RSOLKCVS-LTEDEGERISSRPR-----LTSVQRIVESDNKRKPLVAVWKA-----SDNEHA  
EQTK-IT-YGK-LR-KLDMK-DTESFAGTYGLN-----RSMNKTEDTV-----FVENKFFYH-EVRKALQRAQVFIQD-----LSI  
QOE-LS-FRK-IY-KEL-E-RACC-PGFEKGINMDRAAL-----GGRDDLGNIT-----LAARWKLGLEDRWQELDVTQIQVINFLA-----DLGSPR  
KKE-IT-FSA-IFKKL-KEKALIA-DQLTSSKSLKGNSTRVALA-SALQYPOYHH-----KIKGNET-----KLQKQSL-----GELMDVGL-----DSI  
SSAS-LS-FAA-LKLL-KEKALIA-DQLTSSKSLKGNSTRVALA-SALQYPOYHH-----KIKGNET-----KLQKQSL-----GELMDVGL-----DSI  
THEK-LK-GSD-LKLL-GLSKTYGR-LGEQFKTIGGNTRVELE-RALGNYPDKKR-----LLOFNLOQESSSMVNYETGEIIPM-----ISLSFEQ  
ESKD-FN-FEK-IPKHL-K-IFEKFNFDITV-PACTI-----ISQRLKLF-----PHPVWEE-----  
SKDK-VT-FDQ-MRKAL-CLA-DSNSFNLENRDF-LIGNPT-AVKMRSKN-----REFKLMDE-----IPL  
GKS-LT-FKA-VRKTL-KIL-PHALVNEBAGER-GLDGLATAK-----LLSKPDHY-----CAAWHG-----LSF  
HHHHHHHHHH-HHHE-----HHHHHHHHHH-----HHHHHHHHHH-----HHHHHHHHHH-----HHHHHHHHHHHHHH  
EFLEFANRYOE-TAKANG-LW-FP-ENALLERADLHPMKNLINLVGQALGV-SPAEGTDFEEIWN5-KVKGRSTVRSICNAIENERKTYGPFSEDYK  
RLYGAQNYELLRQKVRAG-IWV-PVKNDDSLN-LEDNSMRLKRCNHPFKRNQIENLAVAGLLGV-KLDEARFAEFEKELWSA-KVGNKALSAYCKNIEELRTHGNTFKIDLE  
VLQIASAYNKE-RSDAAG-IW-FDNFSLCENINPPRQKILLPVLAVLISEDFINNKQWAKFIWNT-HKIGRTSLKSKCKEAREKNSGNMFKIDYE  
TFLDGCARRYRE-ADDARVGLW-FDNADCLLERSDLHPMKNLINLVGQALGV-SPAEGTDFEEIWN5-KVKGRSTVRSICNAIENERKTYGPFSEDYK  
FLLVKVIYDE-TKAREG-LW-FETENKLFKFCGKTPPRKELKSTLISAVLIG-NLSDDEQKFLDEIWRK-QIKGRFTVASCARLETYKVSFGGGENIAN  
GTFELHVLCCYKQ-RQARDSRLYIMPEYRYDKLEKRYNNTGR-FDODNQLLYTCNHPKRYQILLNDLAGVLSVFNFLKDKIGSDDDLFTSKLVEHIRGFKKACEDSLKIQDNRGLLNHKIN

Figure S2 (Continued)



A-AK VS FTA LR KKILPE TTVFVGVK ADEESKLDVVARSGK AAGTARLRISVIVDALGELANGALIC SPEKLDK  
K-ND FN FDV LA KELIEK GSSFCGYKSSKNDFE YWENYKPTDVAACOVAALSKNAIGEDWK TKSEFYQITNSNKEQVSRITVDY  
R-TH FK FED I RKYLEK RLGIIHF SYNDKT INYKDS TVAGCPI TARFRKMLGEWE SFVVEGQKROARSKNN ISEHRVSY  
K-FN FD FED JA KALACK K NYAWIHDKERA YAFNYRTOGVGCP TLAQASIFGDWK TGLAETVTLIQKNGSKSLQKVD  
Q-EK HT HT HT RHLEIPE EMLFTGLPYHRS GSKAEKRLVHLAGI HEIRKALDKGPDPAVWDLRS RRDLIDS  
K-KK LT FKA MR ALLRQGV VDYVGENL EDKAKYELKGNATS IFRDARKNIGSADWL DEEEDS  
K-AK LS FEQ IR KLIKLSG SVQFN GANKEAYRE LEADAKYELKGNATS AALARKEFLGAANSGE DEALQDE  
R-PK CG FEM LR KIVF KVLKLDGERFTLETGVRD TIESEARK ELKGC DTAAKAKVNAIGTRWQAL SLDEQDR  
KKPTK SLSMWLK LPA LA RAIGLR GVKFTAETE AIACDEPLRSPAHP DRFGPWS ILLDADAOW EVISR  
ANVE WPA LR ELLEPP GTGFL ESDRRLKRGDATG RYNGAQAARGFAGNL TEALAPLIFGWSGW DLDRKDR  
RPPKAGRKPQKVSPPFK LR IPYTR LR KLIGDMS PVCFAQ EDRRAALDQPTA ARLSDKKGF NKAWRGF PLEKQDA  
DPDGLAAKDR LR VK LR TLKLPB EARFNL ESDRRAALDQPTA ARLSDKKGF NKAWRGF PLEKQDA  
N-RE VE FRH LR KFLDLS NEIFKGLHYKGPKTAK KREATLDPNEPTEL EFDKVEAKKAWISLRGAALREALGNEFYCRFVALGKRADE  
R-EK LS FRH LR KFLDLS NEIFKGLHYKGPKTAK KREATLDPNEPTEL EFDKVEAKKAWISLRGAALREALGNEFYCRFVALGKRADE  
N-GT LT YKQ TK KLGLSD DYEFGS EGTFTYEFKYEK IK ALGEHNL SODDLNE  
HHH  
EAQXN ENGLT YSK LK LLLDLS DFEFLGLDYS GKNPEKAVFLSLPST FKINKITQ DRKIQDK  
TETSK YKT YKT LK NAFKAGLWGDVRFGLAYPSQADAEK TADPEQDFVKLPAW HELRKAFAAGHEALMQI STPALDGDPTLDDQ  
K-NK MT YD IR KLNLS DHEFKGLLYDPKSS LKQENLRLELDSY HKIRKLENIYVYKDGIRME NETDIDT  
K-NE IK YK IR KLLDIB EILFKABNLTCHKP SGNNEKRRFEMKSY HKIKSTIPTDIWGLHS NKESLON  
R-ST YK IR RQFIDP EYRNLVNYERDP ALYKQGEPAKSKLKNL ELGGEKLLGNALKA-KLDMFG PDWPARPRKQEIHAVHERLMAADYGETPDKKR  
Q-AS MS WPGVSAIK LR KLLGSE QAFKGLRYGKN AEASTMEMKAY HAISHALEKEGLKDKASPLNL SPELQDE  
K-SK LT YAO AB LR KLLGSE QAFKGLRYGKN AEASTMEMKAY HAISHALEKEGLKDKASPLNL SPELQDE  
K-SK LT YAO VR LR KLLGSE QAFKGLRYGKN AEASTMEMKAY HAISHALEKEGLKDKASPLNL SPELQDE  
E-GK LT KAS LE LR KAISRAKGETVNSNYF TLHPDSEALYNPAVEVLQSGIGQLSPSVYRI HAIRKALENOGLKTWODLAR KPDLLE  
HTSES PT WDD VA RKLVEPR HRLGSSRAS LETGGITPPYDDT TVRMSAEVDMLADWDCANDES RGEIMDALSN AANRLRR  
QTERS LT WSD IA LEILKEN ESDLTSVPE EDGSSLAYSQAPF DETSARLAEEFAKRRKIPTEAOMWEO DRISDS  
PSSD IT WSD LC DFLGK PSOLKGVG LFDGGERISSPPR LTVORLYESDKIRKPLVAMKSA SDNEEA  
EQTK IT YGK LR KLLDK DRESFAGLYGLN RBRNKEDTV FVEMKFKY-EVRKALORAGVETQD LSI  
QQE LS FRK IV KEL E RAGCFEGKGLNMDKRAAL GRRDLSGNTT LAARWKLGLEDKQELDEVTOVINFLA DLGQPE  
KKE IT FSA IPKLL KAEFDLR EGIDFLNGMSPKD KIKNET LQLOKSL LLDMELETRMTVOLDESETGETVEREVAV VTDYSVR  
SSAS LS FAA LKLL KRALIA DQUTSKSGLGNSTRVALA SALOPYQYHH LQENLQESSMVNTETGELIPM ISLSEFO  
THEK LK GSD LKLL GLSXYGR LGEQFKGICQNKTRVELE RALGNYPDKR LLQENLQESSMVNTETGELIPM ISLSEFO  
ESKD FN FEK IPKHL K LFEKFRFDITTKV PACTT ISOLRKLK PHVWEE  
SRDK VT FDQ KRAL CLD DSNFNLBNRDF LIGNPT AVKMSKN RFGKLWDE IPL  
GKS LT FKA VRKTL KIL PHALVNFPEAGEK HHHHHHHHH HHHHHHHHH HHHHHHHHH HHHHHHHHH HHHHHHHHH HHHHHHHHH  
HHHHHHHHHH HHHHHH HHHHHHHHHH HHHHHHHHHH HHHHHHHHHH HHHHHHHHHH HHHHHHHHHH HHHHHHHHHH HHHHHHHHHH HHHHHHHHHH  
BELEFANRYOC TAKANG LW FP ENALLERADLHPMKNKILNVIQCALGV SPAEGTDFIEETWNS KVKGSTVRS ICNALENERKTYGPFSEDKYK  
RLYFAGNYELRQKVRAG LW PVKNDLSL LEUNSMKRCNHPKKNQNLHNVAGILGY KLDZAKFAEFKELWASA KVNKKLSAYCKONEELRKHGNTFKIDLE  
VLOIASAYNKE REDAOC IW FDMAFSCELSNINPPRQKILPELVGAILSEDFINNKDWAFAKFIWNT HKIGRTSLAKSKCKETEARKNSSNAFKI DYE  
TELDARRYRE ADDAKV LW FDMADCLLERSDLHPFKKILPELVANILCT DSTGKFLDEIWRK QIKGRETVAASRCARLETVARKSFGGFIANV  
FILLVKNIVDR TREAREG LW FETENKLFKCGKTPPRKIKSTILLSAVLGR NISDDEQSSFTIEEFWKSQTPKIERNVRCRCLASOVOKTYGYLAK EYG  
GFPLHVCYYIQ QRARDRLYIMPEYRYDKLKYNTGR FDDDNOLLTYCNHPRQRQYLLINDLAGVLOYSFNLKDKIGSDDDLFI SKWLVHEHRGFKRACEDSLKIQDRNRLNHLXIN

Figure S2 (Continued)



227494853	<i>Actinomyces coleocanis</i> DSM 15436#	-----AEK-----LTEGTAEEV-----EVAEFLQNL-----
328956315	<i>Coriobacterium glomerans</i> PM2	-----IILW-----NTLF-EDRK-----ILSQRLKEEYGSR-----LS-----
227824983	<i>Acidaminococcus</i> sp. D21	-----IVER-----MTYS-DDTK-----RVRLWNNNYGT-----LT-----
303229466	<i>Veillonella atypica</i> ACS-134-V-Col17a	-----IITD-----ITIFGSK-----MLQTLNKFQSO-----LN-----
34762592	<i>Fusobacterium nucleatum</i> ATCC 49256	-----SILW-----KCIYGDKK-----IFEKKIKNEYGDI-----LN-----
374307738	<i>Filifactor aloocis</i> ATCC 35896	-----IIKW-----ITIVGDKK-----MLKRVIRANYSNQ-----LT-----
320528778	<i>Solobacterium moorei</i> F0204	-----IIKG-----ITIVSDDKS-----MLRRWLKNNIKG-----LS-----
291520705	<i>Coprococcus catus</i> GD-7	-----IVIN-----VWLFGDKK-----LLKQRLSKMYPN-----LT-----
42525843	<i>Treponema denticola</i> ATCC 35405	-----IIRWAT-----IYDEGEKT-----ILKTRIKAEYGY-----CS-----
304438954	<i>Peptoniphilus duerdenii</i> ATCC BAA-1640	-----IIKL-----IVLYEDDKT-----YLAKKIKSAYKND-----FT-----
224543312	<i>Catenibacterium mitsucakai</i> DSM 15897	-----IIYD-----LTVF-EDKK-----IMKRRLRKKYA-----LP-----
116628213	<i>Streptococcus thermophilus</i> LMD-9	-----IIHT-----LTIF-EDRE-----MIRKRLNYSYD-----IF-----
24379809	<i>Streptococcus mutans</i> UA159	-----IVLT-----LTLF-EDRE-----MIERLKTAYH-----LF-----
13622193	<i>Streptococcus pyogenes</i> SF370	-----IVEL-----QTVF-EDKE-----TLLHQLQLEG-----IS-----
310286728	<i>Bifidobacterium bifidum</i> S17	-----ITEL-----QTVF-EDKK-----VIRROLDQLDG-----LS-----
366983953	<i>Oenococcus oeni</i> DSM 17330	-----IIKV-----QTVF-EDKK-----SIKNYLKFQFG-----HLEILD-----
422884106	<i>Streptococcus sanguinis</i> SK49	-----IIEI-----QTVF-EDSK-----IASREL-SKLP-----LD-----
339625081	<i>Fructobacillus fructosus</i> KCTC 3544	-----IIFL-----KTVYGDKR-----FVKEEIVEKYG-----DEID-----
306821691	<i>Eubacterium yuzii</i> ATCC 43715	-----IIER-----LTIF-EDKQ-----ILNEKLSSKY-----SYT-----
336394882	<i>Lactobacillus farciminis</i> KCTC 3681	-----IIOW-----ITIF-EDKK-----ILVOKLKECYP-----ELT-----
323463801	<i>Staphylococcus pseudintermedius</i> ED99	-----IIYW-----IYV-EDRE-----ILHLKIQEKYP-----SIT-----
389815359	<i>Planococcus antarcticus</i> DSM 14505	-----IIEK-----ITHTGNKK-----LKKYIEETYPD-----LS-----
258509199	<i>Lactobacillus rhamnosus</i> GG	-----HHH-----HEE-----HH-----HHHHHH-----
169823755	<i>Finnegoldia magna</i> ATCC 2932#	-----I-----SNI-----LNKE-S-----TIQDRIRILEGYEPEFSNLK-----
Jnet	227501312	-----VDSANL-----KEFS-DSNK-----LPERIQKQKDGFLKFEQ-----
47458868	<i>Mycoplasma mobile</i> 163K	-----LDSIYLAISYSSDUKERNE-----WPKLKLKELYPKIKNNLEIENV-----
284931710	<i>Mycoplasma gallisepticum</i> str. F	-----LDAICILIDREKS-RQDE-----VKKLTKRNIFEVLIKIDREKQLDV-----
71894592	<i>Mycoplasma synoviae</i> 53	-----LDNLYLFLYQKESNKDS-----SIDLFIAKNESINENIKLUKEFL-----
363542550	<i>Mycoplasma ovipneumoniae</i> SC01	-----IGYI-----MTIN-TDRE-----AMAEAFQKSWID-----LS-----
384332286	<i>Mycoplasma canis</i> PG 14	-----LAKV-----LTLN-TERE-----GIENLAFE-----LP-----
238924075	<i>Eubacterium rectale</i> ATCC 33656	-----IAEI-----LTIY-QDKD-----SIKSKLFEIDI-----LL-----
315149830	<i>Enterococcus faecalis</i> TX0012	-----IAEI-----LTKT-KDIE-----GRKKQISE-----LS-----
116627542	<i>Streptococcus thermophilus</i> LMD-9	-----IGIV-----LSQA-QTPK-----RRREKIKALNIG-----LD-----
315659848	<i>Staphylococcus lugdunensis</i> M23590	-----IGEI-----LTYI-KNDD-----SRSSRLKEIG-----LS-----
160915782	<i>Eubacterium dolichum</i> DSM 3991	-----IGTA-----LTYI-SSDK-----RRRYFAEELN-----LP-----
325677756	<i>Ruminococcus albus</i> 8	-----IIEI-----LTFN-KSDK-----TIESNLKLE-----LS-----
225377804	<i>Roseburia inulinivorans</i> DSM 16841	-----LWHI-----LYSI-EDKQ-----EIEKALHSFANKNN-----LS-----
336393381	<i>Lactobacillus coryniformis</i> KCTC 3535	-----LWHL-----DYSNDYADKE-----KTEKSLISSLGNKRNKWEKSKN-----
310780384	<i>Ilyobacter polytropus</i> DSM 2926	-----LWHS-----LYSF-EGDNTPTGNRGLQKMTLEYCF-----
301311869	<i>Bacteroides</i> sp. 20 3	-----LAEI-----LQSS-KTPQ-----EALDRLRALMAGK-----LD-----
385811609	<i>Ignavibacterium album</i> JCM 16511	-----FLYD-----WNSC-PDEK-----LITEKLSNEYH-----LT-----
60683389	<i>Bacteroides fragilis</i> NCTC 9343	-----FFSA-----WINT-PDDK-----RLSKYLKMKHLL-----LT-----
319957206	<i>Nitratifactor saluginis</i> DSM 16511	
187250660	<i>Eubismicrobium minutum</i> Peil91	
325972003	<i>Sphaerochaeta globus</i> str. Buddy	

Figure S2 (Continued)

SDADNEK--LDSI--SILPIC--RAAYSVDLSERUKRMI--ENGE--DLFEAR--  
AEQIKI--ICRK--RFTG--MGRLSKELIGIVQVD--EDSV--SMDVL--  
ADVKU--ISRL--RKHD--FGRLSKMEIGLGVYK--ETGE--RASILD--  
DETIK--LSKI--RYRD--MGRLSKELIGIDGDK--ACN--GAPKT--  
DEIKK--INSF--KFNY--MGRSEKLLGIEFINL--ETGE--CYSSVM--  
EEMKK--ITGF--QYSS--MGNFSKMLGIGSSDV--STGE--TEDIIT--  
ENDVY--LAKL--NYKE--MGRSKULLDIYTFN--EDGE--AGSILD--  
TGOIAG--ICSL--SYQG--MGRSKTELEETVPAP--GTGE--VNIIMT--  
DEIHK--IINI--KFSG--MGRLSRKFLETWTSEMP--GTGE--GSEEV--  
DEIHK--IAAL--NYAD--MGRLSKRELIGIEVDK--TIGE--KGSIIY--  
DDVKO--ILKI--KYKD--MGRSKKLLDGIADNR--FGS--SVTVLD--  
DKSVLK--LSRR--HYTG--MGLSAKLINGIRDEKS--GNLIL--DYLIDD--  
TMEQVK--LERR--HYTG--MGRLSAELIHGIRKES--RKTIL--DYLIDD--  
DDVMQO--LARR--RYTG--MGRLSRKLINGIRKOS--GKYL--DFKSD--  
EADCAL--LVNT--HYTG--MGRLSRKLITWAGECKI--SDFAPR--KHSITE--  
DHNREK--LSRK--HYTG--MGRSKKLLITKIVQAD--KIDNQIDVPRMOSIID--  
EEKIQ--LSKL--RYSG--MGRYSAKLLIDRDED--T--GNLQA--  
DDQVK--LSQT--HYTG--MGRSEKLLDSKI DERG--Q--KVSILD--  
KDKIK--ILGF--KFSN--MGNLSKSFIEGADVGT--G--E--VRSIQC--  
PQIKK--IENN--RYKG--MGRLSKILMDITTEINT--POLLIS--NVSILD--  
SKQNO--LAKI--NYSG--MGRSEKLLIETAY--Q--GHSITE--  
DVORQ--LALV--KLPG--MGRFRLMIDGILDE--Q--GQSVLD--  
PKKINE--LSGI--RYRG--MGRFRLMIDGILKEN--GHTVTC--  
SSQIQ--IINI--KYKD--MGRLSKLLDGIKPKK--ETE--KDTVTI--  
HHHEH--HEH--HHHHHHHHHH--H--HHHHH--  
KDVKSEIYSEIAKLREFS--TSSISFGAYAFI PNL--SEGS--KNYSIT--  
TDKDEK--ILAQ--THSLSRAMELAI TRM--NL--DNEDN--  
KSIFS--NTRK--NFKK--IGNFSLKAIREFLPRWF--EONK--  
LGAGN--EFENH--SK--THSLSKKAIDEILPKLL--DNEGN--  
DDVKQCLINRKTNGALFNK--MGSFLKIMNELI PEMY--AQPK--QNTLLT--  
ELSESVK--LLVLDRYKELSQSISTQSMHRESLKTLLHILPELM--NAT--SQONTI--  
DCFSQKQVDELVQFRKANSSIFGKGWHNYSVKLMWELIPELY--ETSE--QNTIIT--  
NEEDKEN--IAQL--TCYTG--THRLSLKIRLVIEQW--YSSRN--  
SDNEES--VQIAGLTKFTA--YHLSFKALRI INEEML--KTEIN--OMOSIT--  
DGLIN--E--LTKI--KLSS--TANVSYKVMOSIEAFC--EGD--LYGKYQ--  
GGEIDGL--LDLS--PAK--YQVSVLKAWRKQO PYDE--DGL--IYDKAC--  
ABLIE--K--LLPL--NFSK--FGHLSIKSMONII PYLE--MGO--VYSEAT--  
REDIIEI--LSEF--FSS--TIANLSLKAIKRILPYLE--KGI--SYNEAC--  
EEFIE--Q--FKNE--PPPKK--YGSYSAKAIKALPLMR--MKG--YWS IEN--  
YDVFNLEVAKATIANLPLKKE--YGSYSALAIRKALVWR--DGKYWHQFDQIAQOENUSLMLFDKNLIDUTNORVLANKYLITLAEVQKHSITLIKQALNEIENPYKLELWS--  
TDREK--LLEI--FRKRQS--YGSLSAKAIKALPLK--EGN--RYDVAC--  
EPEIDNA--FNEI--VJSSS--TRELSHRVILEALPLFL--EGY--DEKVOQ--  
ENEVYDA--LKTV--SLIGD--YAPIGKSAMLI ILEKIK--NDL--SYTEAV--  
ENEVYDA--LKTV--SLIGD--YGPICKTATQILMKHLE--DGL--IYTEAL--

Figure S2 (Continued)

296446027	<i>Methylosinus trichosporium</i> O83b	IAEV---ISFR-SDIG---RISEGLAAGCNAP---LV---
347536497	<i>Flavobacterium branchiophilum</i> FL-15	---KDLWHL---LAVA-TSDV---YLFEFADKLG---LD---
345885718	<i>Prevotella</i> sp. C561	---SIEDIMHF---CYDA-EEPE---AVLAFQETLR---LE---
282880052	<i>Prevotella timonensis</i> CRIS 5C-B1	---DVMNV---LTFY-KNED---EILPRLESYG---LS---
312879015	<i>Aminomona paucivorans</i> DSM 12260	---FILM---LQDDQKGGD---EVRSLTQOYG---LS---
294086111	<i>Candidatus Puniceispirillum marinum</i> IMCC1322	---IVWQ---LWTE-EGEG---ALIAWLGHTG---VD---
330822845	<i>Allicyclophilius denitrificans</i> K601	---LVCL---LDBG-ENDA---VLADALREHYG---LT---
344171927	<i>Ralstonia syzygii</i> R24	---IRR---VQSD-AEHA---ALLALIGDRGPR---VT---
159042956	<i>Dinoroseobacter shibae</i> DFL 12	---VVEL---LWAAQRDRS---ALLALIGDRGPR---VT---
83591793	<i>Rhodospirillum rubrum</i> ATCC 11170	---LVEL---LWTE-AEPE---RAIAALFARWA---LD---
288957741	<i>Azospirillum</i> sp. B510	---YVRD---VWARGADSA---ALARLLAEGAGHVP-PVA---
427429481	<i>Caenispirillum salinarum</i> AK4	---IVAR---LEET-EDEN---ELIAWLEKCEA---L---
92109262	<i>Nitrobacter hamburgensis</i> X14	---IVAL---LESA-LDEA---ALIASLGTHS---LD---
148255343	<i>Bradyrhizobium</i> sp. BTAl1	---ATKI---LTVY-KDEG---OKRRELTKLP---LE---
34557790	<i>Wolinella succinogenes</i> DSM 1740	---IAKD---ITLI-KDEI---KLKALAKYD---LN---
218563121	<i>Campylobacter jejuni</i> NCTC 11168	---HHH---HH---HH---HHHHHHH------
Jnet	218563121	---IANI---LGAN-KDWE---ALLKELESQ---LS---
291276265	<i>Helicobacter mustelae</i> 12198	---IANI---LGAN-KDWE---ALLKELESQ---LS---
222109285	<i>Acidovorax ebreus</i> TPSY	---IATV---LSVY-KDGA---EVOQLRQLA---LP---
365156657	<i>Bacillus smithii</i> 7 3 47FAA	---FGYA---LTFI-KDDE---DIVAYLQNEYITKNGKRVSNLA---
220930482	<i>Clostridium cellulolyticum</i> H10	---LYC---LTVY-KDNW---EIKDYIQANN---LP---
297182908	uncultured delta proteobact. HF0070 07E19	---AARL---ITLI-KDDE---KLSQDLADL---LP---
154250555	<i>Parvibaculum lavamentivorans</i> DS-1	---VIIL---SS-KDRK---AHREAAASF---VA---
218767588	<i>Neisseria meningitidis</i> 22491	---IGIA---FSLF-KTDE---DITGRUKD-R---IQ---
15602992	<i>Pasteurella multocida</i> str. Pm70	---IGTA---FSLX-KTDE---DIQQYLTN-K---VF---
187736489	<i>Akkermansia muciniphila</i> ATCC BAA-835	---CGG---SEPDVEDE---EVNELISSA---KE---
315605738	<i>Actinomyces</i> sp. oral taxon 180 str. F0310	---LVAA---LADNSIAGE---EEQELIVH---LP---
117829158	<i>Acidothermus cellulolyticus</i> 113	---MIRL---LSNT-VDID---KVEDVAYASAREFIDGLD---
189440764	<i>Bifidobacterium longum</i> DJ010A	---IGWI---LSV---WKS---DDNRKRLSTL---GLS---
403744858	<i>Allicyclobacillus hesperidum</i> URH17-3-68	---QLDQ---DWSI---RFM---GKN---GRPRNFSDFVAFM---
407803669	<i>Alicanivorax</i> sp. W11-5	---NRQLE---LWNI---LYNE-KGNE---YDLTSRSTSKVLEFINKYG---
423317190	<i>Bergeyella zoohelcum</i> ATCC 43767	---KPLYR---LWMI---LYS---IEE---REMRALITQLGME---
402847315	<i>Porphyromonas</i> sp. oral taxon 279 str. F0450	---EPLYR---LWHV---LYS---IDD---REQLQSVLRQKFGIDD---
404487228	<i>Barnesiella intestinihominis</i> YIT 11860	---XREE---IWHC---FYF---YDD---NTLLEKIKQDYALQI---
374384763	<i>Odoribacter laneus</i> YIT 12061	---EEQDL---LIET---IIT---ADE---DDAVEVIKKYD---LI---
384109266	<i>Treponema</i> sp. JC4	---AEKDI---FVGG---LLD---EAD---EERLRRIYV---ENRLS---
402849997	<i>Rhodovulum</i> sp. PH10	---HHH---HHH---HHHHHHH---HHHHH---HHHHHHHH---
Jnet	331001027	---FVK---TALKEGKTEKELSKFAAVI---KVLKMW---SEVVFYKGLR---
331001027	<i>Parasutterella excrementihominis</i> YIT 11859	---ELR---KKDEAELSKEKAKL---RLTDDVI---LNEWSOKIANFFD---
34557932	<i>Wolinella succinogenes</i> DSM 1740	---EAL---NHPEH---SNKALI---ALIQTI---EDIIQAIQSHLG---
54296138	<i>Legionella pneumophila</i> str. Paris	---TAQYREVNKLPRN---AQDKELL---TIRDRV---AETADFTAAANLG---
319941583	<i>Sulterella wadsworthensis</i> 3 1 455	---LQQ---LHKLEAGKK---LDDKPLA---LLYKNS---GLIASKICEALN---
254447899	gamma proteobact. HTCC5015	---IAR---NTKKGCKEKEIFNLICKIE---GSEDKK---GNYKRGLAYELG---
118497352	<i>Francisella novicida</i> D112	-----

Motifs  
informative positions

Figure S2 (Continued)

DALTA--AASDREDDPTG--AGHSSKAARNILSGLR--QGM--TYDKAC  
 EKNAK-A-FSKT--KLRKD--FASLSLSAINKILPYLK--EGL--LYSHAVFVANIENIV--DENIWRDEKQBDYAKTOISEI IENYTLKSRFEIINGLLKXKSENEDECK--RVYYSKEAEQSEFENDI  
 RKAAP-E-LVAIWS-AMPS--YAMLSSQKAIKRNINKILM--LGL--KYTSAVILAKYPELVWDEBELLSTAKDYIYVEAQVNY--DKRINSIVNGLLAKYKVSSEVRFADBNZYLLIDESDEKDI  
 SESAE-K-FAKI--KLSHS--FALSLSKAIKRFPLR--KGM--YITRASFFANLPTIV--GKEIWNREONRKYIENAVYCE--LVFNQPKAREVO  
 PENAR-A-LAPI--SEFS--TAHLSLSALGCKLPHLE--EGK--SYTQAR  
 DVVAEC-LVVR--LPSG--HGSLSKKAIDRILFVLR--DOGL--IYYDAV  
 EARQAQIWDVSI--PEG--YGNLSRKAARIVPALR--AAVI--TYDKAV  
 DAQID-T-LGL--SEFDS--HMRGRSALLRVLDAESGRDEGLPL--SYDKAV  
 RAHAETAHAPL--PDS--YGRIGLTAITRILXQLT--ADVV--TYADAV  
 EDEIYAE-VADAIQIVLPTG--RASLSAKAARAQAAMA--PGI--GYDEAV  
 EAT--AAKLAGAATLPDF--HGRYGRVAVABLLPVLETRCDPGRVPIRUDAV  
 AAAYPAATAAILESIMQPG--RYSYCPWAAZAILDAMANA--PTE--GYDVTGRIGLFGAFGIVIEDLRNRGALLA  
 DCAAAAR-VANT--TLPDS--HCRIGLRAIKKIVPIMQ--DGL--DEDEVA  
 EAAQRA-LSAL--GFS--FKLSLSKAIKRIILPAME--SGA--RYDEAV  
 AEMVE-R-LVXI--EFXD--FKLSLSKAIKRIILPAME--SGA--RYDEAV  
 QNQLD-S-LSKL--EFXD--HLNLSFKALKLVTRILM--EGK--KYDEAC  
 HHHH-H-HH--HHHHHHHHHHHHHH  
 KEQIQIKDAKI--NFSK--HINLSLEALYELLPLMR--EGK--RYDEAV  
 EPAASIVLEKI--SFDK--FSSLSKAIKRIIVPIMQ--SGL--RYDEAV  
 NKVYDKS-LIDELLSFSK--FALSLSKAIKRIILPAME--QGE--IYSKAC  
 LDYLEY-IAKL--PTFNK--FKLSLSKAIKRIILPAME--AGY--KYSKAC  
 SDKAITQ-LCEL--DFTT--AAKISLEAMYLILPBMN--QGM--GFFDVC  
 DEGITGEQAQIQALKPTG--WEPYSIPALNLFALAE--AGE--RFGALV  
 PEILKAL-LKHI--SFDK--FVQISLKAIRIVPIME--QK--RYDEAC  
 NSVINAL-LVSI--NFDK--FIELSLKRIILPAME--QK--RYDOAC  
 ADYADIP-LKPK--YATG--RPFYARVLYKAVVEIL--DGE--DTRPA  
 TAEDMLK-LELLAK-LPSG--RVAYSRLTSGVTRVWR--DDBV--DVHNR  
 DALE-A-LEGL--ALPSG--RVAYSRLTSGVTRVWR--DDBV--DVHNR  
 DDA--LTKLDSVLPDS--RAAYSRLTSGVTRVWR--DDBV--DVHNR  
 DNV--IEELPL--N--GSKFGLSLKAIKRIILP--FLEDGY--SYDVACE  
 NELRMTDGRDLSKMGFEFG--RSSYSIKALKALTEMMIAP--HMRSTP--EPRVDS--EAAIREC  
 NNIVDNEETAIRISKIPE--AAVSSLSKAVRILP--LVRSK--YENDESQQLSKILKILNENVEDPFAKAAQ  
 EDL-DGGLDOLYRLDFV--KPGYGNKSAKIFCILP--QLQGL--GYSEACA--AVGYRHS  
 DEV--LERSAIDL--KAGFNKSKAIRILP--FLOLGM--NYAEACE--AAGYRHS  
 NDL--EKIKKIRL--SESYGNVSLKARRINP--YLRAGY--AYSTAVLGGIRN--SEKRFEPYKYEPEIE  
 QEO--RDFIVANTLQSC--TSMJCKEVSXKLVKLE--ELADL--KYHEAVE--SLCYKFA  
 EDA--ARRCASIPLADG--YCRIGRTANTETLAAALVETDETGPVV--TYAEAVR--RAGERTG  
 HHHH-H-HH--HHHHHHHHHHHHHH  
 LS--DEAQQK--FDNLYSLAQYXLIETER--NGF--SAVLSLAHLENARMT--MFDG  
 ID--DKHQQR--FNNLSMAQLHVITDPR--SGF--SSTCKRKTAEHRFSRSTAFYNDTETGE  
 HN--DSOALI--YHNFESLSQYFYLETAR--DGF--HANCVAFTENYRSOKYIBDPEISY  
 LS--DEQK--FANPFLAQFYFYLETAR--SGF--SANTLAVHLENARMT--IKDAVIN  
 IE--PDEVSR--FASPHSLAQIFNIEGDDV--AGF--KATCRACRYENINMEQERVESLITNQLSE  
 VLLFGEPEASKPE--FDRKLIK--FNSIYSFAQIQOLAFEAER--KGN--ANTCAVCSADNARHQOIKITEPVED  
 \*\*\*\*\*

Figure S2 (Continued)

--REGCPVS  
 VNEFVSDWRPPA  
 GGRGAAVMEILLRDEELGQKVDFFRAFF  
 MNTNDNLMOQLSECTY--FSEITKIQEAYV  
 ELMRDSYHMEILGDKFSMECEHEENAKLA  
 EALREYVNIEMELSSKFFLOESIDNENEMW  
 AMWETDNLMOQLSKKFFMDVDFNSGRV  
 MNTVATIMELISNEVQ--EQNTENYKARNY  
 IMQINDNLMOQLSRNYG--FYNEVESRNTLAK  
 NITTAARSTONIMELISSEFETENIKKINSGETS  
 FMRSYNIEMELMSGHYT--FYEEVEKLNVPEN  
 VLEMSRLMEIINDKDLGVAQMBEANTSQPE  
 GLSNRNFMQLIHDH--ALSFKKKTQKQAI  
 GMSNRNFMQLINDH--ALSFKEBTAKAQV  
 GFANRNFMQLIHDH--SITFFKEDIQKQAV  
 IARAEDRNLMEIITDKQLGFSWTEQENIGAE  
 TYNTKNNIMEIINNAEDDFGVRAMIDKQNTT  
 FLRDEENRNLKILSDNTLSFEKIKDI-QSKST  
 KLSSTSONPMSIINDKYGQVAMITEQ--NTG  
 SLMWETNLMELISSRFTYHDELKRVKLEXP  
 LMAWANNPISIMSDKDYKRYIENH-NLNKRH  
 LRHSDENPBEIITNDVYGFQNFKEE--NQVQ  
 HMEQVSVYKVEVKNRKGEGLEKRIKQMNQVYD  
 ELMSSHNIHQIADET--LAKETVTEINQDKI  
 FLRNSSDNLMOQLIGSONSFNEYDKLKKYI  
 I-----HHHHHHHH-----HHHHHHHH  
 STEKALQONKN--NFSHSNLPEKTYW  
 QKNDKGNFEATKNDQKFDITKNNKLSIJ  
 NYESLKSNEBELAKKTEKALKAQON  
 NSEYLMKDEBEIRKESQKSELGTDKTY  
 NLEALKNVDEEIKNSQIEDNSSILMAKQK  
 EMGVYKQSEEPAG-----LKYTFV  
 LEQQLKSDVRRVRYSEYKLLPT  
 RLKQK-----TTSSENK  
 QMEIFTHLNKPKKI--NLTRAANKIPKAMI  
 IPELKNQNELSVRG  
 AKFNKEIPDIDENARPO--KLPPEKN  
 EAAGYDFRALNDGNKKH--LNGEENALIV  
 INTGYDFPKQIISK  
 EKADYDKNNGITKARGE--LIPWVDK  
 BRVOKAINTLDTIT--HFRALPLWJACYI  
 DQDLKQVLSKSFLEKKNESD--YAKGLKTYQAGYL  
 VYAGYRHSSESLI--REETANKVUKDRLI  
 GFDDRE  
 DYSRVYKSLRHLH  
 EALKEKELKKEQAQIKD--RLPYPGAVIQEST  
 ERMETGEFQELSVWQSSILFYQGLIGTST  
 CAADYDHTASRERG  
 AFDVCGHREALK  
 KKKLVIFYKSN--EIKNEQOETIFNELLPFIQQLKYE--FKIQRLOKQVLI FLKGNQEQICFTEERKGTAEKCKKI

Figure S2 (Continued)

227494853 *Actinomyces coleocanis* DSM 15436#  
 328956315 *Coriobacterium glomerans* FW2  
 27824983 *Acidaminococcus* sp. D21  
 303229466 *Veillonella atypica* ACS-134-V-Dol7a  
 34762592 *Fusobacterium nucleatum* ATCC 49256  
 374307738 *Fillifactor aloocis* ATCC 35896  
 320528778 *Solobacterium moorei* F0204  
 291520705 *Coprococcus catus* GD-7  
 42525843 *Treponema denticola* ATCC 35405  
 304338954 *Peptoniphilus duerdenii* ATCC BAA-1640  
 224543312 *Catenibacterium mitsuckai* DSM 15897  
 116628213 *Streptococcus thermophilus* IMD-9  
 24379809 *Streptococcus mutans* UA159  
 13622193 *Streptococcus pyogenes* SF370  
 310286728 *Bifidobacterium bifidum* S17  
 366983953 *Oenococcus oeni* DSM 17330  
 422884106 *Streptococcus sanguinis* SK49  
 339625081 *Fructobacillus fructosus* KCTC 3544  
 306821691 *Eubacterium yuzii* ATCC 43715  
 336394882 *Lactobacillus farciminis* KCTC 3681  
 323463801 *Staphylococcus pseudintermedius* ED99  
 389815359 *Planococcus antarcticus* DSM 14505  
 258509199 *Lactobacillus rhamnosus* GG  
 169823755 *Finegoldia magna* ATCC 29328  
 227501312  
 Jnet  
 47458868 *Mycoplasma mobile* 163K  
 284931710 *Mycoplasma gallisepticum* str. F  
 71894592 *Mycoplasma synoviae* 53  
 363542550 *Mycoplasma ovipneumoniae* SC01  
 384393286 *Mycoplasma canis* PG 14  
 238924075 *Eubacterium rectale* ATCC 33456  
 315149830 *Enterococcus faecalis* TK0012  
 116627542 *Streptococcus thermophilus* IMD-9  
 315659848 *Staphylococcus lugdunensis* M23590  
 160915782 *Eubacterium dolichum* DSM 3991  
 325677756 *Ruminococcus albus* 8  
 225377804 *Roseburia inulinivorans* DSM 16841  
 336393381 *Lactobacillus coryniformis* KCTC 3535  
 310780384 *Bacteroides* sp. 20 3  
 301311869 *Ilyobacter polytropus* DSM 2926  
 385911609 *Ignivibacterium album* JCM 16511  
 60683389 *Bacteroides fragilis* NCTC 9343  
 319957206 *Nitratifactor saluginis* DSM 16511  
 187250660 *Elusimicrobium minutum* Peil91  
 325972003 *Sphaerochaeta globus* str. Buddy  
 296446027 *Methylosinus trichosporium* OR3b  
 347536497 *Flavobacterium branchiophilum* FL-15

-----EPICARVGNPAVDRVLKAVNRVLMAAEAE-----WG--APL--SVN--IEHVREG  
 -----AENAQALG--VNELPGSPAVRHSINQSIRIVDEIASI-----AGK--APA--NIF--IEVTRDE  
 -----AKAQLSINDF--LDSMYISNAVKRPIYRTIAVNDIRKA-----CGT--APK--RIF--IEMARDG  
 -----QGOVNPBHD--IDELALSPAVKRAVQALRIVDEVVAHI-----KKA--LPS--RIF--VEVARTN  
 -----EVSYRDL--IEESYVSESLKRALIQTLKIVEEIKKI-----TGR--VPK--KVF--IEMARGG  
 -----GKIDKITDST--VKEMFLSPENKRAVQTIQVAEIKKV-----MGC--EPK--KIF--IEMARGG  
 -----DEKQNHHEE--LDDMYISPAARRSIQALRIVDEIVDI-----KKS--APK--KIF--IEMAREK  
 -----ETDLSYKT--VDELYVSPAVKROIWOTLKVKEIKQV-----MGN--APK--RVF--VEMAREK  
 -----DAEQFSYDGL--VKPFLFSPVKKMIWOTLKVKEIKSHI-----TOA--PKK--KIF--IEMAKGA  
 -----RELCEYEM--VDELYLSPVKKRMLWQSLRVVDEIKRI-----IGK--DKP--KIF--IEMARAK  
 -----DGKFTYEE--VERLAGSPALKRGIWQSLQVVEEITKV-----MKC--RPK--YIY--IEFERSE  
 -----IGDEKGNIKEV--VKSLLPGSPAIKKGIIQSIKIVDELVKV-----MGRKPE--SIV--VEMAREN  
 -----IGETD--NINQV--VSDIAGSPAIKKGIIQSLKIVDELVKI-----MG--HQPE--NIV--VEMAREN  
 -----SGQGD--SLHEH--LANLAGSPAIKKGIIQTVKVVDELVKV-----MGRHKPE--NIV--IEMAREN  
 -----NGSIMEV--VDDLRSVPKVRGIQSIIRLIDDI SKA-----VKG--RPS--RIF--LELADDI  
 -----DGDEQDVSJL--IDELAGPKEIKRGIQVSRILDDI IKA-----VGY--APK--RVY--LEFARKT  
 -----JEDDIFDE--IKKLAGSPAIKRGINSIKIVDELVQI-----IGY--PEH--NIV--IEMAREN  
 -----SSKLTPEKVNELTTPANKRGIKQSFVANDIKKA-----MKE--EPR--RVY--LEFARED  
 -----ISEWTTIED--LDDMYLSPVKKMIWQSMKIVDEIQTV-----IGY--APK--RIF--VEMTRSE  
 -----EDONISDL--VNDIHSVPAKRGITQSIKIVQEI VKE-----MGH--APK--HIF--IEVTRTET  
 -----SNKIQHOD--IANLTTSPALKKGIWSTIKVRELTSL-----FGE--PE--KII--MEFATED  
 -----TKKIRYED--IEELAGSPAIKRGITWRSVKIVEELVSI-----FGE--PA--NIV--LEVARED  
 -----KTDDEDVINDAYTSPSNKALQVLRVVEDIKKA-----ANGQDS--WLF--IETADGT  
 -----POEISYEVENLYVSPVKKMIWQVIRVEEITKV-----MGY--DPD--KIF--IEMAKSE  
 -----HHHHHHHHHHHHHHHH-----H-----EEE--EEEE--  
 -----EDLIASPTVKRSRQTMNLIKELIFKY-----SEKNLLEIE--KIV--VEVTRSS  
 -----KONRYLDDREFINDAILSPGVKRRILREATAKVFNAIQK-----FSE--EYDVTKVV--IELAREL  
 -----QKYLKDNELKEALVSLVKTSLVQAIKLFNQI IKN-----FGK--KYEISQVW--IEMAREL  
 -----KYLNPRIFODEIISPQTKNTEQAVIVLNQI IKK-----YSK--ENHIDAIT--IESPREK  
 -----KYLNDNFTKDAIILPNVVKVTPQAIILFNKI IQK-----FSK--DFEIDKVV--IELAREM  
 -----DVSSEDIENPVVRSVRSIFSKTILNVAI LK-----YK--ALD--TIV--IEMPRDR  
 -----KDVLAEIYNPTVKNKTVSQAKVIDALLVK-----YGKEQIR--YIT--IEMPRDD  
 -----TRKYIDEKLLTEEINPVVAKSROAIKIVNAAIKRE-----YG--DEFD--NIV--IEMARET  
 -----DEFILSPVVKRTFQAINLINKI IER-----YG--VPE--DII--IELAREN  
 -----MKNIQADDTAILSPVAKRAQRETEKVVNRLREI-----YG--EPD--SIV--VEMAREK  
 -----EDDCEFFKNPVVRSINETRKLINAI IQK-----YG--YPA--AVN--IETADEL  
 -----NDITNPVVKRSVSTIKVINAITQK-----YG--SQQ--AVN--IELAREM  
 -----DITREIYNPVVRVYTKI KIVEQI IRR-----YG--KPD--GIN--IELAREL  
 -----DLIANPVVTRATSQTRKVNVAI IRR-----YG--TPH--TTH--VEVARDL  
 -----DIDLKSPKQHSRNPIVEQVITETIRIVRDIWQQ-----VG--HID--EIH--IELGREM  
 -----KKLPNNSLRNPIVEQVIRETIFIVRDMVKS-----FG--IID--EIH--IELGREL  
 -----MLLPKNSLHNPVVEKILNQMVVINVI IDI-----YG--KPD--EIR--VELAREL  
 -----LREGNLFERENPINNHAVKSLASWALGLADLSWR-----YG--PPD--EII--LETTROA  
 -----GRIANPVVHQTINELRKLVNEI IDI-----LKG--KPC--EIG--LETAREL  
 -----DEKRYELEYGRIANPVVHQTINELRKLVNEI IDI-----LGA--KPC--EIT--VELAREL  
 -----DEKRYELEYGRIANPVVHQTINELRKLVNEI IDI-----LGA--KPC--EIT--VELAREL  
 -----RILQOERY SREIVGSP TARKALLES IKQVKAIVER-----YG--VPD--RIH--VELARDV  
 -----SIRKPNMAMRALHQLRKLVALILE-----CGQID--EKT--IIH--IEMAREL  
 -----KNRLKCLYHPSDIEKEKKI LKDFEGNEKIVLGSPLTPIKPNMAMRALHQLRKLVALILE

Figure S2 (Continued)

345885718	<i>Prevotella</i> sp. C561	IRQENSLGARWSLMDANEQTDILQKVRDRYQDFRSHERRFVSSPKLGESENYLTKKFPVSE
282880052	<i>Prevotella timonensis</i> CRIS SC-B1	-----GTIEMLKDKLANNFELP-----
312879015	<i>Aminomonas paucivorans</i> DSM 12260	ADAGVAAAPPDRUP-----KLPLLEK-----
294086111	<i>Candidatus Punicispirillum marinum</i> IMCC1322	XKAGLGEANLDPYALSDKLDYKGLAGRY-----
330822845	<i>Alicyclobacillus denitrificans</i> K601	-----EVEDI-----VHPTEGIRSV-----FKQLPYTKALQRHV-----
344171927	<i>Ralstonia syzygii</i> R24	AAQYPAFTADLNGERD-----ALFYYGELLNRYT-----
159042956	<i>Dinoroseobacter shibae</i> DEL 12	KACGWHSDGRTGCEFD-----RUPYYGVELRRHV-----
83591793	<i>Rhodospirillum rubrum</i> ATCC 11170	TLRLGLHSHRPFQERLA-----RUPYYAALDPWG-----
288957741	<i>Azospirillum</i> sp. B510	KLIRGGKDISDFREGALLDALPYGAVLERHV-----
427429481	<i>Caeniispirillum salinarum</i> AK4	ELPKYMAARTPNRAAGQGFPAFESVTFSQL-----
92109262	<i>Nitrobacter hamburgensis</i> X14	AKKRAYDHAKLFTGEGLRUPYYGQHLQDNY-----
148253343	<i>Bradyrhizobium</i> sp. BTAl1	SAAGYDEALLRGCGLSPKLPYLLPYGQWLQNDV-----
34557790	<i>Wolinella succinogenes</i> DSM 1740	-----IMLGVFHEKSAIIPPLAK-----
218563121	<i>Campylobacter jejuni</i> NCTC 11168	ELNLKVA INEDKKO-----FLPANNE-----
Jnet	218563121	HHHHHHHHHHHH-----
291276265	<i>Helicobacter mustelae</i> 12198	ELIQERGLSEKPKQPKMRO-----LPLPSELAK-----
222109285	<i>Acidovorax ebreus</i> TPSY	AOIETFGHSQRIFEGCAKXIMLPFYEAQRKYA-----
365158657	<i>Bacillus smithii</i> 7 3 47FAA	ELAGVNTGPKKEXAL-----LPLVTPM-----
220930482	<i>Clostridium cellulosyticum</i> H10	-----NNAELDFGSS-----KLEKCKLTV-----
297182908	uncultured delta proteobact. HF0070 07519	QOESLPELGVF-----PAGDKVFP-----
154230555	<i>Parvibaculum lavamentivorans</i> DS-1	NGPDKGQWRRTFNPHRNOPTGRIIDKLPSPASKES-----
218767588	<i>Neisseria meningitidis</i> Z2491	ALIIYGHY-CGKQTEKI-YLPPIPA-----
15602392	<i>Pasteurella multocida</i> str. Pa70	REIYGHY-GEANQKTSO-LPAPA-----
187736489	<i>Akkermansia muciniphila</i> ATCC BAA-835	RGCAFDFGELKADDCCLYCLLDVDSVWQKQK-----
315605738	<i>Actinomyces</i> sp. oral taxon 180 str. F0310	IRLIGVDFGAVPTP-----
117929158	<i>Acidothermus cellulolyticus</i> 11B	ATCFGVDNWRKPP-----
189440764	<i>Bifidobacterium longum</i> DJ010A	KTLFNVTDSWRPPA-----
403744858	<i>Alicyclobacillus besperidum</i> URE17-3-68	-----GRTXYKQALLP-----
407803669	<i>Alcanivorax</i> sp. #11-5	ESLAFPAQCGRSKLE-----
423317190	<i>Bergeyella zoohelcum</i> ATCC 43767	NOSVLSFEGVGNLSLMT-----IIVXDMETAKG-----
402847315	<i>Porphyromonas</i> sp. oral taxon 279 str. F0450	-----PFSSELYERTLLEKLP-----
404480228	<i>Barnesiella intestinihominis</i> YIT 11860	-----YTKAEAEARALIDRLP-----
374384763	<i>Odoribacter lanens</i> YIT 12061	ILKEWAGGSEVIRKIKOYLWHNFGFAKNDRAQKLY-----
384109266	<i>Treponema</i> sp. JC4	-----DQIVKDYLLP-----YCKVLFK-----
402849997	<i>Rhodovulum</i> sp. PH10	-----WHSDERDGVILDRLP-----YTGELDQR-----
Jnet	331001027	-----
331001027	<i>Parasutterella excrementihominis</i> YIT 11859	-----
34557932	<i>Wolinella succinogenes</i> DSM 1740	-----
54296138	<i>Legionella pneumophila</i> str. Paris	-----
319941583	<i>Sutterella wadsworthensis</i> 3 1 45B	-----
254447899	gamma proteobact. HCC5015	-----
118497352	<i>Francisella novicida</i> U112	-----

Figure S2 (Continued)

Motifs  
informative positions



```

RQWKLYHPSQITIRPVSYGKDRS--VLRNGNPDIGAIKRNFTVLRVLTNRRVNQLLDD--GVISP-DET--RVV-VETAREL
-AGATDKLYHPSMIETY-PNAQRNEFG--ILQLGSPRTNAINRPMAMRSLHLIRRVNQLLKE--SID-ENT--EVH-VEYAREL
-----ADWRNPFVFRALTQTRKVNVALVRR--YG--PPW--CIH--LETAREL
-----MGASCFEDSDEKRYGTISNPTVHIHALNQRVAVNDELIRL--HG--KPD--EVV-IEIGRDL
-----AFSGCPEDPDEKRYGKIANPTVHIHALNQRVAVNDELIRL--YG--RPT--EVV-IEIGRDL
-----IPGSYHFDLDDITRGRITNPTVHIHALNQRVAVNDELIRL--YG--KEA--QIV-VELARL
-----LDGDPVGPPEADCAAEAYGRIGNLSVHIALMTRKIVNALLAR--HG--KPH--QIV-VELARL
-----AFGTGNPADPEKRYGRVANPTVHIHALNQRVAVNDELIRL--HG--RPE--EIV-IEIARL
-----ITSLRBAKRAADWSAADPEERNPELRTWTGNAATDHIILNQRVAVNDELIRL--YGNRFGWDLPS--RIT-VELAREA
-----VSGDARDQKEKQYQFPNPTVHIHALNQRVAVNDELIRL--YG--PPT--EIS-IEFTRAL
-----VGSDDERTNERWGLPNPTVHIHALNQRVAVNDELIRL--HG--PPA--EIT-VELTROL
-----TYKDEVNPFVLRKALKEYKVLNALLIKK--YG--AFD--NVH-FELAREI
-----HHHHHHHHHHHHHHHH--H--EEE-EEEE--
-----EESYFDIPNFVLRRAISEFRKVNALLK--YG--GFH--YFH-IELTRDV
-----GKGDHIGSQFRDDADI PRNPFVLRALNQRVAVNALLIKK--YG--SPI--AVN-IEMARDL
-----EPIENVTNPFVLRALNQRVAVNALLIKK--YG--SPV--SIH-IEIARL
-----FDEMYNPNVNRVLSQRSKLNVAIDE--YG--LPY--MVN-IEIARL
-----REKISQLRNPTVTRQNEIRKVNALLIKK--YG--MPA--KIR-VELARL
-----DEIRNPFVLRALNQRVAVNALLIKK--YG--KPD--RIR-IEVGRDV
-----QEIRNPFVLRALNQRVAVNALLIKK--YG--SPA--RVH-IEIARL
-----ERRLDYMTNHNLRVRRMLLDRLKLDLIDQ--YG--SPA--RVH-IEIARL
-----APTEAPVGNPFDVLRNQRVAVNALLIKK--FADGDKRISRVCVEVGRKEL
-----PALHEATGHPVDRLNALLIKKFLSSATMR--WG--VPQ--TVN-IEHTREG
-----DPTEGFLGNPFDVLRNQRVAVNALLIKK--WG--PPQ--SIV-VELARGA
-----VTNEVVRALNQRVAVNALLIKK--HG--NPF--SVN-IEHVRS
-----PLGEGE--VTNEVVRALNQRVAVNALLIKK--HG--SPE--SIH-IEIARL
-----PPPEL--TNEVVDVALQVRETNMIDQ--IG--SVEA--QIV-VEMAREM
-----YSHDELYKSYKEINLLKQDLRNPVEQIILNEALVLRDITKN--YG--IKFN--EIR-VELARL
-----LLQRNE--LRQPLAVEKIILNQMIVNALLIKK--YG--ID--EVR-VELAREL
-----ALIKKNE--LRQFVVEKIILNQMIVNALLIKK--YG--REF--EIR-VELAREL
-----HHSQAITTQKQKRLPETGNLRNPTVQOGLNELRRVTKLATCREK--YG--SPK--HIH-VEMGREL
-----STMEIDLSAFEINPEKRYGKISNPTVHVALNQRVAVNALLIKK--YG--KPS--QIA-IELSRDL
-----HVVPGSGEPEKNEARWRGLANPTVHIHALNQRVAVNALLIKK--HG--RPD--QIV-VELAREL
-----EE--HHHHHHHHHHHHHHHHHH--EEE-EEE-EE--
-----SAQCCRLPADCVRFDFGTRKAI DRNSWEVAKRIAEVVKKSVDFT--NG--TVKI--PVA-IEANSFN
-----FHKKATATCQLRPADQRPFSCKIERYIDKGLYEYELAKIKAKKELEGMEA--K--EIKV--PII-LEQNAFE
-----ASRLPADSVRPFDFGVLARMQRILAYELAMAKWEQIKHIPD--NS--SLLI--PII-LEQNRFE
-----GETVRAQCRLPATARPFDCGLVRLVDRQAWELAKRVSTUQSKYDFS--NG--IVDV--SIF-VEENKFE
-----IHGERVPLKSAMCTRLSADSTRFFDCQMASIIEHLARKLAQHKIAGINDVPK--EF--SIDI--PII-IESNQFS
-----NKDKIILLSAKAQRLLPALTRIVDCAVKKAMATI LAKNIVDDNWNQIKQVLS-A--KH--QLHI--PII-TESNAFE

```

RUVCI I \*\*\*\*\*

Figure S2 (Continued)



227494853	<i>Actinomyces coleocanis</i> DSM 15436#	F-----ISKROAVEIDRENQKRY--QRNCAVRSQIADRHINATS-----
328956315	<i>Caribobacterium glomerans</i> PW2	DPK-KKGRFKRRYNDLKDALKAFK--KEDEFLWRELK-----
227824983	<i>Acidaminococcus</i> sp. D21	E----SKKRSVTRREQIKNLYSTR--KDFQOEVDFLEKILENKSJDG-----
303229466	<i>Veillonella atypica</i> ACS-134-V-Col7a	K----SEKKKDSRQKRLSDLYSAL--KDDVFLQSGLDKQKDFGALKSG-----
34762592	<i>Fusobacterium nucleatum</i> ATCC 49256	DES-MKNKTPARQEQKQKLYDSCG--NDTANFSDITKEMKNSLS-----
374307738	<i>Filifactor aloccis</i> ATCC 35896	E----KVKKRKSRAQELLELYACE--EDCHELILKEIE-----
320528778	<i>Solobacterium moorei</i> F0204	KSA-MKKRTESTRKPTILLEYSCK--SQADCFYDELEFEKLSNESNS-----
291520705	<i>Coprococcus catus</i> GD-7	Q----EGKSDSRKQQLVELYRACK--NEEDWITIELN-----
42525843	<i>Treponema denticola</i> ATCC 35405	E--LEPARTKRLKIQLDLYNCK--NDADA'SSEIKDLSGKIE-----
304438954	<i>Peptoniphilus duerdenii</i> ATCC BAA-1640	E--AKNSKRKRKILLEYFKGKAFINEIGEERYNYLLINEIN-----
224543312	<i>Catenibacterium mitsuokai</i> DSM 15897	E----AKRTESEKIKLENYKDLD--EQTKKEYKSVLEELKGFNTK-----
116628213	<i>Streptococcus thermophilus</i> LMD-9	Q-----YT-----NOGKNSQ--QRLKRLKLSLKEIGSSKILKENIPAKLS-----
24379809	<i>Streptococcus mutans</i> UA159	Q-----FT-----QKQKNSR--ERMKRIBEGIKELGSGIILKEH-----
13622193	<i>Streptococcus pyogenes</i> SF370	Q-----PSGRTISRKRIQLDLYRNANLGEFKGIADBLN-----
310286728	<i>Bifidobacterium bifidum</i> S17	Q--ESHLYNSRNQSLTLLKNA--GLSELVTVQVSOY-----
366983953	<i>Oenococcus kitaharae</i> DSM 17330	MTTEEGQKAKTRKTKLESAL--KNIEN--SILENGKV-----
422884106	<i>Streptococcus sanguinis</i> SK49	Q--TSVRSVPRVNLKLEKYQKLSSEAKVLKKTLD-----
339625081	<i>Fructobacillus fructosus</i> KCTC 3544	G--EKVTKSRKDRILKELY--NGIKEDSKQWVKELD-----
306821691	<i>Eubacterium yurii</i> ATCC 43715	K----KSEITTSREKRIKRLS--KLLNKAMDFKPOLRE--YIVENKKIQE-----
336394882	<i>Lactobacillus farciminis</i> KCTC 3681	Q--QKGGKQSRQIWDNTKKNKLSVDEYKYIIDV-----
323463801	<i>Staphylococcus pseudintermedius</i> ED99	G--EKKRTKSRKQWEELTK--TTLKNDPDLKSFICE-----
389815359	<i>Planococcus antarcticus</i> DSM 14505	G--TAGKRTQSRQIQIOTVYANAQOELIDSAVRGELEDKI-----
258509199	<i>Lactobacillus rhamnosus</i> GG	E--EKYTTISRKMLLDLYKAIKKDRDSQYKELLFGLN-----
169823755	<i>Finneyaldia magna</i> ATCC 29328	E-----HHHHHHHHHHHHHHHHHH--HHHH-----
Jnet	227501312	N-----NKHERRKIEGINRYRK--EKYEELKRVYDLP-----
47458868	<i>Mycoplasma mobile</i> 163K	S-----EKELENTKRYKLIK--KNGDKISEGLKALGISEDEIK-----
284931710	<i>Mycoplasma gallisepticum</i> str. F	TKP-NLEKLLNNAI'NSNIKILKEKL--DQTEKDFDPTKKKEID-----
71894592	<i>Mycoplasma synoviae</i> 53	NDK-KTIEEIKK--RNKKGKGTLL--EKLFQILNLENKNGYKLS-----
363542550	<i>Mycoplasma ovipneumoniae</i> SC01	TQD-QENDALKG-IAKAQSKKSLV--EERLEANNIDKSVFN-----
384393286	<i>Mycoplasma canis</i> PG 14	N-----SEEQKRINDSQKLINE--KEMEYIEKKI'AVTYGIKLSPS-----
238924075	<i>Eubacterium rectale</i> ATCC 33656	N-----EEDKKAIOKIOKANK--DEKDAAMKAAQYNGKAE'LPHSV-----
315149830	<i>Enterococcus faecalis</i> TX0012	N-----SKDKQKFINEMQKNE--NTRKRINEIIGKYGN-----
116627542	<i>Streptococcus lugdunensis</i> LMD-9	N-----SEEQKAI'IRERQKFFE--MRNKQVADIGDDR-----
315659848	<i>Staphylococcus lugdunensis</i> M23590	N-----KTFEDRAIDTRKNDNQ--KENDRIVKKEIIECIKQDE-----
160915782	<i>Eubacterium dolichum</i> DSM 3991	S-----KNFQDRVNI'EKMKRQ--QENERAKQOII'ELGKQ-----
325677756	<i>Ruminococcus albus</i> 8	G-----RNFKRGDIQKQDKNR--Q'NDKIAAELTEI'GT-----
225377804	<i>Roseburia inulinivorans</i> DSM 16841	A-----KSYD'RAQTII'ENKRE--LENEKTKR'FISEEFGIK-----
336393381	<i>Lactobacillus coryniformis</i> KCTC 3535	K-----NPADKRAMSQMIKNE--N'NIRKALLTE'FNPEFCIENVRPY-----
310780384	<i>Ilyobacter polytropus</i> DSM 2926	K-----NNSEERK'ISEQK'NF--QEKERAKLLKELNS-----SNFEHY-----
301311869	<i>Bacteroides</i> sp. 20 3	K-----KNAKREELTKSIAQ'IT--KAHEEYKTLQ'IEFGLT-----
385811609	<i>Ignavibacterium album</i> JCM 16511	L-----PEKIRKEIDKAMRERE--KALDKIIGKYYKKEF-----
60683389	<i>Bacteroides fragilis</i> NCIC 9343	K-----KSAEDRSKLSR'EQDNE--SNRNRIYEIYIR'PQQQVIITR-----
319957206	<i>Nitratifractor sahsuginis</i> DSM 16511	
187250660	<i>Elusimicrobium minutum</i> Peil191	

Figure S2 (Continued)

GYRGS DV --- TRY-LAIOR --- QMGE --- CLY --- CGVALTFVN --- SEMDHVPR  
 ETAPNDM --- DERU-SLYFM --- QRGK --- CLY --- SGRAIDHQLSNAG --- YVDHILPRT  
 QIQSDAI --- QUGR --- DMY --- YLYFA --- YDPIKLEHIXQSE --- YNDHYPQS  
 L --- ANYDAAALR --- SKKL-YLYYT --- QMGR --- CAY --- YGNIIDLMLNTDN --- YNDHYPRS  
 SYDNMSIQC --- KKL-YLYYL --- QFCK --- CMY --- TGREIDDLRLQNDT --- YDHIYPRS  
 DRDERDENS --- MKL-FLYYT --- QFCK --- CMY --- SGDDIDINELIRGNSK --- WDRDHYPQS  
 RLRRDQI --- YLYYT --- QMGR --- SMY --- TGRIDDFDKLINDKMT --- YDHIYPRS  
 AQSQQIURS --- DKL-FLYYT --- QMGR --- CAY --- SGETTQLDELMWTK --- YDHIYPQS  
 NEDNLRIS --- DKL-YLYYT --- QMGR --- CAY --- CGRPIEIGHEVFTSN --- YDHIYPQS  
 SEESKFNW --- DML-YLYYT --- QMGR --- CAY --- SLEPIDLADLASNI --- YDQHIYPKS  
 KISSDSI --- FLYFT --- QMGR --- CMY --- SKRIDDISDK --- YDHIYPQS  
 KIDNNALQN --- DRU-YLYYL --- QMGR --- DMY --- TGDHLDLRLSN --- YDHIYPOA  
 PVENSQIQN --- DRU-FLYYT --- QMGR --- DMY --- TCEERLDIYLSQ --- YDHIYPOA  
 ACSDDIQD --- DRU-FLYYT --- QMGR --- DMY --- YDQELDINRLSD --- YDHIYVPS  
 DAAALQN --- DRU-YLYYL --- QMGR --- DMY --- TGERLDLRLSSA --- YDHIYPOA  
 PHSDEIQS --- EKL-YLYYL --- QMGR --- DMY --- TLDKYGSPAPLYLDLQD --- YDHIYVPS  
 G-KNKMSD --- DRY-FLYFQ --- QMGR --- DMY --- TCRPINFERLSQD --- YDHIYVPS  
 SKDESYFRS --- KKW-YLYYL --- QMGR --- CMY --- SCEVELEKLMDDN --- LYDHIYPRS  
 ELKKNLSS --- ERI-MLYFL --- QMGR --- SLY --- SEESLNINKLS --- YVDHILPRT  
 ANKLNQIQD --- EKL-MLYYS --- QMGR --- CMY --- SCOSIDLDALLSPNATKHYEVDHIFPRS  
 IKSQDQFNE --- QRF-MLYVT --- QMGR --- CAY --- TCKALDIQNLSS --- YEVDHILPON  
 ADKASFT --- DRU-VLYFM --- QMGR --- DTY --- TGAFINIDOLSH --- YDHIYVPS  
 KLDDSDIAS --- RKL-YLYYT --- QMGR --- DMY --- TCEKIDDLKLFDSH --- YDHIYVPS  
 BHHH --- HHH-BHHBB --- E --- EEEEE  
 NENTLL --- KKL-MLRQC --- QGGY --- DAY --- SLRKEANDVINRPNW --- YDHIYVPS  
 DILKSPTS --- YXF-ILATQ --- QDHI --- DRY --- SLKETAADDIETKTEK --- FEIDHIYPS  
 KIENSWER --- NKL-FLWFS --- QDRK --- DRY --- TQLDIKINIEDE --- TEIDHVPYS  
 DLETRAKLI --- DRU-REYHQ --- QDGI --- DLY --- TLDKINIDQINGSK --- YEIEHIYPS  
 DKYEKLI --- YKI-FLWIS --- QDFK --- DRY --- TGAQISVNEIYN --- NK --- VEIDHIYPS  
 DFSQKLS --- LKL-KLWNS --- QDGI --- CLY --- SKGTIDNDIINWFQ --- FEIDHIYPS  
 QTI --- QKNRPI --- AKL-LYVYE --- QDGI --- CAY --- TGLPISPELIVSDS --- TEIDHIIPS  
 HGHQIA --- TKI-RLWHQ --- QGER --- CLY --- TERTISHDLINNSQ --- FEVDHIPLS  
 QNAKIV --- EKI-RLHDE --- QGER --- CLY --- SLESIPLEDLNPNHI --- YEVDHIIPRS  
 KNAKLR --- EKL-VLYQE --- QGER --- TAY --- SLEPIDLKLIDDPNA --- FEVDHIPIPS  
 VHARLI --- ERY-KLWEA --- QMGR --- CLY --- SGETTTEMLRDKL --- FEVDHIYPS  
 NPTQDI --- LKY-RLWND --- QGGY --- CLY --- SKRPIELEEDGG --- YDHIYVPS  
 PVNGQI --- IRY-KLHKE --- QNGV --- DRY --- TGDQIPERAFSEC --- YEVDHIYPS  
 NVKGLI --- LAV-RLYQS --- QGER --- CAY --- SRKELSEVILDES --- YDHIYVPS  
 SPSQDLRIYEGVINSILPEPIDIGIILGKFNQDTYKRPTRSEI --- LRY-KLWLE --- QKYR --- SPY --- TCEMIPLSKLFTPA --- YEIEHIIPS  
 DENG --- KLFSSVFNENPSPLDIENKFLWQNSGLTDELNKLDKLEKIPTELEV --- KRY-ILWLT --- QKCR --- SPY --- TCKLIPLSKLFNSV --- YEIEHIIPS  
 NVRTDI --- LRY-KLYES --- LESGGY --- TLY --- SNTYSREKLESKE --- FDEHIIPOA  
 PSDKLI --- ARKL-QLWEX --- QKGL --- DLY --- SKVINLSQLDGS --- ADIEHVPS  
 R --- ENPRVI --- LXP-ELLER --- QKSG --- CPE --- CGGQISNDIINNO --- ADIEHLFPIA

Figure S2 (Continued)

325972003	<i>Sphaerochaeta globus</i> str. Buddy	K	VGAEKREDIIRQQTQOE	KEAVLAYSKYCEPN
296446027	<i>Methylosinus trichosporium</i> OB3b	G	KSIEEREIEITIEGRDNK	RQKDKIRGLFEKEVGRPFQDG
347536497	<i>Flavobacterium branchiophilum</i> FL-15	N	DANKKGIQYQNDKN	KFREDALKEIKKLVFEDCKKEVEP
345865718	<i>Prevotella</i> sp. C961	N	DANKRALADROKEQD	DENEKIKKILEEFPKRDGISTDDIDK
282880052	<i>Prevotella timonensis</i> CRIS 5C-B1	S	QPAKVRRIETEFQANE	KKQQAERSFLDIVGT
312879015	<i>Aminomonas paucivorens</i> DSM 12260	P	MCADCKRELEFQKEGR	AKNERARDELAKLGH
294086111	<i>Candidatus Punicispirillum marinum</i> IMCC1322	K	QSRQKVEAQRQADNQ	ARNARIRRSIAEVLIGEE
330822845	<i>Alicyclophilius denitrificans</i> K601	K	AGLEEKERIKKQQTANL	ERNERIRQKLODAGY
344171927	<i>Ralstonia syzygii</i> R24	K	KSEEQKRADIKRIRDIT	EAAKKEKLELELEI
159042956	<i>Dinoroseobacter shibae</i> DFL 12	K	AGADERKRMIAFQAERE	RENAEIDVERLRSRDMW
83591793	<i>Rhodospirillum rubrum</i> ATCC 11170	K	RSADERRREDKRDQDNQ	KRNEERKULILSGE
288957741	<i>Azospirillum</i> sp. B510	K	LGSEOKAEROREORRNQ	RKESAAALDFCQENTVSWQAGG
427429481	<i>Caenispirillum salinarum</i> AK4	K	LSFRLAELEREQANQ	RNDKRYSLRLKGL
92109262	<i>Nitrobacter hamburgensis</i> X14	N	TKGETIEDIKESQKNE	KERKEAMWIAETSQV
148255343	<i>Bradyrhizobium</i> sp. BT411	G	KNHSQPAKIEKQENY	KAKDAELECEKGL
34557790	<i>Wolinella succinogenes</i> DSM 1740	G	HHHHHHHHHHHHHH	HHHHHHHHHH
218563121	<i>Campylobacter jejuni</i> NCTC 11168	C	KAKSARMQLEKINKNKK	SENDAASQLLEVLGL
Jnet	<i>Helicobacter mustelae</i> 12198	S	RFLDERNKVRAQAEFR	DRNDRARSEFERDEGY
291276265	<i>Acidovorax ebreus</i> TPSY	S	HSFDERKKIQKQYENR	KKNETALKQLIEVELTK
222109285	<i>Bacillus smithii</i> 7 3 47FAA	G	MTRQDRDNLIKBEHNR	KAREKISDLIRQNGK
365156657	<i>Clostridium cellulolyticum</i> H10	G	KGRELRERIKLDQL	DKSQNDQRAEDERAEEFQO
220930482	uncultured delta proteobact. HF0070 07E19	G	KSKRKRKEIQSGIRNE	KORRKAATEDLKNQIA
297182908	<i>Parvibaculum lavamentivorans</i> DS-1	G	KSEKDRKEIEKROENR	KDREKAAAKFREYEPNFVG
154250555	<i>Neisseria meningitidis</i> 22491	G	KSEKREIREIQQOENR	TKRESAVQKFELESDFSS
218767588	<i>Pasteurella multocida</i> str. Pm70	T	TFSAMDSKKIOREITLRO	KSHTDAVNRKRLKPLGK
15602992	<i>Akkermansia muciniphila</i> ATCC BAA-835	L	KSA-SLLEEREKWERF	BARREIROKEMYRKLGISG
187736489	<i>Actinomyces</i> sp. oral taxon 180 str. F0310	S	ESREKQAEKAEARRAHR	KANDRIRAEIRASGLS
315605738	<i>Acidothermus cellulolyticus</i> 11B	F	SSVAFARKKREYEKNN	EKRISIFSSLSQLRADQME
117929158	<i>Bifidobacterium longum</i> DJO10A	S	KNLDERPKIEKQAKENQ	KN-NEQIKDEIRE-ILGS
189440764	<i>Alicyclobacillus hesperidum</i> URH17-3-68	K	GGVTRNDIEQNKREA	SE-RKKAQSTEE-NGK-
403744858	<i>Alicyclobacillus</i> sp. W11-5	K	NSAKERATIHKRNKDNQ	TI-NMKIKETLVK-NKK-
407803669	<i>Alicyclobacillus</i> sp. PH10	K	MSREERERMARNKRE	ER-NGGVAAKIRE-CGL-
423317190	<i>Bergeyella zoohelcum</i> ATCC 43767	K	QSEKERSNTYKSIKNQ	RE-NEQIAKRIVE-YGV
402847315	<i>Porphyromonas</i> sp. oral taxon 279 str. F0450	R	SSKTEREKQSQIRENE	KK-NEAAKVKLAE-YGL-
404487228	<i>Barnesiella intestinihominis</i> YIT 11860	K	NNVEKKAELARKONORA	KENLAINDTISALYETAFFGK
374384763	<i>Odoribacter laneus</i> YIT 12061	K	LNREOKERLDRNKNR	EE-NERTAILAE-HGQ-
384109266	<i>Treponema</i> sp. JCA	F	HHHHH	HHHHH
384109266	<i>Rhodovulum</i> sp. PH10	F	TASLTDLKYIQLK	EOKLKKLEDI
402849997	<i>Parasutterella excrementihominis</i> YIT 11859	Y	EESLRKSKTGSND	RVINSKDRDGKEL
Jnet	<i>Wolinella succinogenes</i> DSM 1740	F	EESFKKIK-GSSS	DKTLEQ
331001027	<i>Wolinella succinogenes</i> DSM 1740	F	SASVADLK	K-NKRVKDKM
34557932	<i>Legionella pneumophila</i> str. Paris	F	TAELEIKRGRGS	AKAKKA
54296138	<i>Legionella pneumophila</i> str. Paris	F	EPALADVKKKSLK	DRRKK
319941583	<i>Sutterella wadsworthensis</i> 3 1 45B			
254447899	<i>Gamma proteobact.</i> HTCC5015			
118497352	<i>Francisella novicida</i> U112			

Motifs  
Informative positions

Figure S2 (Continued)

```

NLDKAYI-----CLHLSVADIAAGR-----ADVDFIFPRD
ANGKSEI-----TDDYLSPQVATDAA-----VOVDHILPWS
TEDDI-----EGRNISIICDILGSRPA-----YDIEHTILPRS
ARYVLDQREVDYFCSKYTKDI-----TCRTUNLSNLPDPN-A-----FDIEHTIPES
TPTDV-----TCEQIGITDFIGSNPK-----FDIEHTIPQS
APCGDI-----CEEYINPTRLARPGY-----AEMDHLIPFS
IDRSER-----TKNMSISDLPDK-----VEIEHLPPFS
RVRSDI-----SGVQISAAMLISDE-----VEVEHILPFS
PDRREN-----SCROISLQRLSND-----CVVDHILPFS
EDGCRN-----TCRTRISAAMI FDGS-----CDVDHILPYS
ANARER-----ISTPIGHADLLGA-----YDIDHVLPLA
RPTFRN-----SCETISMRRMLISEQ-----VDIDHILPFS
LEKAAA-----CAERPKLKATLFPRAE-----TEIDHVLERR
PANPRN-----TCEQISIERLISDE-----VDIDHILPVA
PASTHM-----TCEALIGLRIAVSDO-----VDIDHILPFS
PLTRKN-----TCEDVIELERLPDEGY-----CEIDHILPRS
KINSKN-----SGEKIKISDLQDEKN-----LEIDHIYPFS
HHH-----SEEEEEEE-----SEEEEEEE
PNTYNR-----SSEKETTIDHLKQORA-----LQIDHAFPLS
RPKAAP-----SQQPUDIQRVLDENY-----ACVDHAILPYS
NPTGDI-----SLKPIELERLLEPGY-----VEVDHILPYS
VASGLD-----SGKPIPCVDLLNDSL-----TQIDHIYPFS
NFRDDV-----SGRMIPVNSVLESD-----TQIDHILPIS
RFSKDI-----TQDQIGNALFREG-----YEVEHWPRS
BPKSDI-----SCKEINLGRINEKY-----VEIDHALPFS
ALSANL-----SCKEINIRINEKY-----VEIDHALPFS
PFRSDQ-----TCATYGDHELEN-----LELEHIVPRS
DPSPADL-----CGNEINFOT-----FEVDHILPVR
KVRSDI-----CGAPLSEW-----SELDHIVPRT
AVTGRDI-----CGRTLYFK-----CEMDHIVPRK
PPTPARI-----SCEKIDVTRLEFP-----GYAEVDHILPYG
ELSLAM-----CESNISLQALSG-----AYNPEHILPRT
YTPRPI-----TQOPIPLSDDLEDK-----EKYDVDHILPIS
PTRSRI-----CGRSLEEQCLRE-----GGMVEHILPKS
KAYRDI-----CCQPVYDGFDRG-----FDVEVEHILPKS
SFYPRNDR-----TCRTUNISHITLSSD-----NSVQIEHILPYS
RUTASMK-----CCKGLSGAELFTK-----EIEIEHILPFS
HHHH-----TCRAIGIAELFTS-----EVEIDHILPVS
QANEQCRMLSKS-----E-----EEB-----
AKAKNAEDRLAKKD-----TGRPLDDV-----GEIDHILPRS
ALEKONIQ-----CGDTIGDD-----GEIDHILPRS
LSEAKLETRMLKN-----RKAIGGC-----GEIDHIYPRS
KELGKSKAGWSKT-----TCDRLAEC-----GEIDHILPRS
ALERTSPENIFAKN-----TGAPIGGS-----GEIDHILPRS
NRIKEE-----SCANITDGFDFGAK-----SELDHILPRS

```

```

*****

```

Figure S2 (Continued)

22749853	<i>Actinomyces coleocanis</i> DSM 15436f	GLGSI	-----NTRDNVAVTCERCNK-SKSN-KPFAV	MAAEGSIP-GVSYAEALK	RVDE
328956315	<i>Coriobacterium glomerans</i> Pw2	Y-VKD	-----DSLNNKALVYREMG-RKIDMLLIDP	KIPRR#SGYWR	MLHE
227824983	<i>Acidaminococcus</i> sp. D21	K-VKD	-----DSLNNKALVWQSEING-EKSRYPIDA	ALFNKMPFAD	AYYN
303229466	<i>Vibriomella atypica</i> ACS-134-V-Col1a	L-TKD	-----DSFDNVLVICTANA-EKSDIYFIDA	ELIQAQKPEMA	ELKE
34762592	<i>Fusobacterium nucleatum</i> ATCC 49256	KVTKD	-----DSFDNVLVICTANA-EKSNIEYFVK	EIGERRKSPFR	FLAE
374307738	<i>Filifactor aloccis</i> ATCC 35895	K-IKD	-----DSIDNVLVWVYKINA-KSNELISEB	IQKMSFNU	SJLN
320528778	<i>Solobacterium moorei</i> FU204	K-IKD	-----DSTNNKLVKQKQING-EKDIYPISE	DIQKQOPFKK	LJAE
291520705	<i>Coproccoccus catus</i> GD-7	K-IKD	-----DSLNNKLVKAKTANA-IKSDTYFSL	DIQKQORFVK	KLQQ
42525843	<i>Treponema denticola</i> ATCC 35405	K-IKD	-----DSLNNKLVKAKTANA-IKSDTYFSL	DIQKQORFVK	KLQQ
304438954	<i>Peptoniphilus dwerdenii</i> ATCC BAA-1640	K-ITD	-----DSLNNKLVKAKTANA-IKSDTYFSL	KVIMKAYGFK	LJFD
224543312	<i>Catenibacterium mitsuokai</i> DSM 15887	L-VKD	-----DSFNRKLVVPEMG-RKLDLWVVF	DIDDMVRFVK	LJFD
116628213	<i>Streptococcus thermophilus</i> LMD-9	F-LKD	-----NSIDNKLVSASSNR-EKSDVPSLE	VVAKKATFY	QLLA
24379809	<i>Streptococcus mutans</i> UA159	F-IKD	-----NSIDNKLVSASSNR-EKSDVPSLE	VVAKKATFY	QLLA
13622193	<i>Streptococcus pyogenes</i> SF370	F-IKD	-----DSIDNKLVSASSNR-EKSDVPSLE	VVAKKATFY	QLLA
310286728	<i>Bifidobacterium bifidum</i> S17	V-TQM	-----DSIDNKLVSASSNR-EKSDVPSLE	VVAKKATFY	QLLA
366989933	<i>Oenococcus oeni</i> DSM 17330	Y-TKD	-----NSIDNKLVSASSNR-EKSDVPSLE	VVAKKATFY	QLLA
422884106	<i>Streptococcus sanguinis</i> SK49	F-LPI	-----DSIDNKLVSASSNR-EKSDVPSLE	VVAKKATFY	QLLA
339625081	<i>Fructobacillus fructosus</i> KCTC 3544	F-IKD	-----DSIDNKLVSASSNR-EKSDVPSLE	VVAKKATFY	QLLA
306821691	<i>Eubacterium yuzii</i> ATCC 43715	F-VKD	-----DSLNNKLVKAKTANA-IKSDTYFSL	DIQKQORFVK	KLQQ
336394882	<i>Lactobacillus farcinis</i> KCTC 3681	Y-ITD	-----DSLNNKLVKAKTANA-IKSDTYFSL	DIQKQORFVK	KLQQ
323463801	<i>Staphylococcus pseudintermedius</i> EP99	F-IKD	-----DSLNNKLVKAKTANA-IKSDTYFSL	DIQKQORFVK	KLQQ
389815339	<i>Planococcus antarcticus</i> DSM 14505	F-VKD	-----DSLNNKLVKAKTANA-IKSDTYFSL	DIQKQORFVK	KLQQ
258509199	<i>Lactobacillus rhamnosus</i> GG	L-IKD	-----DSLNNKLVKAKTANA-IKSDTYFSL	DIQKQORFVK	KLQQ
169823735	<i>Finnegoldia magna</i> ATCC 29328	KKKDD	-----SIINNVIVKAKTANA-IKSDTYFSL	DIQKQORFVK	KLQQ
	Jnet		-----HEHEHEHEHEHEHEHEHEHEHE		
47459868	<i>Mycoplasma mobile</i> 163K	I-SFD	-----DSFNNKLVKAKTANA-IKSDTYFSL	DIQKQORFVK	KLQQ
284931710	<i>Mycoplasma gallisepticum</i> str. F	I-SFD	-----DSFNNKLVKAKTANA-IKSDTYFSL	DIQKQORFVK	KLQQ
71894592	<i>Mycoplasma synoviae</i> 53	K-RAD	-----DSFNNKLVKAKTANA-IKSDTYFSL	DIQKQORFVK	KLQQ
363542550	<i>Mycoplasma ovipneumoniae</i> SC01	K-SFD	-----NSQAKKLVKAKTANA-IKSDTYFSL	DIQKQORFVK	KLQQ
384393286	<i>Mycoplasma canis</i> FG 14	L-CFD	-----DSBANKLVKAKTANA-IKSDTYFSL	DIQKQORFVK	KLQQ
238924075	<i>Eubacterium rectale</i> ATCC 33656	I-SFD	-----DANSNKLVVTRSEMG-KGSDQFPYD	MDGSEKILNGKFSMD	DYQK
315149830	<i>Enterococcus faecalis</i> TX0012	I-SLD	-----DSINNKLVVTRSEMG-KGSDQFPYD	MDGSEKILNGKFSMD	DYQK
116627542	<i>Streptococcus thermophilus</i> LMD-9	I-TFD	-----DSLNNKLVVYATANO-EKSDTYFSL	DIQKQORFVK	KLQQ
315659848	<i>Staphylococcus ingrahamensis</i> M23590	V-SFD	-----NSYNNKLVVTRSEMG-KGSDQFPYD	MDGSEKILNGKFSMD	DYQK
160915782	<i>Eubacterium dolichum</i> DSM 3991	I-SLD	-----DSLNNKLVVTRSEMG-KGSDQFPYD	MDGSEKILNGKFSMD	DYQK
325677756	<i>Ruminococcus albus</i> B	L-ILD	-----NTLNKLVVYATANO-EKSDTYFSL	DIQKQORFVK	KLQQ
225377804	<i>Roseburia inulinivorans</i> DSM 16841	I-TFD	-----DSYNNKLVVTRSEMG-KGSDQFPYD	MDGSEKILNGKFSMD	DYQK
336393381	<i>Lactobacillus coryniformis</i> KCTC 3535	I-SFD	-----DSYNNKLVVTRSEMG-KGSDQFPYD	MDGSEKILNGKFSMD	DYQK
310780384	<i>Liyobacter polytropus</i> DSM 2926	K-SFD	-----DSYNNKLVVTRSEMG-KGSDQFPYD	MDGSEKILNGKFSMD	DYQK
301311869	<i>Racteroides</i> sp. 20 3	K-YFD	-----DSLNNKLVVTRSEMG-KGSDQFPYD	MDGSEKILNGKFSMD	DYQK
385811609	<i>Ignavibacterium album</i> JCM 16511	K-MKN	-----DSLNNKLVVTRSEMG-KGSDQFPYD	MDGSEKILNGKFSMD	DYQK
60683389	<i>Bacteroides fragilis</i> KCTC 9343	K-LFD	-----DSFNNKLVVTRSEMG-KGSDQFPYD	MDGSEKILNGKFSMD	DYQK
319957206	<i>Nitratifactor salinus</i> DSM 16511	L-CSI	-----SITNNKLVVTRSEMG-KGSDQFPYD	MDGSEKILNGKFSMD	DYQK
187250660	<i>Kingimicrobium minutum</i> Pe1191	F-SED	-----NSANNKLVVTRSEMG-KGSDQFPYD	MDGSEKILNGKFSMD	DYQK

Figure S2 (Continued)



325972003	<i>Spiseraobacta globus</i> str. Baddy	D-TAD	MFYGRKVVAFHQCD-INGKRTFYAA	-----FNTSANG	-----FDMH
296446027	<i>Methylobium trichosporium</i> OB3b	R-PAD	DSYANKVTCAKAMQ-DKRCRTPYEK	-----FKAKTYEMD	-----AFIV
347536497	<i>Flavobacterium branchiophilum</i> Ft-15	R-SQD	SSDMKLTICDQFRRVAVRQSGPITKAMHLE	-----ILFRJABRLEKEMAMVTRTEITISBIIKAA	-----
345683718	<i>Prevotella</i> sp. C561	I-SFD	SSDMKLTICDQFRRVAVRQSGPITKAMHLE	-----ILFRJABRLEKEMAMVTRTEITISBIIKAA	-----
282880052	<i>Prevotella timonensis</i> CRIS SC-81	V-GSD	STDMKLTICDQFRRVAVRQSGPITKAMHLE	-----ILFRJABRLEKEMAMVTRTEITISBIIKAA	-----
312879015	<i>Aminomonas paucivorans</i> DSM 12260	R-SLD	NSMKNVIVHCKMR-DKGRNTPFEA	-----FGCDTARD	-----KIVA
294086111	<i>Candidatus Punicelapirillum marinus</i> IMCC1322	I-TLD	DSMKNVIVHCKMR-DKGRNTPFEA	-----FGSDTARD	-----KIVA
330822845	<i>Alicyclophilius demitricificans</i> M601	K-TLD	DSMKNVIVHCKMR-DKGRNTPFEA	-----FGSDTARD	-----KIVA
344171927	<i>Ralstonia snyderii</i> R24	R-TLD	DSMKNVIVHCKMR-DKGRNTPFEA	-----FGSDTARD	-----KIVA
159042956	<i>Dinorossobacter shibae</i> DFL 12	R-TLD	DSMKNVIVHCKMR-DKGRNTPFEA	-----FGSDTARD	-----KIVA
83591793	<i>Rhodospirillum rubrum</i> ATCC 11170	R-GER	DSLDMNVLCSDAMK-FYGDRTFPEA	-----FHDDEPNI	-----AGDD
288957741	<i>Azospirillum</i> sp. B510	V-SDD	DSMKNVIVHCKMR-DKGRNTPFEA	-----FGSDTARD	-----KIVA
427429481	<i>Caenilapirillum salinarum</i> AK4	N-G-G	DPDMNVIAHDCM-AGKKTIPHER	-----AGDLDSPALALM	-----GRRKENVADLAKH
92109262	<i>Mitrobacter hamburgensis</i> XI4	K-TLD	DSMKNVIVHCKMR-DKGRNTPFEA	-----FGSDTARD	-----KIVA
148255343	<i>Bradyrhizobium</i> sp. BT411	I-SWD	DSMKNVIVHCKMR-DKGRNTPFEA	-----FGSDTARD	-----KIVA
34557790	<i>Nolanelia succinogenes</i> DSM 1740	R-SAD	DSMKNVIVHCKMR-DKGRNTPFEA	-----FGSDTARD	-----KIVA
218563121	<i>Campylobacter jejuni</i> NCTC 11168	R-SFD	DSMKNVIVHCKMR-DKGRNTPFEA	-----FGSDTARD	-----KIVA
Jnet			-----KHHHHHHH-H	-----HH	-----HHHH
291276265	<i>Halibacter mustalis</i> 12198	R-SLD	DSMKNVIVHCKMR-DKGRNTPFEA	-----FGSDTARD	-----KIVA
222109285	<i>Acidovorax obrevus</i> TPSY	R-SYD	DSMKNVIVHCKMR-DKGRNTPFEA	-----FGSDTARD	-----KIVA
365156657	<i>Bacillus smithii</i> 7 3 47FAA	R-SLD	DSMKNVIVHCKMR-DKGRNTPFEA	-----FGSDTARD	-----KIVA
220930482	<i>Clostridium calliolyticum</i> H10	R-SND	DSMKNVIVHCKMR-DKGRNTPFEA	-----FGSDTARD	-----KIVA
297182908	uncultured delta proteobact. HF0070 07E19	Q-SFD	MSLSNVIVHCKMR-DKGRNTPFEA	-----FGSDTARD	-----KIVA
154250555	<i>Parvibaculum lavamentivorans</i> DS-1	R-SFD	NSPDMNVIAHDCM-AGKKTIPHER	-----AGDLDSPALALM	-----GRRKENVADLAKH
248767588	<i>Neisseria meningitidis</i> 52491	R-TWD	DSMKNVIVHCKMR-DKGRNTPFEA	-----FGSDTARD	-----KIVA
187736489	<i>Pasturella multocida</i> str. Pm70	R-TWD	DSMKNVIVHCKMR-DKGRNTPFEA	-----FGSDTARD	-----KIVA
315605738	<i>Atkermansia muciniphila</i> ATCC BAA-835	F-RQS	MALSSVIVHCKMR-DKGRNTPFEA	-----FGSDTARD	-----KIVA
117929158	<i>Actinomyces</i> sp. oral taxon 180 str. PD310	DASID	SRDMNVIAHDCM-AGKKTIPHER	-----AGDLDSPALALM	-----GRRKENVADLAKH
189440764	<i>Acidithiobaculum caliolyticum</i> 118	D-GGS	MRHMLAVHCKMR-DKGRNTPFEA	-----FGSDTARD	-----KIVA
403744858	<i>Bifidobacterium longum</i> DVO10A	GVGSI	NRDMNVIAHDCM-AGKKTIPHER	-----AGDLDSPALALM	-----GRRKENVADLAKH
407803669	<i>Alicyclobacillus hesperidum</i> URH17-3-68	I-SFD	DSMKNVIVHCKMR-DKGRNTPFEA	-----FGSDTARD	-----KIVA
423317190	<i>Bergeyella zoohelcum</i> ATCC 43767	R-YFD	DSMKNVIVHCKMR-DKGRNTPFEA	-----FGSDTARD	-----KIVA
402847315	<i>Porphyromonas</i> sp. oral taxon 279 str. F0450	V-LMD	DSYGRKVVAFHQCD-INGKRTFYAA	-----FNTSANG	-----FDMH
404487228	<i>Barnesiella intestinihominis</i> YIT 11860	I-YFD	DSMKNVIVHCKMR-DKGRNTPFEA	-----FGSDTARD	-----KIVA
374384763	<i>Odobacter lanens</i> YIT 12061	I-SLD	DSMKNVIVHCKMR-DKGRNTPFEA	-----FGSDTARD	-----KIVA
394109266	<i>Treponema</i> sp. JC4	R-TLL	DASMLVAHSCMA-FKARSPEA	-----FGSNPS	-----GYSWQ
402849997	<i>Rhodovulum</i> sp. PH10	I-TLD	DSMKNVIVHCKMR-DKGRNTPFEA	-----FGSDTARD	-----KIVA
Jnet			-----KSEEE	-----KSEEE	-----KSEEE
331001027	<i>Parasutereella excrementihominis</i> YIT 11859	I-TLKS	SIYNSVNLVYSAQGM-EEKKIYLLS	-----HLAKYLAAVFGTSLDLSQITREISTV	-----
34557932	<i>Nolanelia succinogenes</i> DSM 1740	E-TLKY	GVIVNSVNLVYSAQGM-EEKKIYLLS	-----HLAKYLAAVFGTSLDLSQITREISTV	-----
54296138	<i>Legionella pneumophila</i> str. Paris	I-SKH	GVIVNSVNLVYSAQGM-EEKKIYLLS	-----HLAKYLAAVFGTSLDLSQITREISTV	-----
319941583	<i>Sutereella wadsworthensis</i> 3 1 45B	I-TRD	GVIVNSVNLVYSAQGM-EEKKIYLLS	-----HLAKYLAAVFGTSLDLSQITREISTV	-----
254447899	gamma proteobact. HTCC5015	I-TGK	GVIVNSVNLVYSAQGM-EEKKIYLLS	-----HLAKYLAAVFGTSLDLSQITREISTV	-----
118497352	<i>Francisella novicida</i> D112	-----	-----	-----	-----
Motifs			*****	*****	*****
Informative positions			*****	*****	*****

Figure S2 (Continued)







227494853	<i>Actinomyces coleocanis</i> DSM 15436#	TSARAKAS	GF5
328956315	<i>Coriobacterium glomerans</i> PW2	SHDLRTAA	EIV
227824983	<i>Acidaminococcus</i> sp. D21	SSDFREFF	GLV
303229466	<i>Veillonella atypica</i> ACS-134-V-Col7a	VSDFRHNN	NFI
34762592	<i>Fusobacterium nucleatum</i> ATCC 49256	ASSREMF	DFI
374307738	<i>Filifactor allocis</i> ATCC 35896	VSDFRKPL	NYL
320528778	<i>Solobacterium moorei</i> F0204	VSEFRNKD	KEIP
291520705	<i>Coprococcus catus</i> GD-7	VSNFRQIV	ELL
42525843	<i>Treponema denticola</i> ATCC 35405	VSMFRNAF	DIV
304438954	<i>Peptoniphilus duerdenii</i> ATCC BAA-1640	ASRERQEF	DII
224543312	<i>Catenibacterium mitsuokai</i> DSM 15897	SHEFRVKN	HIY
116628213	<i>Streptococcus thermophilus</i> LMD-9	VSQFRKDF	EIV
24379809	<i>Streptococcus mutans</i> UA159	VSDFRKDF	QVY
13622193	<i>Streptococcus pyogenes</i> S8370	TKEMHRYL	GFS
310286728	<i>Bifidobacterium bifidum</i> S17	TADMRXYV	DIP
366983953	<i>Oenococcus oeni</i> DSM 17330	ITNFRNFF	HIA
422884106	<i>Streptococcus sanguinis</i> SK49	TADMRLIV	GIX
339625081	<i>Fructobacillus fructosus</i> KCTC 3544	VSEFRKFF	ELF
306821691	<i>Eubacterium yurii</i> ATCC 43715	STAFKALSGDDTYHFKHP	EIV
323463801	<i>Lactobacillus farciminis</i> KCTC 3681	VSEFRKFF	DIP
389815359	<i>Staphylococcus pseudintermedius</i> ED99	VSKFRKFS	EIP
258509199	<i>Planococcus antarcticus</i> DSM 14505	SHQLREEL	DFP
169823755	<i>Lactobacillus rhamnosus</i> GG	ASDLRKDM	NIL
Jnet	<i>Finnegoldia magna</i> ATCC 29328	HIIHHHH	
47458868	227501312	TSNLRNQIAFVGLKNNKETEREM-KRPEGFK	
284931710	<i>Mycoplasma mobile</i> 163K	TSFLRKNFD	DSSYA
71894592	<i>Mycoplasma gallisepticum</i> str. F	TSFLRKSMM	RKN
363542550	<i>Mycoplasma synoviae</i> 53	TKYFRAPVQKNGPNEN	LNNNK
384393286	<i>Mycoplasma ovipneumoniae</i> SC01	TSFIRKNN	SYDN
238924075	<i>Mycoplasma canis</i> PG 14	THQMRCL	KLD
315149830	<i>Eubacterium rectale</i> ATCC 33656	THTLRKKW	GAD
116627542	<i>Enterococcus faecalis</i> TX0012	TSQLRRHW	GIE
315659848	<i>Streptococcus thermophilus</i> LMD-9	TDYLRKVM	KFK
160915782	<i>Staphylococcus lugdunensis</i> M23590	TNAFRREI	NLK
325677756	<i>Eubacterium dolichum</i> DSM 3991	TSMFRRW	LN
225377804	<i>Ruminococcus albus</i> 8	TSYLRRKW	GLM
336393381	<i>Roseburia inulinivorans</i> DSM 16841	TSKIRSW	GFI
310780384	<i>Lactobacillus coryniformis</i> KCTC 3535	TAQLRAW	RUN
301311869	<i>Ilyobacter polytropus</i> DSM 2926	TDRLKDM	GIN
385811609	<i>Bacteroides</i> sp. 20 3	TSELKIW	GLN
60683389	<i>Ignavibacterium album</i> JCM 16511	TDKLRDM	OLI
319957206	<i>Bacteroides fragilis</i> NCTC 9343	TSKTRSLI	GLK
187250660	<i>Nitratifactor salisuginis</i> DSM 16511	TAQLRMW	GLQ
325972003	<i>Eubacterium minutum</i> Peil91	TSILRKAW	NLQ
296446027	<i>Sphaerochaeta globus</i> str. Buddy	TDRLRAW	GLQ
	<i>Methylosinus trichosporium</i> OB3b		

Figure S2 (Continued)

SR  
 KC  
 KS  
 KV  
 KS  
 KF  
 KV  
 KC  
 KC  
 KN  
 KV  
 KV  
 KV  
 HK  
 KN  
 K  
 K  
 K  
 K  
 KN  
 KS  
 SI  
 KK  
 EE  
 PE  
 KL  
 KN  
 LD  
 KT  
 KE  
 KD  
 ER  
 KV  
 KV  
 DVWVKVLEPFIRL  
 GVWVKVLEPFIRL  
 DV  
 SK  
 GI  
 GI  
 WW

Figure S2 (Continued)

347536497	<i>Flavobacterium branchiophilum</i> FL-15	VAEERKLV	---	GLI
345885718	<i>Prevotella</i> sp. C561	TAKFRKIL	---	GLI
282880052	<i>Prevotella limonensis</i> CRIS 5C-B1	TAEFRKWM	---	GLI
312879015	<i>Aminomonas paucivorans</i> DSM 12260	TAFLRKQW	---	GLH
294086111	<i>Candidatus Puniceispirillum marinum</i> IMCC1322	TSLLRGFW	---	GLN
330822845	<i>Alicyclophilus denitrificans</i> K601	TALLRGKF	---	GLN
344171927	<i>Ralstonia syzygii</i> R24	TAMLRAKW	---	GLN
159042956	<i>Dinoroseobacter shibae</i> DFL 12	TEMLRRHW	---	GLN
83591793	<i>Rhodospirillum rubrum</i> ATCC 11170	TGLRLAW	---	DIITGAPRDLPTPRDALRDDTAARRFL
288957741	<i>Azospirillum</i> sp. B510	TALLRRRW	---	GID
427429481	<i>Caenispirillum salinarum</i> AK4	ISVQPVN	---	GSVTEMLRQLRQDKNRYQTHHAEDAC
92109262	<i>Nitrobacter hamburgensis</i> X14	TSMLRGKM	---	GLN
148255343	<i>Bradyrhizobium</i> sp. BTAl1	TALLRAIW	---	ELN
34557790	<i>Molinitella succinogenes</i> DSM 1740	TSVLRVQW	---	GLI
218563121	<i>Campylobacter jejuni</i> NCTC 11168	TSALRHITW	---	GFS
Jnet	218563121	HHHHHH	---	
291276265	<i>Helicobacter mustelae</i> 12198	TSMRSEFW	---	GVQ
222109285	<i>Acidovorax ebreus</i> TPSY	TAFLRARW	---	GLT
365156657	<i>Bacillus smithii</i> 7 3 47FAA	TAHLRSRW	---	DFN
220930482	<i>Clostridium cellulolyticum</i> H10	TAQLRSRW	---	GLN
297182908	uncultured delta proteobact. HF0070 07E19	TGYLRKQW	---	GLE
154250555	<i>Parvibaculum lavamentivorans</i> DS-1	TAQLRKLM	---	TLN
218767588	<i>Neisseria meningitidis</i> Z2491	TNLLRGFW	---	GLR
15602992	<i>Pasteurella multocida</i> str. Pa70	TALLRSRW	---	GLI
187736489	<i>Akkermansia muciniphila</i> ATCC BAA-835	TAEVRRGW	---	DVF
315603738	<i>Actinomyces</i> sp. oral taxon 180 str. F0310	TACARRAA	---	HVD
117929158	<i>Acidothermus cellulolyticus</i> 11B	TAEARRWL	---	DISIE
189440764	<i>Bifidobacterium longum</i> DJO10A	TASARRAA	---	GIE
403744858	<i>Alicyclobacillus hesperidum</i> URH17-3-68	TAYRRARW	---	GLT
407803669	<i>Alicanivorax</i> sp. W11-5	TAELRRRW	---	GLD
423317190	<i>Bergeyella zoohelcum</i> ATCC 43767	TDYLRNHW	---	GLT
402847315	<i>Forphyromonas</i> sp. oral taxon 279 str. F0450	TALRSLSW	---	GYG
404487228	<i>Barnesiella intestinihominis</i> YIT 11860	TSFLRBVW	---	GWD
374384763	<i>Odoribacter laneus</i> YIT 12061	TAELRHLW	---	GLN
384109266	<i>Treponema</i> sp. Jc4	TKLLRDKW	---	EMD
402849997	<i>Rhodovulum</i> sp. PH10	TGLLRARM	---	GLN
Jnet	331001027	HHHHHH	---	EEEEEE
331001027	<i>Parasutterella excrementihominis</i> YIT 11859	AWFVRSIT	---	SKVQALAAWQTEIGNELIFDAISVPA
34557932	<i>Molinitella succinogenes</i> DSM 1740	KYLAKKIQ	---	PKHLSFEFILADA
54296138	<i>Legionella pneumophila</i> str. Paris	KFLGKQIM	---	DSKQLQLEFSIKQITA
319941583	<i>Sutterella wadsworthensis</i> S 1 45B	IWN1KNLA	---	NKIREELQNWCKTNNRHLFQAAATNV
254447899	gamma proteobact. HTCC5015	AWLAKKIA	---	SLLAEHL
118497352	<i>Francisella novicida</i> U112	RYFAEVLAA	---	NNIYLRA--RKENLNTDKISFDYFGIPTIGNG

Motifs  
 Informative positions  
 \*\*\*\*\*  
 R

Figure S2 (Continued)

```

--ES--
--SV--
--SE--
--KN--
--ST--
--DV--
--RV--
--SI--
DGLTFPLAKAVEGAVQ
--A--
LLLAGRA
--GI--
--DL--
--SK--
--AK--
--EK--
--XV--
--KN--
--NI--
--XV--
--KA--
--GV--
--KR--
--RI--
--GK--
--XD--
--TVI--PQVRESG-MPVVDEGASL--TPEEKEKRLQWEGHVRVREMTDRR
--DKFKLLKERY--FALLESEKLEAFYDKDFSRKKEBEKEVLFEEQLTREFEIKYENYIRKKNKLIKGRS
--KLLHTLNLDRY--DSMGTERV--SREGATEELHTNWS
--TVLHDINEFDY--KRVGLTEVLEV--NHRGSIIRREQIKDWS
--NLIQSAPIITFPIVSAFENHEREYVITNEQNEVIRLFPKQCEPRTEKGELELLTGEVERKVFACKGQEFQTDVSDCKYWRRIKLSVVTWSPFAPKPIISADGQIVLKGRIEKGVFCVQCQKIKTKTGLPDSYWI SL
--SIL--CRKTEKEVALGLK
--SDSN--
--HHHHHHHHH--KEE
--ADSEMKRFAEYRPERK
--TEVSELRQYARQNPLLAK
--EEVDEHELLSKQBBKLVK
--SDAKNLRKLAQKQDFEK
--WSYKQKMLASAEFTWAK
--RGAETINQYKVDSDIQAQAK

```

Figure S2 (Continued)

227494853	<i>Actinomyces coleocanis</i> DSM 15436f
328956315	<i>Coriobacterium glomerans</i> PW2
227824983	<i>Acidaminococcus</i> sp. D21
303229466	<i>Veillonella atypica</i> ACS-134-V-Col17a
34762592	<i>Fusobacterium nucleatum</i> ATCC 49256
374307738	<i>Filifactor alocis</i> ATCC 35896
320528778	<i>Solobacterium moorei</i> F0204
291520705	<i>Coprococcus catus</i> GD-7
42525843	<i>Treponema denticola</i> ATCC 35405
304438954	<i>Peptoniphilus duerdenii</i> ATCC BAA-1640
224543312	<i>Catenibacterium mitsuokai</i> DSM 15897
116628213	<i>Streptococcus thermophilus</i> LMD-9
24379809	<i>Streptococcus mutans</i> UA159
13622193	<i>Streptococcus pyogenes</i> SF370
310286728	<i>Bifidobacterium bifidum</i> S17
366983953	<i>Oenococcus oeni</i> DSM 17330
422884106	<i>Streptococcus saguinis</i> SK49
339625081	<i>Fructobacillus fructosus</i> KCTC 3544
306821691	<i>Eubacterium yurii</i> ATCC 43715
336394882	<i>Lactobacillus farciminis</i> KCTC 3681
323463801	<i>Staphylococcus pseudintermedius</i> ED99
389815359	<i>Planococcus antarcticus</i> DSM 14505
258509199	<i>Lactobacillus rhamnosus</i> GG
169823755	<i>Finagoldia magna</i> ATCC 29328
Jnet	227501312
4745868	<i>Mycoplasma mobile</i> 163K
284931710	<i>Mycoplasma gallisepticum</i> str. F
71894592	<i>Mycoplasma synoviae</i> 53
363542550	<i>Mycoplasma ovipneumoniae</i> SC01
384393286	<i>Mycoplasma canis</i> PG 14
238924075	<i>Eubacterium rectale</i> ATCC 33656
315149830	<i>Enterococcus faecalis</i> TX0012
116627542	<i>Streptococcus thermophilus</i> LMD-9
315659848	<i>Staphylococcus lugdunensis</i> M23590
160915782	<i>Eubacterium dolichum</i> DSM 3991
325677756	<i>Ruminococcus albus</i> 8
225377804	<i>Roseburia inulinivorans</i> DSM 16841
336393381	<i>Lactobacillus coryniformis</i> KCTC 3535
310780384	<i>Llyobacter polytropus</i> DSM 2326
301311869	<i>Bacteroides</i> sp. 20 3
385811609	<i>Ignavibacterium album</i> JCM 16511
60683389	<i>Bacteroides fragilis</i> NCTC 9343
319957206	<i>Nitratifactor salsauginis</i> DSM 16511
187250660	<i>Elusimicrobium minutum</i> Pe1191

Figure S2 (Continued)

---VFTGGNGKRLDRRHAMDAAVAMRNSVA---KTLVLRG---  
 ---REANDFHHAHDAFLACRVGFIO---KR--HPC---  
 ---RNINDLHAKADAFALAVTGNVYH---ER--FNR---  
 ---RSLNHHAHAKADAYLNIVVGNVYH---EK--FTR---  
 ---RELNDTHHAKDAYLNIVAGNVYH---TK--FTE---  
 ---RRVNDYHHAADAYLNIVVGNVYH---KK--FTS---  
 ---REINDLHAKADAYLNIVVGNVYD---TK--FTE---  
 ---REANLHAKADAYLNIVVGNAYF---VK--FTK---  
 ---REINDFHAHADAYLNIVVGNVYH---TK--FIN---  
 ---RTVNDLHMHADAYLNIVVGNVYH---TK--FTK---  
 ---RDINDYHHAHADAYIVALIGGFMR---DR--YPM---  
 ---REINDFHAHADAYLNIVAVASALL---KK--YPK---  
 ---REINDYHHAHADAYLNIVAVGTALI---GV--YPO---  
 ---RELNHYHHAHADAYLNIVAVGTALI---KK--YPK---  
 ---NRDINDYHHAQDALCVGAGQFAA---NRGFFAD---GE  
 ---RDINDYHHAFDALFSTVQVYTE---NSGLMKK---GQ  
 ---IRELNDYHHAHADAYLAVVVGOTIL---KVYPKLA---PE  
 ---HREINSFHHAFDALIIAAGQYMO---NRYPDRD---  
 ---SELINDFHANDAYLNIVVGNVSY---  
 ---NRVNDFHAQDAYLASFGLTYRL---RRFPTNE---ML  
 ---IROQNDADAHADAYLNIVVYHGAQ---LAYPNVD---  
 ---IRDYNNKHAMADALFAAALIQSIL---GKYCKN---  
 ---RDVNYHHAFDALFARIGTYLIL---KR--YPK---  
 ---REVNDLHHAHADAYLNIVAGDVNN---RE--FTS---  
 ---HHHHHHHHHHHH---HH--H---  
 ---NSNDFLIREEGKNDVKDDVLIKDRSFGHHAADAYFIIISOYFR---SFKRIER---  
 ---DRDNIIHHAVDASIIIFSNETK---TL--FNO---  
 ---QVYRFNFVKDRDQFFHHAVDASIIAIFSLITK---TLYNKLR---  
 ---KIE---KNRENNEHHAVDAAIVAIIGNKNP---QIANLLT---  
 ---RL---KDRSDFSHAYDAIIAIFSNKTK---TYLNLID---  
 ---RDESYSHHAVDAMLIGYSELGYE---AVHKLQG---  
 ---KTRETHHHHAVDATLCVATSFVKV---SRHYAV---KE  
 ---RDYTHHHAVDALIIAASSQLNI---WKK---  
 ---RNHGKHAHADALIIANADEFK---ENKKLKA---  
 ---RDEDYGHHAIDALIIASMPKRLU---LST---  
 ---JWGRYDKA-ELKKLTYLDAADAIIANCRPEYVVLAGEKJLKNK---  
 ---RDEGDLHHAIDATVIAITPKFI---HK--ISR---  
 ---DRSTDHHAADAVIACCTDGM---QQ--VTK---  
 ---RENGDLHALDAVAVTDQKAI---NN--ISN---  
 ---NELTESRFTSINTNNTMIPSEPLEQKGNK---NY--LNN---  
 ---ESII-NRKLIFQEDDPKNKHFDLSINFOLDKEGL-KRDLDRHHAADALIIAATREHV---RY--LNS---  
 ---KELNWEKYKALGEVYFEDRDGROIQRKDWTKRNDHRHHAADALITVATKDFVI---QY--FNN---  
 ---SRETNFHAHADALIISTLTRGWO---NR--LHR---  
 ---MIFPAKQLITEKESSEFNKDVNSNKKIRLDNRHHAADALIVIAYASRGY---NL--LNK---

Figure S2 (Continued)

325972003	<i>Sphaerochaeta globus</i> sLr. Buddy	
296446027	<i>Methylophilus trichosporium</i> OB3b	
347536497	<i>Flavobacterium branchiophilum</i> FL-15	
345885718	<i>Prevotella</i> sp. C561	
282880052	<i>Prevotella timonensis</i> CRIS 5C-B1	
312879015	<i>Aminomonas paucivorans</i> DSM 12260	
294086111	<i>Candidatus Puniceispirillum marlinum</i> IMCC1322	
330822845	<i>Alicyclophilus denitrificans</i> K601	
344171927	<i>Ralstonia syzygii</i> K24	
159042956	<i>Dinorossobacter shibae</i> DEL 12	
83591793	<i>Rhodospirillum rubrum</i> ATCC 11170	
288957741	<i>Azospirillum</i> sp. B510	
427429481	<i>Caeniaspirillum salinarum</i> AK4	
92109262	<i>Nitrobacter hamburgensis</i> X14	
148255343	<i>Bradyrhizobium</i> sp. BTAll	
34557790	<i>Wolinella succinogenes</i> DSM 1740	
218563121	<i>Campylobacter jejuni</i> NCTC 11168	
Jnet	218563121	
291276265	<i>Helicobacter mustelae</i> 12198	
222109285	<i>Acidovorax sbreus</i> TPSY	
365156657	<i>Bacillus smithii</i> 7 3 47FAA	
220930482	<i>Clostridium cellulolyticum</i> H10	
297182908	uncultured delta proteobact. HF0070 07E19	
154250555	<i>Parvibaculum lavamentivorans</i> DS-1	
218767588	<i>Neisseria meningitidis</i> Z2491	
15602992	<i>Pasteurella multocida</i> str. Pm70	
187736489	<i>Akkermansia muciniphila</i> ATCC BAA-835	
315605738	<i>Actinomyces</i> sp. oral taxon 180 str. F0310	
117929158	<i>Acidothermus cellulolyticus</i> 11B	
189440764	<i>Bifidobacterium longum</i> DJ010A	
403744858	<i>Alicyclobacillus hesperidum</i> DRH17-3-68	
407803689	<i>Alicanivorax</i> sp. W11-5	
423317190	<i>Bergeyella zoohelcum</i> ATCC 43767	
402847315	<i>Porphyromonas</i> sp. oral taxon 279 str. F0450	
404487228	<i>Barnesiella intestinihominis</i> YIT 11860	
374384763	<i>Odoribacter lanus</i> YIT 12061	
384109266	<i>Treponema</i> sp. Jc4	
402849897	<i>Rhodovulum</i> sp. PH10	
Jnet	331001027	
331001027	<i>Parasutiferella excrementihominis</i> YIT 11859	
34557932	<i>Wolinella succinogenes</i> DSM 1740	
54296138	<i>Legionella pneumophila</i> str. Paris	
319941583	<i>Sutterella wadsworthensis</i> 3 1 45B	
254447899	gamma proteobact. HTCC5015	
118497352	<i>Francisella novicida</i> U112	

Motifs  
informative positions

Figure S2 (Continued)





227494853	<i>Actinomyces coleocanis</i> DSM 15436†	-----NIRASERAIQ-----AAETWKSFRGENVADRQI-----
328956315	<i>Coriobacterium glomerans</i> PW2	-----VYENPIGISOVNVVRO-----CADLFEKCRITIPGSSGFTVNSFMTSGFDEKTEGELIF
227824983	<i>Acidaminococcus</i> sp. D21	-----RREMNOPY-----SVKTKLFTHSIKNGN-----
303229466	<i>Veillonella atypica</i> ACS-134-V-Col7a	-----NRLFLFKNGAMR-----TYNLAKMENVYDVICTNAQD-----
34762592	<i>Fusobacterium nucleatum</i> ATCC 49256	-----KPYRYLOEIE-----NYDKKIKYNYDI-----
374307738	<i>Filifactor alocis</i> ATCC 35896	-----NPIQMKKQRDI-----NYSLNKVFEBHDVING-----
320528778	<i>Solobacterium moorei</i> F0204	-----KFFNNIRNE-----NYSIKRVDFDSV-----
291520705	<i>Coproccoccus catus</i> GD-7	-----MAAFIRNPGR-----SYNLKRMFEFDIRSG-----
42525843	<i>Peptonema denticola</i> ATCC 35405	-----NPLMFIKDKDNR-----SYNLENMFKYDVRGSS-----
304438954	<i>Peptoniphilus duerdenii</i> ATCC BAA-1640	-----LEPEFVYGDYPKYN-----SFRERKSATEKVYFYSNTIMFVKKISLADGSRVIERPLIEVNESETG-----
224543312	<i>Catenibacterium mitsuokai</i> DSM 15897	-----LEPEFVYGDYPHFH-----GEKE-NKATAKFYFYSNTIMFVKKD-----DVR-----TDKNG-----
116628213	<i>Streptococcus thermophilus</i> IMD-9	-----LESEFVYGDYKVVYDVRKMIKAKSEQEIQKATARYFYFYSNTIMFVKTEITLANGERIKRPLIETNGETG-----
24379809	<i>Streptococcus mutans</i> UA159	-----LDSAGNQYRVYKEMIIHAARLNA-----QSQRVNPGFVVGSMRNPAAPEK-IMPETG-----
13622193	<i>Bifidobacterium bifidum</i> S17	-----LITYGHAFNRHEENKATIRKHLYSN-----IMREFNFDK-----VS-----
310286728	<i>Oenococcus oeni</i> DSM 17330	-----STVYVNEFDRYTNDYLNLRQISSRD-----EVRRLKSGFVGMIRKGNED-----
366983953	<i>Streptococcus kitaharae</i> DSM 17330	-----IMGEYKRFYGVVXELHKKKALPDS-----RANGFLISPLVNSTQ-YDRVITG-----
422884106	<i>Streptococcus sanguinis</i> SK49	-----LDFENKMEVREKKAALGFEN-----TKQKSREL-PEFKLEMEVS-----
339625081	<i>Fructobacillus fructosus</i> KCTC 3544	-----FLAFDLSKDKORQKQVRSKGSN-----KEF-FLFKNFENLRQ-SPVITG-----
306821691	<i>Eubacterium yurii</i> ATCC 43715	-----LAPFFYGEFAYV-----DVKKREFNFIFGALTHAKNKIILAKDTG-----
336394882	<i>Lactobacillus farciminis</i> KCTC 3681	-----NPIINYKVNREGDKV-----KYSLSKDFTRPKSG-----
323463801	<i>Staphylococcus pseudintermedius</i> ED99	-----HHHH-----
389815359	<i>Planococcus antarcticus</i> DSM 14505	-----LNVYRKE-----TRELDLEKNNIKKKE-----
258509199	<i>Lactobacillus rhamnosus</i> GG	-----LIQFADYKLFKN-----TDGSKKIDKTKGWTEVT-----
169823755	<i>Finnegoldia magna</i> ATCC 29328	-----VYESYDQRRBDG-----VYLINKETGEVKKAD-----
Jnet		-----LADNKTDKKFLHIDENYKE-----NITETGEIVRTP-----
47458868	<i>Mycoplasma mobile</i> 163K	-----PSLNG--IISKRSQYVYIE-----DRTGELKELK-----
284931710	<i>Mycoplasma gallisepticum</i> str. F	-----ETGKVVREIDFETGEIVNEMS-----YWEFKSKKYEKTY-----
71894592	<i>Mycoplasma synoviae</i> 53	-----QKWLIVSYSEDO-----LIDETGELISDDEKVESVF-----
363542550	<i>Mycoplasma ovipneumoniae</i> SC01	-----VNSVIERPEIE-----SKOLDIQVDSQDNY-----
384393286	<i>Mycoplasma canis</i> PG 14	-----IFSRVKTEDIY-----DESTGEVPSGDDSMY-----
238924075	<i>Eubacterium rectale</i> ATCC 33656	-----MYHQAGKRITPEYEQSKKACIDNLYKLFMRDRRTAKLILSGHGRUTPIIPNISEEVDKRLMDKNIY-----
315149830	<i>Enterococcus faecalis</i> TX0012	-----YSQHOEVKN-----NQALWHDALKDAEYAAQRMADALFNKIFNCF-----
116627542	<i>Streptococcus thermophilus</i> IMD-9	-----ISRYKEKNCQKDVPSYEHHA-----DEFTGEVYFEEVKDFRF-----
315659848	<i>Staphylococcus lugdunensis</i> M23590	-----VSASKNTKIY-----RRDLQTLILCHKDKTDNNGNYKVI-----
160915782	<i>Eubacterium dolichum</i> DSM 3991	-----LMAADNDE-E-----KREYFSLCNKIRD-----F-----
325677756	<i>Ruminococcus albus</i> 8	-----KRAALDPNAN-----EHALKNKYFQNGRAIA-----
226377804	<i>Roseburia inulinivorans</i> DSM 16841	-----MLRQNYGKSEAE-----LKELMVKMPEHIEGTLADYI-----
336393381	<i>Lactobacillus coryniformis</i> KCTC 3535	-----MAGDYKINYSERNWLSKILLPPN-----
310780384	<i>Ilyobacter polytropus</i> DSM 2926	
301311869	<i>Bacteroides</i> sp. 20 3	
385811609	<i>Ignavibacterium album</i> JCM 16511	
60683389	<i>Bacteroides fragilis</i> NCTC 9343	
319957206	<i>Nitratifactor saluginis</i> DSM 16511	
187250660	<i>Eusimicrobium minutum</i> Peil91	

Figure S2 (Continued)

FESW---SENMRVIVKFKFNI---ALYNDEVSI---FSSIRLQL---GNGKAHDDTI---  
 KDDW---DABAEEVGLRRSL---NFRQCF---ISRMPED---HGVFNDAIYS---  
 FVAV---NGEEDLGLIV-KM---LKNKNATH---FTRFSPDR---KEGLFDIQLP---  
 GKAW---DVKTSMTVK-KM---MASNDVR---VTRRLLEQ---SGALADATIYK---A  
 KNAM---DKENSLEIVKKNM---EKWTVN---ITRFIKEE---KGELEFNLP---IK  
 EVIW---EACTYHEDFN-TY---DGGTLDRKIRKIVERDNL---YTEYAYCE---KGELEFNATI---  
 PGAW---DAKGSITNT IKKY---MAKNPTI---IAFAPYEV---KGELEFDQOI---  
 EIAW---KAGNKGSIIVTKK---VMQKNLIL---VTRKAYEV---KGGLEFDQOI---  
 ITAW---ERKTIITVK-DM---IKRNTPI---YTRQACK---KGELEFNQTI---  
 YTAWLADDSGKVRKATIKK---VRELEGNRYR---FTRMSYIG---TGGLYDQNL---  
 KLIWN---PDLINEIKKC---FYKDCY---CTTKLDQK---SGOLEFNLTV---L  
 ESVW---NKBSDLATVR-RV---LSYQVN---VVKVEEQ---NHGLDRGKPKGLFNALSS---  
 EIIW---KKDEHISNIK-KV---LSYQVN---IVKVEEQ---TGGFSKESI---  
 EIVW---DKGRDFATVR-KV---LSMPQVN---IVKTYEVQ---TGGFSKESI---  
 EIVW---SEEDKDYLRK-VM---NYRRML---VTQKVGDD---FGALYDETRVA---  
 EITP---DENADWSIA-DL---DYLHRV---MNFRTITVTRRLKQD---KQQLYDESIRVP---  
 KDIWD---CNEIDLPIKD-VI---YNSQIN---FVKRTMIK---KGAFFYNQFVG---  
 W---SEENTSILRK-VM---MFKNIL---TTKTKPKD---RGLNKNKTIYS---  
 QSEG---NSGTIVVK-TM---AKNSFI---ITKKEEG---HGSITKETTIVGKELKFRNKVEK---  
 EIIW---NVGFRDKILK-IP---NYHOCN---VTRKTEIK---TGOFYDQTIYS---  
 Q---GERLISIKLI-DM---NHFKIN---YSRKLANI---POOFYNQTAVS---  
 EIVW---SGVEVMKH-VY---FELPQK---TKMKIQTG---DGMFYKESIFS---  
 KVIWTPKGRKLIIVDTL---YNFRRML---ITHEVYFE---TADLPKQTIYA---  
 EE---HHHH---E---EEEEEE---KGELEFNATIAG---  
 KASP---DNFLLINALD-EL---NEKINQMR---FSRWITK---KNTOLEFNELYS---GKY  
 DENW---KQIRVRNOVS-EI---AKVIBKYIQDSNIERKAR---YSRKTENK---FNISLEFNDAIVYSAKRKYVEDQIKRKNLKTLDTHESAK  
 KDYWRDQBNFKIRENAIEI---KNVNNVDFQOVR---YSRKANTK---INTOLEFNELYG---VKEF  
 KFE---VOKLAKVEDLKKII---QEKYEAAKHTAIK---FSRKTRTI---LNGGLSDELYG---FKYDEKEDK  
 KEDWTSIKNNVQARKIAKEI---EYLIIDL---DEVF---FSRKTTRK---TNRQLXNETIYG---IATKIDEDG  
 GKSW---ANIRNEVKA---ERNVK---YWHYVWRK---SNRGLCNQTI---RGTRG  
 QVW---PNF---RE-QL---KPVNLHPRIK---FSSHQVDRK---ANRKLSDATIYS---VREKTEVTKLKS GK  
 KAFY---QHFVDTLXSK-EF---EDSIL---FSYQVDSK---FNKRLSDATI---YATROAKVVG  
 PMFIIIPKQVQ-DI---KDFRNFK---YSHRYDVK---PNRQLINDTL---Y  
 DDY---FAFIASLKA---YVRK---FSHKIDTK---PNRSVADETI---  
 EQFW---KDDKKSCE-EL---YRENVASLYKGDPKKASSLMPV---ISLKDPHK---YRGTITGEEA---  
 PLPW---NSFRDELIRLLNEDPKNFLHADVQRELDYPGWYGEESPIEEGRYNYIRPLF---VSRMPNHK---VTGSAHDATI  
 PLPW---PEFLDELLARISD---NPVEMMKRSWNYTPIETAKIKPVF---VVRLANHK---ISGPAHLDTI---RSAKLF  
 PMFW---SGFDLEIKRRESENPREEFYN---L---LSDKRYLGFNYEE---GFTKLRPFV---VSRMNRG---VKGOAHQETI---  
 DKPW---EFTQD'TALQK---ITVSPKQNLRVINKNTNH---YHRYENKGIIVSNQSGDSMAIRKSMIKETV---HCEVNLRMIXT  
 KLPW---ENFTSEYKSLLS---CVWSYKESKPLLSDFPNKYLKWEYKNGKQKVA---IQIKNDRW---KAVRRSMFKPEI---GTVMIRKIKIYEVSLKEAIIQAIWEEV  
 PMFL---REFRAEAKKHLEN---TLISIKAKKAVITGNLTKRKGK---VKNKQOQI---PRQLHLETI---YSGCKQYLTKEEKVNASF  
 DEAF---RFRMSKEES-LF---YRDMFDTIRS---ISYWYDKK---PLSASHKETE---YSSRHEVPT  
 NIWV---ENIDADLESF-ES---SVKALANAF---ISVKHDS---DNGLVKGHTM---YKIFY

Figure S2 (Continued)

325972003	<i>Sphaerochaeta globus</i> str. Buddy	MSEQGRRAVEIEA
296446027	<i>Methylosinus trichosporium</i> O83b	ATREVQIEDKGLHYDIKVV
347536497	<i>Flavobacterium brachylophilum</i> FL-15	AWTLEQQNK--KEARSIIEA
345885718	<i>Prevotella</i> sp. C561	FAKIBINMKLSPSGSDRGL
282880052	<i>Prevotella timonensis</i> CRIS 5C-81	YRMEYFK--CGRSKRF
312879015	<i>Aminomona paucivorans</i> DSM 12260	HNAWGETPRGRGAENG
294086111	<i>Candidatus Funicleispirillum marinum</i> IMCC1322	AARRSDLDLTRLFGR
330822845	<i>Alicyclophilus denitrificans</i> K601	ASAARGDSLTRVDGM
159042956	<i>Dinoroseobacter shibae</i> DEL 12	LAARAREQDAERLIGM
83591793	<i>Rhodospirillum rubrum</i> ATCC 11170	AAGCGAAGSABLIARDY
288957741	<i>Azospirillum</i> sp. B510	ASACGRILLDRKWRPNE
427429481	<i>Caenispirillum salinarum</i> AK4	ASAREAAAREDNIRVLEGFK
92109262	<i>Nitrobacter hamburgensis</i> X14	ADIVPESDRQDETC
148255343	<i>Bradyrhizobium</i> sp. BTAl1	WMANAYDEEHEKFI
3457790	<i>Wolinella succinogenes</i> DSM 1740	MANAHDTRKLV
218563121	<i>Campylobacter jejuni</i> NCTC 11168	YRFKETHREKERPKL
Jnet	218563121	ZAKKISELDYKMKRKE
291276265	<i>Helicobacter mustelae</i> 12198	HHHHH
222109285	<i>Acidovorax ebreus</i> T2SY	YKDKKETHRLK
365156657	<i>Bacillus smithii</i> 7 3 47FA	SHOKAIILEGDKL
220930482	<i>Clostridium cellulosyticum</i> H10	DPETGELLNPAAFDRARQHF
297182908	uncultured delta proteobact. HF0070 07E19	FYKAEQNKESAKKKEPIF
154250555	<i>Parvibaculum lavamentivorans</i> DS-1	YNERENIMYKRV
218767588	<i>Neisseria meningitidis</i> 22491	YHODIRRYKLGKGR
15602992	<i>Pasteurella multocida</i> str. Pm70	YWLQDDRAEKPAL
187736489	<i>Akkermansia muciniphila</i> ATCC BAA-835	FVRYKEMAFDCKT--IDKTEGVLQKTHF
315605738	<i>Actinomyces</i> sp. oral taxon 180 str. F0310	FIRKVBHPYKLENRYEMVQESGELISP--HF
117929158	<i>Acidothermus cellulosyticus</i> 118	ENGLIRRYLAMRI
189440764	<i>Bifidobacterium longum</i> D7010A	AFESWKNFCSERMQDR
403744858	<i>Alicyclobacillus hesperidum</i> DRH17-3-68	ARSRRV--SAEYWRPFDVRRHSTEEFPQSPA
407803669	<i>Alcanivorax</i> sp. W11-5	RESLRESQRLIGLMP--GERSWKEVYEGYRVES
423317190	<i>Bergeyella zoohelcum</i> ATCC 43767	YDKFKRNALAKRNVYQVSKSEG--ITQYVDKTEGVTWESFDERKFL
402847315	<i>Porphyromonas</i> sp. oral taxon 279 str. F0450	AVKVADEQRHGR--VDVAVEL
404487228	<i>Barnesiella intestinihominis</i> YIT 11860	LKKVLQDWLVEHKESEMPNFGSN--SELLELILSPENERTLFTQLEK
374384763	<i>Odoribacter laneus</i> YIT 12061	ISSFGFRDKKKE--DQFAEQQATETGRISNLERWLT
384109266	<i>Treponema</i> sp. JCA	LRAEEGPPFNKMS--LERYIQ
402849997	<i>Rhodovulum</i> sp. PH10	YVARBNKRGLD--SEHF
Jnet	331001027	KNSHKGNRIE
331001027	<i>Parasutterella excrementihominis</i> YIT 11859	DARAEQDLDRV--FRDV--EEK
34557932	<i>Wolinella succinogenes</i> DSM 1740	EEEE
54296138	<i>Legionella pneumophila</i> str. Paris	DKRMSOLAIYEPINFDN--LFTGSCOVIONTPRNESDKNTIANS
319941583	<i>Sutterella wadsworthensis</i> 3 1 45B	NADEVAKLMLDSWNPASNEPLT
254447899	gamma proteobact. FTCC5015	KEFPQFSQDLNSWFLNH
118497352	<i>Francisella novicida</i> U112	AERDONGFYLDQKTVLG--LYPQCEVILQAKPQEKSHQSV
		QPHYASRLKTISSSTFEKYGWRS--ALPDLIKVDALDRRPAFRRYNIGST
		KMYSLYPLDKKNTGEVTKDIFSQ--IKITDNEFSDDKLVKKALEGFNIER

Motifs  
informative positions

Figure S2 (Continued)

MIPI---PCYASEENLSFEA---QRELFRRKILFEMDLHAF---VSMKTDMND---ANGALLKQTV---Y  
TPFW---PGFREQAVEA---VEKVF---VARERRR---ARGAHDATI---  
SKPW---KTFKEDLLK---IEEELIV---SHYTPMVK---KOAKKIVRVGRKQKQVAVAEVDVNGKA---VPKASGKTIYK  
IQBL---EGLKNKLOMEVKVCRIHNVSEIGTF---INDNLIIV---NHHIKNQA---LFPVRRRLRKKGY---VGGVDN  
SKPW---ATFTEVDLIV---IYKNLIV---VHDTNNM---PHTTKVQVT---PHTTKVQVT---SICK  
SLEP---PAFRSEVIARLCE---TREILILRDOGGVGYDEAFNGIRPVF---VSRAPSRR---LRGKAHMETL  
IDFW---DGFREDEVKKH---IDALI---VSHRPRK---SQCALHNDTA  
PMFW---PYRDHVERA---VRIHW---VSHRPDHG---PEGAMMEETS  
PTFW---PNFLEDVRAA---LDRIL---VSHRADHGRIDHARQGRDSTAGOLHQETA---YGIVAGPF  
PEPW---PFRRAEVMKQ---WDHIIH---PSIRFAHR---DGSLSHAATV  
EPEW---DGFRAELERR---ARTIV---VSHRPEHG---IGGALHKETA  
RVHW---RLTRAGLKRIDDLTRNC---VILSR---PRPSETG---TPGALHNATH---YGRREITVDGRJDT  
LLEW---PTFRIDLETR---LEKML---VSHKPDHG---LOARLEHDTA  
AVPL---ANFRDAVEEATRI---ENTETVKEGVEVKRIL---ISRPPRAR---VTGOAHEQTA  
FEFF---SGFRQKVLDK---H---EE---EEE---PSGALHEETE  
SLEW---PMSNEFKDKIQES---IQNII---PSHVYSHK---VTGELLEQETV  
PEPW---THFAHELKARLFT---DDLAALREDMORLGSYTTEDIGRLRILF---VSRAPQRR---SGGAVEKETI---Y  
PQPW---PHEADELKARLSK---FPQESIEAFALGNVDRKLESLEPVE---VSRMPKRS---VTGAAHQETL---RRC  
PLPW---HSFRQDIMEI---LAGVF---ISRAPRK---ITGPAADETI  
PEPW---PETFRQDVLV---EEEIF---ITROPKVV---SGGIQTKDTL  
TPFW---DTIRADAOKA---VSRIV---VSHVRNKK---VSGPLHKEETV  
PQSW---EFAQAVMIRVFCRKPDCRPEEEADTPEKL---RTLLAEKLSRPEAVEYVTFV---VSRAPNRK---MSGQGHMETV  
PEPW---AYFRQEVNIRVFD---NHPD---TVLKEMLPDRPQANHOVQPLF---VSRAPTRK---MSGQGHMETI  
PEKL---IGDVEVACDRLINE---LIDAKLIPV---TENLRK---NSGKLHADQP---ESLKKARRGSK  
YRQWKECSGLGILLIS---TAARDSIA---VAAPLRAR---PTGALHEETL  
FHLWL---DNMDVLELLELINDA---LDNDRUAV---MQSORVVI---CNSIAHDATIHPIL  
LEPW---FFRELLARLSDDPKSKNIRAIGLLTYSE---TEQ---IDPFLVSR---MPTRVK---TGAAHKETIRSP---RI  
PMPIL---TIRDIA---LEA---VRSVRISH---KPDYRP---DGRFFEATAYGI---AQ  
EMFWK---GFPEQV---EOK---LKEILISH---KPKDKLLOXNKAGRQIKL---RGQIHEGTYGI  
ORPHE---SVRTV---SDK---VAELLISY---RPGORAVTRGRNIYRKKMADGREVSCVORGVLVPRGELMEASFYK---IL  
SQPHE---SVAQV---REA---VDRLLVSF---RAGKRAVTPGKRYIRKNNKRLISVQSVLIF---RGALSEESVYGV---IH  
PSWPW---GFAQDV---RQS---VWPLLVSY---KONPKTLCKISKTLYKDKGKLIHSCGNVAV---RGQLHKEIVYGO---RT  
---PEF---PILRSDI---IEK---VKNIVVSF---KPDGGA---EGKLSKETLLGK---IK  
PVPFE---DFRDHV---RER---VSTITVAV---KPEHGK---EEE---GGALHEETS YGL---VP  
---PIFKSTIY---AERTDIIVSRG---EIFIGYP---SNMPEEK---PNRISIGGKD  
YUK---SIFGENAL---GERYKPIVQEG---CYYIGYPAIVKKG YELK---NCKVVTSKND  
---PLFKDSLY---AERTFVVRKGE---TFAIGFS---EKDLFEIK---PSNKEK  
---ALFKEGIY---AEQFLPIFTLINE---KIMIGYE---TLNKA GER---CGALEVSGKQ  
---SLFKDGIY---AERTFPIIDEN---GIMAGYD---IDNLSLAK---GADVVFESLS  
---QMTRDGIY---AENYLPILHKEINELNEVRKGYI---WKNSEEIKLFFKGGKYDIQIUNNLVYCLKEVDKPIIS

H

Figure S2 (Continued)

-TKLOHRKVGDAWSHTEIDRAST-----PALWCAIATR-OPDFTMKDGL-----  
 -----FRAKXIAALPLKQGLN-----PSYGSFSRQOFAYFFIYKARPRK-----  
 -----KASTGINPRKAGLD-----WKYGGYDKSTAAYVIAVRFRTLED-----  
 SVAAKADGAYIGMNTKYSVFAD-----VTYGGMTKIKMAYSIIVQYTKK-----  
 KGETSNEIISIRPKLYDGRDKL-----NEKYGYTSLAKAYFIYVEBEKKN-----  
 -----QNKNGNSTVSLAKGLD-----VKYGGYFSANTSYFSLIEF-----  
 -----VPKGGQPIKQGLD-----TEKYGGNKUSSAFLFAVEYKGRAR-----  
 -----MKKGQVPIKGNDRERLAD-----IEKYGGYKKAAGTYFMVAKSLDKKGG-----  
 -----MKKGLQHPKAKRGGPFSN-----ISKYGGYKRVSAAYTTLIEYEEK-----  
 -----MRKGGQIPKRENTMKN-----IEKYGGYKASSAYFALIESDKAG-----  
 SNIDADAGVTAKAVVPRKNSD-----VHKYGGFSQYTYIVAHEGQKRRK-----  
 -----KFPNSNENLVGAKYLD-----PKYGGYAGISNFSVIVKGTIEKG-----  
 -----LPKNSDKLIPRKTKTMD-----TKYGGFDSPIVAYSIIIVADIEKG-----  
 -----LPRNSDKLIARKKMD-----PKYGGFDSPIVAYSIIIVAVKVEKG-----  
 -----ATDPKINGIIPDGAQD-----TSYGGFS SAKPAAVAVLIESG-----  
 -----SVLHDAKSAKINFDKEHP-----VDIYGGFSSAKPAYAALIIFKN-----  
 -----KTKQLAANNRYPKYKALCLD-----TSYGGYGFANSAISIIIIAERFNE-----  
 -----PKS-----GKLLIPLNKRSID-----TALYGGYSNYYSAYMIVAVRANG-----  
 -ADKTPKPNLQAYRPIKTS-DEKJCN-ILRYGGRFSISIGYCIIVEYK-----  
 -----PKPKYKLIQAQKD-MD-----PNLYGGFSGNKSSITIVKIDN-----  
 -----PKYALAYESKSN-----EVVYNGITPYTYVVAIRKSNK-----  
 -----PKVQAKYVSPKTE-----KPVHDEVKHNSICLIVETFPK-----  
 --AKDSKERGSKOLIPKQGY-----TQVYGGYDSSGYNALVAVAE-----  
 --KDYKKGKIYLPKAKDRUQD-----VSKYGGYKALNGAFFIIVEHTKS-----  
 -----EEEE-----EEEEEE-EE-----  
 DKGNITIKVKEKINLIDNRTKI-----KKLEEFFDEDKIKENEIKIIFNH-----  
 ENNSKVAQGFYRKLWVYSLN-----NKLADLFAEKEDILAVRANPWVINAQIIFNEYTE-----  
 ENNFYKLEKVMLFPSRKLDRKFL-----EDLNESSEKKNKNGSKRRLIT-----  
 -----YFKIKKKLVTSK-----NEELKYFENPFKKAQDGESEYTVMAQSHLSEFNKLEKLEFF-----  
 ITNYYKKEKFSILDKDIYLRIL-----RE-REKVFINSQNP-----VIDQIIEIIESYKKNIPRDEA-----  
 YDKQYXTMRLDIRTEGKIVFA-----KLAFSKSDRERLLVYLN-----  
 QKITDEYTI GKIKDIYLDGNE-----AFKKQDKLAKDL-----  
 KKADETYVLGKIKDIYDQGYD-----AFMKIYKDKSKFIMYRH-----  
 STRKDNSTYIVQTIKDIYAKDN-----TTLKQFDKSPERFIMYQH-----  
 --STRVIDEKEKVVKKYKDIY-----DPRFTALAEIDLNNAYQEKYLMALH-----  
 IRVXIDSKLHKRKSISEITA-----ESINSIYTD-----  
 DEKGIIVLSRVSTIKKINRQV-----ATGDGIYDPSNSNG-----  
 -----RSSKLSIQIANS-----KRPINSIKLADLENQGRDT-----  
 VSFNEALKPOAIVENDLAKKIL-----AMLELGYDTKRIKRYFENKDTWODINPS-----  
 KNDP-VRKRKEYITDYDAQVI-----AKTYQELISSSNRGQDEKIAKFIENKRVSAQVKKMLNYYTKFI-----  
 DMRKIGTVSKSAYRALLKRLYENDNDPKAFAGKNSLDKQPIWLDKQMRKV-----  
 LRUNLEAFDSLNVKINDRHKLT-----EEFMKRYDKETROKMLHRIGT-----

Actinomyces coleocanis DSM 15436#  
 Coriobacterium glomerans FW2  
 Acidaminococcus sp. D21  
 Veillonella atypica ACS-134-V-Co17a  
 Fusobacterium nucleatum ATCC 49256  
 Filifactor alocis ATCC 35896  
 Solobacterium moorei F0204  
 Coprococcus catus GD-7  
 Treponema denticola ATCC 35405  
 Peptoniphilus duerdenii ATCC BAA-1640  
 Catenibacterium mitsunakai DSM 15897  
 Streptococcus thermophilus LMD-9  
 Streptococcus mutans UA159  
 Streptococcus pyogenes SF370  
 Bifidobacterium bifidum S17  
 Oenococcus oeni DSM 17330  
 Streptococcus sanguinis SK49  
 Fructobacillus fructosus KCTC 3544  
 Eubacterium yurii ATCC 43715  
 Lactobacillus farciminis KCTC 3681  
 Staphylococcus pseudintermedius ED99  
 Planococcus antarcticus DSM 14505  
 Lactobacillus rhamnosus GG  
 Finegoldia magna ATCC 29328  
 Jnet 227501312  
 Mycoplasma mobile 163K  
 Mycoplasma gallisepticum str. F  
 Mycoplasma synoviae 53  
 Mycoplasma ovipneumoniae SC01  
 Mycoplasma canis PG 14  
 Eubacterium rectale ATCC 33656  
 Enterococcus faecalis TX0012  
 Streptococcus thermophilus LMD-9  
 Staphylococcus lugdunensis M23590  
 Eubacterium dolichum DSM 3991  
 Ruminococcus albus 8  
 Roseburia inulinivorans DSM 16841  
 Lactobacillus coryniformis KCTC 3535  
 Eubacterium polytropus DSM 2926  
 Bacteroides sp. 20 3  
 Ignavibacterium album JCM 16511  
 Bacteroides fragilis NCTC 9343  
 Nitratifactor saalsuginis DSM 16511

Figure S2 (Continued)

PANEDRILVNGTHYGLDKVGLF-----GKAAASLVRGG-----SVDIGSA  
 RQTLFFFAQVPRVLSAQIRODNA-----LERYARELAKD-----QCLEFIR  
 KKTQHKLMI-----PVGLYKARID-----HKREFTDYA-----QTTISEI  
 GEEIKELVPLPYLNNRATDLEL-----IDYKSVIEKARDISIKYR-----KLCINQL  
 KVKTFERITRIDSILRNEKMLIK-----YLVSQKLLNPKIHKKI-----YKEQTLI  
 EDKKGDRARH-----IIGVPIYIANN-----LEHSPSAFL-----EYCEQKG  
 ERSLETVYIK-----DVELYLODPK-----YCESV-----JGLRFPQ  
 EIRTIIEVPL-----YLNQOIEINHE-----SAIQYLAQERGLNS-----PEILLKX  
 GKNRISLETIPLYLVKDIQKQDV-----LKSYLTDLLGKKE-----FKILVFK  
 RERTJETIPIMVYNQEKYGNTEAV-----DKYLKDNLELQD-----PKILKDK  
 KTELVKISGVPVHLKKAASINEKI-----NYIEREGLSDVRI-----IKDNIPV  
 AKKKITNVLEFQGISLIDRINYRK-----DKLNFLEKGYK-----DI---ELI  
 KSKKLTAKVAVGTIHEKMTFER-----DPVAFLEKGYR-----NVQENI  
 KSKKLSVKELGITIMERSSEFK-----NPIDFLEAGYK-----EVKDDI  
 KTRLVNWTVQ-----EYSLIGDRPSD-----DELKRVLAKKSE-----YAKANIL  
 KERLVNWRQ-----WYSDKNSEDI-----LEQIRGK-----YPKAEMV  
 KKGKIEVKEFH-DIFIDYKFN-----NNPQFVNTPTSENGFLKKNIN-----RVLGFY  
 ANLLIKIPISA-NQIEVGNLKN-----D-----YIVNPAIKFEK-----ILISAL-  
 KCKTIRSELAIP-VYLRKDSISE-----EKLNVFRVINDGGKDS-----VSDIRLC  
 NKIK-----PVAIP-IRLI-NDLKD-----KTLQNWLEENVKHKASIQ-----IKNVY-  
 KGEKMEYQMD-HYVDFYKFN-----GNEKELALYLAQENKDE-----VILDAQI-  
 KEKEVQETKFLD-LAVIEHQFLK-----EPESQAKFLAETNSP-----IHHARI-  
 DTTAYQVIKISAQNASKIASANIK-----SREKGGQLINEIVVQQLAKR-----KRWKFS  
 KKRIRSELEFPHLHLSKFEYDKNT-----VLDYAINVLQ-----IQDPKLI  
 -----EEEEEE-----HHH-----HHHHHHHH-----  
 DRNLYETIKIINNEVKLEIKRNL-----NEKNYKYFVNKKLQEGKISFN-----EWPFLDNDFKI  
 NKKIKSQNVFKYMLDITKEFPEK-----FSEFLVKSMLRNTAITYDD-----KKNIVHR  
 EKYVDEILQILENERFKDSKSDI-----NAINKYMDSLPSKFS-----EFFSQDF  
 -----KVMGFSNKTG-----NAFVYVNDLALKEP-----I-LKAEI  
 INIKYTKNKINYN-----LYLQYMRSLTKSLD-----Q-FSEEF  
 DRRTFDDLCKIYEDYSDAANFPVQ-----YEKGTDIRKYS-----KKNNGEP  
 DEKTYERLLSTAETTPDFQEVEEK-----NGKVKRVKRSPEAVYCEENDIPAQKYA-----KKNNGPL  
 DPQTFEKVIEPILEYNPKQINEK-----GKEVFCNPFYKHEHGYIRKYS-----KKNNGPI  
 DPRTFEKLEV-----IMQYANEKNP-----LAKYHEETGEYLTKYS-----KKNNGPI  
 DPQTFQIVKVVNYFPEMSKSEK-----YFTKDKKGRIKISGMNPLSLYRDEHGMKRYIS-----KKGDPGA  
 DKLLIDSIKT-----IFEQADYKDVG-----DYLKKTNQHEF-----TTSSEGR  
 DRLLYQALVR-----OLLHGNDRK-----AFADDFHKEPA-----DGTEGFR  
 DKVYSAIRQALEAHNGSGELAFP-----DGYLEYV-----DHGTRKJ  
 DRKLYEALKN-----RLEEYDDKPEK-----AFAPFVKPTN-----SGKRGPL  
 KIKVYFTKFTKORVAVRKPITDTSFKKLIKESIT-----DTGQIQIMLRHLETKDNDFLAFS-----PDGIDEM  
 YNLEGFYKLEVAEYVLYKAKMPLMKRYEIKLSL-----QRMFNDFNFLEKSLDNYPEILKELSDMKYIIEPHKKNNEVNRLLLEHILEYHNNPKAFS-----TEGLEKJ  
 PEKVIWTLERAIYTIKREISPDLK-----VDKVIDGVGRKILIDRLINEYGNDAKKAFAFSLMDKNPFWL-----NKEKGIS  
 NDESTRAVEERATOLAQILLTRYQL-----MDAQNDRKIEIDKFOALKEKITSP-----IEVTCKJ

Figure S2 (Continued)

187250660	<i>Elusimicrobium minutum</i> Pei191	SEGVTLTYTKLSALKLTDPOK-----KTKPKDFLETAALKFKGRESEMKNKIKSAIEN-----
325972003	<i>Sphaerochaeta globus</i> str. Buddy	SILGADTQGEELVFWAKKIKDI-----GVKIGDYEVAASAINGRIT-----
296446027	<i>Methylosinus trichosporium</i> CB3b	-----RHIAVREGQVYVRRRVA-----EKLDLDRVDAER-----
347536497	<i>Flavobacterium brachiophilum</i> FL-15	LDGEGKLRLOQGGPIRGSLHO-----DSIYCAI--KPLNWTDE-----IKVVRKDL-----
345885718	<i>Prevotella</i> sp. C561	-----FRWGTGDAIAGEIHK-----ASYYCAI--TQFAMDDGKVLMEGEPQNPYIKFVIRREIKYKKS-----
282880052	<i>Prevotella timonensis</i> CRIS 5C-B1	-----VLAQGTARGSLHL-----DTYYCAI-----ERDGE-----IRVYVRRPL-----
312879015	<i>Aminomonas paucivorens</i> DSM 12260	-----RSPKMKDHPGPRFAS-----RPLKADIANLEKLERVNGKDR-----
294086111	<i>Candidatus Puniceispirillum marlinum</i> IMCC1322	-----YGIIVEHAENGASTVHRVVP-----ITSIGKQSDIEKVR-----
330822845	<i>Allicyclophilius denitrificans</i> K601	-----YGIKQDG-----SIKORRKADSSAGR-----
344171827	<i>Ralstonia syzygii</i> R24	EDGRVVRVRSVSLFLAKGDLSN-----VRCDAFLQAELEPIEQD-----
159042956	<i>Dinoroseobacter shibae</i> DFL 12	-----YSIVDIIHVASRTDLS-----LKPQALLDEPRGSGQVR-----
83591793	<i>Rhodospirillum rubrum</i> ATCC 11170	FGVNRPDARVLAQRKPVKELW-----DANAKPLFADKLAELIDGFA-----
288957741	<i>Azospirillum</i> sp. B510	-----YGFVDDPEGPNLVV-----RKPFDGLSKDEINSVR-----
427429481	<i>Caenospirillum salinarum</i> AK4	VVIQRMARDVVALIDNAKIVFAARLDAAGDYLKKEICTEADRDKRVVDEPSTHARWISARLAALVPAHA-----
92109262	<i>Nitrobacter hamburgensis</i> X14	-----YGFVKPDAVGLKEEAGNLVY-----RKAIESINENEVDRIIR-----
148255343	<i>Bradyrhizobium</i> sp. BTA11	-----YGTVEHPETEDGANLVY-----RKTFFVDISEKELDIR-----
34557790	<i>Wolinella succinogenes</i> DSM 1740	-----KPYPRIKQVKKKKWRLLAPI-----
218563121	<i>Campylobacter jejuni</i> NCTC 11168	-----RKEEFP-----YQSYGG-----
Jnet	218563121	-----
291276265	<i>Helicobacter mustelae</i> 12198	-----RTKEFY-----YQAFGG-----
22109285	<i>Acidovorax ebreus</i> TPSY	AQPSLKGQGVIEKILTSKLI-----QDFDKLIMPESNDHFVEHR-----
365156657	<i>Bacillus smithii</i> 7 3 47FAA	VGIDQSGKIQITAVTKLSDIK-----LDQDGHFPYOKES-----
220930482	<i>Clostridium cellulolyticum</i> H10	-----RSPKHFKGILTSVKIPLT-----TVTLKLELTVKNTGGGIS-----
297182908	uncultured delta proteobact. HF0070 07E19	-----RKRKSKP--DRQRVALT-----KVLADLERIVERKDA--S-----
154250555	<i>Parvibaculum lavamentivorans</i> DS-1	-----YDGTGDLTKTSGTYRQFVT-----RKKLESLSKGEIDR-----
218767588	<i>Neisseria meningitidis</i> 22491	-----KSAKLDGVSVL-----RVPLTQLKLDLERNVWV--RER-----
15602992	<i>Pasteurella multocida</i> str. Pm70	-----KSAKLAEGISVL-----RIPLETQLKPLENNVWV--KER-----
187736489	<i>Akkermansia muciniphila</i> ATCC BAA-835	-----SASLKENIRQMLEORVIOVVP-----ADMGCALLKEMORVLSV-----
315605738	<i>Actinomyces</i> sp. oral taxon 180 str. F0310	RPRQRVYLGALPADVLRVVD-----PGLWALVR--APGFDQSLGL-----
117929158	<i>Acidothermus cellulolyticus</i> 11B	-----RAFSEFTVGAAMGAELRIVE-----PEVYAAFTALADPGRGFRFKVSPS-----
189440764	<i>Bifidobacterium longum</i> DJ010A	KVPLGAMSADLIRRASTPPLMCA-----LTRLDPDYDEKEGL-----
403744858	<i>Allicyclobacillus hesperidum</i> URH17-3-68	VKVDNRKGTETIQ---VWVSKVALTE---LKLTKQGEI-----KDYFRP-----EDD-----
407803669	<i>Alcanivorax</i> sp. W11-5	RLDERSGKQVD---MVSRSKSLTD---LAPKKSID---VSKVRA-----NIS-----
423317190	<i>Bergeyella zoohelcum</i> ATCC 43767	GKEYA-----RIPLTK---FGSKFAT-----EKNLQK-----IVS-----
402847315	<i>Porphyromonas</i> sp. oral taxon 279 str. F0450	SQGRV-----RIVKRYELHD---IK-----GE-----VVD-----
404487228	<i>Barnesiella intestinihominis</i> YIT 11860	VWEKDEQGHVIOKORAVMKYPLTS---INRE-----MLDKKK-----VVD-----
374384763	<i>Odeoribacter lanens</i> YIT 12061	APGATEK---SYHTRKDIRE---LKT-----SKHIGK-----VVD-----
384109266	<i>Treponema</i> sp. JCA	LHGKE-----TFVCRENIVS---LS-----EKNLDD-----IVD-----
402849897	<i>Rhodovulum</i> sp. PH10	DTDPNAALG---NLAVRKPFRS---LT-----AGEVDR-----VRD-----
Jnet	331001027	HHHHHHHHHHH---EEEEEE---HHHHHHHHHHH---HHHHHH---HHHHH-----
331001027	<i>Parasutterella excrementihominis</i> YIT 11859	PFSTLSVLGAYLIDKAPSSKEKELIYRVV---KNAKAFELFSKVAG---SKFTAKEIKAAKI---LEALHF-----
34557932	<i>Wolinella succinogenes</i> DSM 1740	IAKLEKII---KQDLSIKENQYIKFISINKQFISELSNRYFNMYKMLVERDKEIVGLLEFIVENCRY-----
54296138	<i>Legionella pneumophila</i> str. Paris	LFTLLKTYSTKMPGSEIQELQAKSKAKWL---YFFINRKTALAEFLHHYFH---KEIVTDDTIVCFH---INSIARY-----
319941583	<i>Sutterella wadsworthensis</i> 3 1 45B	PKELLEMALPFFNK--PVGDLSAHATYRIL---KKPAYEFLAKAAL---OPLSAKEKRLAAL---LDAIARY-----
254447899	gamma proteobact. HTCC5015	PFLLFK---GEVFGQSLSDWQRINDGRVY---YMSIDUKVAFDYIQEVG---EADIABEL---LMSIHF-----
118497352	<i>Francisella novicida</i> U112	IDIQISTEELRNLTWNIAATAEYTYI---NLKTKQLREYIENYIN---TALGYKRYKSEMEF---LRSLAY-----

Matifs informative positions

Figure S2 (Continued)



NKRLFDVIQIMLEKAKKLLFEENE-----KSAEGKKEKNINDASIIYQ-----KAISLSG  
 DKQPKYFPMKDKLEQLQSKNEA-----ALQKYKESLVQMAAVLESNRKLEISGKPKIQ-----PKGDF  
 NARLIEKIRNWIETAGSPKDDPPLS-----LSSNAQKNKLVGTVMW---NEE-----KRLA  
 ---EST---KGSDES---IVDEVVKEKIKEALANKVLL-----KFACEGGYMIKKKNGMPD-----IKLHE  
 AADSEFASWDDLKALIVDELEFALMKGQFPAETSF-----KQALAEFIYM---NEE-----KGIL  
 ---SSTTKPELEN---IVDEVVKTITKEALADKNF-----AFVAFERKFC-----RSEGGFL  
 DRKLYALRE-----RLAAFGNGKK-----Q-----KWCADNS  
 DFLIKSALLMETAGLSGSEFENAV-----ELMSVLPFR-----RGEQFLA  
 ---EISMLIRIH---EATQFLRHGVS-----NALYFASRGGFYQ-----ADGQFLA  
 DARAREVALYALAEYRORVWLE-----ALALAVARDA-----RCEDEGT  
 DEQLRKAIRV---ATGKTKGDFE-----ALAKAEDIA-----A  
 SPRMAKFKALLARYOAAHEVFP-----ALAKAEDIA-----FGRGMT  
 DPRIRRALIDRIALIRRDANDPAT-----ALAKAEDIA-----AGPARG  
 ZAVARDIAELADIDALADADKTFEQEAR-----LADALQLOAFSDYF-----I  
 DIQARTIVRD---HVNVKTKGVA-----LADALQLOAFSDYF-----QFKHG  
 DRRLDLVRA---HVAGERGGKI-----LKAAYLSEQRD-----IAGEPNS  
 DEEKFEFKA-----DRVASAN  
 KEGVLKAL-----ELGKIRKVNKG-----  
 -----EEEE-----  
 EGVKKAAL---KFKKIREINOG-----AFQDNLEHKPKDN-----NOPTGPV  
 NERLYAARIQ---RLEQFGSRADK-----AFQEPYKPKK-----NGTGPV  
 DPTYEARIQ---ELLEHNNDPKK-----AFAEKIYKPKL-----NGINGAI  
 DKAVYVNLKN---RLIEHNKPKL-----AFKAPYMP-----SGPE  
 NNLIEHKKQ---CLEESGDPK-----AFFPYFC-----VSPGGE  
 DERIKETVAA---HVAGRGDPK-----AFAEPYKYDK-----AGNRTOQ  
 EPKLYEALKA---RLEAHKDPK-----AFATPYK---Q-----GG---QQ  
 EPALYAGLKA---RLAEFNQDPK-----KASLIVGVFF-----EGFSK  
 DSGGEDAMVSLSRKDKRKEKNQV-----PTDSPALAVOQGYV-----GLE  
 PADLNRGLKRGKRI SADFPIDYF-----VKLFEDDRGSIRVAG-----GAAYIAS  
 EDVLPADENRHIVLSDRVLSRDR-----ASQAQJAVQEG-----SADIGSA  
 PEDSHREIRVDTYSADDEMGFF-----KEPVYKISKD-----GSVRTP  
 PRLYNTLRELU---VQFGDAKAAF-----POALREPIEF-----QONI  
 RIVGSAIRLHISNIFEKRVSKGHT-----AEGINDNNKLAQYRNEK-----GELRPHIP  
 PFLSGFIANHL---KEYNKKKEAFS-----IPPLCLDKDK-----KQE  
 PHLRHILSRL---AOYNDNPKCAF-----AKFVYIDKEC-----RIP  
 KRIRHILSRL---AOYNDNPKCAF-----IPSNAFKQGV---YRIELP-----NKHGEPVP  
 ITIRQMLIKHLQBYHIDITQEFN-----LSDF-----SKENG  
 EKIKSKVDYV---AKHKGQKLEAV-----AGLALALEAF-----GAENG  
 RALRALRGALA---APFRDESGRVR-----HHH---HHHHHHHHH  
 ---EEEEEEEEHHH-----HHH---HHHHHHHHH  
 ---LAKQGCCLKKVEYSREKFKKOSLIIPAAVWGWKVLWVFKENTABELKD  
 VTKQDVAATVSDLIKSKKELSKDSIEN-----EVEE-----HHHHHHHHH  
 YTKRSDVWKA---PKYIBTKPYFYD-----LNEYQPDKNRYKLD---VD  
 YTKRESITVKI-----LKE-----PMPVLSVKFESKKNVLSGSKHTIALPATKDWERLFNH PNF LALKANPAP  
 CTSRSKISMSLFMA---ANGKSLKRRDVLK-----FKINGSUULPVKQDWLRICDSEPELADAFKPCS  
 TORUKLRAKFS---DSGKMKTLDAIRK-----SIKLTIVVWEIGKREKCGFSTIGIPAKSAWENLLEDEFILETYGTQMP  
 RSERVNIKS-----IDDVKAQVLDKDSNF---IIGKTIUPEFKKEMQRLYREWQNTTIKDD-----

Figure S2 (Continued)



IHH-----ARTY-----RIAGKRP-----TYGVRVFPAPDLR-----  
 IER-----SKILKNQILLEIDGDRLC-----ITGKEVRNACEL-----AFAQDEMEVIRMLVSEK  
 LOK-----DRQKVINI-----MFPMTGR-----HIKINSMISIDGFLYSI  
 VKV-----NGFYLLGG-----KTKDKIYIDN-----AIELVWPHDIATYKLL  
 IDS-----YPTFTGVDSSNKVELKTKQ-----LYLEKRYEQ-----ITKRNALKEPVRDNOGEPE  
 YQN-----VRLIVE-----KIKKNSJ-----LILNGYPLARIRGNEVE  
 IIK-----PKIIMGSLFSIMNKKLAVVT-----GRSGKYVYCHHYYQLSI  
 IKI-----DTLEKV-----DGFKM-----NL-SGRGNOI-----IFKGANQLLISHQEAAL  
 IKI-----NSLKI-----NGFPC-----HTKTKND-----SFLRPAVQFCCSNNEV  
 IKI-----NSLKI-----DGFLY-----NIKKTGD-----SLSIAGSVQVLIVNKEEQ  
 NOM-----JEMDGGEYLLTSPTE-----VYNARQIV-----INKEKCALIADIYN-----  
 IEL-----PKYSLF-----ELSDGSRMLAS IL-----SUNNRGEIHKGNQIFLSC  
 IKL-----PRYSLF-----KLENGRKRLLASA-----RELQKNEIVLZN  
 IKL-----PKYSLF-----ELENGRKRLLASA-----GELQKNEIALPS  
 IRVPRMQLIRYGGGLMVIKSAGEIANN-----AQLWLPYEEYCYFDDI  
 LSHIPYQVLVKKDGAIVTSSATELHN-----FEQLWLPLADYKILNLI  
 RIP-----KYSIMOKTDGTRMLFESKSN-----LHK-----ATQFKLTKTQWELFFHM  
 -----PLQIIVNEDGNLIVLAGNEY-----RHN-----AKQWLSTTDADKIASI  
 LPFTSTNHWKIDGYYLGGKND-----RIQIYN-----AYQLMKKKEVEYIRKI  
 -----PIGQIIVSKVGLULSNDRE-----VAN-----RQQLILPPEHSALLRL  
 VYSLNKGDLIYINHPCTVSRKE-----VTN-----AKQFELTVEQQLSLYNV  
 IRTIPYQKQIWIHEFPYFISTRE-----LHN-----AKQFEISYELMEKVKQL  
 ANS-----FKVIP-----RFGMTLQFN-----AKYGLFWNSDFTYRN-----  
 IDK-----INVRTEIIDNFSYLIS-----TKSNDGSIIVKRPQMY  
 -----EEEE-----E-----E-----EEEE-----E-----E  
 IRK-----IRYIKFSSEKETDEIIFSQSNFL-----KIDQRNFS-----FHNFLYVQIWIYKQK  
 IKR-----LKMLSSEKKNKLSNVTI-----RSKQSGTKLS-----YQDTINSLALMIRSIL  
 INKCKENSLITFDALKINDPKVKIKIKNKLFREDAIKNKO-----AVHKDSKNQIKSFEYSY  
 ESAKVERKLLYNEKPDQFYHDNINNESFKRYKNIIRIIEKYSPIK-----FKIILSKHGGKSEFKDTL  
 INQMIANKTFVL-----YNPFKNTNR-----KIK-----FLRUVNDVKINDIRKNOVI-----NRKFNKNEPKAFYENI  
 IDK-----JKYDQ-----EVCACIDISHKY-----GFEKGSKRV-----ILSILVYRMDVYKKEE  
 IRG-----JKYDQ-----K-LGNHIDI-----TPKDSNNKY-----VLQSVSPWRADYVFNKI  
 IKS-----JKYDQ-----KILKHINI TKDSQGRPVEKTRGRKV-----KSSTKKL-----VKLSIKYRFDVYLTDK  
 ITQ-----MKYEDG-----VLGNHIDISAHY-----QVRDKKLV-----VLQQISPYRTDFYSKE  
 VNK-----VTVLEK-----VPSRWLRK-----EIDNNNFS-----LLNDSYYICIELYKDSK  
 VRK-----VKIEKK-----QTSGV-----MVRGG-----JTGIAANGEMVRIDVFERENG  
 VRK-----VRVAKKVELFVRLKKA-----AAUNGSWVRIDVFNTEK  
 VRG-----IKVPEEK-----QNVGV-----YVNGG-----QASNGSWVRIDVFERKNG  
 NRN-----LILNKKKKHQPIYKRVYKAEKFTVQGNKTKFV-----ZAAKGTNLFPAIYETEE  
 NKK-----AI-----NKIGKPKYIIRLDG-----DIMEEERFQAVF-----ETDKGSWYFVMEZ  
 IKR-----VTISGISNAQSLHVKKDKDQK-----PILBENGRNIPVDFVFNHNVAVYRVP  
 LRK-----MRFYVD-----KLNAM-----QIDRGL-----VEIDKNMLGHIHISKQPN  
 DKY-----VQISKK-----EPGKFFAISK-----PTPTTYG-----GYDTGDSLCLVDLYYDNK  
 LSE-----KTSIKALELVGGYYLILSNKRIKTFVVKFQSNVAKGF-----AFDTGSLCLDFYHDAG  
 IFK-----VRUVTKSKVNIALDT-----GNPKRPG-----TVDRGEMARVDVFKAS

Actinomyces coleocanis DSM 15436#  
 Cariobacterium glomerans FW2  
 Acidaminococcus sp. D21  
 Veillonella atypica ACS-134-V-Col1a  
 Fusobacterium nucleatum ATCC 49256  
 Filifactor alocis ATCC 35896  
 Solobacterium moorei F0204  
 Coprococcus catus GD-7  
 Treponema denticola ATCC 35405  
 Peptoniphilus duerdenii ATCC BAA-1640  
 Catenibacterium mitsuokai DSM 15897  
 Streptococcus thermophilus LMD-9  
 Streptococcus mutans UA159  
 Streptococcus pyogenes SF370  
 Bifidobacterium bifidum S17  
 Onococcus kitaharae DSM 17330  
 Streptococcus sanguinis SK49  
 Fructobacillus fructosus KCTC 3544  
 Eubacterium yuzii ATCC 43715  
 Lactobacillus farciminius KCTC 3681  
 Staphylococcus pseudintermedius ED99  
 Planococcus antarcticus DSM 14505  
 Lactobacillus rhamnosus GG  
 Finegoldia magna ATCC 29328  
 227501312  
 Jnet  
 Mycoplasma mobile 163K  
 Mycoplasma gallisepticum str. F  
 Mycoplasma synoviae 53  
 Mycoplasma ovipneumoniae SC01  
 Mycoplasma canis PG 14  
 Eubacterium rectale ATCC 33656  
 Enterococcus faecalis TK0012  
 Streptococcus thermophilus LMD-9  
 Staphylococcus lugdunensis M23590  
 Eubacterium dolichum DSM 3991  
 Ruminococcus albus 8  
 Roseburia inulinivorans DSM 16841  
 Lactobacillus coryniformis KCTC 3535  
 Llyobacter polytropus DSM 2926  
 Bacteroides sp. 20 3  
 Ignavibacterium album JCM 16511  
 Bacteroides fragilis NCTC 9343  
 Nitratifactor salsauginis DSM 16511  
 Elusimicrobium minutum Peil91  
 Sphaerochaeta globus str. Buddy  
 Methylosinus trichosporium OB3b

227494853  
 328956315  
 227824983  
 303229466  
 34762592  
 374307738  
 320528778  
 291520705  
 42325843  
 304438954  
 224543312  
 116628213  
 24379809  
 13622193  
 310286728  
 368983953  
 422884106  
 339625981  
 306821691  
 336394882  
 323463801  
 389815359  
 258509199  
 169823755  
 Jnet  
 47458868  
 284931710  
 71894592  
 363542550  
 384393286  
 238924075  
 315149830  
 116627592  
 315659848  
 160915782  
 325677756  
 225377804  
 336393381  
 310780384  
 301311869  
 385811689  
 60683389  
 319957206  
 187250660  
 325972003  
 296446027

Figure S2 (Continued)

YRNE DLNVELEPP-----QSVSMRYAEKPVREAIR-----EGKAEYLGWLVGDELLDLDSSETSGQIA-----ELQODEPFGTT  
PVSRECVIS-----LFRNLI LHGQASRLI-----SKQIKLALISEAFSEASDNVQRN-----VWLGILAI FN  
GGKSKGKSVLCEAMVPLIVPHK-----IECYIKAMEFARKFKENKLRIVVEKFKDTIV-----EDNLNL-----YELFLOKIQWPNKFFS-----TQFDVLTNGR  
DK-----YDLRKENKILKA-----SSTTSTYINTSTVVSLSKNGVLDVDFYF-----MSKLRTPYMKMGKNGRVDLSST-----GSKFKTKMTL  
TS-----ENKFIYILKRBAN-----NEKNETIDAVKERYNIEFN-----EMYDKFL-----EKLSSKDYKNYINN-----KLYTNELNSK  
NDSDSQVILKIAK-----KRAIQIKLDQRN-----YELVRNTEKFLKYEKKGKGYPI DENRDRHITH-----EMNQL-----YEVLLSKMKKNKMGWADPS-----DRIEKSXPKF  
LGVVAVNRKNE-----LKVAVVNRKNE-----NDAKLSERDQGTBEEL-----LOLYDTF-----LDKLSNTVYSIRLS-----ACIKTLTEKR  
LYFKKILRFSEIR-----KLKMKDKFLVK-----KDKDKIVTSFDK-----SOBEKIGKTI SPYEDLSFSYIKENIKRKTNDIEIG-----KEFYDLLQKNLEIYMLTRHKDITLYKKEPNSATIDILVCKK  
ALYKQYDN-----ALYKQYDN-----LDDIIMIQIYI-----KNNKMKVLYPAVYGIARKE-----ELTNKMKVLYPAVYGIARKE-----SNVENVVIS  
KPVKLIYHAKRIS-----INENKRYVENK-----NT-----INENKRYVENK-----KEFEELFYILFENENKRYVENK-----KLNLSAFQSW  
HUGTLIYHAKNTH-----KV-----DEPKHLDVYDKR-----DEFKELLDVVSNEFSKRYTLAEGNL-----EKIKELIYAQN  
RYVNFYLAHYE-----KLVGSPEDNEQQLFVQHK-----KLV-----KLVGSPEDNEQQLFVQHK-----HYLDEI IEQISEFSKRVILADANL-----DKVLSAYNKH  
QKGSLEKDDIHK-----LIDSILGSVQC-----LYPWHRETE-----EELADLVAFADKIPEDERKKNVIT-----GIVSALHADA  
KPKEDNLVDLHNRDLDPMTIESAFY-----KAEDSILSFAFN-----RYALHQNAL-----VKQAHRDDFNALNYEDKQOTLE-----RILDALHASP  
RLLTKSNIMDL-----KKSATIKESQNFIL-----KHEEFTNISQOISA-----FSQKMLGN-----TSLKNLKIGYNERKIREIDIRDETIRYFY-----DNFINKFSFV  
ENSSDELEEA-----YDILISENVKRNFP-----FPKKD-----IDKLSQVRDEFIDS-----DKRIAVIQ-----TILIRGHQIDA  
EAVSKEFDEIDREKVPVLTAEKNIE-----LYNKIQDKFENTVFSKMSIAVKYKNDLSF-----FYKGE-----GDFLKNKSKFEEIDL-----EKQCKVIV-----MIIENLSNK  
IPDELDQI-----LAFYDKNIIIVE-----LI-----EYDFIAEK-----VINEYH-----HYLNSKUK-----EKRVRTFFSESQCH-----EDFIKALD-----ELEKVVYASA  
NPK-----ERNEVEKL-----L-----PGLHLDQ-----MNDNY-----LYTKS-----SIQDRVQKFPDQLDYF-----KSFIEGFE-----ELKRAVAANA  
YQELMISRENQKL-----LAKGFSIKYKTYQMHIDALQVYK-----LAKGFSIKYKTYQMHIDALQVYK-----AIIIDQVERKFKLYDINQFRAKLS-----DAIEREFKLP  
WRVDEISLNRKIE-----NRYKQDAILTEEDRKIME-----NRYKQDAILTEEDRKIME-----SYTDKIYQOFKAGYKNNRRTDT-----LIEKVEIIDL  
E-----HHHHHHHH-----HHHH-----HHHH-----B-----HHHHHHHH-----HHHH-----HHHHHHHH  
YCFISIDARNSKF-----EKDEIKINYEKIKTQEKIQL-----EKDEIKINYEKIKTQEKIQL-----INEBPI-----LKINKGDLFENE-----EKELFYIVGR  
YRVPNTANLHL-----GDHDFDLHNDAYLKKPKFVYKLAN-----GDHDFDLHNDAYLKKPKFVYKLAN-----EIGDPEKPRVLTSGTLLIHKKD-----EKLMYTSFQ  
SLSLVYK-----VYENG-----KESYKSTPVTSQMRNFGIDEFDLDEN-----KESYKSTPVTSQMRNFGIDEFDLDEN-----LYNKEKI-----DLYKSDFAKPIP-----VCKPVEVFK  
NSLGAIVK-----NS-----ANNFKTLISINTQIATFGKRWIDEDFK-----ANNFKTLISINTQIATFGKRWIDEDFK-----TYNMEKI-----EYKKEIYGID-----KTYNPHSFTF  
YLVGVKQSDIKE-----BGRNVI DEAYARILVNERMI QSGSRAD-----BGRNVI DEAYARILVNERMI QSGSRAD-----LENLGKFKLSFKVKNDIIEY-----EKDGTIYTER  
TKA-----YTVQIYSDIRF-----VEGKYGITTEKYNKVAEQTK-----VEGKYGITTEKYNKVAEQTK-----GQVVRFC-----FSLQKNDGLEIEMK-----DSQRYDVRFY  
YELGLYADIQF-----EKGTVYKTSQRYNDIKKKEGV-----EKGTVYKTSQRYNDIKKKEGV-----DSDSEEK-----FTLYKNDLILVQDTEP-----KEQQLFRFLS  
YKFTIYLRDVRW-----KDNYYYIPEQKYDKLIKLGKAI-----KDNYYYIPEQKYDKLIKLGKAI-----DKNAKFI-----ASFYKNDLILKLD-----GEIYKILGVN  
NLIQIAMSIVH-----DRKFKGILRPFNYP-----DRKFKGILRPFNYP-----DDYTHV-----MYLPPGYLRIKSTSKSGEQI-----KPEGYFISVK  
YFVYVYVYADVR-----KVLNRAATHWPK-----YSERAVM-----KVLNRAATHWPK-----DDANFV-----FSLYSRDIHV-----KSKKDIKTN  
FVFPYIYKDFVE-----QVLPNKATARGKS-----LHWQI-----QVLPNKATARGKS-----TESDQFC-----FSLYPPGMVHTESKT-----GIRPKYSKNE  
YVTVPLVHQTL-----KELPNRATNGRYV-----KWRDLI-----KELPNRATNGRYV-----DGSFEEL-----YSFVNDLIEIEF-----GKSKSILKND  
IDKRVKVIKRSYSTIPLNVJIERQ-----KQGLSAPEDBNGLPKYILSPNDLVVYPTQ-----KQGLSAPEDBNGLPKYILSPNDLVVYPTQ-----EEINKGEVMPIDRRIYKAVDSS-----GITANFIPAS  
NNQKREFLKPNESI SVLKALEBK-----NKIDFEAP-----NRLGESRIILSPGDLVYVPTNDQYVLKDNSSNETLINDDNEF-----SNRLYQVKKFT-----GNSCVELKND  
IDKRGQIADVDSAGNPKAYELEEVV-----VSFEEAVTRANLGLPIIDKDYKTEGWOPL-----VSFEEAVTRANLGLPIIDKDYKTEGWOPL-----FSMKNONE-----YFVFPNKEGTENKFEIDL-----DVENYGLISP  
EK-----LIFERRMDVANAHE-----LQKERSGLCYLN-----LQKERSGLCYLN-----EML-----FIENKGLIHY-----GCLRSYLEKG  
LGGETIKRIDAQQ-----GK-----KNPJRYK-----EGQFTLF-----ERYGGDLEVDF-----DIHSDKNSER  
LGGELIKIQAMN-----GK-----PSYKPAYM-----KQCYSLY-----VRLYQGVJCELRAASDLTE-----AESNLAKTTH  
KKGKYE-----YVLVPIYPHDIAI-----MKTPIRVAQYKPEDEWPEM-----DSSTYFC-----WSLVPMTYLVQVSSKGE-----IPEGYTRGMN

Figure S2 (Continued)

347536497	<i>Flavobacterium branchiophilum</i> FL-15	IRK	VRIYAMSVKPLHLKHSLL	SKSRVHKQKV	YCONDENYAMAIE
345885718	<i>Prevotella</i> sp. C561	IRH	VRC--ZAFQSGIKKEQT	YKSEREYKRYF	YAAVDLIYAMCCYT
282880052	<i>Prevotella limonensis</i> CRIS 5C-B1	IKK	VRCFAKSVKQPINRQHR	DISRREYKQOY	HVMENNYLLAIYGL-
312879015	<i>Aminomonas peucivorans</i> DSM 12260	VRS	LRLFDGSGYSGVE	LRDGGVYA	VADHESMVRVDVYAKKN
294086111	<i>Candidatus Punicispirillum marinum</i> IMCC1322	IKS	LRIIVETVSI	PITDKGCVAYK	GKCGDCMAYMDIYQDPT
330822845	<i>Alicyclopholus dentrificans</i> K601	YK	YVGGSSNYCIETIYNDK	YVGGSSNYCIETIYNDK	YVGGSSNYCIETIYNDK
344171927	<i>Ralstonia syzygii</i> R24	ISP	YVGGSSNYCIETIYNDK	YVGGSSNYCIETIYNDK	YVGGSSNYCIETIYNDK
159042956	<i>Dinoroseobacter shibae</i> DEL 12	IRH	VRI IKPLQARVPV	SAPYK	AYGSDSNICYELFINER
83591793	<i>Rhodospirillum rubrum</i> ATCC 11170	ANT	VIAGSDGSDGACLIIT	PAQDPTK	AYGSGNHLEIWKLP
289357741	<i>Azospirillum</i> sp. B510	IRR	VRVLKESNPIRVEHGG	NPSGRSGGPFHLLLAGVHVDVALRAD	VRVLKESNPIRVEHGG
427429781	<i>Caenispirillum salinarum</i> AK4	MEAR	PSLIVRIRAKMTRDA	FGRPVFDAAVWVKTDGNAVSQLWRLTSSVTDGRLPLPKEKRIE	PSLIVRIRAKMTRDA
92109262	<i>Nitrobacter hamburgensis</i> X14	IRH	VRI LMK	EKGDIYV	PIANRAGVAYK
148255343	<i>Bradyrhizobium</i> sp. BTail	IRH	YRLIYKS	IKPDYIV	FIRDAGRIYK
34557790	<i>Wolonia succinogenes</i> DSM 1740	OKN	YVETSTIPRVDDVYKKG	YVETSTIPRVDDVYKKG	YVETSTIPRVDDVYKKG
218563121	<i>Campylobacter jejuni</i> NCTC 11168	OKN	YVETSTIPRVDDVYKKG	YVETSTIPRVDDVYKKG	YVETSTIPRVDDVYKKG
Jnet	218563121		EEEEEEEE	EEEEEEEE	EEEEEEEE
291276265	<i>Helicobacter mustelae</i> 12198	VRS	IKLVRG	KQTCI	IVDNCAMVRVDIEKSK
222109285	<i>Acidovorax sbreus</i> TPSY	IRT	VKLIPT	KNKW	HLGSKT
363156657	<i>Bacillus smithii</i> 7 3 47EAA	IKS	IRVETP	SYTGV	FRNEKGG
220930982	<i>Glostridium cellulolyticum</i> H10	AKQ	RPILSKVILLREKPEP	KQTLKSGC	RYDSDMACGLDIYKYE
297182908	uncultured delta proteobact. HF0070 07E19	IRK	VKLFK	QQML	MAQNG
154250555	<i>Parvibaculum lavamentivorans</i> DS-1	VKA	VRVEGV	QKIGV	WVRNNG
218767588	<i>Meissneria meningitidis</i> Z2491	VKA	IRVEGV	QKSGV	IVRENG
15602992	<i>Pasteurella multocida</i> str. Pm70	FHH	ALYRIIGPK	EKV	KYALLRVCALDCC
187366489	<i>Akkermania muciniphila</i> ATCC BAA-835	FHH	ARVFRGSSHSPS FALL	RVSLADL	AVAGLLRDGVDVFTAL
315605738	<i>Actinomyces</i> sp. oral taxon 180 str. F0310	IHH	ARVRC	WKNAGVKY	FYGMIRVCFDLELRACH
117929158	<i>Acidothermus cellulolyticus</i> 11B	VAK	VKIQEKLTL	GV	FVHGGGGAENG
189440764	<i>Bifidobacterium longum</i> DJ010A	LAK	VRCFYSKADCV	RI	EHSSNRGHHYKMLNDG
403744858	<i>Alicyclobacillus hesperidum</i> DRE17-3-68	IST	VKIYKDFPKNKKKDEEDLSLQKL	DRE	KAFNEKLYVKIG-DNYL
407803669	<i>Alcanivorax</i> sp. W11-5	VRS	VRCYAKTSLDKAIPMC	FD	EKGEPTAFVKS
423317190	<i>Bergeyella zoohelcum</i> ATCC 43767	IRY	VRCFAKPAINTLIVPLK	KD	DKGSPVAVWVNG
402847315	<i>Porphyromonas</i> sp. oral taxon 279 str. F0450	IKK	VRVNRVQPTI	EI	TSGKLSRYSLE-DIYA
404487228	<i>Barnesiella intestinihominis</i> YIT 11860	IRH	VRILKPDASVTTIA	DR	RTGVYRAVAVG
374384763	<i>Odoribacter laneus</i> YIT 12061	IKK	VRILKPDASVTTIA	DR	RTGVYRAVAVG
384109266	<i>Treponema</i> sp. JC4	IRH	VRILKPDASVTTIA	DR	RTGVYRAVAVG
402849997	<i>Rhodovulum</i> sp. PH10	IRH	VRILKPDASVTTIA	DR	RTGVYRAVAVG
Jnet	331001027		HHHHHHHH	HHHHHH	HHHHHH
331001027	<i>Parasutterella excrementihominis</i> YIT 11859	ENAL	RKALEAWPESFGYR	NHLSKARVFLPVPV	NHLSKARVFLPVPV
34557932	<i>Wolonia succinogenes</i> DSM 1740	QQP	WDDFCQRYFLDRYKAND	KKSFLKARKTFLSIAE	KKSFLKARKTFLSIAE
54296138	<i>Legionella pneumophila</i> str. Paris	NPK	NEFIRKYLSDNNPNSDIPN	NGHNKPKQKAVKRVFLPVI	NGHNKPKQKAVKRVFLPVI
319941583	<i>Sutterella wadsworthensis</i> 3 1 45B	ADL	TSKLARLWKPVRD	LAHA	VYRREFFSLPAL
254447899	gamma proteobact. HTCC5015	POE	WEKVYAKHFRNTIPN	QAHKVKKDFSLPVI	QAHKVKKDFSLPVI
118497352	<i>Francisella novicida</i> J112		YEFKLSFFNVKSI	KLHKVKKDFSLP-I	KLHKVKKDFSLP-I

Motifs  
Informative positions

Figure S2 (Continued)



227494853	<i>Actinomyces coleocanis</i> DSM 15436#	HMTVAGFFPSR
328956315	<i>Coriobacterium glomerans</i> PW2	G---STNM---
227824983	<i>Acidaminococcus</i> sp. D21	STFKLSPEQVQTL
303229466	<i>Veillonella atypica</i> ACS-134-V-Col7a	-----EQS---
34762592	<i>Fusobacterium nucleatum</i> ATCC 49256	EKPKKIKLMEK
374307738	<i>Filifactor aloecis</i> ATCC 35896	IKLEDLIDKIV
320528778	<i>Solobacterium moorei</i> F0204	EKFSELDILEQC
291520705	<i>Coproccoccus catus</i> GD-7	AKFIGLSNEDQC
42525843	<i>Treponema denticola</i> ATCC 35405	EKPKSLIENQF
304438954	<i>Peptoniphilus cherdanii</i> ATCC BAA-1640	DKPKKSIIEKI
224543312	<i>Catenibacterium mitsuokai</i> DSM 15897	KEERKANIQQML
116628213	<i>Streptococcus thermophilus</i> IMD-9	QNHSDIELCSSFTIGTSERK
24379809	<i>Streptococcus mutans</i> UA159	NGEDLKELASFF
13622193	<i>Streptococcus pyogenes</i> SF370	RDKPIREQAENI
310286728	<i>Bifidobacterium bifidum</i> S17	XTANLSIVGTC
366983953	<i>Oenococcus oeni</i> DSM 17330	A5SDLKKINLSS
422884106	<i>Streptococcus sanguinis</i> SK49	XSGAPKDINDEFNKCTV
339625081	<i>Fructobacillus fructosus</i> KCTC 3544	AYQAPVKIISK-KVSDW
306821691	<i>Lactobacillus yuzii</i> ATCC 43715	E---VDLSDIGGSKST
336394882	<i>Lactobacillus farciminis</i> KCTC 3681	T-SAHLIFENNI-EKKAFG
323463801	<i>Staphylococcus pseudintermedius</i> ED99	FRSD-----KIGSRK
389815359	<i>Planococcus antarcticus</i> DSM 14505	Q-----RSDTFG
258509199	<i>Lactobacillus rhamnosus</i> GG	INTDGNKIGKTE
169823755	<i>Finagaldia magna</i> ATCC 29328	DILDNKOLYQLL
Jnet	227501312	-----HEHHH---
47458868	<i>Mycoplasma mobile</i> 163K	DE---KPKK---
284931710	<i>Mycoplasma gallisepticum</i> str. F	N---LNDV---
71894592	<i>Mycoplasma synoviae</i> 53	FGALLLNFGQQ
363542550	<i>Mycoplasma ovipneumoniae</i> SC01	KGSILKKKSLDIDDKFKETKTEEGN
384393286	<i>Mycoplasma canis</i> PG 14	PGTILLDKONKE
238924075	<i>Eubacterium rectale</i> ATCC 33656	LVSRTPKQKQNY
315149830	<i>Enterococcus faecalis</i> TX0012	NFQ-----SANS
116627542	<i>Streptococcus thermophilus</i> LMD-9	RTMPK---QKHY
315659848	<i>Staphylococcus lugdunensis</i> M23590	SD-----TRNM
160915782	<i>Eubacterium dolichum</i> DSM 3991	HNENFFFHAGET
325677756	<i>Ruminococcus albus</i> 8	NV-----NENS
225377804	<i>Roseburia inulinivorans</i> DSM 16841	LVNGGILLQ
336393381	<i>Lactobacillus coryniformis</i> KCTC 3535	NNTSVVPIKNFY
310780384	<i>Ilyobacter polytropus</i> DSM 2926	KLTKTEIPEVNI
301311869	<i>Bacteroides</i> sp. 20 3	TANLIFALPKAT
385811609	<i>Ignavibacterium albus</i> JCM 16511	IASLILSYSASN
60683389	<i>Bacteroides fragilis</i> NCTC 9343	NLFRVQKFSILKN
319957206	<i>Nitratifractor salisuginis</i> DSM 16511	QGSKYIALFNPR
187250660	<i>Elusimicrobium minutum</i> Peil191	NNTGSAPENRVF

Figure S2 (Continued)

LRPRVYLAQEGLEDVSESGKSIAG---QCRPVAVKVFGSAMPEVTRRDGLGR-KRRFYSGLFVSMQC  
 VNLSDIGGSKFAGVVRVYKKEKELASPK---VNVHLI-DQSVTCM-FERRTKIGL---  
 LNILSIFKTCRSRGGDLKSIINGSQAQA---RIMSADLNGLSKYSKDIRLV-EQASGL-FVSKSONLLEYL  
 IYLVEVMLLNSKTTFDVKKPLGTGS---NSTIGVKIHLDEFKII-NESIYGL-YSNEVTIV  
 LILREPLKIFKNTYKVEIKDSOTKE---KLFSPFDEGRIRLQSSICGKNKLELESVTEL-FVKKIKL  
 INMIMLRCDMDTKADLSIELPANA---GSFVVKNTIGSKILLV-NQSVTEL-YENREEL  
 NLLIQLLKAFKCNRESSNLEKLNKKQ---AGVIVPHLFTKSVKVRHQSVTGL-FEKEMDLAK-  
 IVLNEIIMFQCSGSANIKLIGGFGS---AGILVNNNITACKQISVINQSPFI-YEKEIDLKIL  
 EVILEILKLSAENVSDLOHIGSSKY---GVAKIGNKISSLNCILIQSVTGL-FEKRIDLLKV  
 DVNLQIILLFQSYNNGCNIKSIKLSAK---TGVVPIP-KKLNYKECALINQSVTGL-FENEVDLLNL  
 IVWHRPQNGNIVYDDFKLSDRIGLIK---TKNHLNANNIVFISQSPFI-YEYKVKYK  
 GLFKLTSRGSAADEFELGKIPRYRDY---TPSSLLKDATLHQSVTGL-YETRIDLAKIGEG  
 INLLFTYAGAPATFRFEDKNIDRRY---TSTTEILNATLHQSVTGL-YETRIDLAKIGEG  
 IHFLTUNIGAPAAFKYFDTTIDRRY---  
 SHRRANKSGYTFSEDEFQSPSGL-FERKVTVGBELKRRKAKAVEVNSKYRTNEK  
 EPCRLFPSHFLLADTDEFQSPSGL-FSTQKTVAGLYOETK  
 ARMRPDKKL-LNATLHQSVTGL-YETRIDLAKIGEG  
 HKLQSGGKILSDNSEMIYQSVNGI-FETRVKISDILL  
 GKCRKNITNYKEFKLIQSVTGL-YSCERKDLMTI  
 RK---THGLTL-NTDFIYQSVTGL-YEARIHIE  
 NSMTRAFGLRQKDRVITATYSGLTKTPKSPKSLFSLAESNEL  
 SRISKPK---KPEEVALIGYESITGLKYRKRSPVVTKR  
 IGLQVSGGKILDKDQTIQVQSPSGL-FKRRIPLADL  
 EPEE---EPEE---EPEE---EPEE  
 E---E---E---E---E---E  
 NKVV-NITYMGE-I FKK  
 FLMLSTILNDYILLDANDNFDLIGL-SKNRIDEILNSKLGDKIVA  
 VSKKITNEDEYKISHVNPFI-NLWNTKALENN  
 AEPSTNPIKEYPPI-HLDELGN-EYVVKIHEHTDDEKLMCTIX  
 QDTSKTPKRLMFGIKSMMNYEQ-DISFFGI-NKALFE  
 GLGKTKFKKY-RYDILGN-KYSCSEKFTSFC  
 NLNIAKYKREGKLRKP-NTDILGN-QHLIKNEGDKPKLDF  
 QCKKGLKSNISIIYKV-RTDVIGN-VTINTQYTKPQLFKRGN  
 TIGKVNISIEKI-TDVLGN-VTINTQYTKPQLFKRGN  
 MRIMPTISKLVIRIDKY-ATDVGN-LYKVKNTLKEFD  
 AVDLGKVOGNGGKISCEPIJSL-LAKEN  
 RGLIQSLLEIFEK-QVDYFGR-YKRVHEKRLQFVKARDE  
 GLAGLLKFEKY-QVDYFGR-YKRVHEKRLQFVKARDE  
 MRIGIKVTANKKY-QVDVLGN-YKVRKXRQTF  
 ITGEMVKEICPPI-KVDLGN-IIVQVSCILLN  
 INKVCIKI-RVDLGN-VRL  
 SKGLDTIKV-RVNHIGQ-IVSVGEY  
 GIIKAKDLDEVRSEYVFTGTFEGS-IEMFKSESGYRVEDDPHH  
 QDDFTINSMGOYPRKI-ILSSGEF-IKRVSPILKKEG

Figure S2 (Continued)

325972003	<i>Sphaerochaeta globus</i> str. Bddy	VALENAKPRTF
296446027	<i>Methylobacterium trichosporium</i> 083b	R---SVGA
347536497	<i>Flavobacterium branchiophilum</i> FL-15	RIVIEGLSIQRIV-RPSGKVDYGV
34885718	<i>Prevotella</i> sp. C561	RKYKTRFESQ---SN
282880052	<i>Prevotella timonensis</i> CRIS 5C-B1	RLYKVVGTEDK---GR
312879015	<i>Aminomones paucivorans</i> DSM 12260	R---GDGR
294086111	<i>Candidatus Punicelaspirillum marinum</i> IMCC1322	L---SGSK
330822845	<i>Allicyclophilus denitrificans</i> K601	I5---GSNGQ
344171927	<i>Ralstonia sylvii</i> R24	M---SENK
159042956	<i>Dinoroseobacter shibae</i> DFL 12	IANG
83591793	<i>Rhodospirillum rubrum</i> ATCC 11170	VDGRLLMPARL
288957741	<i>Azospirillum</i> sp. B510	LEP---SSNS
427429481	<i>Caenispirillum salinarum</i> AK4	LDA---AAGR
92109262	<i>Nitrobacter hamburgensis</i> X14	LSP---SNL
148255343	<i>Bradyrhizobium</i> sp. BT411	GAMVLLSSINNSPCEGFTCT---P
34557790	<i>Molinitella succinogenes</i> DSM 1740	SSIVBLVSKED
218563121	<i>Campylobacter jejuni</i> NCTC 11168	---EEEE
Unet	218563121	---EEEE
291276265	<i>Helicobacter mustelae</i> 12198	KSTNSAKATIEL
222109285	<i>Acidovorax sibiricus</i> TPSY	R---STGA
385156657	<i>Bacillus smithii</i> 7 3 47EAA	-S---TNEE
220930482	<i>Clostridium cellulolyticum</i> H10	R---GTGS
297182908	uncultured delta proteobact. HF0070 07E19	I---ANGQ
194250555	<i>Parvibaculum lavamentivorans</i> DS-1	VM---ASGQ
218767588	<i>Neisseria meningitidis</i> Z2491	R---GIGN
15602992	<i>Pasteurella multocida</i> str. Pm70	R---ATGN
187736489	<i>Akermansia muciniphila</i> ATCC BAA-835	DP---KHAG
315605738	<i>Actinomyces</i> sp. oral taxon 180 str. P0310	SWRVSALDTPSK
117929158	<i>Acidothermus cellulolyticus</i> 11B	SAEQAEVLRTER
189440764	<i>Bifidobacterium longum</i> DJ010A	IRIRPRYLAAG
403744858	<i>Allicyclobacillus hesperidum</i> URH17-3-68	RIH---
407803669	<i>Alcanivorax</i> sp. W11-5	---YFVSN
423317190	<i>Bergeyella zoohelcum</i> ATCC 43767	---QLIGDG
402847315	<i>Porphyromonas</i> sp. oral taxon 279 str. P0450	GKNIYVQKFSKQ---IYFKET
404448728	<i>Barnesiella intestinihominis</i> YIT 11860	SEHLXVQRMSYS---YVFRYH
374384763	<i>Oderibacter laneus</i> YIT 12061	MKLYRVQKLSKSD---YSEFRYH
384109266	<i>Treponema</i> sp. JCA	SKHLXRVQKLSGMY---YTFRHH
402849997	<i>Rhodovulum</i> sp. PH10	KLDIRPV---YAVSYC
Unet	331001027	MRV---RLSPHN
331001027	<i>Parasutterella excrementihominis</i> YIT 11859	---EEEE---HHHHHH
34557932	<i>Molinitella succinogenes</i> DSM 1740	SRRLVEMFGKFLAMFGE
54296138	<i>Legionella pneumophila</i> str. Paris	PISAIKRSKRRIIFRALFEKKEKRRK
319941583	<i>Sutterella wadsworthensis</i> 3 1 45B	TRYRITLOSADFIATIDE
254447899	gamma proteobact. HTCC5015	GRRYRVETFIQASHWFEQSVENNA
118497352	<i>Francisella novicida</i> D112	NRFYIRFTVDSQFERHF
		SRPKVRVKLDYVIDDS

Motifs  
informative positions

Figure S2 (Continued)

VLIITFTEGSGYQIVFSNLAKSKGQ-----DSTFTLTIKGYDVRKV-QLSAGL-VRYVSPDLLVDKIEKOEVALCGE-----  
IQLSAHSSSDVVOGI-----GARTLTERKF-NVDRFGR-KEEVERELRTWGTETWRGKAVI-----  
IMLYFKEARMA-DLQDNFAPDGVFKLG-----ENKPTRKNNHM-OFAFVEGLDFKVLPSGK---FEKI-----  
IWMTHHLSYSEB---RGRSLGRTVYQNL-----PEIERSVSKSL-NFLIMGNRDFVKN-GKILFNHR-----  
IKFKYHQARVCGGLFIFSTYKWBDDY-----APIFQSNMI-NILVDCI---DFIIDLKRYTLKE-----  
LILDRHRMPRESDCGTFYVS-----TRADVLSHKKY-QVDPIGE-IRLVGSEKPPFVL-----  
ILMAPHTEANVDARORDKNDFKLFSK-----SPKQLQASARKV-HISPTGL-LREG-----  
IFMAPIHEANVDARNTDKDAFTYTSKYA-----GSIQKARTRRV-TISPIGE-VRDEGFKG-----  
ITLBBHFEGGTLKORDADKDDPFKYLITL-----KPGCALRDLGARRI-FVDLIGR-VLDGKIGKD-----  
LFIYPHNEANADPRNNDKSDDFKWTQI-----GAPALASGIRRV-SVDEIGR-LRDGGTRPI-----  
ATGKATALYAQLSCPENLNLNGDQGYCV-----QSAEGRKIKRIT-SCIALGR-LRLSKKAT-----  
VVVYEPHQVKTDRSKVKKISC-----DOLRARGARRV-TVDPHGR-VRVHAPGARVIGGDAGRTAMEPARDI-----  
FKLADHNETGNLDRKHAATNNDIDPRM-----VCDLEVARGD-TVKDGKTKGV---ITQGIQGMFVDAAGSARPTP-----  
LYLVHEHQAGVQTRHDDPESF-----LMASTYTKLKAAPV-RVDEIGR-VRVVMPN-----  
VSMOKKHKCKLCPENRIAGRCIQGFLDWSQ-----RNLFSADKRENNALV-NIDTIGQ-PWRKNGLETGSEDAIRLIGTRFKKWF-----  
NKETLSKQKILFKNANEKVIAKSI-----EELRPRKPEFCQKALDVKKY-QIDPLGY-YVEVKQKRLGTTI POMSAKKLXK-----  
-----HEHHH-----EEEE-EE-----E-----  
EHLKYLAKNEDEKMTYDDEKNNKT-----MTRESQIQLKVFQKV-KLSVLGE-VLEHKFRNRQNIALKTTFKXV-----  
MNLMAHRAASVGEKGLIRGI-----GVKALSVEKP-NVDVIGR-IYLAPETRSGLA-----  
LILINDLFAKTYTIDSAAGLELISHDR-----NFSLRGVGSKTLRKFVKY-QVDVLGN-IHKVGEKXRVGLAAPTNOKKGKTVDBL-----  
LKSITYLEGROQDFE-----GVRPAISIEKY-NVDLIGN-KSIVAGEPRGSEKYNKSKN-----  
VLEKDTADHSTYTRMP-----GANRHAFAKV-QVNLGK-VIK-----  
INIRIHDLDKIKGNGLIGI-----NPIKLDMAKV-SIDPIGR-VRPSND-----  
LSLSEHDGELISEGKGVV-V-----GVKALSFOKY-QIDEIGK-EIRFCRLKXRPVVR-----  
VWRLSIOSSGGVKLDLQARCAVPK-----GVKLALSEKY-QVDEIGK-NROICRPOQRPVR-----  
ITFLPRLSMEPLKTRVGGHESDLVVAECVEKINK-----KHTHCNREVDLISLKK-YQMBRYPTSYTTPR-----  
SDREPTLEAGEILAQEFFRCWRATVA-----KVLCHPGLVI-RRTALGO-PRRRGHELFPYSWRPWSADPWBGGTP-----  
LAKAFSDVVPDGVOKIVTQGMLEPPV-----NTASKTAVIV-RNNAFGE-PLLSAHEMPCSNWRHE-----  
LABASIS-----GIQGLEVEKY-VVGLIGD-THPVYKERMPFRVERKSN-----  
GGKLLIATP-----VTEKPADL-LSARWGR-LKVGGRN-IHLLRLCAE-----  
TADILKK-----EGRSIK-ENCFKL-EIDRIGN-IVKVIKR-----  
LETSVAD-----DWTSGRIPEFRVQSLKAY-----KER---NIRKV-RVDLIGR-ITSLJ-----  
TETSVED-----KYDGKFNKLSMOKGLKRV-----SIXSLG--LNPHKV-HISVAGE-IKEIS-----  
LASTLNN-----EREERIOGLEAW-----KRA---MPVKV-QIDEIGR-ITFLNGPLC-----  
ADWLNSTN-----ETMLGYNKPTPTQNVSVN-----VLEDKOKARLV-TVSPIGR-VFRK-----  
DGCKIQD-----RHADADDPFRMDLATIPLL-----KDR---GCVAV-RVDPIGV-VTLRRSNV-----  
-----EE-----EHHHHH-----EE-EEEE-EEEEEE-----EHHHHHHHH-----  
-----NSISLGSSEKFSVSEVD---NKIYTERAEFTKFLPKPREDMKGN-----GTTI-FEELVG---PRVIFNYIVGGAASSLKEIFSEACKER-----  
KETTILSFKESGFQDYLKVF-LKKI-----VKI-QLRDTGSISNIVVRKMAADFLSFRSEHLQRLK-----  
-----ALMKFSDSIDDPLAMP---NEIVCKNKLFGNELAPRD-----GRM-KVSTG---KIVTEFEESDSTQWIOQLYVYQLKQK-----  
ITSPULSPASFKVQKPAEFO---KAV---CTELSELLQPR-----SEI-FIENVGNAKHIREWYIVVSNKKNKONESTYNVSKS-----  
-----KSALRVNGIQDLDTVWNTFDNRSRIPSLIIPR-----SNL-FELLEFG---OKITFEYIANGANAFAVAKAYSILRA-----  
-----KINYPNHSLSLKSRY-----RVL-BILK-----QSTIIEFESSGFNKTIKEMLCKMLAGI-----  
-----PD-----

G

Figure S2 (Continued)



16130662	<i>Escherichia coli</i> str. K-12 substr. MGI655	-----MTWLPINPIPKD
11500011	<i>Archaeoglobus fulgidus</i> DSM 4304	-----MRK
18977490	<i>Pyrococcus furiosus</i> DSM 3638	-----MRK
46447796	<i>Desulfovibrio vulgaris</i> str. Hildenborough	-----MKKLL
54296139	<i>Legionella pneumophila</i> str. Paris	-----M
331001028	<i>Parasutterella excrementihominis</i> YIT 11859	-----M
319941582	<i>Sutterella wadsworthensis</i> 3 1 45B	-----M
118497353	<i>Francisella novicida</i> U112	-----M
34557933	<i>Wolinella succinogenes</i> DSM 1740	-----MT
187736488	<i>Akkermansia muciniphila</i> ATCC BAA-835	-----MSY
407803668	<i>Alcanivorax</i> sp. W11-5	-----MSEQ
325972002	<i>Sphaerochaeta globus</i> str. Buddy	-----MW
187250661	<i>Elusimicrobium minutum</i> Pe1191	-----MW
47458867	<i>Mycoplasma mobile</i> 163K	-----MGW
363542551	<i>Mycoplasma ovipneumoniae</i> SC01	-----MK
71894593	<i>Mycoplasma synoviae</i> 53	-----MAAK
384393287	<i>Mycoplasma canis</i> PG 14	-----MSK
284931711	<i>Mycoplasma gallisepticum</i> str. F	-----MQLPPEHLCSGFLFMSK
117929157	<i>Acidothermus cellulolyticus</i> 11B	-----MTGPM
317482065	<i>Bifidobacterium</i> sp. 12 1 47BFAA	-----MQNQM
315605739	<i>Actinomyces</i> sp. oral taxon 180 str. F0310	-----MNSQM
294086112	<i>Candidatus Puniceispirillum marinum</i> IMCC1322	-----MLK
282880053	<i>Prevotella timonensis</i> CRIS 5C-B1	-----MLK
374384762	<i>Odoribacter laneus</i> YIT 12061	-----MLY
423317188	<i>Bergeyella zoohelcum</i> ATCC 43767	-----MIK
385811610	<i>Ignavibacterium album</i> JCM 16511	-----MKK
402847305	<i>Porphyromonas</i> sp. oral taxon 279 str. F0450	-----MYK
345885719	<i>Prevotella</i> sp. C561	-----MIK
404487227	<i>Barnesiella intestinihominis</i> YIT 11860	-----MIK
301311870	<i>Bacteroides</i> sp. 20 3	-----MIK
60683388	<i>Bacteroides fragilis</i> NCTC 9343	-----MLG
402849996	<i>Rhodovulum</i> sp. PH10	-----MIR
154250554	<i>Parvibaculum lavamentivorans</i> DS-1	-----MD
159042957	<i>Dinoroseobacter shibae</i> DFL 12	-----MTG
286957740	<i>Azospirillum</i> sp. B510	-----MIG
330822846	<i>Alicyclophilius denitrificans</i> K601	-----MIG
344171926	<i>Ralstonia syzygii</i> R24	-----MAAN
319957207	<i>Mitratifactor salsuginis</i> DSM 16511	-----MAAN

Figure S3

RWSMFLQGVQIDVIDGAFVLDKNT-----GIRHHPVGSVACIMLE-FGRTRVSHAARVLAQAQVGLVWVGEAGRVVAVSQCGG-ARSDKLYAOAKLADLDELDRLKV-VRMFEFLRGEFAP-----  
 KNYLVSQG-KLRBHEWNTYFENE-----DKRPPINSIYALYALG-SLSITSKASILLAKEGVCHFFWRYGYTGSFPRESLVSVEVLDQAEHLLDSERLHL-ARAFVEGAILMARVLK--AEQD  
 KSLTFSQK-TILARENTLYFENY-----NGRPLAIEGIDYIYIG-HWNTSQALHYIAQKGLIHFHNYGYTGTTFPRELISGDLIQAQAEHYLDKRRLEL-AKSFVUGAKNMEKMLK--WGIS  
 NPLVYTIQGTYLAGEGECIVWRGD-----EVRLRVPHSLGGVCFG-QVSCSFLMGEFAERGLSEFLREHREGLARVQGPVS-ENVLRRQYERADSPESABEV-ARSIVSAGVNVNARGVLOK--NRD  
 KHLTSEQSGCKQGERLWFKND-----QWVEGTARLAKTISIVKQGVITSLMIVACAMRGIOPLIDYSKAVCSLQG--G-LERTAKVRESFKYIYKESNMAARFILAKIKKOSAVLKY--FAKA  
 RHLTVSGDFLVRGNRNLIVRNGD-----NVILEALSARTRID-KSVSASSMILLACAAARGIRYVFDIMKGVGVVEVSGHR--RHAVVGVNRSQFECLSIQOQLLAKELIFSKINQRRAVLY--FSKY  
 RYLTVSSGTAIGLSGERLWHEAD-----GRWETALSARTRIEKKGISVSTHMLACAAARGIRLFDVDRGVGVVALSGLHQ-HAVVSTRBAQFLAHSSQGRTI-BAEMIAKIRINQRVILY--FWKY  
 KHLIYSEYIYLGIESGRLVYK--DOKYPLNRLATLSIARAGVSPSDVQFSLRGLKFFLDFRVAESMIVGANO-HAVVQARI-NQYRYIDRNALTL-SIKLIAAKIKNQBATLNY--FNKE  
 KHLIYDEYGVFGTKSERLTVSQG-----ELKAEIPLSRKSWNLIKGGISLSSDIIEACSLRGTKIFFLDFRVAHTATISTAN-HAVANVRKAQFLTESNHGREI-ARTLIVKLNQRATIQVFYKYLK  
 HILSIDAYTCHLSCDQKQRCANDE-----NSPRTIPIEDVAVLSSSEKATIMLTELAKRRIGFVLCSEYR-PAVILLAPAD--RSTDTGLRLHLDAMPALRNL-WQTLDAKGNQTALAQ--WPHR  
 HILVSPC-ALNHLIECLVSRKQ-----MEPLRLLPDSIAVLCIHHHTVQLSIVLRAKLETAGAAIIVYDDEHQIPAGMLQDYG-RPNVAVLROQJIALESQSDRDLVWQVLSWKLATQSRLLG--LERR  
 RVDLGGTYLIVANKLIVEAD-----GEESLFPDDIHSIICHGIGFRYSDFKFSCLRHHRVYVTFCKEKIPIAGMLPMLQ-HKDTSRMDIQJAKASLPRKQA-HQOIIITEKLNQDFLQK--LQIP  
 RVDLPGDGYHLVKNNSAVKDR-----EEXLHCLFDIDINSIILVGNNTISNTC IQKCLEHRVPIVFCDKTVNPAAMLSST-TWYGRRLQIQINASKPQJXA-HQOIIITSKLNQAEVLKR--FDTL  
 KIVEINTDE-VYHLYLNNLYIKK--NEKILANTRIDITLLENNOYISIHALLFLAKNNVINIFWNEKNEPNSYLIPIEG-NHNSLKVVAEOVWTKYKAIL-WKDIKKNKIRNOKNLLK--WKLK  
 KIIIEISE-VYVLFANHTYIKD-----SEKVFPIPTVQVLIFENDRANISIPVINELEKKNVILICKHL-POSLEIPIYG-YVNN-KIPQEQIKMDIPIKKT-NOEIIKIKIORSIVLKS--IQK  
 KWELIKET-YLPHIMSLIVKKE-----NNKIVLPSLIDITLISNPTYSVPLINAVSNINLICKNDFEPVQLSISG-YVSN-KNPLSQIANTQEFKDKT-MEKIILKAKTNYVNIYF--FGLL  
 KVLEISQSE-YISYFMSLIVKSN-----EKRIIVPISQIDTVVSNPRCHISVPLINELVAQNVNIIICDSKTKPVOJSPIQG-YVSN-KNFLAQIWNKDFKGFV-NREIITKIVNYTKLIY--LDIT  
 KVLVSEYE-FISLHLDLIVRKE-----GARVTLKLTDLITLITNPTCHISVPLANKIVENIINLIVCNKQPTWOJLPISS-YVSN-KYLMQIRWTDQKQI-NKQLVWKLKIINYARLLNH--LNT  
 RVDLSELSGEVHAOGGALVGD-----RVLVDVA-HMLTGPVLSHGSDVDRMAANGVGVVHCDMRGVPVAAIIPMST-HNRVAARHRAQAELSIPRQNA-MNVIVTKIINQAALVRA--LRD  
 RVDICTLDGSIATERGRVWRNDT-----GECDVPLAQJAVVLL-GLWRVCSAVLHEMARYGVSMCDMRGVPVDAALHAWTDMPTVTTTQLQASNSLPRKSA-WARIVWAKIISGQAACLDS--SKLE  
 RVIDLGGFETLRSTRGVVECAQS-----GLTYPVIAELAVVY-GAKWLSAAVHLRAEAADVLECDMRGIPSGCYSMSD-HGRVAARHQAQAEVTLPRKNA-WARILIRAKIISGQASVLEN--IKFR  
 -MIESLQPRHSVRRGAIIVSINE-----TELQIDLDQVLSVIVTSRGAVTTPMITEAARNIPIIICNDREQPVYSLSPVQ-HSDQTRRFEIQAQAKGIGKRI-WOXIIGKGVNQAQZLVZ--YQSA  
 RSVVFSHPA-SLSLRYNQLVTKDLP-----DERTTPIEDIGLAVDNPVHSVYVPLINALTENVVSVFCQKGPYSMLFNLDN-HNTQGETLRNQLSASQSNFQL-WKQIIECKIRNOQALLEK--IGRE  
 RTLFFGPS-RLSVKMQLVIEIKET-----GELHRAPTEIDGVTWENQOISVSLPQGFNSNVAVFCDSASHPMSMLINDG-HHLSQEMFRFOINASEPLKKNL-WKQIIEAKIRNOQALLDX--SGRN  
 RSTYIGPA-YLKLKQDKMTCVPEK-----AEGSVPEDEIGLMLDHFQIITISQLIQWAGNVVILISCDALHPLHGMPLPHG-NAYSQRLKQIASEPLKQJ-WKQIIECKIANQANVLER--LGT  
 RTLYFGPA-YLSLRNMQVLRPEVSEKAE-LPDIK-----TTRVTTPIEDIGVWLDHPCITITTHALLGALENNCAVTCSATMPSGLLPLDG-HTLQTEAFRAQLDASLPLRQL-WOQTVAKIINQAHALHT--VLGE  
 RTLYFGPA-YLSLRNMQVLRPEVSEKAE-LPDIK-----QETIRVTPIEDIGVWLDHPCITITTHALLGALENNCAVTCSATMPSGLLPLDG-HTLQTEAFRAQLDASLPLRQL-WOQTVAKIINQAHALHT--VLGE  
 RTLYFGPA-YLSLRNMQVLRPEVSEKAE-LPDIK-----KEARTTPIEDIGVWLDHPCITITTHALLGALENNCAVTCSATMPSGLLPLDG-HTLQTEAFRAQLDASLPLRQL-WOQTVAKIINQAHALHT--VLGE  
 RTLYFGPA-YLSLRNMQVLRPEVSEKAE-LPDIK-----RQSEITRPIEDIAVLDNKQIITTOGLLEALENNCAVTCDSRSMKPVGLMPLYG-NTTQNERFRQLDASLPLRQL-WOQTVAKIINQAHALHT--VLGE  
 KTLVFGPA-YLSLRNMQVLRPEVSEKAE-LPDIK-----KQAEVTPIEDIGVWLDNKQIITTOGLLEALENNCAVTCDSRSMKPVGLMPLYG-NTTQNERFRQLDASLPLRQL-WOQTVAKIINQAHALHT--VLGE  
 RTLYFGPA-YLSLRNMQVLRPEVSEKAE-LPDIK-----GLLAKTPIEDIGVWLDNKQIITTOGLLEALENNCAVTCDSRSMKPVGLMPLYG-NTTQNERFRQLDASLPLRQL-WOQTVAKIINQAHALHT--VLGE  
 RTLYFGPA-YLSLRNMQVLRPEVSEKAE-LPDIK-----LPKATPIEDIGVWLDNKQIITTOGLLEALENNCAVTCDSRSMKPVGLMPLYG-NTTQNERFRQLDASLPLRQL-WOQTVAKIINQAHALHT--VLGE  
 RTLYFGPA-YLSLRNMQVLRPEVSEKAE-LPDIK-----REIGRPLDQIAGVYHAGHTWTLSLTELADRGAPVVLGPNHNAKMLPDLG-HHIGQERHRAQVSEPTKARA-WQALIRSKIAQOQIVLAH--FTGE  
 RTLYFGPA-YLSLRNMQVLRPEVSEKAE-LPDIK-----REIGRPLDQIAGVYHAGHTWTLSLTELADRGAPVVLGPNHNAKMLPDLG-HHIGQERHRAQVSEPTKARA-WQALIRSKIAQOQIVLAH--FTGE  
 RTLYFGPA-YLSLRNMQVLRPEVSEKAE-LPDIK-----RELGRVPLDQIAGVYHAGHTWTLSLTELADRGAPVVLGPNHNAKMLPDLG-HHIGQERHRAQVSEPTKARA-WQALIRSKIAQOQIVLAH--FTGE  
 RTLYFGPA-YLSLRNMQVLRPEVSEKAE-LPDIK-----RELGRVPLDQIAGVYHAGHTWTLSLTELADRGAPVVLGPNHNAKMLPDLG-HHIGQERHRAQVSEPTKARA-WQALIRSKIAQOQIVLAH--FTGE  
 RTLYFGPA-YLSLRNMQVLRPEVSEKAE-LPDIK-----RELGRVPLDQIAGVYHAGHTWTLSLTELADRGAPVVLGPNHNAKMLPDLG-HHIGQERHRAQVSEPTKARA-WQALIRSKIAQOQIVLAH--FTGE  
 RTLYFGPA-YLSLRNMQVLRPEVSEKAE-LPDIK-----EYVGRVPLDQIAGVYHAGHTWTLSLTELADRGAPVVLGPNHNAKMLPDLG-HHIGQERHRAQVSEPTKARA-WQALIRSKIAQOQIVLAH--FTGE  
 RTLYFGPA-YLSLRNMQVLRPEVSEKAE-LPDIK-----KVRKLSLSDTOSIIFEGDRFTLSARVLAALSRHKVATLFCDEYYNTPALHPHQ-SSLATETLKAQKLSLDDLDRL-WQEIIRVKILNQASVLSH--FAKK

Figure S3 (Continued)

315149831	<i>Enterococcus faecalis</i> TX0012	-----MLLGF-----
116627543	<i>Streptococcus thermophilus</i> IMD-9	-----MTW-----
238924076	<i>Eubacterium rectale</i> ATCC 33656	-----MAF-----
218563120	<i>Campylobacter jejuni</i> NCTC 11168	-----MSYDEAF-----
291276264	<i>Helicobacter mustelae</i> 12198	-----MFDEAF-----
325677757	<i>Ruminococcus albus</i> 8	-----MM-----
296446028	<i>Methylosinus trichosporium</i> OR3b	-----MAAM-----
34557789	<i>Molinitella succinogenes</i> DSM 1740	-----MYM-----
222109284	<i>Acidovorax ebureus</i> TFSY	-----MAF-----
336393380	<i>Lactobacillus coryniformis</i> KCTC 3535	-----MTM-----
218767587	<i>Neisseria meningitidis</i> Z2491	-----MMSVKIDKPADLSKDNLCVKLYHPDFLYIGNRDFLGVIMSN-----
15602991	<i>Pasteurella multocida</i> Pm70	-----MGN-----
310780383	<i>Rlyobacter polytropus</i> DSM 2926	-----MGM-----
312879014	<i>Aminomonas paucivorans</i> DSM 12260	-----MSH-----
403744859	<i>Alicyclobacillus hesperidum</i> URH17-3-68	-----MGN-----
421874296	<i>Brevibacillus laterosporus</i> GI-9	-----MGF-----
220930481	<i>Clostridium cellulolyticum</i> H10	-----MSW-----
225377803	<i>Roseburia inulinivorans</i> DSM 16841	-----MSW-----
315659847	<i>Staphylococcus lugdunensis</i> M23590	-----MSF-----
160915783	<i>Eubacterium dolichum</i> DSM 3991	-----MSW-----
323463802	<i>Staphylococcus pseudintermedius</i> ED99	-----MAF-----
389815358	<i>Planococcus antarcticus</i> DSM 14505	-----MGN-----
328956316	<i>Coriobacterium glomerans</i> PW2	-----MAGN-----
422884107	<i>Streptococcus sanguinis</i> SK49	-----MGN-----
116628212	<i>Streptococcus thermophilus</i> IMD-9	-----MAGN-----
24379808	<i>Streptococcus mutans</i> UA159	-----MSY-----
13622194	<i>Streptococcus pyogenes</i> SF370	-----MGN-----
227824982	<i>Acidaminococcus</i> sp. D21	-----MGN-----
169823756	<i>Finogoldia magna</i> ATCC 29328	-----MGN-----
320528779	<i>Solobacterium moorei</i> F0204	-----MSW-----
303229394	<i>Veillonella atypica</i> ACS-134-V-Col7a	-----MTW-----
304438953	<i>Peptoniphilus duerdenii</i> ATCC BAA-1640	-----MSN-----
374307737	<i>Filifactor alocis</i> ATCC 35896	-----MSGW-----
256845020	<i>Fusobacterium</i> sp. 3 1 36A2	-----MSW-----
306821690	<i>Eubacterium yurii</i> ATCC 43715	-----MSW-----
42525844	<i>Treponema denticola</i> ATCC 35405	-----MSW-----
291520706	<i>Coprococcus catus</i> GD-7	-----MGN-----
258509198	<i>Lactobacillus rhamnosus</i> GG	-----MGN-----
336394883	<i>Lactobacillus farciminis</i> KCTC 3681	-----MGN-----
339625080	<i>Fructobacillus fructosus</i> KCTC 3544	-----MAAM-----
310286727	<i>Bifidobacterium bifidum</i> S17	-----MAAM-----
366983954	<i>Oenococcus oeni</i> DSM 17330	-----MAAM-----

informative positions

Figure S3 (Continued)

RTVFRNGE-KLAVKDNLEVIKE-----GNVTYIPLDIESVILEGQDVTITRLLAKAAGHHIDTVICDNTFMPVGFPLGIGQ-YHRSAKRAIWSNLEKEQKVA-WCEIIVTQKIQNQAIVAKY---LSTD  
RWVVSQSE-NRRLKLDNHLVQRH-----GQFTVPLSDISIIVAEQDVTVLRILSALSALSKYNIALWVCDNEHLPTGIVHSQNG-HFRAYRKLQEDWISQKDKKA-WQIVTYVYKINQEDVILAM---FEKS  
RVILIEVEV-TIKVKLJNLIITRK-----GEDIWIPDDISHVILNDLNSLFRMHCQAEGGIGLMLQHSHTLPTGYSYDN-HRSASVLTGQIDKEQDYGKL-WQIVYKIKENQAKAYQI---WTRD  
KTLIISNA-KLNLNHLVTKQD-----ENAKFLKDNINIVLESLSQISSALFNAAKYKILILLTCDNQHSLNFTPLG-HFSQKTAKEQMVSAQKAIL-WQIKIKNKINQAFILKK---ENKT  
RTLFPNSPA-KLSHKNLIIHGR-----EDIG-IPLRDHTIVTDFQITLTSKHLAKYKIVFVCDSDSHLPSGFSPLA-HYSSALLDQJHFSKQNSIL-WQIKIQAKIQNQTNLMLQ---ENKT  
RTVIVTQGE-KLTLRDNHLVVSVD-----NNEQCVPLGDIYSVINDRBAUVSVMALAEANIHVIFCDSRHTTALMLPMT-HYRFGVIRKQLGLSDEFKAVL-WQIVKQKLTNOIICLK---AGIN  
KGLHLRPA-RLNSFMQIVVQD-----DCEARLPEDVAYITLDPHATLSTLISACHAGVAVTSDARFTNGALPFS-HRQAGVAAQVLLSEPFKRC-WQGVVAVAKTINQAALLES---LGRN  
RTVVIATSC-HLRVDMQIVLESNE-----GERTPLADIGVIVENRQVTLSSALISACQGTALMSCDSSHLDGVPFPGT-HSRBTARARQAVWSEPFKRC-WQIVRAKYSQGAELVR---VGAE  
RSIVINRA-KLRBEHALVVEQE-----Q-SARVPEFEDIVVNHREITLVSALISACQGTALMSCDSSHLDGVPFPGT-HSRBTARARQAVWSEPFKRC-WQIVRAKYSQGAELVR---VGAE  
RNLVIQMPA-RLSISRSQVTOPE-----ERYTVVEDISSIMEHKQCTIAPTLSFLAEHQVAVTCDQKHLPCAVVPLGQ-YRKLMALEAQLAVRKTARL-WQKLMKQKTEQAIVLEL---NHCK  
RSLI QNGG-KLSLQRQLIQN-----GESHTVPLEDIAVI I ENRETLI TAPLSALAEHAGTILTCDEQFLPCGQWLPVQ-YRQLM TLKQLNISEPLKQOL-WQIVRQKILNQAFLVADE---TQND  
RSILI SQGG-KLSLQRKMLIQOS-----NNSFTVPLEDIAVIVIESKETITAPLSALAQYGVTLITCDEQFLPCGQWLPVQ-YRQLM TLKQLNISEPLKQOL-WQIVRQKILNQAFLVADE---TQND  
RNFITST-KISTROGGLITKE-----GSEVNTPLEDISIILDNNSVITSRVLEETASRDI VLLTSDERKHPNGIFQYGR-HRQLAVAKAQNINSVPRKRL-WQIVRQKILNQAFLVADE---TQND  
RSVFTMPA-RLSTGGRLVQOE-----GEAAVPLEDELGALVLESFQVSLVPLARELAHRVPLFCVDRHLPGVLLPFA-HCRQEVIRRQNGLSRPFKRC-WQIVRKRKILNQAALRF---LDLP  
RHVAVTSA-KLSCRMQLVVRQDESSGE-----TCEYI TPLEDIRTLEMDRQSIITSAALARAEFRIVFTCDEKHLNGLVLLPCQ-HSRQPKTMSQLEKREPLVRL-WQAVVRKKTENQAALCLTL---CGCS  
RNIITVSNPT-KLTKQNNLWVEQ-----EEVVTIPEDIASIVVEENOWLTAALSRCSEVIGLTYCDEKHLNGLVLLPFCQ-HSRQPKTMSQLEKREPLVRL-WQAVVRKKTENQAALCLTL---CGCS  
RNIITVSNPT-KLTKQNNLWVEQ-----MSIRLMLVVKQ-----Q-SARVPEFEDIVVNHREITLVSALISACQGTALMSCDSSHLDGVPFPGT-HSRBTARARQAVWSEPFKRC-WQIVRAKYSQGAELVR---VGAE  
RNIITVSNPT-KLTKQNNLWVEQ-----GSEVHTPLEDINCILLIENQSVTL SAYLQKVADEGIALVCDKHLNGLVLLPFCQ-HSRQPKTMSQLEKREPLVRL-WQAVVRKKTENQAALCLTL---CGCS  
RNIITVSNPT-KLTKQNNLWVEQ-----DLETKIPLADIPALVIEDIVTITSRGLVLSNVLVFCNQKHLPECSLQPSG-HFNQYIQMQLAWDEARWEL-HSAIVRKHQTECMKY---LDDI  
RIVVSDW-HMSLNLSLAKIKG-----QGDIFPDISIQILVIVDRVSLVPIVNLKTENNVCIIGIDHLPKSYLPMNG-HFAQSGNINQKQITMSNEIKQIL-HQOIVRKAIFNQTIELKT---NOCK  
RVLYEQY-HLQYLDHLVETS-----NLSRIPLNEILCVIEHPNWSMTGHPNALSOQVITLCTNKHLPNTLALPLNG-HRQARLIQNNKWRKMGOL-WQLVVKHKTINQKVIQE---CFKE  
RTVILAKA-KLTVNGHLVSGE-----YNRVPLEISEIFLLENPTQVSHLINALSHHIVTLICDANKKPSVVNSYVG-HRQARLIQNNKWRKMGOL-WQLVVKHKTINQKVIQE---CFKE  
RVLLESAC-KCTYQGYLVVRE-----DAAKIHLEISEISVTLQSTOVFSAYLISELAKAKITVIGSEIKRNPVYLPFYG-AHNTSKRIGESI SWEPEIKRRV-WQIVRQKILNQAALCLTL---LDK  
RTVVNTHS-KLSYKNNHLIFKDA-----SKTELIHLESEVILLETDDIVLSTMLIKRUVENILVIFCDKRLPTALMPPYG-RHDSLSLQSRQITWNETRQAV-WTIIIAKILNQAALCLTL---LDK  
RTVVNTHS-KLSYKNNHLIFKDA-----YKTEMIHLESEIDILLETDDIVLSTMLIKRUVENILVIFCDKRLPTALMPPYG-RHDSLSLQSRQITWNETRQAV-WTIIIAKILNQAALCLTL---LDK  
RTVVNTHS-KLSYKNNHLIFKDA-----YKTEMIHLESEIDILLETDDIVLSTMLIKRUVENILVIFCDKRLPTALMPPYG-RHDSLSLQSRQITWNETRQAV-WTIIIAKILNQAALCLTL---LDK  
RTVITQRC-KLDFCNYVEVQTA-----VSKKRIFIDEIKTLIENGVAITAYLISELAKRNLWILFCDAHPFOSLPLIHA-RFDSIRKIKQMTWQEI KDEV-WDCIVAKIRQALFLDE---LSKK  
RTVLI SGRA-KLDKLANHLVTRK-----BEIVKIHIFIAVLIIDSTWSTFALIEKLAEKVITFCGDHNTIGEIVNYYN-KHNSPLMIRKQISWNNKSKQT-WREITREKINRQALLKF---IGNE  
RTVVISRS-KLDFRNNYIIRKD-----NOVSRVHLESEISLIVETPVSFTALINELVKNKIKLIFCDEKRYKIPASEVILPHG-SHNSKRCREQADMSKDVQAYI-WTEIVRKTMMQAEILKY---KELK  
RVVWDMRA-KLELKLHLVVRQD-----FETKIYIDEIYMLVETANSLSLAVLNELIKKVAIVFCDEKRNPASEVILPHG-SHNSKRCREQADMSKDVQAYI-WTEIVRKTMMQAEILKY---KELK  
REVITIGHA-KLDYKNDYLVRKQ-----NKVKIYKADISHIIVNTTEASITFALINELVKNKIKLIFCDEKRYKIPASEVILPHG-SHNSKRCREQADMSKDVQAYI-WTEIVRKTMMQAEILKY---KELK  
RVVITGHS-KLDRYNSISIRRD-----NGTDFHIGEVNLTILETTISITPAALCELKOKKVVIFCDEKRNPASEVILPHG-SHNSKRCREQADMSKDVQAYI-WTEIVRKTMMQAEILKY---KELK  
RTVVISRA-KLDYKNDYLVRKQ-----EKTORVFEISVLIETAVSITPAALNELIKOKKVVIFCDEKRNPASEVILPHG-SHNSKRCREQADMSKDVQAYI-WTEIVRKTMMQAEILKY---KELK  
RIVISKRA-KLDLQLGRMVRSD-----EITK-IVLISEISITLIESPANSITASLISELAKRINIVFCDEKRNPASEVILPHG-SHNSKRCREQADMSKDVQAYI-WTEIVRKTMMQAEILKY---KELK  
RSFIITQHC-KVTKQKSLVQZN-----GEVYMPIDDINGVITTRALISADALADANAKVIFESGRQVPTETHKLYS-DRRAEQVRLVQVWPLDILETV-WTKIVAAKTINQAALCLTL---CGHD  
RTVITQKA-KLSYKNNHLIQN-----MDIKIPFPQINCILIAITQVGTGLSKLVTEINIKVIFCAEHPNWSMTGHPNALSOQVITLCTNKHLPNTLALPLNG-HRQARLIQNNKWRKMGOL-WQLVVKHKTINQKVIQE---CFKE  
RTVFNDEA-KINTKMDIMIQTK-----EGGRTPIDDIRHILGHTVTVTSYAVNELQKKNKVICNCDKGEPEGEELS YCD-NSRONAYIWKQVFAENRKNIL-WQIVQKTKINQKVIQE---CFKE  
RNVVTRHC-KLSTKNNHLVQTD-----DQVQOLPMDIGKMLIETRAVITCALADLLKRDVKIVFCDDKHLPTIGETSPYET-ESSRNNYI IQQIWDDEARDDL-WQIITREKTAHQVSVLEK-NCDL  
RNVVTRHC-KLSTKNNHLVQTD-----TDVHOIPEVEDIRLVIATRAVITTYAVNELLKRVQVILFTDDKAFP IGEINVLG-GENRKNVIOQVQVDPKERQNL-WAEIVRKTMMQAEILKY---KELK

Figure S3 (Continued)



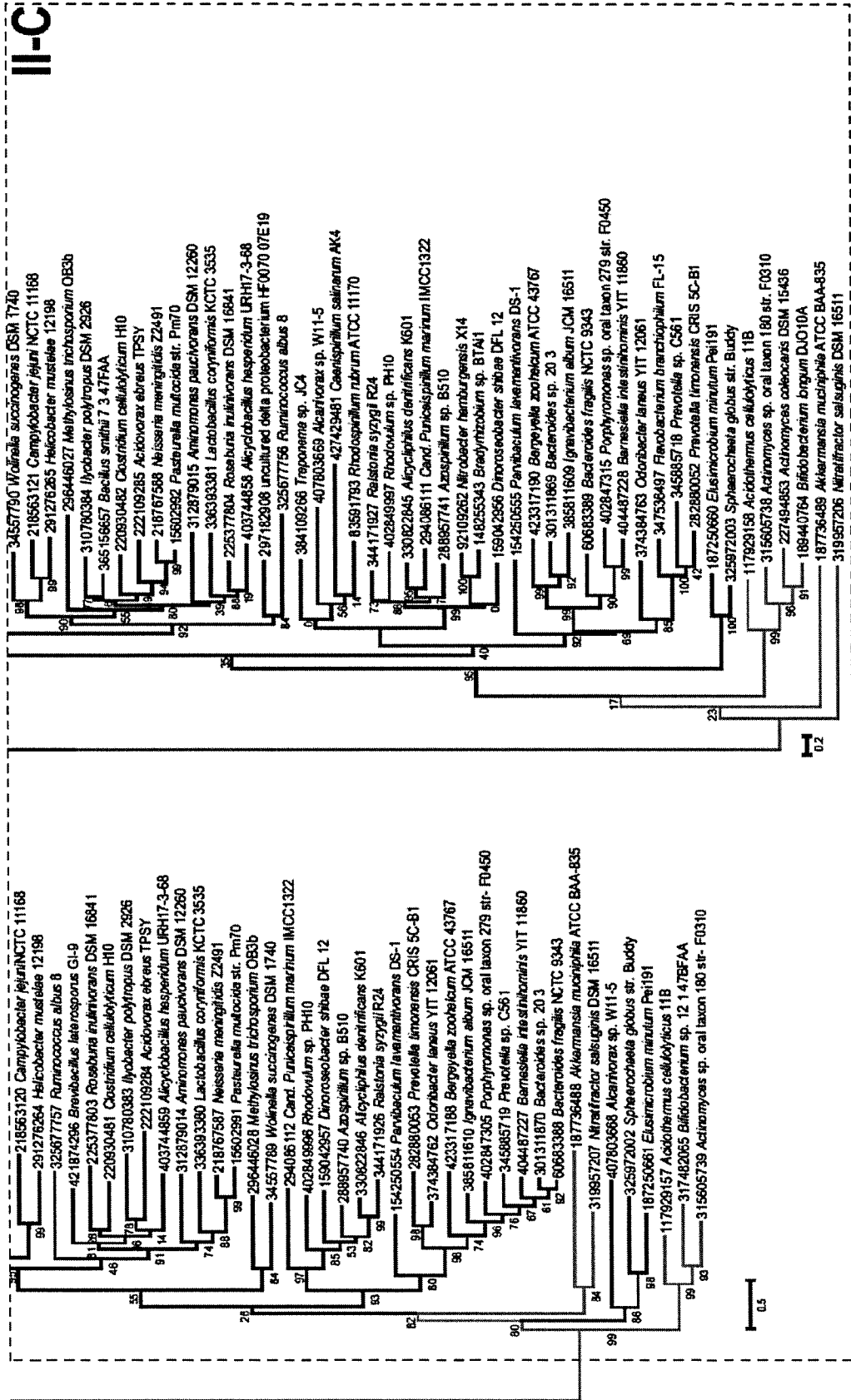


Figure. S4  
(Continued)

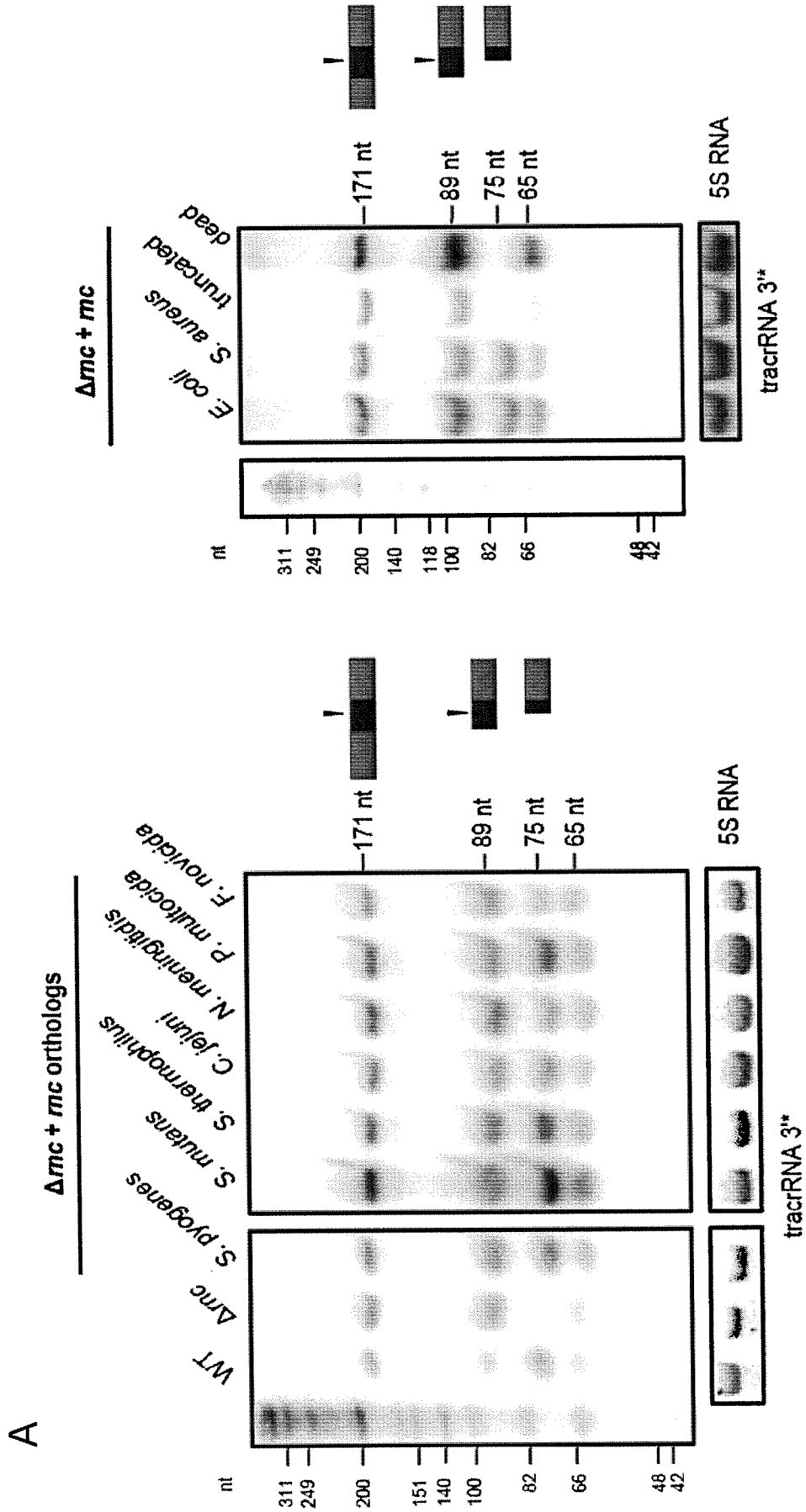


Figure S5

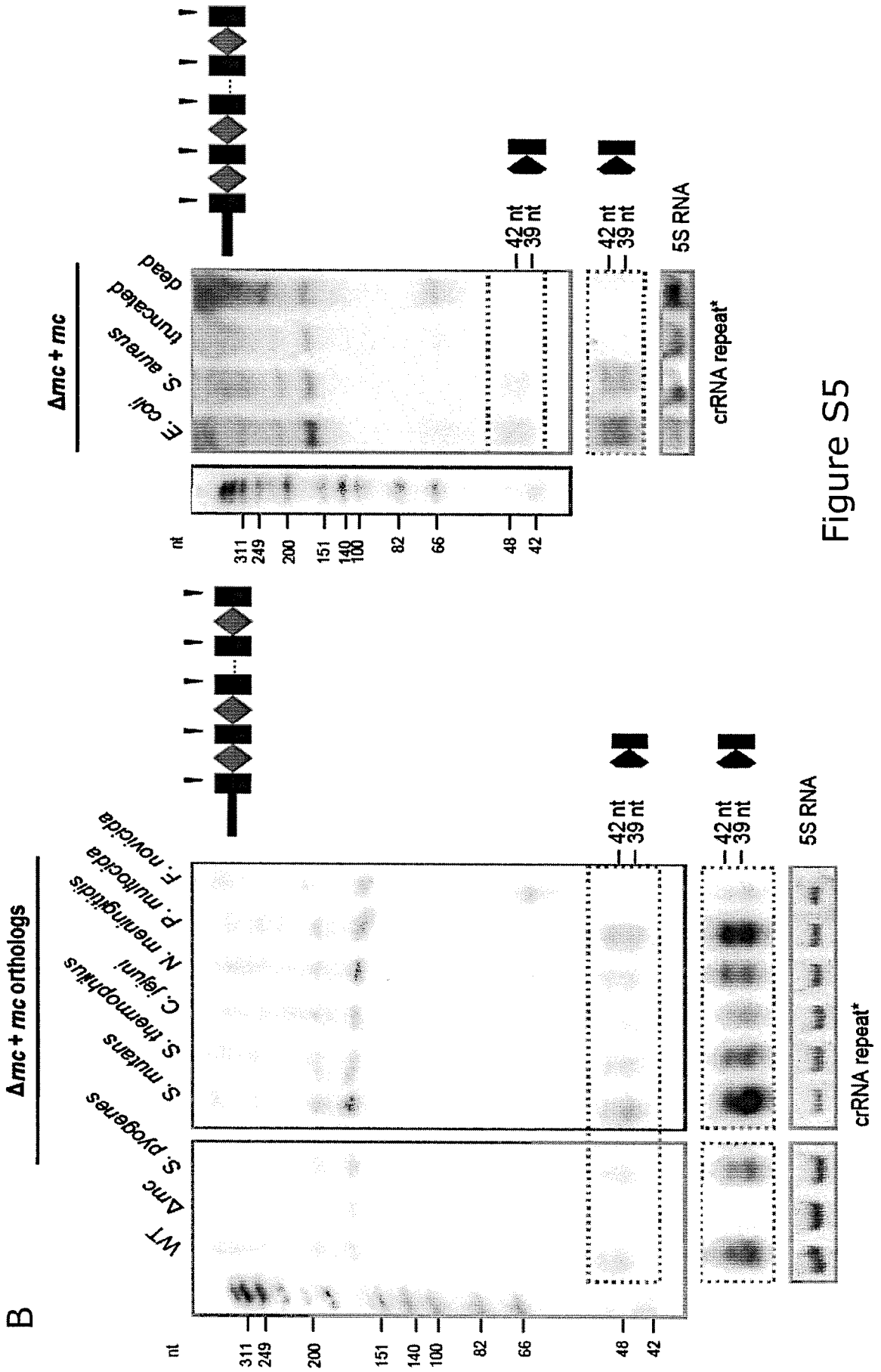


Figure S5



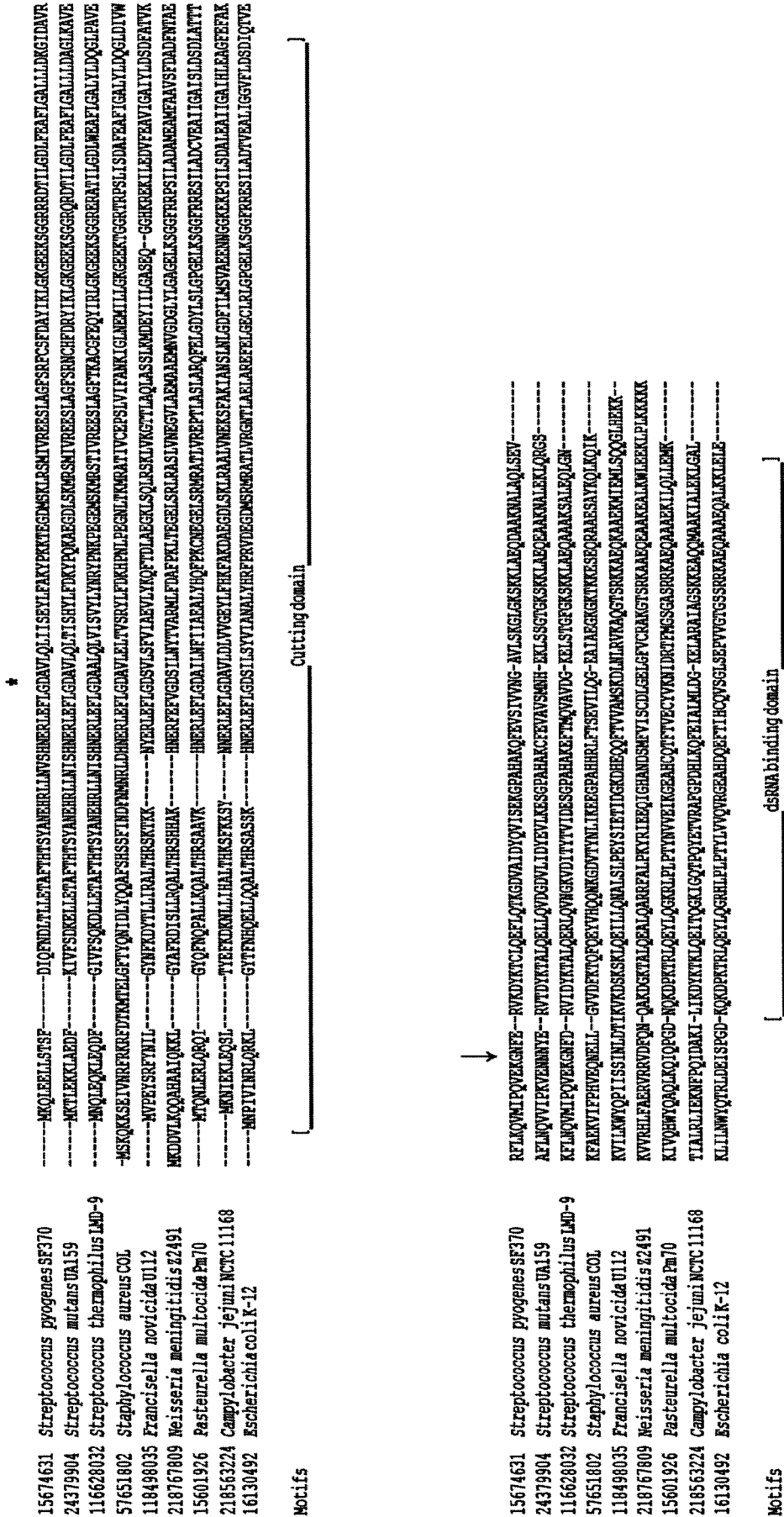


Figure S6

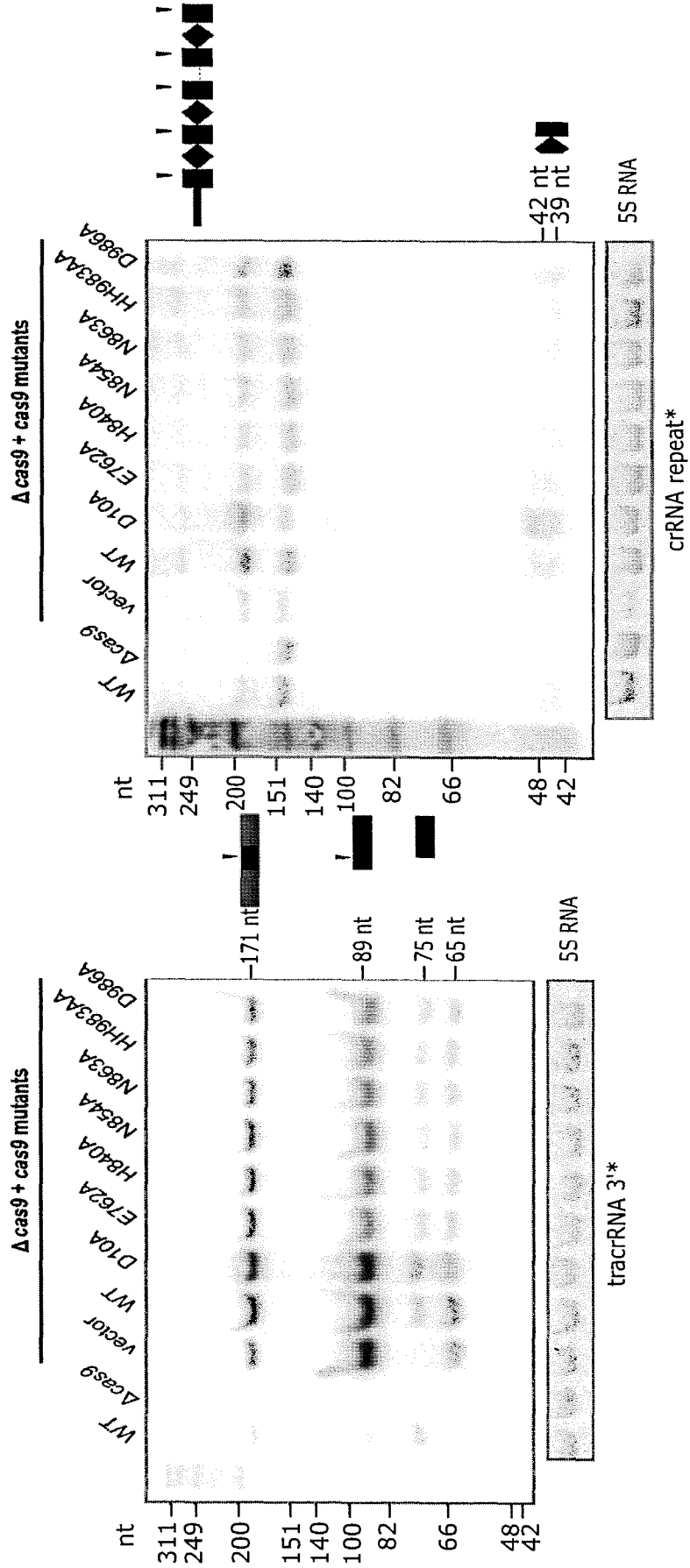


Figure S7

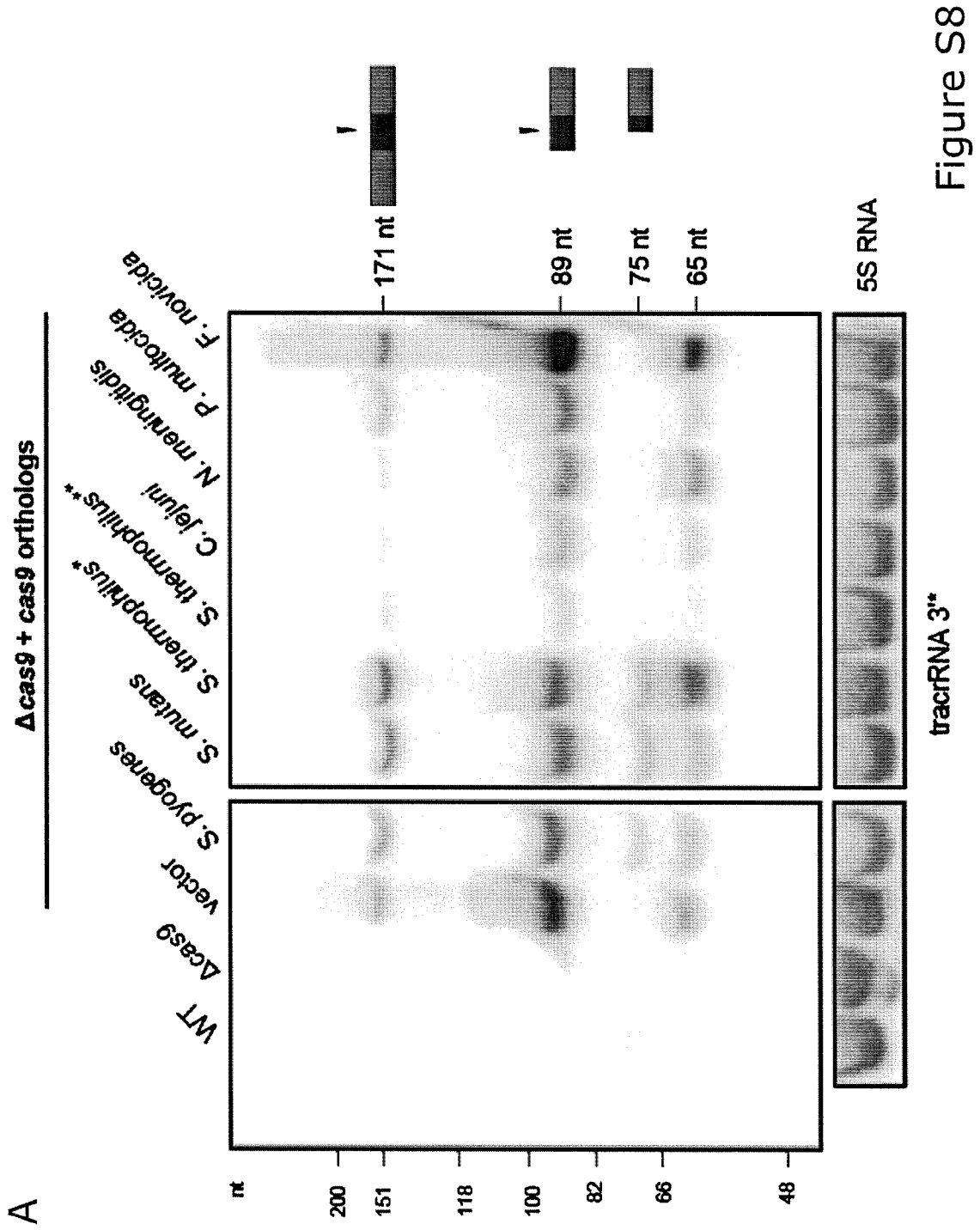
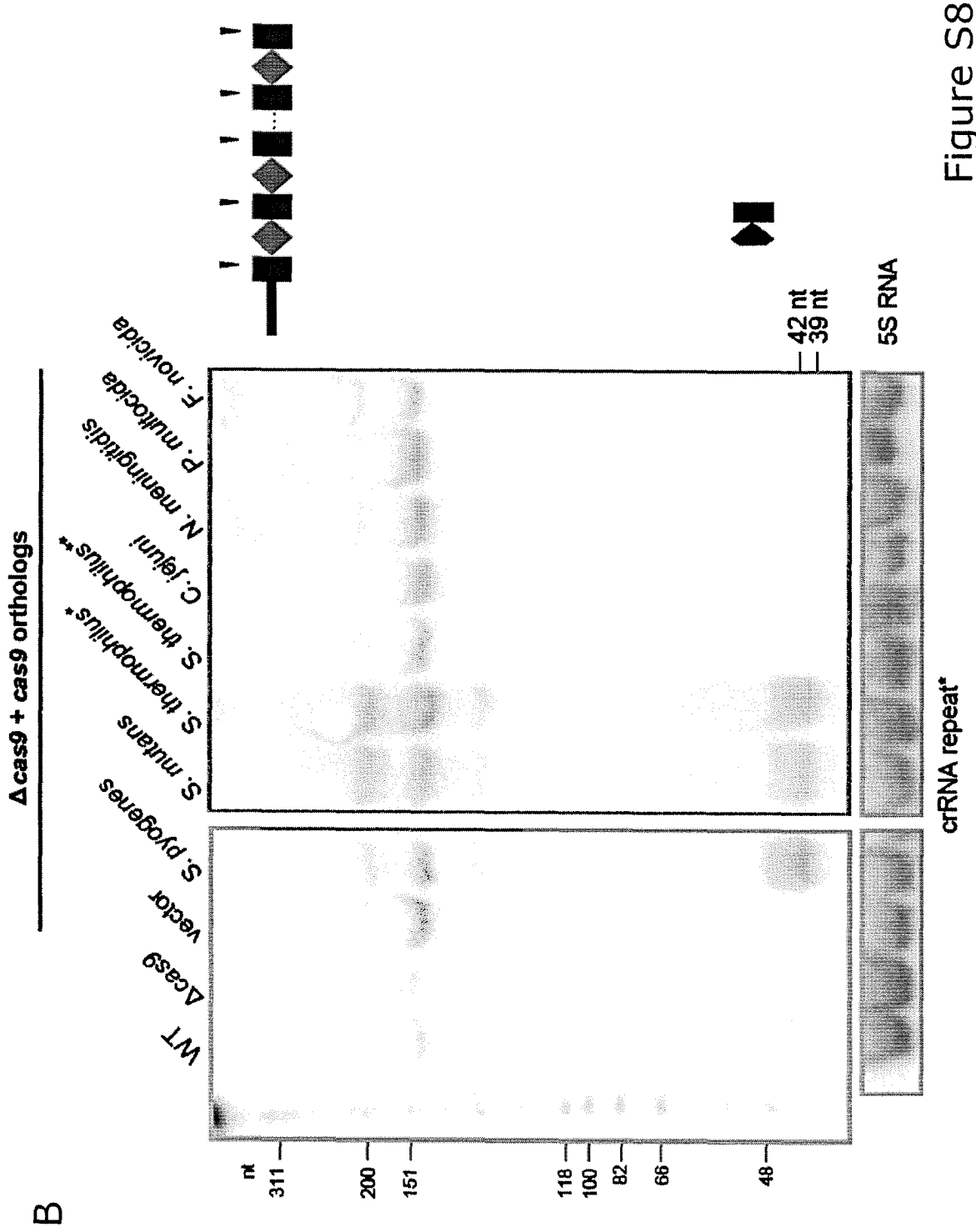
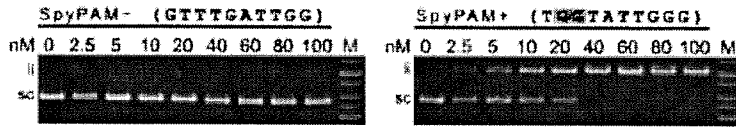


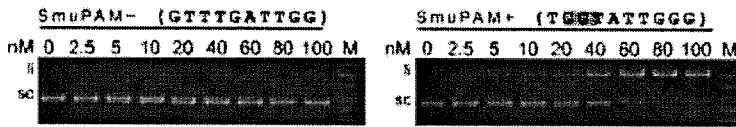
Figure S8



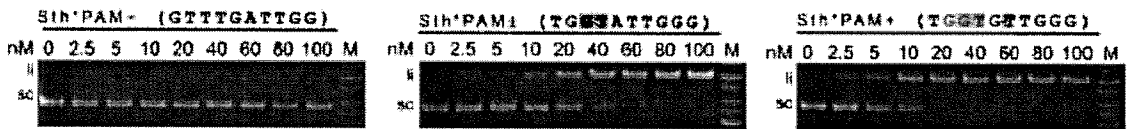
■ *S. pyogenes* dual-RNA:Cas9



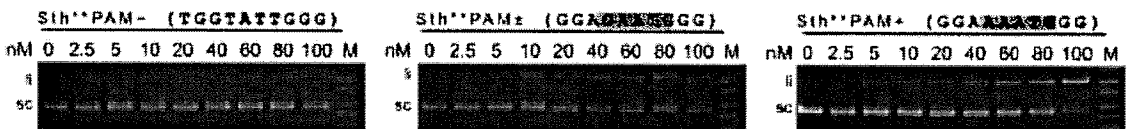
■ *S. mutans* dual-RNA:Cas9



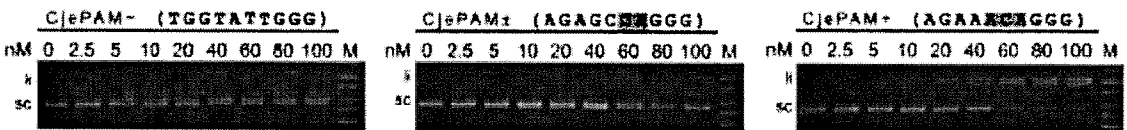
■ *S. thermophilus\** dual-RNA:Cas9



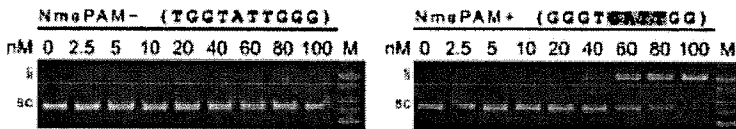
■ *S. thermophilus\*\** dual-RNA:Cas9



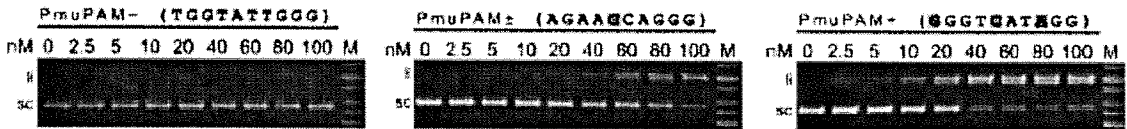
■ *C. jejuni* dual-RNA:Cas9



■ *N. meningitidis* dual-RNA:Cas9



■ *P. multocida* dual-RNA:Cas9



■ *F. novicida* dual-RNA:Cas9

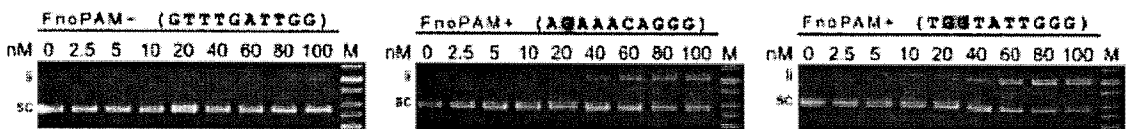


Figure S9

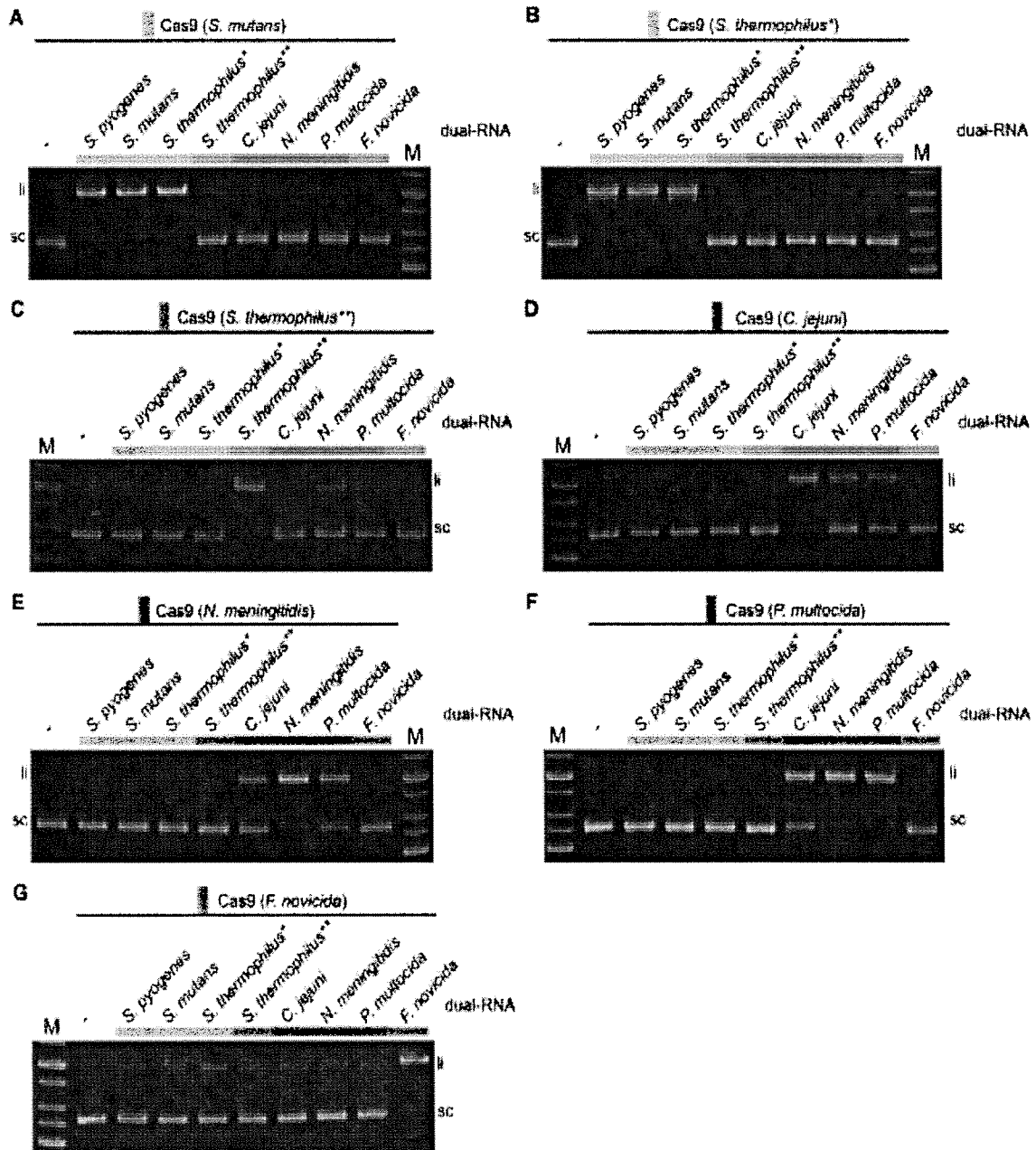


Figure S10

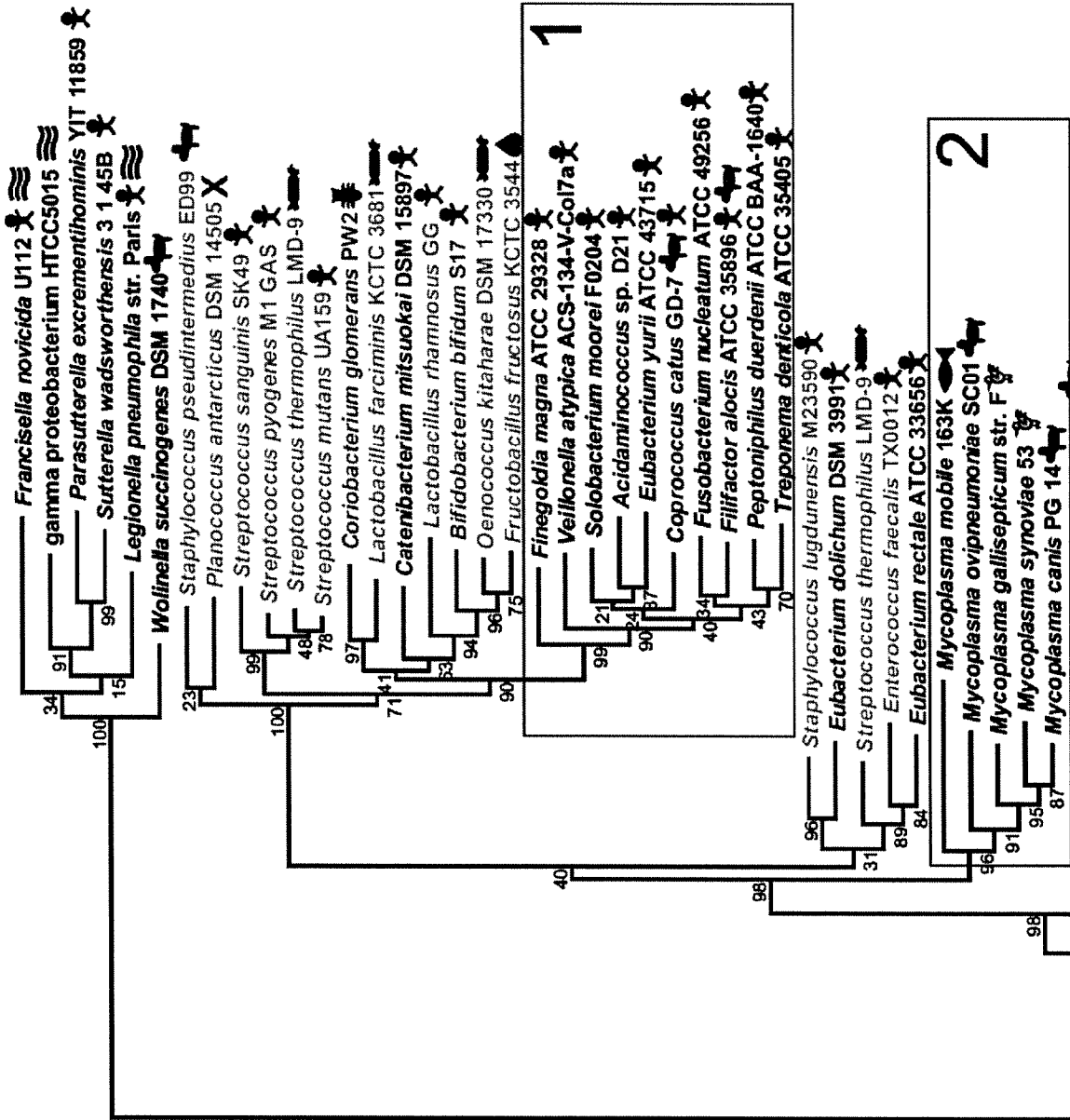
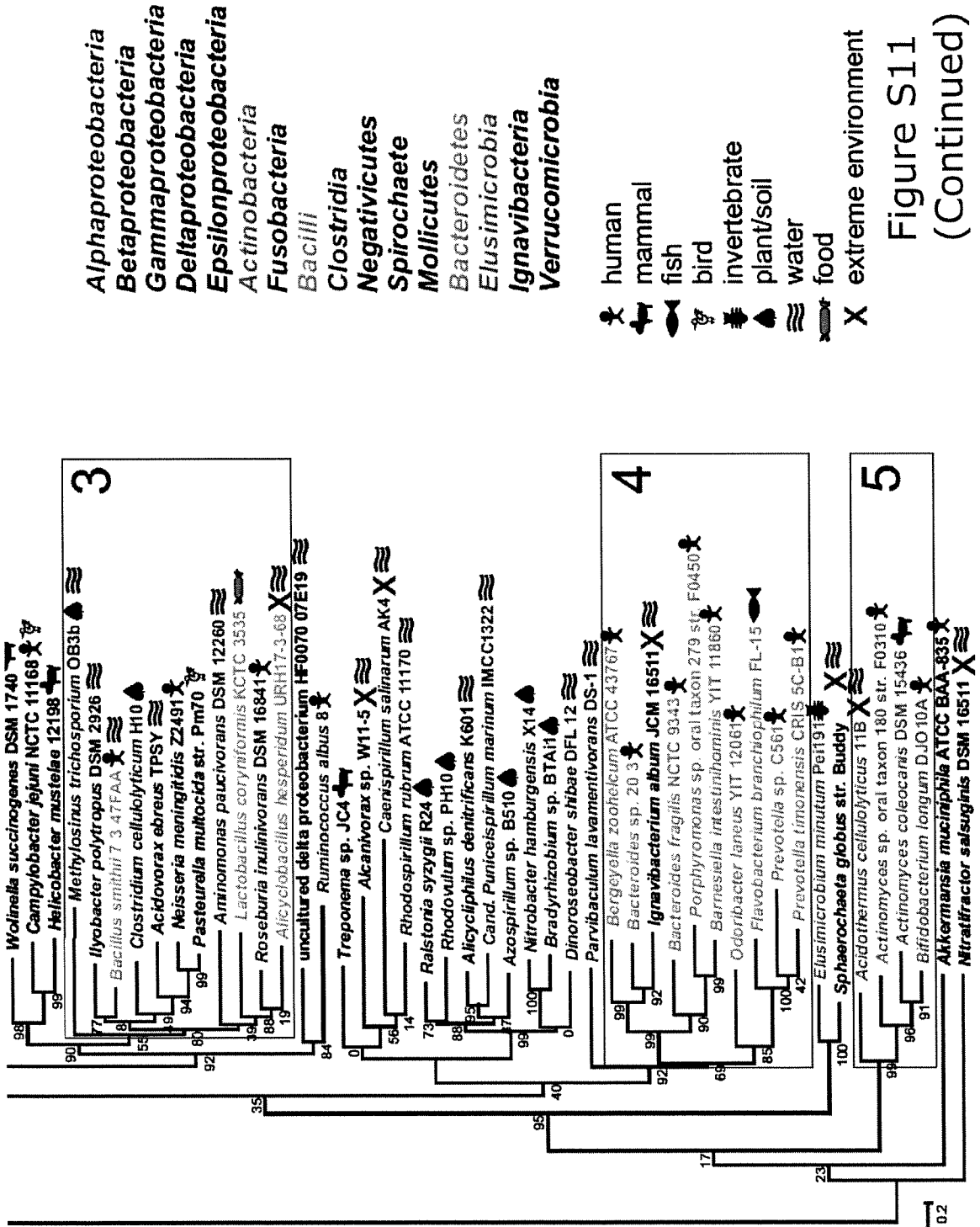
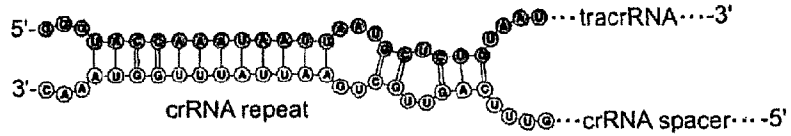


Figure S11

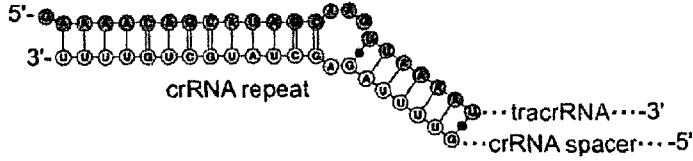




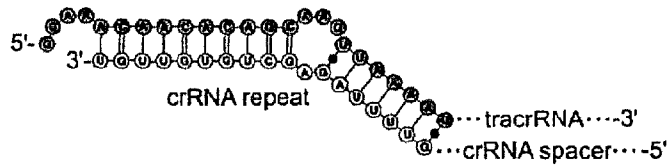
*F. novicida*



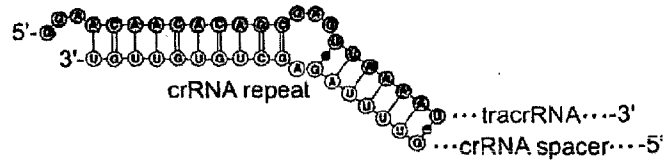
*S. pyogenes*



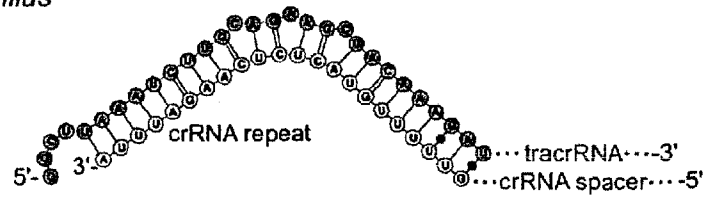
*S. mutans*



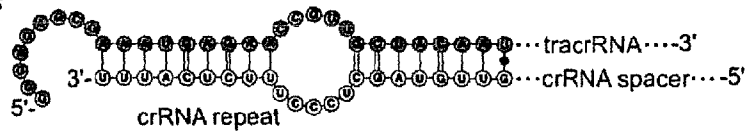
*S. thermophilus\**



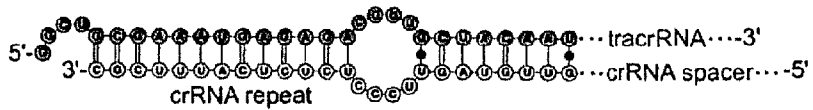
*S. thermophilus\*\**



*N. meningitidis*



*P. multocida*



*C. jejuni*

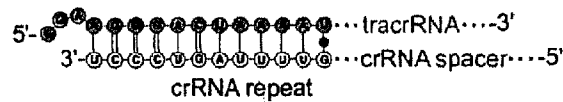


Figure S12